

# Acute and Sub-Chronic Toxicity Evaluation of the Crude Methanolic Bark Extract of *Bridelia micrantha* (Hochst.) Baill. (Phyllanthaceae) and Its Fraction

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## Abstract

Bridelia micrantha, commonly known as coastal golden leaf, is a member of the family Phyllanthaceae. In preliminary studies nine fractions, named F<sub>1</sub> -F<sub>9</sub>, were obtained by fractionating the crude methanol extract of the stem bark of Bridelia micrantha using column chromatographic techniques. The fraction F<sub>6</sub> was the most active when tested for antibacterial activity. Thus, toxicity of this fraction was investigated for further use. The present study evaluated the acute and sub-chronic toxicity of the crude methanolic bark extract of Bridelia micrantha and its fraction. The acute toxicity was carried out according to the experimental protocol of Organization for Economic Cooperation and Development (OECD). The plant extract or the fraction F<sub>6</sub> was administered orally to female mice at a single dose of 2000 mg/kg and the animals were observed for any behavioral changes or mortality for 14 days. In the sub-chronic toxicity study, the extract and fraction were administered orally at 200, 400 and 800 mg/kg bw/day for 28 days to healthy Wistar rats. The general behavior and body weight of the rats were recorded daily. At the end of the experimental period, hematological and biochemical analyses, changes in vital organ weight (liver, lung, heart, spleen and kidney), and histopathological examination of the liver and kidney were performed. No mortality or adverse effects were noted at the 2000 mg/kg dose during the oral acute toxicity test. In the sub-chronic study, the crude methanolic bark extract of Bridelia micrantha and the fraction F<sub>6</sub> induced no mortality or treatment-related adverse effects on body weight, general behavior, relative organ

weights, hematological and biochemical parameters. Histopathological examination of the liver and kidney showed normal architecture suggesting no morphological alterations. In conclusion, the oral administration of the crude methanolic bark extract of *Bridelia micrantha* and the fraction  $F_6$  for 28 days at a dosage of up to 800 mg/kg did not induce toxicological damage in rats. From acute toxicity study, the median lethal dose (LD<sub>50</sub>) of the crude methanolic bark extract of *Bridelia micrantha* and the fraction  $F_6$  was estimated to be more than 2000 mg/kg.

## **Keywords**

*Bridelia micrantha*, Acute Toxicity, Sub-Chronic Toxicity, Biochemical Parameters, Hematological Parameters

# **1. Introduction**

The use of medicinal plants is on the rise worldwide, because of the enormous expansion of traditional medicine and a growing curiosity in herbal therapies. Plants are used in treatment to preserve and improve physical, emotional, and moral fitness or to cure particular diseases and disorders [1]. According to the World Health Organization (WHO), up to 80% of the world's population uses medicinal plants for their primary health care. This is sometimes the only affordable and accessible source of care, especially for the poorest patients [2]. There are many types of plants in Africa in general and Cameroon in particular, used in traditional medicine to treat diseases of various origins [3]. However, despite the importance of the medicinal plant sector for the national and regional economy, their exploitation is carried out under artisanal conditions [4]. Most herbalists do not indicate the protocol usage that goes with their products; they also do not know the side effects and the toxicity of the plants used. Therefore, in-depth scientific studies on the benefits and risks of these plants could allow their efficient use to prevent producing adverse effects on the population's health [5].

Bridelia micrantha, commonly known as the coastal golden leaf, is a member of the family Phyllanthaceae [6]. It is a plant widely used across Africa and in Cameroon. Ethnobotanical knowledge reveals that a decoction of the bark of the trunk is generally used in Cameroon against several ailments such as amoebic dysentery, cough, eye pain, diarrhoea, gastric ulcers, infertility [7]. Previous studies on this plant showed anti-cancer [8], antidiabetic [9], antidiarrheal, antiplasmodial, antioxidant properties [10]. In precedent studies, we confirmed the antibacterial activity of the stem bark methanol extract of this plant [11]. They revealed a significant increase of this activity achieved with an active fraction  $F_6$  following a partition of this methanol extract and column chromatography on silica gel of the dichloromethane (DCM) portion [11]. HPLC chemical analysis of the fraction  $F_6$  highlighted the presence of almost 180 identified compounds from various secondary metabolites, including alkaloids, flavonoids, phenols, quinones, steroids and terpenoids [12]. Thus, further studies are necessary to continue exploring the antimicrobial effect of this stem bark methanol extract and its fraction. Hence, assessing their toxicity profiles is crucial; toxicity studies concerning *B. micrantha* extracts are still scarce, and most results are preliminary [13]. This study investigated the acute and subacute toxicity of the stem bark methanol extract of *B. micrantha* and the  $F_6$  fraction.

# 2. Materials and Methods

## 2.1. Plant Material

Fresh barks of *B. micrantha* used in this experiment were collected in January 2017 in the Centre Region of Cameroon at Mount Kalla. The plant was identified at the Cameroon National Herbarium, where a voucher specimen N° 5714 HNC (YA) was deposited.

## 2.2. Experimental Animals

Female and non-pregnant mice (*Mus musculus* nulliparous strain, 8 - 12 weeks old and 12 - 30 g) were used for the acute toxicity test, according to the Organization for Economic Cooperation and Development [14]. Nulliparous and non-pregnant adult rats (Wistar albino strain, 8 - 10 weeks and 100 - 180 g) were used for sub-chronic toxicity tests. Animals were obtained from the Animal Biology Laboratory, University of Yaoundé I. They were placed in plastic cages under normal laboratory conditions (12 hours light/dark cycle,  $25^{\circ}C \pm 2^{\circ}C$ ) for an acclimatization period of 7 days before the experiments. They received daily a standard food ration and water. Experiments were conducted according to internationally accepted guidelines for evaluating the safety and efficacy of herbal medicines [15].

## 2.3. Plant Extraction

*Bridelia micrantha* barks were collected and dried for 21 days under shade and ground into powder. The powdered plant material (2.5 kg) was soaked in 10 L of methanol for 3 days. The mixture was filtered using a Whatman No.1 filter paper, and the residue was re-extracted four times as previously described. The total methanol extract was concentrated using a rotatory evaporator (Heidolph). The extract was dried in an oven (VENTI-Line) at 45°C for 24 h.

## 2.4. Fractionation of the Crude Extract

The extract (600 g) was dissolved in a mixture of methanol (2 L), distilled water (1 L) and dichloromethane (DCM) (2 L). The DCM phase and the methanol/water phase were separated. Each phase was concentrated using a rotatory evaporator. The DCM phase (150 g) was chromatographed through a silica gel (250 - 300 Mesh) as described previously [11]. Nine fractions labelled  $F_1$  to  $F_9$  were obtained. The fraction  $F_6$  (24 g) was used in this study.

## 2.5. Acute Oral Toxicity

The acute toxicity study was conducted according to OECD guidelines No. 425 [14]. Eighteen mice were divided into three groups of six animals each. They were fasted for 4 h and then weighed before the experiment. Control group received only distilled water (10 ml/kg), while experimental groups received the *B. micrantha* methanol extract solution and fraction  $F_6$  solution, prepared in distilled water at a single dose of 2000 mg/kg body weight (bw). Animals received substances by gavage using a probe. All animals were observed for the first 3 h for toxicity signs (pain sensitivity after tail nipping, noise sensitivity after this time, animals were supplied with food and water, and then observed for 14 days. During this period, animal weights were recorded once per week.

## 2.6. Sub-Chronic Oral Toxicity Study

#### 2.6.1. Experimental Design

This study was conducted according to OECD [15]. Thirty-five rats were divided into seven groups of 5 animals. Group 1 (control) received distilled water. Groups 2, 3, and 4 received the extract at 200, 400 and 800 mg/kg bw respectively; also groups 5, 6 and 7 received fraction  $F_6$  at 200, 400 and 800 mg/kg doses. The administration was once daily over 28 days by gavage. During the experiment, body weight was noted once every two days, and toxicity signs of the individual animals or in the community were observed every week (mobility, tremor, social interaction and sensitivity to noise).

On day 28, all animals were subjected to a 24-hour food and water fast. Animals were then weighed, anesthetized intraperitoneally with ketamine (50 mg/kg) and sacrificed. Blood was collected in EDTA and dry tubes by cardiac puncture. The blood contained in EDTA tubes was transported to the hospital for the complete blood count, while the blood contained in dry tubes was left to rest for 6 hours and then centrifuged at 3000 rpm for 15 min. The serum was collected and stored at  $-80^{\circ}$ C to determine biochemical toxicity parameters.

Animals were dissected and vital organs such as liver, heart, kidneys, lungs, spleen, and uterus were removed. These organs were degreased, rinsed in a 0.9% NaCl solution and drained. Each organ was weighed and the organ relative weight compared to fasted animal weight was calculated. Organs were then preserved in a 0.9% Formalin-NaCl solution to produce histological sections.

#### 2.6.2. Blood Analysis

Hematological parameters such as white blood cell (WBC) count, red blood cell (RBC) count, platelet (PLT) count, hematocrit (HCT), hemoglobin (HGB), mean corpuscular volume (MCV), mean cell hemoglobin (MCH) and mean corpuscular hemoglobin concentration (MCHC) were determined using an automatic analyzer (System H1, Bayer Diagnostics). The serum was analyzed for total protein, total cholesterol, HDL cholesterol, creatinine, urea, triglycerides, glucose, and alanine transaminase (ALT)/aspartate aminotransferase (AST) ac-

tivities using specific commercial diagnostic kits (Fortress Diagnostics, London, UK).

## 2.6.3. Histopathological Analysis

Kidney and liver samples were preserved in 0.9% Formalin-NaCl solution and processed by conventional techniques. Before microscopic examination, Paraffin sections (5 µm thick) were stained with hematoxylin and eosin.

#### 2.6.4. Ethical Considerations

Experimental protocols used in this study strictly aligned with the internationally accepted standard ethical guidelines for laboratory animal use and care as described in the European Community guidelines, EEC Directive 86/609/EEC of the 24<sup>th</sup> November 1986 [16]. This study was approved by the Institutional Animal Ethics Committee (Registration No. 778/PO/a/03/CPCSEA; 03.09.).

#### 2.6.5. Statistical Analysis

Data were statistically analyzed using SPSS software 16.0. Quantitative data were expressed as mean  $\pm$  SD. One-way ANOVA followed by Newman-Keuls post hoc test was used for multiple comparisons, and the values of p < 0.05 were considered statistically significant.

## **3. Results**

## **3.1. Acute Oral Toxicity**

The acute toxicity test using an oral limit dose of 2000 mg/kg of either the crude methanolic bark extract of *Bridelia micrantha* or the  $F_6$  fraction caused no adverse reactions or death in mice throughout the 14 days study period. Therefore, the oral LD<sub>50</sub> of the extract and the  $F_6$  fraction is considered greater than 2000 mg/kg in mice.

#### 3.2. Sub-Chronic Oral Toxicity

#### 3.2.1. Weight Gain

The extract or the  $F_6$  fraction did not significantly affect the animals' food intake. Moreover, the animal weight generally increased over time (Figure 1).

## 3.2.2. Relative Organ Weights

No significant changes in organ weight (liver, kidney, heart, and spleen) were observed between the treated and control rats after oral administration of the crude methanolic bark extract of *Bridelia micrantha* or fraction  $F_6$  over 28 days (Table 1).

## **3.2.3. Biochemical Parameters**

The effects of administration of *B. micrantha* and fraction  $F_6$  after 28 days on plasma biochemical parameters in experimental rats were presented in **Table 2**. No significant difference (p > 0.05) was found in albumin, total protein, uric acid, and HDL-cholesterol levels. No significant difference (p > 0.05) was noted in



**Figure 1.** (a) Evolution of the animals' weight during 28 days of administration of the crude methanolic bark extract of *Bridelia micrantha*; (b) Evolution of the animals' weight during 28 days of administration of the fraction  $F_6$ .

**Table 1.** Relative organ weights after 28 days of oral administration in female rats according to doses of the crude methanolic bark extract of *Bridelia micrantha* or fraction  $F_6$ 

	Dose	Relative organ weights (g/100g bw)								
	(mg/kg)	Heart	Liver	Lungs	Spleen	Kidney	Uterus			
	0	$068\pm0.02$	$5.38\pm0.01$	$1.52\pm0.17$	$0.97\pm0.09$	$1.23\pm0.03$	$0.46 \pm 0.01$			
Ema	200	$0.62\pm0.03$	$5.34\pm0.28$	$1.46\pm0.21$	$1.03\pm0.14$	$1.19\pm0.03$	$0.44\pm0.04$			
Em	400	$0.63\pm0.04$	$5.26\pm0.25$	$1.43\pm0.12$	$0.99\pm0.11$	$1.18\pm0.09$	$0.42\pm0.04$			
	800	$0.63\pm0.03$	$5.15 \pm 0.22$	$1.44\pm0.05$	$0.94\pm0.07$	$1.19\pm0.04$	$0.42\pm0.05$			
	0	$0.68\pm0.02$	$5.38\pm0.01$	$1.52 \pm 0.17$	$0.97\pm0.09$	$1.23\pm0.03$	$0.46 \pm 0.01$			
Б	200	$0.65\pm0.04$	$5.23\pm0.33$	$1.49\pm0.18$	$1.00\pm0.08$	$1.21\pm0.07$	$0.43\pm0.03$			
г <sub>б</sub>	400	$0.62\pm0.04$	$5.00\pm0.37$	$1.46\pm0.06$	$1.01\pm0.11$	$1.15\pm0.13$	$0.41\pm0.04$			
	800	$0.67\pm0.02$	$5.28\pm0.03$	$1.51\pm0.14$	$1.00\pm0.04$	$1.21\pm0.02$	$0.40\pm0.03$			

Data are expressed as mean  $\pm$  SD, n = 5. No statistically significant difference was observed between the test and control groups. Em: crude methanolic bark extract of *Bridelia micrantha*; F<sub>6</sub>: fraction F<sub>6</sub>.

**Table 2.** Biochemical parameters in experimental animals after 28 days of oral administration in female rats according to doses of the crude methanolic bark extract of *Bridelia micrantha* or  $F_6$  fraction.

	Dose (mg/kg)	ALT (U/L)	AST (U/L)	Total proteins (mg/dl)	Albumin (g/l)	Creatinine (mg/dl)	Uric acid (mg/dl)	Glucose (mg/dl)	Total cholesterol (mg/dl)	HDL cholesterol (mg/dl)	LDL cholesterol (mg/dl)	Triglycérides (mg/dl)
	0	56.90 ± 1.54	75.70 ± 0.62	0.76 ± 0.02	2.24 ± 0.07	0.97 ± 0.02	3.14 ± 0.14	100.52 ± 2.35	51.00 ± 3.52	36.30 ± 3.00	15.40 ± 0.43	19.90 ± 2.08
Em	200	17.90 ± 1.88*	60.75 ± 1.26*	0.72 ± 0.02	2.27 ± 0.15	0.80 ± 0.02	2.85 ± 0.14	80.28 ± 1.82*	41.90 ± 3.40*	34.44 ± 2.78	13.20 ± 0.18*	15.80 ± 1.54*
	400	21.60 ± 1.94*	68.20 ± 1.21*	0.77 ± 0.06	2.20 ± 0.05	0.90 ± 0.06	3.32 ± 0.12	84.76 ± 1.48*	40.80 ± 3.77*	34.00 ± 2.43	13.50 ± 0.36*	16.00 ± 1.26*

	800	20.20 ± 1.42*	67.00 ± 1.18*	0.77 ± 0.02	2.25 ± 0.10	0.81 ± 0.06	3.17 ± 0.09	82.00 ± 1.52*	39.20 ± 4.12*	33.50 ± 2.02	12.00 ± 0.60*	16.90 ± 1.27*
F <sub>6</sub>	0	56.90 ± 1.54	75.70 ± 0.62	0.76 ± 0.02	2.24 ± 0.07	0.97 ± 0.02	3.14 ± 0.14	100.52 ± 2.35	51.00 ± 3.52	36.30 ± 3.00	15.40 ± 0.43	19.90 ± 2.08
	200	18.40 ± 1.50*	63.60 ± 1.50*	0.70 ± 0.02	2.22 ± 0.05	0.71 ± 0.03*	2.84 ± 0.05	81.50 ± 2.26*	43.20 ± 3.80*	37.00 ± 2.46	12.90 ± 0.40*	11.20 ± 0.22*
	400	20.00 ± 1.24*	66.40 ± 1.31*	0.73 ± 0.03	2.25 ± 0.06	0.75 ± 0.04*	3.57 ± 0.14	86.76 ± 3.79*	42.10 ± 3.90*	38.80 ± 2.68	10.80 ± 0.40*	16.40 ± 0.58*
	800	28.30 ± 1.59*	66.56 ± 1.74*	0.75 ± 0.06	2.21 ± 0.06	0.72 ± 0.03*	3.27 ± 0.09	84.87 ± 2.19*	40.30 ± 4.34*	38.30 ± 3.91	12.50 ± 0.36*	$19.70\pm0.31$

Data are expressed as mean  $\pm$  SD, n = 5. \*Indicate a significant difference at p < 0.05 compared to the appropriate control group (Newman-Keuls). Em: crude methanolic bark extract of *Bridelia micrantha*; F<sub>6</sub>: fraction F<sub>6</sub>.

**Table 3.** Hematological parameters in experimental animals after 28 days of oral administration according to doses of the crude methanolic bark extract of *Bridelia micrantha* or  $F_6$  fraction.

	Dose (mg/kg)	WBC (10 <sup>3</sup> /mm <sup>3</sup> )	RBC (10 <sup>6</sup> /mm <sup>3</sup> )	HGB (g/dl)	HCT (%)	MCV (µm³)	MCH (pg)	MCHC (g/dl)	PLT (10 <sup>3</sup> /mm <sup>3</sup> )	MON (10 <sup>3</sup> /mm <sup>3</sup> )	LYM (10 <sup>3</sup> /mm <sup>3</sup> )	NEUT (10 <sup>3</sup> /mm <sup>3</sup> )
Em	0	8.05 ± 0.09	8.03 ± 0.25	13.50 ± 0.54	42.10 ± 1.12	55.60 ± 0.70	17.30 ± 0.20	31.00 ± 0.32	618.00 ± 35.20	1.04 ± 0.26	4.50 ± 0.42	1.72 ± 0.16
	200	8.03 ± 0.31	8.00 ± 0.13	14.20 ± 1.02	44.50 ± 0.53	55.40 ± 1.03	16.80 ± 0.33	30.50 ± 0.12	647.00 ± 19.70	0.91 ± 0.09	4.65 ± 0.18	1.62 ± 0.06
	400	7.96 ± 0.33	8.50 ± 0.24	14.40 ± 0.52	43.90 ± 0.63	54.60 ± 1.03	16.90 ± 0.29	31.00 ± 0.18	649.00 ± 28.20	1.01 ± 0.14	4.62 ± 0.38	1.53 ± 0.11
	800	8.21 ± 0.28	8.91 ± 0.27	15.00 ± 0.41	47.40 ± 0.86*	54.30 ± 0.54	16.90 ± 0.12	31.10 ± 0.14	802.00 ± 49.40*	1.19 ± 0.11	4.60 ± 0.12	1.73 ± 0.21
F <sub>6</sub>	0	8.05 ± 0.09	8.03 ± 0.25	13.50 ± 0.54	42.10 ± 1.12	55.60 ± 0.70	17.30 ± 0.20	31.00 ± 0.32	618.00 ± 35.20	1.04 ± 0.26	4.50 ± 0.42	1.72 ± 0.16
	200	8.14 ± 0.26	8.80 ± 0.33	13.70 ± 0.84	49.50 ± 0.54*	53.10 ± 0.88	16.30 ± 0.29	30.60 ± 0.29	652.00 ± 14.00	1.09 ± 0.26	4.69 ± 0.11	1.84 ± 0.26
	400	8.12 ± 0.20	8.45 ± 0.31	14.60 ± 0.82	43.80 ± 0.49	54.00 ± 0.61	16.40 ± 0.16	30.40 ± 0.26	654.00 ± 38.50	1.13 ± 0.13	4.65 ± 0.11	$1.72 \pm 0.02$
	800	8.22 ± 0.32	8.33 ± 0.45	13.80 ± 0.45	43.70 ± 0.57	54.40 ± 0.95	16.60 ± 0.38	30.60 ± 0.22	699.00 ± 20.70	1.07 ± 0.09	4.57 ± 0.11	1.60 ± 0.14

WBC: White Blood Cell Count, RBC: Red Blood Cell Count, HGB: Hemoglobin, HCT: Hematocrit, MCV: Mean Corpuscular Volume, MCH: Mean Cell Hemoglobin, MCHC: Mean Corpuscular Hemoglobin Concentration, PLT: Platelet Count, MON: Monocytes, LYM: Lymphocytes, NEUT: Neutrophils. Data are expressed as mean  $\pm$  SD, n = 5. \*Indicate a significant difference at p < 0.05 compared to the appropriate control group (Newman-Keuls). Em: crude methanolic bark extract of *Bridelia micrantha*; F<sub>6</sub>: fraction F<sub>6</sub>.

animals treated with the extract compared to the controls regarding the creatinine level. The fraction  $F_6$  at doses of 200, 400 and 800 mg/kg caused a significant (p < 0.05) decrease in creatinine level. Administration of the extract or fraction  $F_6$  in repeated doses for 28 days resulted in a significant reduction (p < 0.05) in serum ALT, AST, glucose concentrations. Similarly, compared to controls, there was a significant decrease (p < 0.05) in total cholesterol, LDL-cholesterol and triglyceride levels in animals given the extract or fraction  $F_6$ .

### 3.2.4. Hematological Parameters

There was no significant variation (p > 0.05) independently of the dose, compared to the control group, in the animals having received the extract or fraction  $F_6$ , concerning the number of erythrocytes (RBC), leukocyte count (WBC), hemoglobin count (HGB), mean corpuscular hemoglobin count (MCHC), neutrophile count (NEU), lymphocyte count (LYM), monocytes count (MON), mean cell hemoglobin (MCH), and mean corpuscular volume (MCV). Similarly, no significant difference was observed concerning the hematocrit level (HCT) in all treatments, except for groups receiving the extract at 800 mg/kg and the fraction  $F_6$  at 200 mg/kg, for which there was a significant increase (p < 0.05). There was also a significant increase (p < 0.05) in blood platelet number (PLT) in the group having received the extract at 800 mg/kg compared to the control group.

## **3.2.5. Liver and KIDNEY HISTOPATHOLOGY**

Histological studies showed no abnormalities in the liver and kidney tissue from the rat treated with the crude methanolic bark extract of *Bridelia micrantha* or fraction  $F_6$  for 28 days compared to controls (Figure 2 and Figure 3).

# 4. Discussion

The extract or the  $F_6$  fraction administered orally at a dose of 2000 mg/kg did not cause death among the selected mice throughout the study. The extract and fraction  $F_6$  have a toxicity index of 5, according to the toxicity scale of chemical



ED: control rat that received distilled water; Em 200, 400, 800, Fi 200, 400 and 800: rats that received the extract and the  $F_6$  fraction at the respective doses of 200, 400 and 800 mg/kg. Vc = centrilobular vein; He = Hepatocyte; Cs = Sinusoidal capillary.

**Figure 2.** Histological sections of the rat liver after 28 days of treatment at different doses of the extract and fraction  $F_6$  (Hematoxylin-eosin ×100).



ED: control rat given distilled water; Em 200, 400, 800, Fi 200, 400 and 800: rats that received the extract and the  $F_6$  fraction at the respective doses of 200, 400, and 800 mg/kg. G = Glomerulus; Eu = Urinary space; Tcd = distal convoluted tubule; Tcp = Proximal convoluted tubule.

**Figure 3.** Histological sections of the rat kidney after 28 days of treatment at different doses of the extract and fraction  $F_6$  (hematoxylin-eosin ×100).

substances depending on the  $LD_{50}$  and administration route [14]. In addition, no toxicity sign was observed during the 4 h after the extract or fraction  $F_6$  administration, particularly the decrease in sensitivity to the stimulus (pain and noise), mobility and feces softening. Onoja *et al.* [17] revealed that hydroalcoholic bark extract did not cause toxic effects in mice at the same dose. Some investigators, using shrimp as an animal model suggested that the plant would be toxic because of the low lethal concentrations that these authors obtained [18] [19]. However, since rodents (mice, rats) are model organisms with multiple advantages linked to their physiology, almost similar to humans [20] [21], the results are more reliable.

The weight evolution in animals that received the extract or fraction  $F_6$  remained insignificant compared to the control during the 28 days of administration. Thus, neither the extract, much less the fraction  $F_6$ , adversely affected the animals' food intake. No significant differences in relative organ weights were found between the control and test groups. Furthermore, neither the extract nor the fraction  $F_6$  adversely affected organ weights. Aminotransferases, ALT and AST describe the cellular integrity of the liver. These are enzymes mainly synthesized in the cytoplasm of hepatic cells and discharged into the circulation when these cells are damaged. Thus, high levels of AST and ALT are frequently attributed to different drugs' metabolic and toxic effects [22] [23]. Our results showed a significant decrease in ALT and AST levels at all doses, indicating that the extract and fraction  $F_6$  could not cause liver damage but have a hepatoprotective action. In addition, Nwaehujor and Udeh [24] showed that ethyl acetate extract from the leaves of *B. micrantha* had a more pronounced hepatoprotective effect than silymarin, a hepatoprotective reference molecule. Proteins are the

building blocks of the human body. A high serum protein level could indicate a damaged tissue or organ. No significant difference in serum total proteins of treated animals compared to controls was observed, suggesting that the extract and fraction  $F_6$  did not cause liver damage [25]. Albumin is the most abundant protein (60%) in blood. In the body, it is produced by hepatocytes. A drop in albumin level may indicate liver failure [26] [27]. There was no significant difference in albumin levels in animals receiving the extract and fraction  $F_6$  at the different doses compared to the control group, suggesting no liver affection. Analysis of all these liver markers revealed that prolonged use (28 days) of the extract or fraction  $F_6$  had no adverse effect on liver function. This was confirmed by the histopathological examination of the liver, which showed no alteration in its structure.

Creatinine and uric acid are markers of impaired kidney function. Creatinine results from the breakdown of creatine, a constituent of muscle. Creatine can be transformed into ATP, an energy source for cells. Creatinine is excreted by the kidneys. In case of renal insufficiency, it is retained in the blood, along with uric acid, a waste product of the metabolism of purine bases. Thus, a high level of creatinine, uric acid could indicate renal failure [28] [29]. A significant decrease in creatinine levels in animals treated with the fraction  $F_6$  was observed and the values of uric acid levels did not vary significantly indicating that neither the extract nor the  $F_6$  fraction adversely affected kidney function. The histological sections of the kidneys, which present a normal architecture of the renal parenchyma, support this assertion.

The extract and fraction  $F_6$  caused a significant decrease in serum glucose levels in treated animals. This result suggests a hypoglycemic action of this extract and fraction  $F_6$ . Adika *et al.* [30] demonstrated that the methanol extract from the leaves of this plant had an antidiabetic activity. These observations approve this plant's use in treating diabetes in traditional medicine in several African countries [31].

Lipids are essential for the proper body functioning. But if they are in excess, they increase the risk of cardiovascular diseases. The lipid profile helps to monitor cholesterol levels (LDL-cholesterol and HDL-cholesterol) and triglycerides [32] [33] A high LDL-cholesterol (bad cholesterol), triglycerides and low HDL-cholesterol constitute lipid abnormalities (dyslipidemias). A significant decrease in total cholesterol, triglycerides and LDL cholesterol was observed, and there was no significant difference in HDL cholesterol in treated animals. These results suggest a hypolipidemic effect of the extract and fraction  $F_6$  and could help combat cardiovascular diseases.

The analysis of blood parameters is very useful for determining abnormalities caused by plant extracts [34] [35]. It can also provide information on the mechanism of toxicity or defense of a therapeutic agent [36]. It was shown that the drop in some parameters, such as RBC, VGM, CCMH and HGB, could induce anemia [37]. The present study did not reveal any significant difference in these parameters between treated and control animals. This result indicates that neither the extract nor fraction  $F_6$  can cause anemia and has no adverse effect on the hematopoietic system.

# **5.** Conclusion

This study aimed to assess the toxicity of the stem bark methanol extract of *B. micrantha* and fraction  $F_6$ . Findings revealed that they did not adversely affect overall mortality and behavior in mice at tested doses. Therefore, the oral  $LD_{50}$  is greater than 2000 mg/kg and generally considered safe. Acute and sub-chronic toxicity investigations showed that they do not adversely affect body weight, liver and kidney, hematological and biochemical parameters. No toxicity sign was observed in the treated mice. Thus, these results showed that the stem bark methanol extract of *B. micrantha* and fraction  $F_6$  can be safely used in humans, and further therapeutic studies can be performed.

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# **Conflicts of Interest**

The authors declare no conflicts of interest regarding the publication of this paper.

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