

Anticandidal Activity of Moroccan Medicinal Plants

Khadija El Assraoui¹, Tarik Rochd²

¹Department of Removable Prosthodontics, Faculty of Dentistry, Hassan II University, Casablanca, Morocco

²Laboratory of Oral Biology, Faculty of Dentistry, Hassan II University, Casablanca, Morocco

Email: Khadijaa.ela@gmail.com

How to cite this paper: El Assraoui, K. and Rochd, T. (2023) Anticandidal Activity of Moroccan Medicinal Plants. *Journal of Biosciences and Medicines*, 11, 79-95.
<https://doi.org/10.4236/jbm.2023.117008>

Received: May 17, 2023

Accepted: July 11, 2023

Published: July 14, 2023

Copyright © 2023 by author(s) and Scientific Research Publishing Inc. This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

The prevalence of oral candidiasis has increased over recent years in patients with the compromised general condition. There are a large number of plant species that have been traditionally used for oral diseases by the Moroccan population. These species could provide a source for discovering new active principles of natural origin against *Candida albicans* involved in the appearance of candidiasis infections. Information was acquired using the electronic databases Web of Science, Scopus and PubMed. In the first part, this review presents the medicinal plants used by the Moroccan population for the treatment of oral disorders (Toothaches, gingivitis, ulcers, stomatitis, abscesses, ...). The second part describes different studies regarding the antifungal activity of essential oils and organic extracts of some Moroccan species against *Candida albicans*. Inhibition zone diameter (IZD), Minimum inhibitory concentration (MIC) and minimum fungicidal concentration (MFC) values of essential oils and plant's extracts were noted. The third part concerns Moroccan plants with the antifungal activity that can be used in medical and agricultural fields. Several species of Moroccan plant proved their effectiveness against *Candida albicans* and can be used for the treatment of oral candidiasis. Further research should be undertaken in the future to investigate the toxicity of essential oils and plant extracts with antifungal properties for clinical use. This review provided an update on Moroccan plants presenting antifungal activity and especially those active against *Candida albicans*.

Keywords

Oral Candidiasis, Antifungal, Moroccan Plants, *Candida albicans*, Essential Oil, Plant Extracts

1. Introduction

Oral health is an integral part of general human health. Oral health disorders can

have consequences on mastication, phonation, and aesthetics, which can have an impact on psychosocial well-being [1].

Among the most common opportunistic infections in several high-risk patient groups is oropharyngeal candidiasis. Cancer radiation and/or chemotherapy, immunosuppression, immunodeficiency, diabetes, vitamin deficiency, hyposalivation, denture wear, patients under frequent topical antibiotic, glucocorticoid treatment and patients with anemia are predisposing factors for oropharyngeal candidiasis [2] [3] [4]. The pathogenic mechanism of *Candida* spp. depend on both *Candida* virulence and human host conditions. There has been an increased virulence of *Candida* spp. colonized in several groups of cancer and non-cancer population groups, such as diabetic patients, HIV-infected patients, hospitalized patients, and denture wearers. *Candida albicans* is the most common opportunistic fungal pathogen in humans. It causes superficial mucosal candidiasis or even disseminated candidiasis [2] [3] [4]. Antifungal drugs are the indicated treatment in the case of fungal infection. They are administered systemically or topically [4]. However, their usage is limited due to their side effects, in addition to the fact that *Candida* is becoming increasingly resistant [3]. That's why the search for new, less toxic, more effective and environment-friendly therapeutic approaches is becoming a necessity. Therefore, several types of research have been conducted on natural plant-based antimicrobials [5].

Plants have been utilized for centuries in folk medicine and there is a strong belief that herbal medicines are healthier and safer than drugs. Owing to their antimicrobial activities, efficacy, higher safety margins, ease of access, and economic feasibility, medicinal plants are used in developed as well as developing countries for primary healthcare.

Morocco, due to its geographical location and the variety of climates and microclimates, offers a rich and diversified vegetation [6] [7]. 4200 among 5000 species and subspecies listed in North Africa grow in Morocco of which at least 500 are potent medicinal and 250 are currently used [8] [9]. Its population is familiar with the use of plants [10]. Infusion, decoction, powder, and maceration of various parts of plants (Roots, flowers, fruits, seeds, leaves), and the whole plants have been used in the treatment of several diseases.

Morocco is one of the providers in the international market of aromatic and medicinal plants and their products, especially essential oils [11].

Essential oils plant-derived have proven their values as a source of molecules with antimicrobial effects [12]. Numerous studies have proven their antifungal effects [5] [13].

The action mechanism of essential oils is difficult to explain due to its complexity. It may be possible that every single component has its mechanism of action. The essential oil efficacy depends on the most abundant compound described [14]. According to Boukssaim *et al.* (2013), the inhibition of germs by essential oils is dependent on their chemical profiles and the structure of the cell membrane [11]. They are distinguished by their hydrophobicity, which helps

them to penetrate the membrane of cells and mitochondria, disrupting the cell structure and making it more permeable, resulting in cell death [15]. Many researchers have also explored their combinational effect with drugs to overcome microbial resistance and to decrease the minimum effective dose of conventional antimicrobial drugs [16] [17]. The knowledge of plants and the study of their properties, and their therapeutic effects are primordial, to discover new active principles [6] [15].

The aim of this article is to review Moroccan plants used for oral disorders and the anti-candidal activities of Moroccan plant essential oils and extracts.

2. Methodology

The literature on Moroccan plants and essential oil and plant's extract with antifungal activities from 2000 to 2022 were collected, and summarized in this review. The electronic databases Web of Science, Scopus, Pubmed were used to explore the published papers. The search terms used were antifungal, Moroccan plants, *Candida albicans*, and Essential oil. All published work in different languages (French or English) were cited in this review. Data provided in editorial/letters, symposiums, case reports, and conference papers were excluded.

The scientific names of all plants mentioned in this article were updated and verified using <https://wfoplantlist.org/plant-list>.

3. Results and Discussion

3.1. Plants Traditionally Used in Morocco for the Treatment of Oral Disease

The data presented in **Table 1** show several species of plants, their local name and their method of use, frequently used in Moroccan for the treatment of oral diseases. Lamiaceae are widely used in traditional medicine by the Moroccan population to treat several diseases [18].

Thymus saturejoides [18], *Thymus Capitatus* [19], *Marrubium vulgare* L [20], *Rosmarinus officinalis* L. *Ficus carica* L, *Olea europaea* L, *Hyoscyamus* sp, *Peganum harmala* L [1], *Daphne gnidium* [21], *Pinus halepensis* L [8], *Allium sativum* L, *Pistacia lentiscus* L, *Hedera helix* L, *Tetraclinis articulata* (Vahl) Masters, *Anacyclus pyrethrum* (L.) Link [21], *Ammi visnaga* (L.) Lam, *Pimpinella anisum* [20], *Cedrus atlantica* (Endl.) Manettiex Carrière, *Populus nigra* L, *Acacia nilotica* L, *Juglans regia* L. *Euphorbia officinarum* subsp. *Echinus* (Hook. F. &Coss.) Vindt, *Commiphora africana* (A. Rich.) Engl., *Heliotropium curassavicum* L, *Rhus albida schousboe* [22] are used to heal toothache.

Thymus saturejoides Coss. [18], *Thymus willdenowii* Boiss, *Origanum compactum* Benth, *Marrubium vulgare* L, *Laurus nobilis* L, *Ficus carica* L, *Olea europaea* L, *Ruta montana* L, *Peganum harmala* L [1], *Calendula arvensis* L. [21], *Melissa officinalis*, *Laurus nobilis* L, *Cinnamomum zeylanicum*, *Ammi visnaga* (L.) Lam, *Coriandrum sativum* L. *Chamaemelum nobile* L, *Artemisia herba alba*

Table 1. Plants used by the Moroccan population for the treatment of oral pathology.

Plant family	Plant species	Local name	Used parts	Indication according to the population	The method of preparation (Mode of use)	References
Lamiaceae	<i>Thymus maroccanus</i> Ball	Azukni	Leave and Flowers	Mouth affections	Infusion and maceration	[21]
	<i>Thymus saturejoides</i> Coss.	Tazouknit	Leave and Flowers	Gum, Toothache Gingivitis Mouth ulcer	Infusion, Decoction	[18]
	<i>Thymus willdenowii</i> Boiss	Zaïtra	Leaves Flowers	Mouth infection Mouth ulcer Gingivitis Gingival bleeding Bad breath	Decoction (Gargle, Masticatory)	[1]
	<i>Thymus Capitatus</i>		Aerial parts	Toothache	Infusion, decoction (external Application, massage, washings)	[19]
	<i>Marrubium vulgare</i> L.	Mariouta	Leaves	Toothache Gingival bleeding Bad breath Gingivitis	Crude (Masticatory)	[1] [20]
	<i>Origanum compactum</i> Benth	Zaatar tadlaoui	Leaves Flowers	Mouth ulcers Gingivitis Dental carie	Decoction (Gargle)	[1]
	<i>Origanum vulgare</i> L.	Zaater	Sap	Stomatitis Oral lesions	Crude (Touching the lesions with a cotton pad soaked in fresh sap)	[1] [20]
	<i>Rosmarinus officinalis</i> L.	Azir	Leaves Flowers	Toothache	Fumigation (Inhalation)	[1] [20]
	<i>Ocimum basilicum</i> L.	Hbak	Leaves	Oral ulcer	Decoction (Mouthwash)	[20]
	<i>Melissa officinalis</i>	Naanaa soufi	Leaves	Gum	Decoction (Mouthwash)	[20]
Lauraceae	<i>Laurus nobilis</i> L.	Wrak moussa	Leaves	Ulcers Gingivitis Bad breath	Decoction (Gargle)	[1]
	<i>Cinnamomum zeylanicum</i>	Karfa	Bark of the trunk	Gum Mouth ulcer	Decoction, Essential oil (Mouthwash, Direct application)	[20]
Apiaceae	<i>Ammi visnaga</i> (L.) Lam.	Bechnikha	Fruits	Gum disease Toothache	Raw/Decoction (Mouthwash, Direct application)	[20]
	<i>Coriandrum sativum</i> L.	Kozbore	Leaves	Toothache	Raw (Direct application)	[20]
	<i>Pimpinella anisum</i>	Nafae	Seeds	Gum disease	Decoction/ Infusion (Mouthwash)	[20]
Moraceae	<i>Ficus carica</i> L.	Karmous	Fruit	Dental abscesses Toothache Gingivitis	Decoction Cataplasm (Gargle, Direct Application)	[1]

Continued

Portulacaceae	<i>Portulaca oleracea</i> L	Rajla	Leaves	Dental abscesses	Cataplasm (Direct Application)	[1]
Oleaceae	<i>Olea europaea</i> L.	Zitoune	Leaves Branches	Gingival bleeding Bad breath Gingivitis Toothache Mouth ulcer Herpes Stomatitis	Decoction, Infusion, Crude (Gargle Brushing)	[1] [20]
Rosaceae	<i>Sanguisorba Minor</i> Scop.	Faggass laklab	Roots	Gum infectious	Powder	[21]
	<i>Rosa canina</i> L.	Nisrine	Bark Fruit	Gingival bleeding	Decoction (Rinsing, Oral use)	[1]
	<i>Rubus ulmifolius</i> Schott.,	Tût azzarb Serrmû	Leaves	Mouth ulcer	Decoction Crude (Masticatory, Rinsing Oral use)	[2]
	<i>Eriobotryajaponica</i> (Thunb.) Lindl.	Mzah	Leaves/Bark of the stem	Mouth ulcer	Decoction (Direct application)	[20]
Rutaceae	<i>Ruta montana</i> L.	Figel	Leaves	Gingivitis	Decoction (Rinsing)	[1]
Solanaceae	<i>Hyoscyamus</i> sp	Sikran	Seeds	Toothache	Decoction (Gargle)	[1]
Thymeleaceae	<i>Daphne gnidium</i> L.	Alezz^az	Leaves	Toothache	Decoction (Gargle)	[1]
	<i>Daphne gnidium</i> L.	Lazzaz	Leaves	Toothache	Decoction	[1] [21]
Urticaceae	<i>Urtica urens</i>	Hariga	Leaves	Mouth ulcer	Decoction (Rinsing)	[1] [21]
Nitrariaceae	<i>Peganum harmala</i> L.	Lharmel	Seeds	Gingivitis Toothache Mouth ulcer Herpes Bad breath Stomatitis	Maceration in vinegar Decoction (Gargle)	[1]
Lythraceae	<i>Lawsonia inermis</i>	Henna	Leaves	Mouth ulcer	Raw (Direct application)	[20]
Pinaceae	<i>Pinus halepensis</i> L.	Taydâ	Leaf	Antifungal Toothache	Decoction	[8]
	<i>Cedrus atlantica</i> (Endl.) Manettiex Carrière	Kdran	Resin	Toothache	Raw (Applied externally)	[22]
Amaryllidaceae	<i>Allium sativum</i> L.	Touma/ Tishert	Bulbs	Gum disease Toothache	Cataplasm (Direct application)	[20] [21]
Anacardiaceae	<i>Pistacia lentiscus</i> L	Drou	Leaves Barks Root	Toothache Gum	Decoction, Infusion Cataplasm (Mouthwash)	[20] [21]
Araliaceae	<i>Hedera helix</i> L	Lwwaya	Leaves	Toothache	Infusion, Cataplasm Decoction	[21]
Asteraceae	<i>Anacyclus pyrethrum</i> (L.) Link	Tiguentest	Wholeplant	Toothaches	Decoction Infusion	[21]

Continued

	<i>Calendula Arvensis</i> L.	Jamra	Flowers	Gingivitis	Infusion, Compress (Gargles)	[21]
	<i>Chamaemelum nobile</i> L.	Baboneje	Flower	Gum	Decoction (Mouthwash)	[20]
	<i>Artemisia herba alba</i> Asso	Chih	Leaves/ Flower bud	Gum	Decoction (Mouthwash)	[20]
	<i>Bubonium graveolens</i> (Forssk) Maire	Tafsa	Leaf	Toothache	Powder (Applied externally)	[22]
Salicaceae	<i>Populus nigra</i> L.	Safsaf	Leaves	Gum	Decoction (Mouthwash)	[20]
Cupressaceae	<i>Tetraclinis articulata</i> (Vahl) Masters	Azougaâ El arâar	Leaves Fruits	Toothache	Cataplasm, Compress, Maceration	[21]
	<i>Thuja occidentalis</i> L.	Afsa	Leaves	Gum	Decoction (Mouthwash)	[20]
Fabaceae	<i>Retama monosperma</i> (L.) Boiss	Rtam	Stems Roots Leaves	Gum	Infusion, Decoction	[21]
	<i>Acacia nilotica</i> L.	Sllaha	Fruit	Toothache Gingivitis	Powder (Applied externally, Rinsing)	[22]
Myrtaceae	<i>Eugenia caryophyllata</i> Thunb	Qronfel	Cloves	Gum	Maceration	[21]
Iridaceae	<i>Crocus sativus</i> L.	Zaafrahor	Stigma	Gum	Decoction (Mouthwash)	[20]
Juglandaceae	<i>Juglans regia</i> L.	Souak	Bark of the root	Gum	Raw/Cataplasm(Brushing/Gum)	[20] [22]
Burseraceae	<i>Commiphora africana</i> (A. Rich.) Engl.	Oum nas	Gum	Toothache	Powder (Applied externally)	[22]
Euphorbiaceae	<i>Euphorbia officinarum</i> subsp. Echinus (Hook. F. &Coss.) Vindt	Dghmouss	Stem	Toothache	Powder (Applied externally)	[22]
Boraginaceae	<i>Heliotropium curassavicum</i> L.	Lehbalia	Leaf	Toothache	Powder (Appliedexternally)	[22]
Anacardiaceae	<i>Rhus albida</i> schousboe	Zewaya	Bark	Toothache	Decoction	[22]

Asso, *Bubonium graveolens* (Forssk) Maire, *Thuja occidentalis* L. *Acacia nilotica* L, *Crocus sativus* L, *Juglans regia* L. [20] are used to heal gingivitis and gum diseases.

Many species have been used in different regions of Morocco to treat mouth ulcers and stomatitis: *Thymus saturejoides* [18], *Origanum vulgare* L S. [20], *Thymus willdenowii* Boiss, *Origanum compactum* Benth, *Laurus nobilis*, *Olea*

europaea L., *Rubus ulmifolius* Schott., *Peganum harmala* L. [1], *Urtica urens* [21], *Ocimum basilicum* L., *Cinnamomum zeylanicum*, *Eriobotrya japonica* (Thunb.) Lindl., *Lawsonia inermis* S. [20].

According to our review report, the preparation methods used by the Moroccan population were infusion, maceration, decoction, fumigation, cataplasm, and powder. Regarding the mode of use, it can be either by mastication, inhalation, brushing, gargling, or direct application.

3.2. Moroccan Plants with Anticandidal Activity

Several studies have demonstrated the existence of antifungal activity in several plant species belonging to different families [23]. Research studies that explored the anticandidal activity of essential oils (EO) and plant extracts obtained from different regions of Morocco are listed in **Table 2**. Antifungal activity against *Candida albicans* was assessed by the disc diffusion method, minimum inhibitory concentration (MIC), and minimum fungicidal concentration (MFC) values.

In all reported studies, inhibition zone diameter (IZD) was assessed including the disc diameter of 6 mm determined by the agar disc-diffusion method at a concentration of 10 μ l of oil/disc except Abdelghani Aboukhalaf *et al.* [24] who used 20 μ l of oil/disc, and Jeldi *et al.* 2 μ l of oil/disc [25]. MIC represents the lowest essential oil/plant extracts concentration that completely inhibits the growth of *Candida albicans*. It was determined using macro/microdilution methods in mg/ml. MFC corresponds to the lowest concentration at which the incubated microorganism was completely killed [26].

The most cited plant family in the present work was Lamiaceae. Other families that were also mentioned are Myrtaceae, Geraniaceae, Apocynaceae, Cistaceae, Asteraceae, Amaranthaceae, Rutaceae, Cannabaceae Ranunculaceae, Euphorbiaceae, Papaveraceae, Apiaceae, Fagaceae, Lauraceae.

When the inhibition zone diameter is 8 mm or more, the crude extracts have good antimicrobial activity. It is moderate if IZD is 6 - 7 mm; low if it is 4 - 5 mm; very low if it is 2 - 3 mm, or without antimicrobial activity [24]. Regarding essential oils, they are considered active if the IZD is greater than or equal to 15 mm [6]. In this review, the values for IZD of plants extract vary between 6 and 13 mm and those for essential oil from 7 to 85 mm.

The antimicrobial activity of plants extracts is considered significant when $MIC \leq 0.1$ mg/mL, moderate when $0.1 \leq MIC \leq 0.5$ mg/ml, low when $0.5 \leq MIC \leq 1$ mg/ml, and inactive when the value of MIC is higher than 1 mg/ml [27]. In this review, the lowest value of MIC of plants extracts was that found by Benoutman *et al.* (MIC: 0.63 mg/ml. Acetonic extract of *Thymus capitatus*). However, a higher concentration of crude methanol extract of *Cistus monspeliensis* was needed to inhibit the growth of *Candida albicans* (MIC: 200 mg/ml).

The EO isolated from *Mentha suaveolens* exhibited the highest activity against *Candida albicans* (MIC: 0.00069 mg/ml) reported by Oumzil *et al.* (2002) [28]. 20.00 mg/ml of *Laurus nobilis* EO was needed to inhibit *Candida albicans*

Table 2. Moroccan plants with antifungal activity against *Candida albicans*.

Family	Species (Local name)	IZD mm	MIC (mg/ml)	MFC (mg/ml)	References
Lamiaceae	<i>Thymus broussonetii</i>	38.5 ± 0.70 EO	0.25 EO	–	[30]
	<i>T. broussonetii</i>	50.00 ± 1.00 W 49.67 ± 1.15 C EO	0.45 W; 0.46 C EO	0.45 W 0.46 C EO	[34]
	<i>T. broussonetii</i>	50.00 ± 1.00 EO	0.45 EO	0.45 EO	[36]
	<i>Thymus maroccanus</i>	44.5 ± 0.35 EO	0.25 EO	–	[36]
	<i>T. maroccanus</i>	52.33 ± 1.15 EO	0.46 EO	0.46 EO	[36]
	<i>T. maroccanus</i>	52.3 ± 1.2 W 41.3 ± 0.6 CWtF 41.0 ± 1.0 CWF EO	0.16 W 0.14 CWtF 0.03 CWF EO	0.16 W 0.14 CWtF 0.03 CWF EO	[33]
	<i>T. maroccanus</i>	52.33 ± 1.15 W 51.00 ± 1.00 C EO	0.46 W; 0.48 C EO	0.46 W; 0.48 C EO	[34]
	<i>T. maroccanus</i>	31 ± 0.1 EO	–	–	[37]
	<i>Thymus saturejoides</i> (Tazouknit)	42.00 ± 1.00 W 41.33 ± 0.76 C EO	0.89 W; 0.90 C EO	0.89 W; 0.90 C EO	[34]
	<i>T. saturejoides</i>	42.00 ± 1.00 EO	0.89 EO	0.89 EO	[36]
	<i>T. saturejoides</i>	–	0.0059 EO		[38]
	<i>T. saturejoides</i> (Tazouknit)	53 EO	0.9062 EO	0.9062 EO	[39]
	<i>T. saturejoides</i>	–	2.5 EO	5 EO	[40]
	<i>Thymus vulgaris</i> (Ziitra)	–	0.6 EO	0.6 EO	[40]
	<i>Thymus zygis</i> subsp. Gracilis/Timahdite	–	0.15 EO	0.3 EO	[40]
	<i>Thymus zygis</i> subsp. Gracilis/Ain Aghbal	–	1.2 EO	1.2 EO	[40]
	<i>Thymus zygis</i> subsp. Gracilis/Tigrigra	–	0.6 EO	1.2 EO	[40]
	<i>Thymus zygis</i> subsp. Gracilis/Bensmim	–	0.3 EO	0.3 EO	[40]
	<i>Thymus serpyllum</i>	17.33 ± 1.15 EO	3.52 EO	3.52 EO	[36]
	<i>Thymus willdenowii</i>	32 WP; 39 L 32S 27 I EO	0.0069WP 0.0138 L 0.0069 S 0.0138 I EO	–	[31]
	<i>Thymus ciliatus</i>	48.00 ± 1.32 EO	0.43 EO	0.43 EO	[36]

Continued

	<i>Thymus pallidus</i>	–	0.0214 EO	–	[38]
	<i>T. pallidus</i> (Ajellabi)	37.67 ± 0.58 EO	0.90 EO	0.90 EO	[36]
	<i>T. pallidus</i>	85 EO	0.7837 EO	0.7837 EO	[39]
	<i>Thymus leptobotrys</i> (Azoukni)	50.00 ± 0.57 EO	0.23 EO	0.46 EO	[36]
	<i>T. leptobotrys</i>	85 EO	0.33 EO	0.33 EO	[39]
	<i>T. leptobotrys</i>	50.0 ± 0.6 W 41.0 ± 1.0 CWtF 43.0 ± 1.0 CWF EO	0.13 W 0.14CWtF 0.05 CWF EO	0.26W 0.14CWtF 0.05 CWF EO	[33]
	<i>T. leptobotrys</i>	–	0.0022 EO	–	[38]
	<i>T. leptobotrys</i>	27.00 ± 1.00 EO	1.25 EO	1.25 EO	[29]
	<i>Thymus capitatus</i>	–	0.0025 EO 0.63 AE 2.5 ME	0.00375 EO 2.08 ± 0.54 AE 3.75 ± 1.37 ME	[41]
	<i>Mentha * piperita</i>	–	0.0057 EO	–	[38]
	<i>Mentha pulegium</i> (Fliou)	–	0.016 EO	–	[38]
	<i>Mentha spicata</i>	–	0.0093 EO	–	[38]
	<i>Mentha suaveolens</i> (Timija)	–	EO-Pul 0.00069	–	[28]
	<i>Mentha rotundifolia</i>	32 EO	0.63 EO	–	[6]
	<i>Rosmarinus officinalis</i> L. (Azir)		0.0228 EO		[38]
	<i>Origanum compactum</i> (Za'tar)	36.0 ± 1.7 EO	0.216 EO	0.288 EO	[25]
	<i>Origanum compactum</i>	34.0 ± 1.7 EO	0.216 EO	0.288 EO	[25]
	<i>Majorana hortensis</i> (Merdedouche)	35 EO	0.63 EO	0.63 EO	[6]
	<i>Vitex Angus-castus</i> L. (Anguerf)	50 EO	0.53 EO	1.06 EO	[5]
	<i>Salvia officinalis</i> (Salmia)	15 EO	0.63 EO	–	[6]
	<i>Lavandula stoechas</i> (Halhal)	8 EO	1.25 EO	–	[6]
	<i>Lavandula angustifolia</i> (Khzamafassiya)	30 EO	0.63 EO	12.5 EO	[6]
Geraniaceae	<i>Pelargonium graveolens</i>	–	0.0045 EO	–	[38]
	<i>Pelargonium graveolens</i>	45 EO	1.25 EO	–	[6]
	<i>Pelargonium graveolens</i>	13.60 ± 0.17 ME	0.47 ME	–	[27]
		9.42 ± 0.37 DCM	1.87 DCM	–	
		7.53 ± 0.32 HX	3.75 HX	–	
Rutaceae	<i>Citrus limon</i>	–	0.0855 EO	–	[38]

Continued

Cistaceae	<i>Cistus villosus</i>	-	6.25 Crude ME 50 ETAC 3.125 B	-	[32]
	<i>Cistus monspeliensis</i> (Tuzzalabéda)	-	200 Crude ME 100 ETAC 50 B	-	[32]
Cannabaceae	<i>Cannabis sativa</i> L. (Lkif)	12.0 ± 0.7 EO	9.5 EO	-	[17]
Apocynaceae	<i>Caralluma europaea</i> (Guss.) (Daghmous, Zakkum, Tikiwt)	14.50 ± 0.35 EO	3.75 EO	7.5 EO	[42]
	<i>Periploca laevigata</i> Aiton (elhallaba)	12.50 ± 0.35 EO	0.937 EO	1.875 EO	[16]
Ranunculaceae	<i>Nigella sativa</i> (Black cumin, Habbatul baraka, alhabbaassaouda,sanouje)	-	0.008 EO	-	[14]
Amaranthaceae	<i>Chenopodium mural</i> L. (Berremram)	NI EO	-	-	[24]
	<i>Dysphania ambrosioides</i> (L.) Mosyakin & Clemants (Mkhinza)	15 EO	0.075 EO	-	[15]
Myrtaceae	<i>Myrtus communis</i> (Rihàn)	22 EO	5 EO	-	[6]
	<i>Syzygium aromaticum</i> (Qronfel)	14.00 ± 1.00 EO	5 EO	5.00 EO	[29]
	<i>Psidium guajava</i> (Guava)	16 ± 1.73 EO			[43]
Asteraceae	<i>Scolymus hispanicus</i> L. (El guernina)	NI	-	-	[24]
	<i>Senecioanteuphorbium</i> (Achbartou)	22.00 ± 0.17 CA (CCMM L61) 27.00 ± 1.00 CA (CCMM L4) EO	1.024 CA (CCMM L61) 2.048 CA (CCMM L4) EO	4.096 CA (CCMM L61) 4.096 CA (CCMM L4) EO	[12]
	<i>Dittrichia viscosa</i> L. (Trehla or Magramane)	7 ± 0.05 EtOH 10.6 ± 0.03 AE	0.93 EtOH 0.87 AE	1.75 EtOH 1.75 ACE	[10]
Euphorbiaceae	<i>Mercurialis annua</i> L. (Horrigalmalssa)	NI Crude extracts	-	-	[24]
Papaveraceae	<i>Papaver rhoeas</i> L. (Belaaman)	6 ± 0.3 Crude extracts	-	-	[24]
Apiaceae	<i>Foeniculum vulgare</i> Mill (Besbas beldi)	8 ± 0.15 Crude extracts	-	-	[24]
Fagaceae	<i>Quercus suber</i> (fernan)	-	12.5 ME (Bark)	-	[44]
Lauraceae	<i>Laurus nobilis</i>	7.00 EO	20.00 EO	20.00 EO	[29]

IZD: Inhibition zone diameter. MIC: Minimum inhibitory concentration. MFC: Minimum fungicidal concentration, W: wild; C: cultivated, CWtF: Cultivated without Fertilizer; CWF: Cultivated with Fertilizer; WP: Whole Plant; L: Leaves; S: Stems, I: Inflorescences, EO: Essential oil; AE: Acetonic extract; EtOH: ethanolic extract; ME: Methanolic extract; EO-Pul: Essential oil rich in pulegone, DCM: Dichloromethane, HX: Hexane. ETAC: ethyl acetate; B: Butanol; NI: No inhibition; CA: *Candida albicans*.

growth [29].

These findings suggest that EO and plant extract can be considered as a potential source of natural antimicrobials. Therefore, their utilization in addition to antifungal drugs for the treatment of some candidiasis due to *C. albicans* is possible. It may constitute a promising strategy to overcome the intense use of antifungal drugs and reduce the minimum effective dose, thus minimizing their toxic side effects and the treatment cost [30].

The effectiveness of essential oils is influenced by the plant's origin and the parts used in their preparation [10]. Essential oil of *Thymus willdenowii* stems (MIC = 0.0069 mg/ml) was more active than that of leaves (MIC = 0.0138 mg/ml) and inflorescences (MIC = 0.0138 mg/ml) [31].

The type of organic extract (Hexane, methanol, dichloromethane, ethanol, ethyl acetate, butanol, acetonic/extract) also affects the antifungal activity as shown by the studies of S. El Aanachi *et al.*, H. Bouamama *et al.* and I. Mssillou *et al.* [10] [27] [32].

The activity of essential oils and plant extracts also depends on whether the plants are wild or cultivated with or without fertilizers [33] [34].

In this review, *Thymus* is one of the most cited genera. 21 species represent the genus *Thymus* (Lamiaceae) in Morocco. IMC values vary from 0.0022 mg/ml to 0.9 mg/ml. This variation may depend on the chemical composition of thyme essential oils, which in turn depends on several factors such as species, genetic heritage, origin, environmental influences, and growth stage [35].

3.3. Moroccan Plants with Antifungal Activities

In recent years, several studies have been devoted to the study of plant extracts to develop new antifungal compounds that can be used in medical fields to treat various infections and in agricultural fields to control post-harvest diseases of fruits and vegetables.

A promising antifungal activity for several plants used in Morocco against many fungal species involved in plant or human diseases has been evidenced. According to studies, this activity exists whether it was in essential oils, plant extracts, or powders. These plants are listed in **Table 3**.

These results may provide support for further studies to evaluate the antifungal action of Moroccan plants against *Candida albicans*. The effectiveness of phytotherapy is well-proven. It is currently one of the main health care in Morocco. However, numerous studies show that medicinal plants can be carcinogenic, teratogenic, or even endanger life conditions. The toxicity of a plant is related to several factors, including the type and quantity of chemical compounds present in the plants, the quantity consumed, the exposure time, the part used (Seeds, leaves, oil, bark, stem, root), the climate and the soil, body chemistry, genetic and the preparation method used (Extraction: Solvent types, essential oils). Guidelines must be established to protect medicinal and aromatic plants and to regulate their local use by the population [56] [57].

Table 3. Moroccan plants with antifungal activities.

Family	Species	Forme	References
Rosaceae	<i>Rubus ulmifolius</i> Schott	Powder	[45]
Amaranthaceae	<i>Hammada scoparia</i>	Powder	[45]
Cistaceae	<i>Halimium antiatlanticum</i>	Powder	[45]
	<i>Halimium umbellatum</i>	Powder	[45]
	<i>Cistus Creticus</i>	EO	[46]
Anacardiaceae	<i>Pistacia atlantica</i>	Powder	[45]
Fabaceae	<i>Ceratoniasiliqua</i>	Powder	[45]
Cupressaceae	<i>Cupressus atlantica</i>	EO	[11]
Cupressaceae	<i>Juniperus phoenicea</i> (Ar'ar)	EO	[47]
Cyperaceae	<i>Cyperus longus</i>	EO	
Lamiaceae	<i>Mentha viridis</i> (Naanaa)	EO	[19]
	<i>Thymus bleicherianus</i>	EO	[35]
	<i>Origanum compactum</i>	EO	[23]
	<i>Thymus glandulosus</i>	EO	[23]
	<i>Lavandula dentata</i>	Organic extract	[23]
	<i>Origanum vulgare</i>	Organic extract	[23]
Pinaceae	<i>Pinus halepensis</i> Mill	Extracts and EO	[48]
Gentianaceae	<i>Centaurium erythraea</i> Rafn (Korsathaya)	Extracts and EO	[49]
Zingiberaceae	<i>Zingiber officinale</i> extracts roscoe	Ethyl acetate extracts Ethanol extracts Water extracts	[50]
Rutaceae	<i>Citrus limonum</i>	EO	[23]
Asteraceae	<i>Anvillea radiata</i>	Organic extract	[23]
	<i>Silybum marianum</i> (L.) Gaertn	Decoction	[51]
	<i>Ighermia pinifolia</i>	Powder	[45]
	<i>Inula viscosa</i>	Powder	[45]
	<i>Artemisia absinthium</i> L. (<i>Chiba</i>)	Decoction	[8]
	<i>Anthemis tenuisecta</i>	EO	[52]
Myrtaceae	<i>Artemisia campestris</i> L.	Extracts and EO	[53]
	<i>Eucalyptu scitriodora</i> L. (kelitto)	Powder	[8]
	<i>Eucalyptus globulus</i> L. (Kalitouse)	Organic extract	[23]
Lauraceae	<i>Cinnamomum zeylanicum</i>	EO	[23]
Apiaceae	<i>Cuminum cyminum</i> L. (Kammun)	Cooking	[8]
Amarillydaceae	<i>Allium sativum</i> L. (Touma)	Cooking	[8]
	<i>Allium cepa</i> L. (Bsal)	Maceration	[8]

Continued

Apocynaceae	<i>Nerium oleander</i> L. (Dafla)	Infusion	[8]
Salicaceae	<i>Salix alba</i> L. (Ud el-mâ)	Decoction	[8]
Vitaceae	<i>Vitis vinifera</i> L. (La'anb)	Powder	[8]
Arecaceae	<i>Phoenix dactylifera</i> L. (Tamer)	Extract	[54]
Urticaceae	<i>Urtica dioica</i> L. (Hourriga, al quarâs)	Powder, Infusions Decoction	[55]

4. Conclusion

The research of new active principles extracted from medicinal and aromatic plants is nowadays a priority for many countries to face the increasing resistance of the human body against drugs. In this article, a review of existing knowledge on the antifungal activity of different plants of Moroccan origin was carried out. First, plants used by the Moroccan population for oral diseases were cited. Then, studies that discussed the anti-candidal activity of essential oils and extracts from Moroccan plants were reported. This review represents the unique comprehensive overview of Moroccan plants active against *Candida albicans*. Finally, the plants presenting an antifungal action against fungal species involved in plant or human diseases were listed. The anticandidal activity has been proven for several Moroccan plants, and this review will supply a baseline to identify this activity for other Moroccan plants. Further investigation should be undertaken in the future to study the toxicity of essential oils and plant extracts and to determine the optimal concentrations for clinical use. This article will provide supporting data and perspectives for future research studies on the antifungal activities of Moroccan plants.

Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] Najem, M., Harouak, H., Ibijbijen, J. and Nassiri, L. (2020) Oral Disorders and Ethnobotanical Treatments: A Field Study in the Central Middle Atlas (Morocco). *Heliyon*, **6**, e04707. <https://doi.org/10.1016/j.heliyon.2020.e04707>
- [2] Fan, F., *et al.* (2022) *Candida Albicans* Biofilms: Antifungal Resistance, Immune Evasion, and Emerging Therapeutic Strategies. *International Journal of Antimicrobial Agents*, **60**, Article ID: 106673. <https://doi.org/10.1016/j.ijantimicag.2022.106673>
- [3] Khan, H., *et al.* (2017) Plant Bioactive Molecules Bearing Glycosides as Lead Compounds for the Treatment of Fungal Infection: A Review. *Biomedicine & Pharmacotherapy*, **93**, 498-509. <https://doi.org/10.1016/j.biopha.2017.06.077>
- [4] Xiao, Y., *et al.* (2022) Comparison of Topical Antifungal Agents for Oral Candidiasis Treatment: A Systematic Review and Meta-Analysis. *Oral Surgery, Oral Medicine, Oral Pathology and Oral Radiology*, **133**, 282-291. <https://doi.org/10.1016/j.oooo.2021.10.023>

- [5] Asdadi, A., *et al.* (2015) Study on Chemical Analysis, Antioxidant and *in Vitro* Antifungal Activities of Essential Oil from Wild *Vitex agnus-castus* L. Seeds Growing in Area of Argan Tree of Morocco against Clinical Strains of *Candida* Responsible for Nosocomial Infections. *Journal of Medical Mycology*, **25**, e118-e127. <https://doi.org/10.1016/j.mycmed.2015.10.005>
- [6] Chebaibi, A., Marouf, Z., Rhazi-Filali, F., Fahim, M. and Ed-Dra, A. (2016) Évaluation du pouvoir antimicrobien des huiles essentielles de sept plantes médicinales récoltées au Maroc. *Phytothérapie*, **14**, 355-362. <https://doi.org/10.1007/s10298-015-0996-1>
- [7] Ouraéni, D., *et al.* (2005) Approche thérapeutique des dermatophyties par les huiles essentielles de plantes aromatiques marocaines. *Phytothérapie*, **3**, 3-12. <https://doi.org/10.1007/s10298-005-0058-1>
- [8] Bouyahya, A., Abrini, J., Et-Touys, A., Bakri, Y. and Dakka, N. (2017) Indigenous Knowledge of the Use of Medicinal Plants in the North-West of Morocco and Their Biological Activities. *European Journal of Integrative Medicine*, **13**, 9-25. <https://doi.org/10.1016/j.eujim.2017.06.004>
- [9] Lahkimi, A., Louaste, B., Nechad, I., Chaouch, M. and Eloutassi, N. (2020) Antibacterial, Antifungal and Antioxidant Activity of *Lavandula Angustifolia* of the Middle Atlas Central (Morocco). *Moroccan Journal of Chemistry*, **8**, 905-918.
- [10] Mssillou, I., *et al.* (2022) Phytochemical Characterization, Antioxidant Activity, and *in Vitro* Investigation of Antimicrobial Potential of *Dittrichia viscosa* L. Leaf Extracts against Nosocomial Infections. *Acta Ecologica Sinica*, **42**, 661-669. <https://doi.org/10.1016/j.chnaes.2021.09.021>
- [11] Boukssaim, H., *et al.* (2013) Caractérisation chimique et microbiologique des huiles essentielles des rameaux, des cônes et du bois de *Cupressus atlantica*, arbre forestier endémique du Maroc. *Phytothérapie*, **11**, 294-300. <https://doi.org/10.1007/s10298-013-0803-9>
- [12] Elhidar, N., *et al.* (2019) Chemical Composition, Antimicrobial Activities and Synergistic Effects of Essential Oil from *Senecio anteuphorbium*, a Moroccan Endemic Plant. *Industrial Crops and Products*, **130**, 310-315. <https://doi.org/10.1016/j.indcrop.2018.12.097>
- [13] Abudunia, A.M., *et al.* (2017) Evaluation of Essential Oils for Antimicrobial Activity from Some Moroccan Aromatic Plants Medicinal. *Journal of Materials and Environmental Science*, **8**, 4240-4245. <https://doi.org/10.26872/jmes.2017.8.12.446>
- [14] Tiji, S., Rokni, Y., Benayad, O., Laaraj, N., Asehraou, A. and Mimouni, M. (2021) Chemical Composition Related to Antimicrobial Activity of Moroccan *Nigella sativa* L. Extracts and Isolated Fractions. *Evidence-Based Complementary and Alternative Medicine*, **2021**, Article ID: 8308050. <https://doi.org/10.1155/2021/8308050>
- [15] Ait Sidi Brahim, M., *et al.* (2015) *Chenopodium ambrosioides* var. *ambrosioides* Used in Moroccan Traditional Medicine Can Enhance the Antimicrobial Activity of Conventional Antibiotics. *Industrial Crops and Products*, **71**, 37-43. <https://doi.org/10.1016/j.indcrop.2015.03.067>
- [16] Ait Dra, L., *et al.* (2017) Chemical Composition, Antioxidant and Evidence Antimicrobial Synergistic Effects of *Periploca laevigata* Essential Oil with Conventional Antibiotics. *Industrial Crops and Products*, **109**, 746-752. <https://doi.org/10.1016/j.indcrop.2017.09.028>
- [17] Nafis, A., *et al.* (2019) Antioxidant Activity and Evidence for Synergism of *Cannabis sativa* (L.) Essential Oil with Antimicrobial Standards. *Industrial Crops and Products*, **137**, 396-400. <https://doi.org/10.1016/j.indcrop.2019.05.032>

- [18] El Yaagoubi, M., *et al.* (2021) A Review on Moroccan Thymus Species: Traditional Uses, Essential Oils Chemical Composition and Biological Effects. *Journal of Ethnopharmacology*, **278**, Article ID: 114205. <https://doi.org/10.1016/j.jep.2021.114205>
- [19] Bouyahya, A., *et al.* (2020) Ethnomedicinal Use, Phytochemistry, Pharmacology, and Food Benefits of *Thymus capitatus*. *Journal of Ethnopharmacology*, **259**, Article ID: 112925. <https://doi.org/10.1016/j.jep.2020.112925>
- [20] Zougagh, S., Belghiti, A., Rochd, T., Zerdani, I. and Mouslim, J. (2019) Medicinal and Aromatic Plants Used in Traditional Treatment of the Oral Pathology: The Ethnobotanical Survey in the Economic Capital Casablanca, Morocco (North Africa). *Natural Products and Bioprospecting*, **9**, 35-48. <https://doi.org/10.1007/s13659-018-0194-6>
- [21] Belhaj, S., Dahmani, J., Belahbib, N. and Zidane, L. (2020) Ethnopharmacological and Ethnobotanical Study of Medicinal Plants in the High Atlas Central, Morocco. *Ethnobotany Research and Applications*, **20**, 1-40. <https://doi.org/10.32859/era.20.18.1-40>
- [22] Idm'hand, E., Msanda, F. and Cherifi, K. (2020) Ethnobotanical Study and Biodiversity of Medicinal Plants Used in the Tarfaya Province, Morocco. *Acta Ecologica Sinica*, **40**, 134-144. <https://doi.org/10.1016/j.chnaes.2020.01.002>
- [23] Lagrouh, F., Dakka, N. and Bakri, Y. (2017) The Antifungal Activity of Moroccan Plants and the Mechanism of Action of Secondary Metabolites from Plants. *Journal of Medical Mycology*, **27**, 303-311. <https://doi.org/10.1016/j.mycmed.2017.04.008>
- [24] Aboukhalaf, A., El Amraoui, B., Tabatou, M., Ferreira da Rocha, J.M. and Belahsen, R. (2020) Screening of the Antimicrobial Activity of Some Extracts of Edible Wild Plants in Morocco. *Functional Foods in Health and Disease*, **10**, 265-273. <https://doi.org/10.31989/ffhd.v10i6.718>
- [25] Jeldi, L., Taarabt, K.O., Mazri, M.A., Ouahmane, L. and Alfeddy, M.N. (2022) Chemical Composition, Antifungal and Antioxidant Activities of Wild and Cultivated *Origanum compactum* Essential Oils from the Municipality of Chaoun, Morocco. *South African Journal of Botany*, **147**, 852-858. <https://doi.org/10.1016/j.sajb.2022.03.034>
- [26] Chraibi, M., Farah, A., Lebrazi, S., El Amine, O., Iraqi Houssaini, M. and Fikri-Benbrahim, K. (2016) Antimycobacterial Natural Products from Moroccan Medicinal Plants: Chemical Composition, Bacteriostatic and Bactericidal Profile of *Thymus satureioides* and *Mentha pulegium* Essential Oils. *Asian Pacific Journal of Tropical Biomedicine*, **6**, 836-840. <https://doi.org/10.1016/j.apjtb.2016.08.002>
- [27] El Aanachi, S., Gali, L., Nacer, S.N., Bensouici, C., Dari, K. and Aassila, H. (2020) Phenolic Contents and *in Vitro* Investigation of the Antioxidant, Enzyme Inhibitory, Photoprotective, and Antimicrobial Effects of the Organic Extracts of *Pelargonium graveolens* Growing in Morocco. *Biocatalysis and Agricultural Biotechnology*, **29**, Article ID: 101819. <https://doi.org/10.1016/j.bcab.2020.101819>
- [28] Oumzil, H., *et al.* (2002) Antibacterial and Antifungal Activity of Essential Oils of *Mentha suaveolens*. *Phytotherapy Research*, **16**, 727-731. <https://doi.org/10.1002/ptr.1045>
- [29] Razzouk, S., Mazri, M.A., Jeldi, L., Mnasri, B., Ouahmane, L. and Alfeddy, M.N. (2022) Chemical Composition and Antimicrobial Activity of Essential Oils from Three Mediterranean Plants against Eighteen Pathogenic Bacteria and Fungi. *Pharmaceutics*, **14**, Article No. 1608. <https://doi.org/10.3390/pharmaceutics14081608>
- [30] Saad, A., Fadli, M., Bouaziz, M., Benharref, A., Mezrioui, N.-E. and Hassani, L. (2010) Anticandidal Activity of the Essential Oils of *Thymus maroccanus* and

- Thymus broussonetii* and Their Synergism with Amphotericin B and Fluconazol. *Phytomedicine*, **17**, 1057-1060. <https://doi.org/10.1016/j.phymed.2010.03.020>
- [31] Ouknin, M., Romane, A., Costa, J. and Majidi, L. (2019) Comparative Study of the Chemical Profiling, Antioxidant and Antimicrobial Activities of Essential Oils of Different Parts of *Thymus willdenowii* Boiss & Reut. *Natural Product Research*, **33**, 2398-2401. <https://doi.org/10.1080/14786419.2018.1443089>
- [32] Bouamama, H., Noël, T., Villard, J., Benharref, A. and Jana, M. (2006) Antimicrobial Activities of the Leaf Extracts of Two Moroccan Cistus L. Species. *Journal of Ethnopharmacology*, **104**, 104-107. <https://doi.org/10.1016/j.jep.2005.08.062>
- [33] Alaoui Jamali, C., *et al.* (2014) Cultivation and the Application of Inorganic Fertilizer Modifies Essential Oil Composition in Two Moroccan Species of Thymus. *Industrial Crops and Products*, **62**, 113-118. <https://doi.org/10.1016/j.indcrop.2014.08.017>
- [34] El Bouzidi, L., *et al.* (2013) Chemical Composition, Antioxidant and Antimicrobial Activities of Essential Oils Obtained from Wild and Cultivated Moroccan Thymus Species. *Industrial Crops and Products*, **43**, 450-456. <https://doi.org/10.1016/j.indcrop.2012.07.063>
- [35] Amarti, F., *et al.* (2008) Composition chimique et activité antimicrobienne des huiles essentielles de *Thymus capitatus* et de *Thymus bleicherianus* du Maroc. *Phytothérapie*, **6**, 342-347. <https://doi.org/10.1007/s10298-008-0346-7>
- [36] Jamali, C.A., *et al.* (2012) Chemical Composition and Antioxidant and Anticandidal Activities of Essential Oils from Different Wild Moroccan Thymus Species. *Chemistry & Biodiversity*, **9**, 1188-1197. <https://doi.org/10.1002/cbdv.201200041>
- [37] Belaqziz, R., Bahri, F., Romane, A., Antoniotti, S., Fernandez, X. and Duñach, E. (2013) Essential Oil Composition and Antibacterial Activity of the Different Parts of *Thymus maroccanus* Ball: An Endemic Species in Morocco. *Natural Product Research*, **27**, 1700-1704. <https://doi.org/10.1080/14786419.2013.768989>
- [38] El Asbahani, A., *et al.* (2015) Chemical Composition and Antimicrobial Activity of Nine Essential Oils Obtained by Steam Distillation of Plants from the Souss-Massa Region (Morocco). *Journal of Essential Oil Research*, **27**, 34-44. <https://doi.org/10.1080/10412905.2014.964426>
- [39] Asdadi, A., *et al.* (2014) Chemical Composition and Anticandidal Effect of Three Thymus Species Essential Oils from Southwest of Morocco against the Emerging Nosocomial Fluconazole-Resistant Strains. *Journal of Biology, Agriculture and Healthcare*, **4**, 16-26.
- [40] Drioiche, A., *et al.* (2022) Correlation between the Chemical Composition and the Antimicrobial Properties of Seven Samples of Essential Oils of Endemic Thymes in Morocco against Multi-Resistant Bacteria and Pathogenic Fungi. *Saudi Pharmaceutical Journal*, **30**, 1200-1214. <https://doi.org/10.1016/j.jsps.2022.06.022>
- [41] Benoutman, A., *et al.* (2022) Phytochemical Composition, Antioxidant and Antifungal Activity of *Thymus capitatus*, a Medicinal Plant Collected from Northern Morocco. *Antibiotics*, **11**, Article No. 681. <https://doi.org/10.3390/antibiotics11050681>
- [42] Dra, L.A., *et al.* (2019) Chemical Characterization and *in Vitro* Antimicrobial Activity of *Caralluma europaea* Essential Oil and Its Synergistic Potential with Conventional Antibiotics. *Journal of Advances in Medical and Pharmaceutical Sciences*, **19**, 1-11. <https://doi.org/10.9734/jamps/2018/v19i430094>
- [43] Lahlou, Y. (2022) Chemical Composition, Antioxidant and Antimicrobial Activities of Moroccan Species of *Psidium guajava* Extracts. *Roczniki Panstwowego Zakladu*

- Higieny*, **73**, 65-77. <https://doi.org/10.32394/rpzh.2022.0199>
- [44] Hassikou, R., Oulladi, H. and Arahou, M. (2014) Activité antimycosique des extraits du chêne-liège *Quercus suber* sur *Trichophyton rubrum* et *Candida albicans*. *Phytothérapie*, **12**, 206-212. <https://doi.org/10.1007/s10298-014-0874-2>
- [45] Talibi, I., *et al.* (2012) Antifungal Activity of Some Moroccan Plants against *Geotrichum candidum*, the Causal Agent of Postharvest Citrus Sour Rot. *Crop Protection*, **35**, 41-46. <https://doi.org/10.1016/j.cropro.2011.12.016>
- [46] Ait Lahcen, S., *et al.* (2020) Chemical Composition, Antioxidant, Antimicrobial and Antifungal Activity of Moroccan *Cistus creticus* Leaves. *Chemical Data Collections*, **26**, Article ID: 100346. <https://doi.org/10.1016/j.cdc.2020.100346>
- [47] Ait-Ouazzou, A., *et al.* (2012) Evaluation of the Chemical Composition and Antimicrobial Activity of *Mentha pulegium*, *Juniperus phoenicea*, and *Cyperus longus* Essential Oils from Morocco. *Food Research International*, **45**, 313-319. <https://doi.org/10.1016/j.foodres.2011.09.004>
- [48] El Omari, N., *et al.* (2021) Phytochemical and Biological Activities of *Pinus halepensis* Mill. and Their Ethnomedicinal Use. *Journal of Ethnopharmacology*, **268**, Article ID: 113661. <https://doi.org/10.1016/j.jep.2020.113661>
- [49] El Menyiy, N., *et al.* (2021) Phytochemical Properties, Biological Activities and Medicinal Use of *Centaureum erythraea* Rafn. *Journal of Ethnopharmacology*, **276**, Article ID: 114171. <https://doi.org/10.1016/j.jep.2021.114171>
- [50] Yousfi, F., Abrigach, F., Petrovic, J.D., Sokovic, M. and Ramdani, M. (2021) Phytochemical Screening and Evaluation of the Antioxidant and Antibacterial Potential of *Zingiber officinale* Extracts. *South African Journal of Botany*, **142**, 433-440. <https://doi.org/10.1016/j.sajb.2021.07.010>
- [51] Marmouzi, I., Bouyahya, A., Ezzat, S.M., El Jemli, M. and Kharbach, M. (2021) The Food Plant *Silybum marianum* (L.) Gaertn: Phytochemistry, Ethnopharmacology and Clinical Evidence. *Journal of Ethnopharmacology*, **265**, Article ID: 113303. <https://doi.org/10.1016/j.jep.2020.113303>
- [52] Mziouid, A., Chebli, B., Berrabah, M., Heimeur, N. and Mayad, E.H. (2020) Antifungal Activities of Essential Oil from Moroccan Endemic *Anthemis tenuisecta* and Seed Emergence. *Materials Today: Proceedings*, **27**, 3108-3113. <https://doi.org/10.1016/j.matpr.2020.03.686>
- [53] Dib, I., Angenot, L., Mihamou, A., Ziyat, A. and Tits, M. (2017) *Artemisia campestris* L.: Ethnomedicinal, Phytochemical and Pharmacological Review. *Journal of Herbal Medicine*, **7**, 1-10. <https://doi.org/10.1016/j.hermed.2016.10.005>
- [54] Taleb, H., Maddocks, S.E., Morris, R.K. and Kanekanian, A.D. (2016) Chemical Characterisation and the Anti-Inflammatory, Anti-Angiogenic and Antibacterial Properties of Date Fruit (*Phoenix dactylifera* L.). *Journal of Ethnopharmacology*, **194**, 457-468. <https://doi.org/10.1016/j.jep.2016.10.032>
- [55] Ait Haj Said, A., Sbai El Otmani, I., Derfoufi, S. and Benmoussa, A. (2016) Mise en valeur du potentiel nutritionnel et thérapeutique de l'ortie dioïque (*Urtica dioica* L.). *Hegel*, **3**, 280-292. <https://doi.org/10.4267/2042/61406>
- [56] El Hassani, F.Z. (2020) Characterization, Activities, and Ethnobotanical Uses of *Mentha* Species in Morocco. *Heliyon*, **6**, e05480. <https://doi.org/10.1016/j.heliyon.2020.e05480>
- [57] Kharchoufa, L., Merrouni, I.A., Yamani, A. and Elachouri, M. (2018) Profile on Medicinal Plants Used by the People of North Eastern Morocco: Toxicity Concerns. *Toxicon*, **154**, 90-113. <https://doi.org/10.1016/j.toxicon.2018.09.003>