

Hydro-Ethanol Extract of *Persea americana* Fruit Pulp Ameliorates Dyslipidaemia and Cardiotoxicity Exerted by Alloxan-Induced Diabetes Mellitus

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Abstract

Diabetes-induced dyslipidaemia has been associated with an increased risk of atherosclerosis and coronary heart diseases. Persea americana fruit has been reported to possess anti-diabetic properties. Therefore, this study assessed the lipid profile and likely cardio-protective effects of hydroethanolic extracts of P. americana fruits in alloxan-induced diabetic Wistar rats. Thirty-five male rats (150 \pm 30 g) were divided into 5 groups (n = 7) and treated orally as follows; groups I-II were normal animals treated with distilled water (0.3 ml/day) and P. americana (300 mg/kg) only respectively. Animals in groups III-V were made diabetic using alloxan monohydrate (100 mg/kg i.p.) and treated orally with distilled water (0.3 ml/day), P. americana (300 mg/kg) and glibenclamide (5 mg/kg) respectively for 21 days. Fasting blood glucose level was monitored prior to, after induction of diabetes mellitus, and on day 21 post-treatment, respectively. Thereafter, retro-orbital blood samples were collected after anaesthesia and analysed for insulin, total cholesterol (TC), triglyceride (TG), low density lipoprotein (LDL), high density lipoprotein (HDL) levels, apolipoproteins A1 and B, superoxide dismutase (SOD) and catalase activities, reduced glutathione (GSH), Vitamin C and malondialdehyde levels, respectively. VLDL, atherogenic index (AI) and ApoB/A1 ratio were estimated mathematically. Pancreatic and cardiac structures were also investigated using Haematoxylin and Eosin stains. Treatment with P. ame*riacana* extracts reduced (p < 0.05) fasting blood glucose and increased (p < 0.05) 0.05) insulin level compared to the diabetic untreated group. Values of TG, TC, LDL, Apo-B, AI, Apo-B/A1 ratio decreased significantly (p < 0.05) while

HDL increased significantly with extract treatment compared to diabetic untreated group. Serum malondialdehyde levels were significantly reduced in the extract treated diabetic group, while SOD and GSH significantly increased compared to diabetic untreated. Histological studies showed partial attenuation of diabetic-induced pathologies in the *P. americana* treated diabetic group. The hydro-ethanol fruit extract of *Persea americana* attenuates diabetes induced dyslipidaemia and reduces the susceptibility to cardiac impairment in experimental diabetes mellitus.

Keywords

Persea americana, Diabetes Mellitus, Dyslipidaemia and Alloxan

1. Introduction

Diabetes mellitus is a chronic metabolic disorder that occurs either when the pancreas does not produce enough insulin or when the body cannot effectively use the insulin it produces [1]. Cardiovascular risk factors such as obesity, hypertension and dyslipidaemia are common in patients with diabetes mellitus and this places them at increased risk for cardiac events [2] [3]. In fact, cardiovascular diseases have been described as the major cause of morbidity and mortality (accounting for about 50% - 70% of deaths) in diabetic patients [4]. Hence, one of the primary goals in the management of diabetes mellitus, aside from glycaemic control, is to reduce the development of cardiovascular risk factors such as dyslipidaemia and hypertension in diabetic patients. Diabetic dyslipidaemia, a condition characterised by elevated fasting and postprandial triglycerides, reduced high density lipoprotein (HDL)-cholesterol, and elevated low-density lipoprotein (LDL)-cholesterol, is often described as one of the major risk factors for the development of cardiovascular diseases in diabetes mellitus [2] [5]. The lipid changes associated with diabetes mellitus have been attributed to an increase in the mobilisation of free fatty acid from adipose tissue stores secondary to insulin resistance [6].

Despite the availability of various pharmaceuticals designed to lower blood glucose level and ameliorate dyslipidaemia in diabetes mellitus, mortality from coronary heart disease arising from serum cholesterol levels has been observed to be increased [7] [8]. Aside from the undesirable side effects such as excessively low blood sugar, stomach upset, skin rash or itching, weight gain, kidney complications, bloating, diarrhoea etc, the high cost of these pharmaceuticals in low to middle income countries has led to a preference for hypoglycemics and serum cholesterol regulating agents (anticholesteremics) that are of plant origin [9] [10]. Furthermore, the phytochemicals in these medicinal plants (e.g., *Artemisia pallens, Bidens pilosa, Bixa orellana, Teramnus labialis, Cinnamomum zeylanicum, Croton cajucara, Allium spp., Musa sapientum*, etc) have been observed to exert a multifactorial and multifaceted approach in the management of

diabetic mellitus [9].

One of such medicinal plants is *Persea americana*, commonly known as avocado, a flowering plant from the Lauracea family that is native to south central Mexico and cultivated in tropical and Mediterranean climes throughout the world [11]. Various parts of *P. americana* ranging from the leaf, seed, and fruit pulp have been extracted with different solvents and reported to be rich in bioactive compounds with anti-diabetic and antioxidant [12], hypolidaemic, anti-aging [13], and anti-cancer [14] activities. Its effect on dyslipidaemia and cardio-toxicity in diabetes mellitus is however yet to be fully elucidated. This study was therefore designed to evaluate the likely ameliorative potential of hydro-ethanol extract of *Persea americana* fruit pulp on dyslipidaemia and cardiotoxicity in diabetes mellitus using the alloxan-induced diabetic rat model.

2. Material and Method

2.1. Collection and Preparation of Persea americana Fruit Extract

Fresh fruits of *Persea Americana* were purchased from the local market in Ibadan, Nigeria and identified at the University of Ibadan Herbarium (UIH-22767). The fresh fruits were washed, peeled and seeds removed. The edible fruit pulp was chopped into small pieces, air dried for 24 hrs, and subsequently oven dried at 30° C - 40° C for 4hrs. Thereafter, the dried fruit pulp was ground into powder and 850g of it was weighed and macerated in 5000 ml of 70% ethanol for 72 hours with constant stirring. The hydro-ethanol filtrate was collected using a muslin bag and Whatman No. 1 filter paper and concentrated using a rotary evaporator. The hydro-ethanol extract of *P. americana* fruit obtained was then stored at 4°C until use.

2.2. Animals and Induction of Diabetes Mellitus

Twenty-five male Wistar rats $(150 \pm 30 \text{ g})$ were housed in well-aerated laboratory cages, maintained on standard rat chow and water *ad libitum*. They were exposed to natural alternating day and night cycles in the animal house. Animals were kept in accordance with the ethics and guideline for the use of experimental animals in the laboratory of the University of Ibadan Animal Care and Use Research Ethics Committee (UI-ACUREC) and that of the Guide for the Care and Use of Laboratory Animals (1996, published by National Academy Press, 2101 Constitution Ave. NW, Washington, DC 20055, USA). Experimental diabetes mellitus was induced using a single intra-peritoneal administration of alloxan monohydrate (100 mg/kg) to overnight fasted animals [15]. The blood glucose level of each experimental animals was thereafter monitored (glucose oxidase method) at intervals of 24 hours for 72 hours using the tail tipping method [16]. Animals that presented with relatively constant fasting blood glucose (FBG) values above 250 mg/dL were considered as being diabetic and recruited for the study [17].

2.3. Animal Grouping and Experimental Protocol

The animals were randomly grouped (n = 5) as follows; group I, control was

treated orally with distilled water (0.3 ml/day). Animals in group II were treated with *Persea americana* (300 mg/kg) only [12] while animals in groups III – V were made diabetic and treated with distilled water (0.3 ml/day), *P. americana* (300 mg/kg) and glibenclamide (5 mg/kg) [18], respectively. All treatments were carried out orally for 21 days using an orogastric tube.

2.4. Biochemical Assays

Blood samples were collected from the retro-orbital sinus after light anesthesia (ketamine: zylazine) using a heparinized capillary tube into plain sample bottles. The samples collected were allowed to stand at room temperature for 10 minutes and then centrifuged for 10 minutes at 3500 rpm. The clear serum obtained was analyzed for total cholesterol, high density lipoprotein (HDL), triglycerides, insulin, apolipoprotein A-1 (Apo A-1), and apolipoprotein B using commercially available laboratory kits. Low density lipoproteins (LDL) and very low-density lipoproteins (VLDL) were estimated using Friedewalds equation [19] while atherogenic index was calculated using the formula = log(TG/HDL-C). Serum antioxidants (superoxide dismutase (SOD) [20], catalase [21], reduced glutathione (GSH) [22], ascorbic acid [23] and lipid peroxidation (thiobabituric acid reactive substances (TBARS) [24] were also analyzed in the serum samples.

2.5. Histological Studies

The heart and pancreas of each animal were also excised, fixed in 10% formalin and evaluated for structure and architectural integrity using Haematoxylin and Eosin staining techniques [25] [26].

2.6. Statistical Analysis

Data obtained are presented as mean \pm standard error of mean (SEM) and statistical evaluation was carried out using Graph Pad Prism 7.0 (GraphPad Software Inc., USA). Statistical significance was assessed using one-way analysis of variance (ANOVA) and pairwise comparisons were conducted using Newman Keul post-hoc test.

3. Results

3.1. Effect of *Persea americana* on Fasting Blood Glucose Level in Control and Experimental Groups

Fasting blood glucose level (mg/dL) in group I, the control group, and group II (*P. americana* only) was relatively constant throughout the duration of the study. Values obtained in the groups III (75.8 \pm 4.5 vs. 312.8 \pm 15.7), IV (72.5 \pm 8.2 vs. 305.4 \pm 20.5) and V (68.8 \pm 7.2 vs. 325.5 \pm 15.5) on day 0 after the induction of diabetes mellitus were significantly increased (p < 0.05) compared to initial blood glucose values obtained in these experimental groups respectively. On day 21, values in group III where 12.0% reduced compared to values on day 0 while values obtained in groups IV and V were reduced by 52.4% and 65.39% respec-

tively compared to values obtained within these groups on day 0. Fasting blood values obtained in groups IV and V on day 21, though reduced compared to diabetic untreated (group II), were still increased (p < 0.05) compared to control (**Table 1**).

3.2. Effect of *P. americana* on Insulin Levels in Control and Experimental Groups

Insulin levels (μ U/mL) at the end of the study were reduced in diabetic untreated group (group III) compared to group I (control) and group II (*P. americana* only) respectively. Values obtained in group IV (diabetic + *P. americana*) were significantly increased compared to groups I, II and III respectively. Insulin values obtained in group V (diabetic + glibenclamide) were comparable to group III and significantly reduced compared to groups I, II and IV respectively (**Figure 1**).

Table 1. Blood glucose level in control and diabetic treated groups.

	Groups							
	Ι	II	III	IV	V			
	Fasting blood glucose levels (mg/dL)							
Initial values	70.5 ± 3.2	65.7 ± 7.5	75.8 ± 4.5	72.5 ± 8.2	68.8 ± 7.2			
Day 0 (Post diabetic induction)	75.3 ± 4.1	70.4 ± 6.4	312.8 ± 15.7°	* 305.4 ± 20.5*	325.5 ± 15.5*			
Day 21	72.8 ± 3.2	80.8 ± 5.2	$275.5 \pm 20.2^{\circ}$	*145.5 ± 10.6* [#]	125.8 ± 10.5*#			

Values are expressed as Mean \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

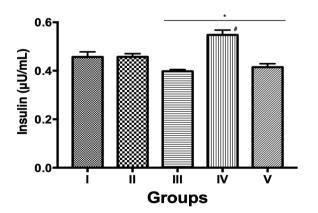


Figure 1. Insulin level in normal and diabetic treated groups. *Values are expressed as* $Mean \pm SEM$; n = 5; *indicates values that are significantly different from group 1 (control), while *indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

3.3. Effect of *P. americana* on the Lipid Profile in Control and Experimental Groups

Evaluation of the lipid profile (all in mg/dL) shows a reduction (p < 0.05) in triglycerides (49.3%), total cholesterol (62.6%), high density lipoprotein (HDL) (63.2%), low density lipoprotein (LDL) (64.9%) and very low density lipoproteins (VLDL) (49.3%) in group II (P. americana only) compared to group I, the control group. Triglycerides (83.32 ± 0.23 vs. 77.92 ± 0.57), total cholesterol (119.14 ± 3.55 vs. 107.23 ± 2.44), LDL (100.25 ± 3.68 vs. 85.52 ± 2.84) and VLDL $(16.66 \pm 0.04 \text{ vs.} 15.8 \pm 0.11)$ levels were elevated while HDL $(2.24 \pm 0.11 \text{ vs.} 6.09)$ \pm 0.31) values was reduced in group III (diabetic untreated) compared to control animals. Animals in group IV (diabetic + P. americana) exhibited significant reductions (p < 0.05) in triglycerides, total cholesterol, LDL and VLDL values compared to diabetic untreated and control. HDL value in this treatment group (group IV) was also increased compared to diabetic untreated but comparable to controls. The triglyceride (66.21 \pm 0.95 vs. 83.32 \pm 0.23) and VLDL (13.24 \pm 0.19 vs. 16.66 \pm 0.04) values observed in the diabetic animals treated with glibenclamide (group V) were reduced (p < 0.05) while HDL (5.57 \pm 0.13 vs. 2.24 \pm 0.11) values were elevated compared to group III, respectively (Figures 2-6). Atherogenic index (AI) was significantly increased (p < 0.05) in group III compared to control and all other treatment groups (Table 2). Apoliporotein A-1 (Apo A-1) was reduced in groups III and IV compared to groups I and V while apolipoprotein B (Apo-B) valueincreased in group III and V compared to group I. Values for Apo-B in groups II and IV were significantly reduced (p < 0.05) compared to group III, the diabetic untreated group. The ratio of Apo-B to Apo-A1 (Apo-B/Apo-A1), a marker of cardiovascular risk, was reduced (p < 0.05) in groups II and IV compared to control and all other treatment groups (Table 2).

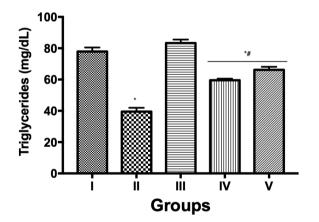


Figure 2. Triglyceride level in normal and diabetic treated groups. *Values are expressed as Mean* \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

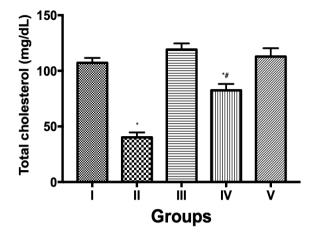


Figure 3. Total cholesterol level in normal and diabetic treated groups. *Values are expressed as Mean* \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

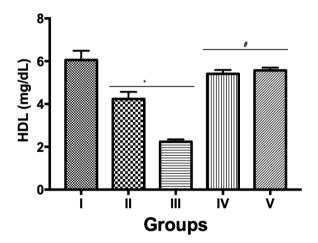


Figure 4. High Density Lipoprotein (HDL) level in normal and diabetic treated groups. *Values are expressed as Mean* \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

3.4. Effect of *P. americana* on Oxidative Stress Status in Control and Experimental Groups

Superoxide dismutase (SOD) levels (μ /mL) were significantly reduced (p < 0.05) in group III compared to all other experimental groups. catalase (CAT) (μ /mL/min), reduced glutathione (GSH) (mM), and vitamin C (Vit-C) (mg/mL) levels were significantly reduced in groups II and III compared to all other

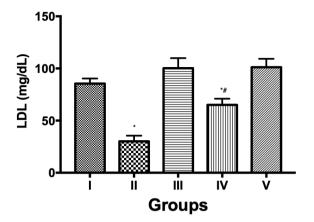


Figure 5. Low Density Lipoprotein (LDL) level in normal and diabetic treated groups. *Values are expressed as Mean* \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while "indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

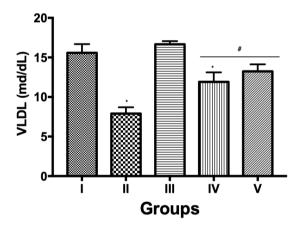


Figure 6. Very Low Density Lipoprotein (VLDL) level in normal and diabetic treated groups. *Values are expressed as Mean* \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while # indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide.

experimental groups. Malondialdehyde (MDA) (μ M), a marker of lipid peroxidation, was elevated (p < 0.05) in group III compared to other experimental groups (Table 3).

3.5. Effects of *P. americana* on the Histology of the Heart and Pancreas

The cardiac photomicrographs obtained from animals in group I (control)

Parameters	Ι	II	III	IV	V
AI	1.25 ± 0.02	1.27 ± 0.06	1.72 ± 0.01	1.18 ± 0.03	1.33 ± 0.01
Apo-A1 (mg/dl)	31.12 ± 0.32	29.76 ± 0.28	28.41 ± 0.15	29.5 ± 0.15	32.16 ± 0.39
Apo-B (mg/dl)	9.20 ± 0.25	6.79 ± 0.55	10.69 ± 0.36	7.64 ± 0.77	12.02 ± 0.28
Apo-B/Apo-1	0.32 ± 0.02	0.23 ± 0.02	0.37 ± 0.01	0.26 ± 0.03	0.38 ± 0.01

 Table 2. Effect of *P. americana* on cardiovascular disease risk factors in control and diabetic treated groups.

Values are expressed as Mean \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide. AI = Atherogenic index, Apo-A1 = Apoliporotein A-1, Apo-B = Apolipoprotein B.

 Table 3. Effect of *P. americana* on markers of oxidative stress in control and diabetic treated groups.

Parameters	Ι	II	III	IV	V
SOD (µ/mL)	0.57 ± 0.04	0.61 ± 0.02	0.23 ± 0.04	0.49 ± 0.06	0.40 ± 0.03
CAT (µ/mL/min)	27.51 ± 0.5	25.95 ± 0.23	25.3 ± 0.2	26.67 ± 0.44	28.79 ± 0.34
GSH (mM)	1.27 ± 0.09	$0.81\pm0.04^{\circ}$	$0.94\pm0.02^{\circ}$	1.83 ± 0.11°*	$1.24\pm0.08^{*}$
Vit-C (mg/ml)	0.23 ± 0.01	$0.17\pm0.01^{\circ}$	$0.19\pm0.00^{\rm\ c}$	0.2 ± 0.01	0.21 ± 0.01
MDA (µM)	5.63 ± 0.25	$5.38\pm0.23^{\ast}$	$6.67 \pm 0.07^{\circ}$	$5.57\pm0.13^{*}$	$6.04\pm0.2^{*}$

Values are expressed as Mean \pm *SEM*; n = 5; *indicates values that are significantly different from group 1 (control), while [#]indicates values that are significantly different from group III (diabetic untreated). I = Control, II = *P. americana* only, III = Diabetic untreated, IV = Diabetic + *P. americana*, V = Diabetic + Glibenclamide. SOD = Superoxide dismutase, CAT = Catalase, GSH = Reduced glutathione, Vit-C = Vitamin C, MDA = Malondialdehyde.

(Figure 7(a)) showed myofibres that were arranged in parallel with normal nuclei orientation. Photomicrographs from group II (Figure 7(b)) also showed myofibres that were arranged in parallel; however, there is also some evidence of coagulative necrosis. Cardiac photomicrographs from group III (diabetic untreated) (Figure 7(c)) exhibited myofibres with atrophy and patchy degeneration. In group IV (diabetic + *P. americana*) (Figure 7(d)), the photomicrographs displayed normal myofibres as well as a few myofibres with lost nuclei. Cardiac photomicrographs obtained from group V (diabetic + glibenclamide) (Figure 7(e)) showed normal myofibres and some evidence focal hyperplasia. Pancreatic photomicrographs obtained from groups I (control) (Figure 8(a)), II (*P. americana* only) (Figure 8(b)) and IV (diabetic + *P. americana*) (Figure 8(d)) showed no observable lesion. While the photomicrographs from the diabetic untreated group (group III) (Figure 8(c)) exhibited atrophy of pancreatic acini and islet

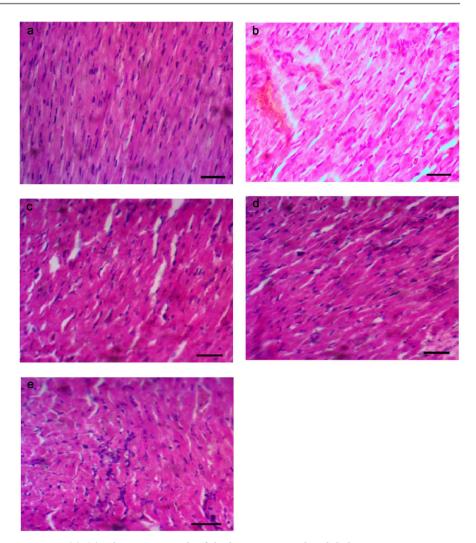


Figure 7. (a)-(e). Photomicrograph of the heart in control and diabetic treatment groups (×400, Scale bar 50 μ m). Sections in the control group (a) indicate s myofibres arranged in parallel with normal oriented nuclei. No observable lesion was seen. Sections in group II (b) also show myofibres arranged in parallel, however some coagulative necrosis is observed. Group III samples (c) exhibited atrophy and patchy degeneration of the myofibres while group IV (d) shows normal myofibres as well as myofibres with lost nuclei. Sections in group V (e) normal myofibres and some evidence focal hyperplasia.

cells. Sample from the diabetic animals treated with glibenclamide (**Figure 8(e)**) exhibited moderate diffuse acinar atrophy.

4. Discussion and Conclusions

Hyperglycaemia in diabetes mellitus has been reported to result in abnormalities in lipid metabolism often referred to as diabetic dyslipidaemia with changes in lipoprotein activity that can either be quantitative (increased triglyceride and decreased HDL levels), qualitative (increased VLDL and small, dense LDLs, as well as increased triacylglycerol content of LDL and HDL, glycation of apolipoproteins and increased susceptibility of LDL to oxidation) and or kinetic (increased VLDL production, decreased VLDL catabolism and increased HDL

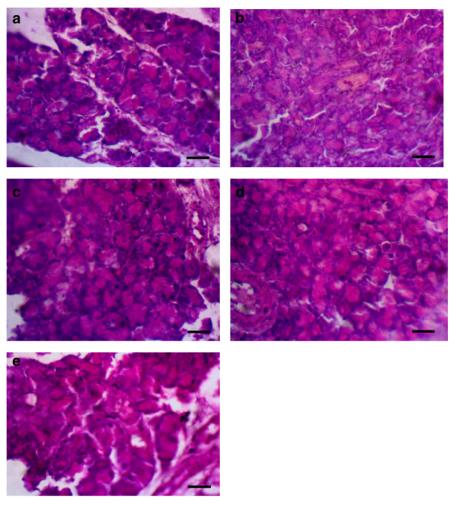


Figure 8. (a)-(e). Photomicrograph of pancreas in control and diabetic treatment groups (\times 400, Scale bar 50 µm). Sections from the groups I (a) and II (b) show no observable lesions while sections from group III (c) exhibited atrophy of pancreatic acini and islet cells. Group IV samples (d) showed no observable lesions while samples from group V (e) showed moderate diffuse acinar atrophy.

catabolism) [27] [28] [29]. These lipoprotein abnormalities result in a shift towards a more atherogenic lipid profile with severe consequences for the cardiovascular system.

Sustained hyperglycaemia, lipid abnormalities consistent with diabetic dyslipidaemia (increased triglyceride, total cholesterol, LDL VLDL, and reduced HDL) were observed in the diabetic untreated group in this study which is consistent with other reports using alloxan-induced diabetes experimental model to investigate lipid abnormalities in diabetes mellitus [30]. The hyperglycaemia observed in these diabetic untreated animals (group III) may be attributed to the effects of alloxan monohydrate which is known to selectively target pancreatic beta cells resulting in necrosis [17] [31] and hence result in inadequate or reduced insulin secretion which was also observed in this experimental group (**Figure 1** and **Figure 8(c)**). In addition to facilitating glucose entry in adipocytes, insulin also inhibits the breakdown of fat in adipose tissue by inhibiting

the activity of intracellular hormone sensitive lipase which hydrolyses triglycerides to release fatty acids [32]. Insulin also directly inhibits hepatic VLDL production and promotes the clearance of LDL by increasing LDL-receptor expression and activity [32]. Hence, when insulin secretion is inadequate or deficient, these aforementioned actions of insulin are impeded resulting in dyslipidaemia as observed in the diabetic untreated group in this study. The observed dyslipidaemia noted could also have resulted in micro and macrovascular complications and hence, account for the patchy degeneration and atrophy of cardiac myofibres observed in the diabetic untreated animals (group III) (**Figure 7(c)**). Also, this group exhibited a higher atherogenic index (**Table 2**), thus suggesting an increased risk for the development of cardiovascular complications in this group.

Persea americana has been reported to possess a number of phytochemicals such as alkaloids, glycosides, oxalate, phytate, phenol, saponins, steroid, and tannins which have been reported to be effective against hepato-toxicity, inflammation, cancer, hypertension amongst other disease conditions [33]. Gas Chromatography-Mass Spectrometry (GCMS) of various parts of P. americana has been evaluated and bioactive compounds seen have been reported to demonstrate analgesic, anti-inflammatory, antipyretic, and antihyperglycemic properties [34] [35]. Therefore, the reduction of hyperglycaemia observed in diabetic animals treated with P. americana which was also comparable to that obtained with glibenclamide treatment (group IV), in accordance with previous studies [12]. This hypoglycaemic activity, though not as potent as that of glibenclamide, may be ascribed to the bioactive phytocompounds in the hydro-ethanol pulp extract and the ability of the extract to stimulate regeneration of the pancreatic islet cells as observed in the histological evaluation of pancreas in this study (Figure **8(d)**). The regenerated pancreatic islet may have also accounted for the increase in insulin secretion seen with *P. americana* treatment (group V) compared to diabetic untreated (Figure 1). Treatment with the extract also ameliorated diabetes induced dyslipidaemia, perhaps via increased insulin mediated lipid regulatory mechanisms [32], and impeded cardiac fibre degeneration and atrophy (Figure 7(d)), thereby suggesting the potentials of the hydro-ethanol extracts of P. americana fruit pulp in preventing diabetes mellitus related atherosclerosis and heart diseases [36]. However, P. americana only treated group showed decreased levels of triglycerides, total cholesterol, HDL, LDL and VLDL compared to control level which aligns with the observation of Dreher et al. [37] who stated that consumption of P. americana significantly reduces total cholesterol, low density lipoprotein cholesterol (LDL-C), triglycerides and high-density lipoprotein cholesterol (HDL-C) respectively in the blood. It is also noteworthy that though cardiac fibres seen in the P. americana group were normal, some level of necrosis was also seen (Figure 7(b)).

Apolipoprotein (Apo), a protein component of plasma lipoprotein synthesized mainly in the liver, binds to and transports blood lipids to various tissues of the body for metabolism and utilization [38]. While Apolipoprotein A1 (ApoA1) is the primary protein component of high-density lipoprotein (HDL), Apolipoprotein B (ApoB) is the primary protein component of low-density lipoprotein (LDL) [39]. The ratio of ApoB/apoA1 has been described as an effective predictor for the risk of coronary heart disease development with increased values indicating an increased risk of atherosclerotic cardiovascular disease, independently of LDL and HDL cholesterol concentrations [39]. In this study, ApoB/A1 was significantly reduced in the *P. americana* treated diabetic group compared to diabetic untreated (Table 2) suggesting that the extract may mitigate the development of atherosclerosis and coronary disease often associated with diabetes mellitus. Furthermore, the atherogenic index, a novel marker for assessing the risk of atherogenicity and cardio-metabolic health, was reduced in the diabetic group treated with extract compared to diabetic untreated which reiterates the likely potential in the consumption of the hydro-ethanolic extract of P. americana fruit in diabetes mellitus. Interestingly, diabetic animals treated with glibenclamide also had elevated ApoB/A1 and atherogenic index compared to P. americana treated diabetic animals (Table 2). This may likely be due to the persistence of elevated triglyceride, total cholesterol and LDL despite increased HDL levels following treatment with a standard anti-diabetic agent, glibenclamide (Figures 2-5).

Hyperglycaemia as it occurs in diabetes mellitus, has also been reported to impair intracellular antioxidant defence mechanism resulting in over-production of free radicals by non-enzymatic glycation of serum and cellular proteins thereby rendering cells vulnerable to oxidative stress [40]. In this study, the significant alterations in oxidant-antioxidant status (increased MDA and reduced GSH, Vitamin C level, SOD and CAT activity) that occurred following diabetes were mitigated by treatment of diabetic rats with *P. americana* fruit hydro-ethanol extract. The antioxidant effect of the extract is in accordance with other studies which had also ascribed this to the presence of the bioactive compound in *P. americana* that is capable of scavenging and mopping up excessive free radicals produced during diabetes mellitus [12].

In conclusion, this study suggests that in experimental diabetes mellitus, treatment with hydro-ethanol extract of *Persea americana* fruit pulp may mitigate hyperglycaemia, oxidative stress, diabetic-induced dyslipidaemia and cardiotoxicity. The effects of the extract are mediated by the reported phytochemicals present in the plant, *Persea americana*.

Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] World Health Organization (2016) Global Report on Diabetes. Geneva.
- [2] Leon, B.M. and Maddox, T.M. (2015) Diabetes and Cardiovascular Disease: Epidemiology, Biological Mechanisms, Treatment Recommendations and Future Research.

World Journal of Diabetes, 6, 1246-1258. https://doi.org/10.4239/wjd.v6.i13.1246

- [3] Piché, M.-E., Tchernof, A. and Després, J.-P. (2020) Obesity Phenotypes, Diabetes, and Cardiovascular Diseases. *Circulation Research*, **126**, 1477-1500. <u>https://doi.org/10.1161/CIRCRESAHA.120.316101</u>
- [4] Raghavan, S., Vassy, J.L., Ho, Y.-L., et al. (2019) Diabetes Mellitus-Related All-Cause and Cardiovascular Mortality in a National Cohort of Adults. *Journal of the Ameri*can Heart Association, 8, e011295. https://doi.org/10.1161/JAHA.118.011295
- [5] Roman, G. and Stoian, A.P. (2021) Cardiovascular Risk/Disease in Type 2 Diabetes Mellitus. In: Stoian, A.P., Ed., *Type 2 Diabetes—From Pathophysiology to Cyber Systems*, Intech Open, London. <u>https://doi.org/10.5772/intechopen.97422</u>
- [6] Mooradian, A. (2009) Dyslipidemia in Type 2 Diabetes Mellitus. *Nature Reviews Endocrinology*, 5, 150-159. <u>https://doi.org/10.1038/ncpendmet1066</u>
- [7] Nagasawa, S.Y., Okamura, T., Iso, H., et al. (2012) Evidence for Cardiovascular Prevention from Observational Cohorts in Japan (EPOCH-JAPAN) Research Group. Relation between Serum Total Cholesterol Level and Cardiovascular Disease Stratified by Sex and Age Group: A Pooled Analysis of 65594 Individuals from 10 Cohort Studies in Japan. *Journal of the American Heart Association*, 1, e001974. https://doi.org/10.1161/JAHA.112.001974
- [8] Jung, E., Kong, S.Y., Ro, Y.S., Ryu, H.H. and Shin, S.D. (2022) Serum Cholesterol Levels and Risk of Cardiovascular Death: A Systematic Review and a Dose-Response Meta-Analysis of Prospective Cohort Studies. *International Journal of Environmental Research and Public Health*, **19**, 8272. https://doi.org/10.3390/ijerph19148272
- Kayarohanam, S. and Kavimani, S. (2015) Current Trends of Plants Having Antidiabetic Activity: A Review. *Journal of Bioanalysis and Biomedicine*, 7, 55-65. <u>https://doi.org/10.4172/1948-593X.1000124</u>
- [10] Siddiqui, S., Khatoon, A., Ahmad, K., *et al.* (2022) Traditional Islamic Herbal Medicine and Complementary Therapies. In: Bernardo-Filho, M., *et al.*, Eds., *Complementary Therapies*, Intech Open, London. https://doi.org/10.5772/intechopen.101927
- [11] Morton, J. (1987) Avocado, Fruits of Warm Climates. Creative Resource Systems, Inc., Winterville, NC and Center for New Crops & Plant Products, Department of Horticulture and Landscape Architecture, Purdue University, West Lafayette, 91-102.
- [12] Mahadeva, U.S. and Adinew, B. (2011) Remnant β-Cell-Stimulative and Anti-Oxidative Effects of Persea Americana Fruit Extract Studied in Rats Introduced into Streptozotocin-Induced Hyperglycaemic. African Journal of Traditional, Complementary and Alternative Medicines, 8, 210-217. https://doi.org/10.4314/ajtcam.v8i3.65277
- [13] Haegele, A.D., Gillette, C., O'Neill, C., et al. (2000) Plasma Xanthophyll Carotenoids Correlate Inversely with Indices of Oxidative DNA Damage and Lipid Peroxidation AMC Cancer. Cancer Epidemiology, Biomarkers & Prevention, 9, 421-425.
- [14] Flagg, E.W., Coates, R.J., Jones, D.P., *et al.* (1994) Dietary Glutathione Intake and Risk of Oral and Pharyngeal Cancer. *American Journal of Epidemiology*, 139, 453-465. https://doi.org/10.1093/oxfordjournals.aje.a117028
- [15] Ojewole, J.A. (2006) Hypoglycaemic and Hypotensive Effects of *Harpephyllum caf-frum* Bernh ex CF Krauss (Anacardiaceae) Stem-Bark Aqueous Extract in Rats. *Cardiovascular Journal of South Africa*, **17**, 67-72.
- [16] Benedé-Ubieto, R., Estévez-Vázquez, O., Ramadori, P., et al. (2020) Guidelines and Considerations for Metabolic Tolerance Tests in Mice. Diabetes, Metabolic Syn-

drome and Obesity, 13, 439-450. https://doi.org/10.2147/DMSO.S234665

- [17] Adewoye, E.O. and Adele, B.O. (2015) Effect of Methanol Extract of Musa sapientum Leaves on Protein Glycation and Erythrocyte Antioxidant Status in Alloxan-Induced Diabetic Wistar Rats. African Journal of Medicine and Medical Sciences, 44, 261-268.
- [18] Adewoye, E.O. and Ige, A.O. (2016) Lipid Profile and Electrolyte Composition in Diabetic Rats Treated with Leaf Extract of *Musa sapientum. Journal of Dietary Supplements*, 13, 106-117. <u>https://doi.org/10.3109/19390211.2014.965866</u>
- [19] Friedewald, W.T., Levy, R.I. and Fredrickson, D.S. (1972) Estimation of the Concentration of Low-Density Lipoprotein Cholesterol in Plasma, without Use of the Preparative Ultracentrifuge. *Clinical Chemistry*, 18, 499-502. https://doi.org/10.1093/clinchem/18.6.499
- [20] Marklund, S. and Marklund, G. (1974) Involvement of the Superoxide Anion Radical in the Autoxidation of Pyrogallol and a Convenient Assay for Superoxide Dismutase. *European Journal of Biochemistry*, **47**, 469-474. https://doi.org/10.1111/j.1432-1033.1974.tb03714.x
- [21] Clairborne, A. (1985) Catalase Activity. In: Greenwald, R.A., Ed., CRC Handbook of Methods for Oxygen Radical Research, CRC Press, Boca Raton, 283-284.
- [22] Sedlak, J. and Lindsay, R.H. (1968) Estimation of Total, Protein-Bound, and Nonprotein Sulfhydryl Groups in Tissue with Ellman's Reagent. *Analytical Biochemistry*, 25, 192-205. https://doi.org/10.1016/0003-2697(68)90092-4
- [23] Omaye, S.T., Turnbull, J.D. and Sauberlich, H.E. (1979) Selected Methods for the Determination of Ascorbic Acid in Animal Cells, Tissues, and Fluids. *Methods in Enzymology*, 62, 3-11. <u>https://doi.org/10.1016/0076-6879(79)62181-X</u>
- [24] Ohkawa, H., Ohishi, N. and Yagi, K. (1979) Assay for Lipid Peroxides in Animal Tissues by Thiobarbituric Acid Reaction. *Analytical Biochemistry*, 95, 351-358. <u>https://doi.org/10.1016/0003-2697(79)90738-3</u>
- [25] Culling, C.F.A. (1974) Handbook of Histopathological and Histochemical Techniques. 3rd Edition, Butterworths, London. <u>https://doi.org/10.1016/B978-0-407-72901-8.50008-1</u>
- [26] Avwioro, O.G. (2010) Histochemistry and Tissue Pathology, Principles and Techniques. Claverianum Press, Bodija-Ibadan.
- [27] Muačević-Katanec, D. and Reiner, Ž. (2011) Diabetic Dyslipidaemia or "Diabetes Lipidus"? *Expert Review of Cardiovascular Therapy*, 9, 341-348. <u>https://doi.org/10.1586/erc.11.17</u>
- [28] Wu, L. and Parhofer, K.G. (2014) Diabetic Dyslipidaemia. *Metabolism*, 63, 1469-1479. https://doi.org/10.1016/j.metabol.2014.08.010
- [29] Thambiah, S.C. and Lai, L.C. (2021) Diabetic Dyslipidaemia. Practical Laboratory Medicine, 26, e00248. https://doi.org/10.1016/j.plabm.2021.e00248
- [30] Ojiako, O.A., Chikezie, P.C. and Ogbuji, A.C. (2015) Blood Glucose Level and Lipid Profile of Alloxan-Induced Hyperglycemic Rats Treated with Single and Combinatorial Herbal Formulations. *Journal of Traditional and Complementary Medicine*, 6, 184-192. <u>https://doi.org/10.1016/j.jtcme.2014.12.005</u>
- [31] Szkudelski, T. (2001) The Mechanism of Alloxan and Streptozotocin Action in Beta Cells of the Rat Pancreas. *Physiological Research*, **50**, 536-546.
- [32] Santoro, A., McGraw, T.E. and Kahn, B.B. (2021) Insulin Action in Adipocytes, Adipose Remodelling, and Systemic Effects. *Cell Metabolism*, **33**, 748-757. https://doi.org/10.1016/j.cmet.2021.03.019

- [33] Yasir, M., Das, S. and Kharya, M.D. (2010) The Phytochemical and Pharmacological Profile of *Persea americana* Mill. *Pharmacognosy Reviews*, 4, 77-84. <u>https://doi.org/10.4103/0973-7847.65332</u>
- [34] Mahmoud, A.H., Samy, M.N., Wanas, A.S. and Kamel, M.S. (2021) Gas Chromatography-Mass Spectrometry Profiling and Analgesic, Anti-Inflammatory, Antipyretic, and Antihyperglycemic Potentials of *Persea americana. Iranian Journal of Basic Medical Sciences*, 24, 641-649.
- [35] Ogunmoyole, T., Alfonso, O.G., Johnson, O.D. and Yusuff, A.A. (2022) In Vitro Antioxidant Properties and GC-MS Analysis of Solvent Extracts of Persea americana Leaf. Journal of Science Letters, 3, 7-14.
- [36] Chapman, M.J., Ginsberg, H.N. and Amarenco, P. (2011) Triglyceride-Rich Lipoproteins and High-Density Lipoprotein Cholesterol in Patients at High Risk of Cardiovascular Disease: Evidence and Guidance for Management. *European Heart Journal*, 32, 1345-1361. <u>https://doi.org/10.1016/S1567-5688(11)70033-2</u>
- [37] Dreher, M.L. and Davenport, A.J. (2013) Hass Avocado Composition and Potential Health Effects. *Critical Reviews in Food Science and Nutrition*, 53, 738-750. <u>https://doi.org/10.1080/10408398.2011.556759</u>
- [38] Parolini, C. (2020) A Compendium of the Biological Effects of Apolipoprotein A-I_{Milano.} Journal of Pharmacology and Experimental Therapeutics, **372**, 54-62. https://doi.org/10.1124/jpet.119.261719
- [39] Florvall, G., Basu, S. and Larsson, A. (2006) Apolipoprotein A1 Is a Stronger Prognostic Marker than Are HDL and LDL Cholesterol for Cardiovascular Disease and Mortality in Elderly Men. *The Journals of Gerontology: Series A*, 61, 1262-1266. https://doi.org/10.1093/gerona/61.12.1262
- [40] Brownlee, M., Vlassara, H. and Cerami, A. (1984) Non-Enzymatic Glycosylation and the Pathogenesis of Diabetic Complications. *Annals of Internal Medicine*, **101**, 527-537. https://doi.org/10.7326/0003-4819-101-4-527