

# Demographic Expansion and Contraction in a Neotropical Fish during the Late Pleistocene-Holocene

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## Abstract

Demographic changes during the late Pleistocene-Holocene left signatures in the DNA of contemporary populations. These signatures reveal demographic phenomena like the increase or decrease in effective population size. In this paper we searched for signatures of demographic change in the DNA of the Neotropical freshwater fish *Poecilia vivipara*. Also, we investigated whether demographic changes are correlated with palaeoclimatic events of the late Pleistocene-Holocene, in particular, if changes in effective population size are correlated with expansion and contraction of available habitats, induced by global ice-volume changes and sea-level fluctuations. We used Bayesian Sky-line Plot (BSP) analysis with sequences from the mitochondrial gene cytochrome *b* to estimate the ancestral demography of the Neotropical freshwater fish *P. vivipara*. To test the assumptions of neutrality and absence of population structure we used Tajima's *D* and Spatial Analysis of Molecular Variance (SAMOVA), respectively. Effective population size of *P. vivipara* remained stable until 75,000 years ago, increased by 10-fold reaching a maximum at approximately 25,000 years ago, then suddenly declined at the Pleistocene-Holocene boundary. Variation in effective population size in *P. vivipara* correlates with expansion and contraction of habitats induced by sea-level fluctuations, caused by the advance and retreat of ice sheets during the Last

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Glacial Maximum (LGM).

## Keywords

Ancestral Population Dynamics, Coalescence, Bayesian Skyline Plot, Mitochondrial DNA, Last Glacial Maximum

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## 1. Introduction

Late Pleistocene-Holocene global climatic oscillations markedly influenced the demographic history of Earth's biota [1] [2] [3]. From the standpoint of demography, departures from stability, experienced as expansion or contraction in population size, are relevant events [4] [5]. Signatures of changes in effective population size are written in the DNA of contemporary populations because of ancestor-descendant transmission of genetic material and the processes of mutation and substitution [6] [7] [8]. Consequently, from evolutionary and ecological perspectives, it is important to assess whether temporal changes in effective population size are correlated with episodic palaeoclimatic events [9]. Of the several known cases, the steppe bison is probably one of the richest for the complexity of its demographic response to the Pleistocene-Holocene climate changes [10] [11] [12] [13]. Bison effective population size increased from 150 thousand years before present (*ka BP*), reaching a maximum between 30 - 45 *ka BP* [11]. After the maximum, effective population size declined precipitously, reaching a minimum at approximately 10 *ka BP* [11]. Following the minimum, effective population size increased and levelled off, and then decreased again [11]. Minimum and maximum effective population sizes in the bison correlate, among other factors, with the contraction and expansion of available habitat, driven by the advance and retreat of ice sheets during the Last Glacial Maximum (LGM) and Holocene, respectively [11]. As demonstrated by coalescent simulation, effective population size correlates with the amount of habitat available [14]. Therefore, a causal association between demographic expansion and contraction and palaeoclimatic cyclic events is tenable.

The expansion and contraction of habitats in continental areas in the Northern hemisphere is primarily a consequence of the cyclical retreat and advance of ice sheets [15]. In non-glaciated regions, however, the dynamics of habitat availability is locally induced by glacio-eustatic sea-level changes [16]. Sea-level low stands, caused by the advance of ice sheets, expose the continental shelf increasing habitat availability. Conversely, in non-glaciated regions the retreat of ice sheets induces an inverse effect on habitat availability. Such glacio-eustatic sea-level changes did have a major impact on coastal areas of South America [17] [18] [19]. Here, we investigate the potential consequences of the late Pleistocene-Holocene climatic oscillations for the historical demography of a poeciliid fish, *Poecilia vivipara*, inhabiting a non-glaciated region in South America. Poeciliids are well-known model organisms [20] [21], and *P. vivipara* is a fresh-

water species that also colonizes brackish and salt-water environments near the mouth of rivers [22]. *Poecilia vivipara* thus has the potential to track environmental changes such as the expansion or contraction of habitats associated with glacio-eustatic sea-level changes. Given the environmental changes of the late Pleistocene-Holocene and the ecological and life history attributes of *P. vivipara*, we expect contemporary populations of *P. vivipara* will harbour signatures of their historical demography. We used sequences of the mitochondrial gene cytochrome *b* (*cyt-b*), sampled from southeastern Brazil, and the coalescent-based Bayesian Skyline Plot (BSP) [11] to explore an association between historical variation in effective population size in *P. vivipara* and climatic oscillations during the late Pleistocene-Holocene. We believe our results do contribute relevant information to the ongoing research on the interplay between late Pleistocene-Holocene climatic oscillations and ecological and evolutionary processes in South America.

## 2. Methods

### 2.1. Study System

We sampled a total of 206 individuals of *P. vivipara* using a fine mesh net from 20 lagoons, which are hydrogeologically associated with the Paraíba do Sul River in northern Rio de Janeiro state in southeastern Brazil (Table 1). These lagoons vary in salinity from freshwater to brackish and saltwater [22].

### 2.2. DNA Processing

DNA was extracted from the caudal peduncle using the salting-out method [23]. The entire mitochondrial *cyt-b* (1140 base pairs) gene was amplified using primers and reaction parameters defined in Schories *et al.* [24]. Purified Polymerase Chain Reaction (PCR) products were outsourced to Macrogen Inc. (South Korea) for sequencing (using the BigDye™ 158 terminator kit and run on an ABI 3730XL). PCR products were sequenced in both directions with the same primers used for PCR amplification and subjected to Basic Local Alignment Search Tool (BLAST) in GenBank to verify that the desired sequences had been amplified. The sequences were deposited in GenBank (accession numbers: MG266475-MG266680).

### 2.3. Tests of Neutrality and Population Structure

We used Tajima's *D* statistic [25] to test for the departure from the mutation-drift equilibrium and to evaluate whether the *cyt-b* gene conformed to neutral expectations using program DNA Sequence Polymorphism (DnaSP) v4.50.02 [26]. We estimated the genealogical relationships among haplotypes using the statistical parsimony method, developed by Templeton, Crandall and Sing (TCS) [27], with a 95% probability that no multiple substitutions have occurred, as implemented in the software TCS v1.21 [28]. The assumption of a panmictic population was tested with Spatial Analysis of Molecular Variance

**Table 1.** Geographical locations of the 20 sampled populations of *Poecilia vivipara*. The number of individuals ( $n$ ) analysed for *cyt-b* mitochondrial DNA (mtDNA) marker and the number of haplotypes ( $h$ ) are given for each location.

<i>ID</i>	<i>Sample</i>	<i>Latitude S</i>	<i>Latitude W</i>	<i>n</i>	<i>h</i>
1	Açu	21.99416°	41.00001°	11	6
2	Atafona	21.62836°	41.02417°	10	5
3	Cabiunas	22.30140°	41.69300°	10	1
4	Cacimba	21.63940°	41.09540°	10	4
5	Catingosa	22.18850°	41.41540°	10	4
6	Campelo	21.66463°	41.20097°	10	3
7	Cima	21.78693°	41.51622°	11	3
8	Comercio	21.57530°	41.06740°	11	7
9	Feia	21.94553°	41.34551°	10	7
10	Barra do Furado	22.08086°	41.11850°	10	5
11	Garça	22.12140°	41.49220°	10	4
12	Grussaí	21.71979°	41.03783°	10	7
13	Heliporto	22.03078°	41.06392°	11	8
14	Iquipari	21.80019°	41.02122°	10	8
15	Lagamar	22.06027°	41.08317°	10	7
16	Praia I	21.57534°	41.06043°	10	5
17	Praia II-Gargau	21.57440°	41.05867°	10	6
18	Santa Clara	21.52043°	41.07912°	10	4
19	Tai Pequeno	21.80620°	41.12870°	12	6
20	Três Vendas	21.68240°	41.33580°	10	2

(SAMOVA) [29], which requires no *a priori* definition of groups of populations. SAMOVA iteratively searches for the partitions of  $K$  groups of populations that maximize the  $F_{CT}$  index, which is the proportion of the total genetic variance due to difference between groups of populations for any number of putative groups. SAMOVA was performed with 18 partitions for the 20 population samples for  $K$  varying from two to 19 groups of population samples. The  $F_{CT}$  index was computed for each partition. The cases in which all population samples belong to the same group ( $K = 1$ ) or each population is itself a group ( $K = 20$ ) were not included in the analysis. For each  $K$  value, 10,000 simulated annealing steps were executed starting from 200 sets of initial conditions.

#### 2.4. Demographic Analysis

We estimated historical variation in the effective population size of *P. vivipara* using sequences of the *cyt-b* gene and the BSP method, which is based on the variable population size coalescent model [11]. Starting with a sequence alignment from a sample, the BSP method co-estimates the phylogeny and the effective population size through time [11]. This method is based on assumptions that samples come from a panmictic population and that sequences are orthologous, non-recombining, and evolve neutrally [11]. We used the software Bayesian

Evolutionary Analysis Utility (BEAUti) [30] to generate the XML input files for the Bayesian Evolutionary Analysis Sampling Trees software (BEAST v.1.8.2) [30]. We used unlinked strict clock rates and the General Time Reversible (GTR) model of nucleotide substitution, which best fitted the *cyt-b* sequences according to jModelTestv.2.1.10 [31]. We ran 400 million Markov chain Monte Carlo (MCMC) iterations, sampling every 40,000 runs. The first 10% runs were discarded as burn-in. We verified convergence to a stationary distribution with Tracer v.1.5 [32]. Effective sample size for parameter estimates was always >200. Demographic history through time was reconstructed also using Tracer v.1.5 [32]. Estimation of absolute ages of demographic events was based on a substitution rate of  $0.9 \times 10^{-8}$  per site per year, which we calculated using estimates of absolute divergence times derived by Meredith *et al.* [33].

### 3. Results

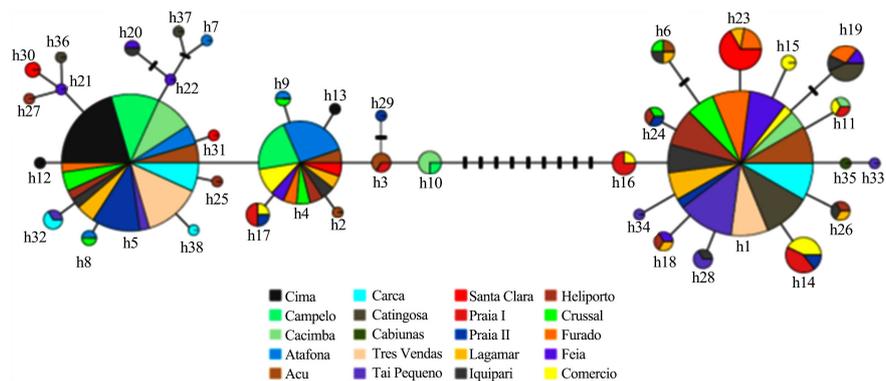
The inference of ancestral effective population size using DNA sequences and the BSP technique require testing the assumptions of neutrality and absence of population structure [11]. The assumption of neutrality was tested with Tajima's *D* statistics [25], which was not significantly different from zero ( $D = -0.1523$ ;  $P > 0.10$ ). This result indicates that the assumption of mutation-drift is not violated and that the *cyt-b* gene is evolving neutrally in the sampled populations of *P. vivipara*. We used the statistical parsimony network [27] and the spatial analysis of molecular variance [29] to test for population structure. Thirty-eight unique haplotypes were identified among the 206 *cyt-b* sequences analysed. All haplotypes were connected into a single network, which has an overall star-like pattern (Figure 1). The network is characterized by three high-frequency haplotypes, several intermediate, and rare haplotypes. The haplotypes are divided in two main groups, with each group connected by a maximum of three mutational steps, whereas a minimum of nine steps separates each group. In SAMOVA, the  $F_{CT}$  statistics estimates the total genetic variance due to differences between groups of samples for partitions of *K* groups of populations.  $F_{CT}$  values decrease smoothly as *K* increases from 2 to 8 and then levels off for values of *K* from 9 through 19 (Figure 2). This result indicates that no single value of *K* is maximized, indicating a lack of population structure in the populations sampled. An example of group composition for  $K = 5$  resulted from the simulated annealing steps is shown in Table 2.

Modelling the effective population size over time with the BSP using the *cyt-b* sequences revealed a complex demographic history for *P. vivipara* (Figure 3(a)). In this plot, the *y*-axis is expressed as *female effective population size* ( $N_e$ )  $\times$  *generation time* and the *x*-axis is *calendar time* in years. Effective population size remained stable approximately until 75 *ka* BP and is followed by a steep, approximately 10-fold increase reaching a maximum value at the LGM (approximately 25 *ka* BP). There follows a sharp and sudden decline in effective population size starting around the Pleistocene-Holocene boundary, a trend that pers-

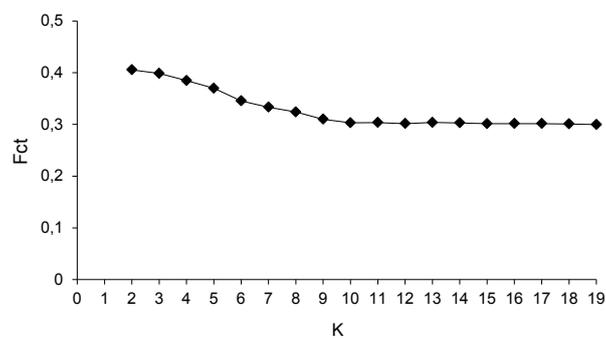
ists to the present. A sample of the MCMC iterations is shown in **Figure 4**. The trace is stable indicating good parameter mixing and convergence of the likelihood estimation (**Figure 4(a)**). Also the plot of the marginal density for the likelihood estimation shows a high degree of symmetry (**Figure 4(b)**).

**Table 2.** Group composition of sampled populations of *Poecilia vivipara* in one of the 10,000 simulated annealing steps for  $K = 5$  in SAMOVA analysis.

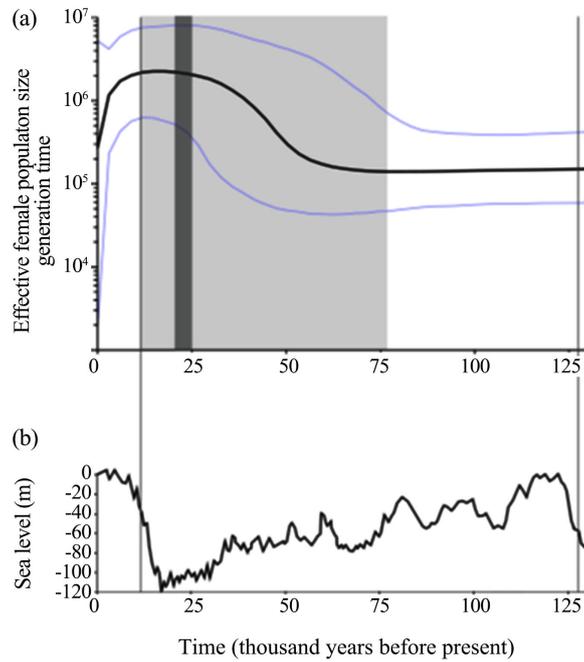
Group	Number of populations	Lagoons
1	16	Açu, Cacimba, Comércio, Feia, Furado, Grussaí, Heliporto, Iquipari, Lagamar, Praia I, Praia II, Santa Clara, Tai Pequeno Três Vendas, Catingosa, Garça,
2	1	Cima
3	1	Campelo
4	1	Cabiunas
5	1	Atafona



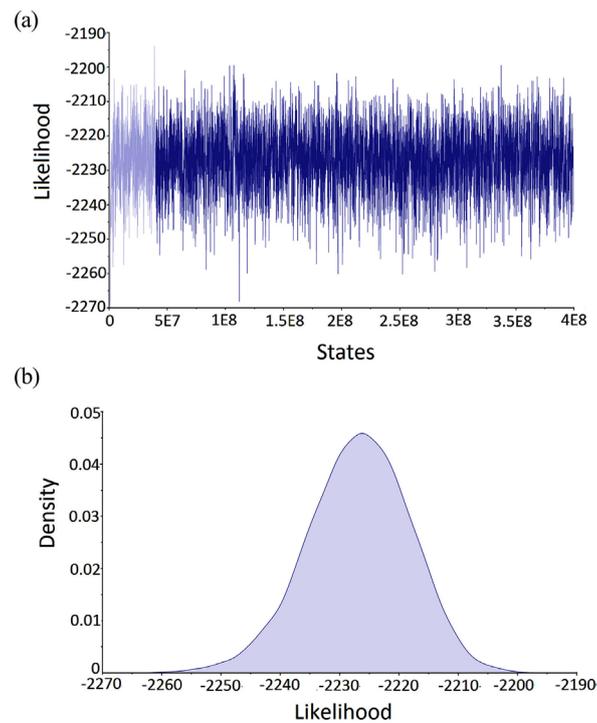
**Figure 1.** Statistical parsimony network, as implement in the software TCS v1.21, showing genealogical relationships among the 38 mtDNA haplotypes found ( $h_1$  to  $h_{38}$ ). Colors represent samples of different lagoons. Size of circles is proportional to haplotype frequencies. Black bars e lines denote mutational steps separating haplotypes.



**Figure 2.** Spatial analysis of molecular variance (SAMOVA) of the 20 populations samples of *Poecilia vivipara*. The proportion of the total genetic variance explained by the among-group variation ( $F_{CT}$  index) for any number of putative groups ( $K = 2$  to  $K = 17$ ) was lower (mean value of 33%) than the within-sample variation (mean value of 67.8%), implying no evidence of structure in the sample.



**Figure 3.** (a) Bayesian skyline plot derived from a sample ( $n = 206$ ) of the cytochrome *b* sequences from *P. vivipara*. The *x*-axis is *calendar time* in years. The *y*-axis is equal to *female effective population size* ( $N_e$ )  $\times$  *generation time*. (b) Eustatic sea-level curve since 130 *ka* BP (based on bathymetric data from Pillans *et al.* [46] and Nicol [47]).



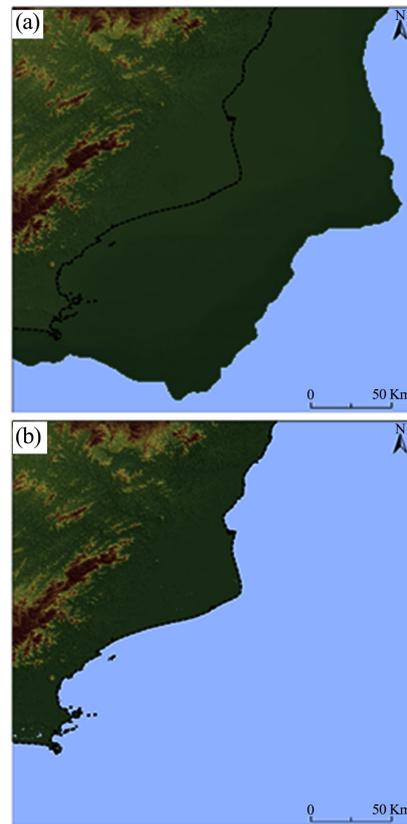
**Figure 4.** Samples of MCMC iterations visualized in Tracer. (a) Trace of likelihood, showing a convergence. The clearer blue line represent the first 10% runs which were discarded as burn-in. (b) The marginal density for the likelihood estimation, showing a high degree of symmetry.

## 4. Discussion

The *cyt-b* sequences of the sampled populations of *P. vivipara* do have variation, which allows the estimation of variation in ancestral effective population size over time. However, it is well established that population structure can influence the estimation of effective population size [34] [35]. In our study, no evidence of geographic structure can be gleaned from the haplotype network, that is, no discernable association between haplotypes and locations is evident. Furthermore, the evidence from the spatial analysis of molecular variance points to the absence of structure in the populations of *P. vivipara*. Equally relevant, the sequences of the *cyt-b* gene appear to be evolving neutrally. Therefore, fundamental assumptions of the BSP method [11] appear not to be violated by our samples.

The BSP revealed a marked increase in the effective population size of *P. vivipara*, which reached a maximum at approximately 25 ka BP (Figure 3(a)). This demographic event coincides approximately with the LGM, which is dated between 26.5 ka BP and 19 - 20 ka BP [36] [37]. During the LGM, the sea level of the coast of Brazil was ~120 meters below its current level [37] (Figure 3(b)) exposing, as a consequence, much of the continental shelf (Figure 5(a)), arguably increasing the availability of both freshwater and brackish habitats for *P. vivipara*. A second striking feature is the sharp demographic decrease after the maximum effective population size (Figure 3(a)). This demographic trend is conceivably associated with a sea-level rising after the LGM (Figure 3(b)), which probably induced a contraction of habitats previously available during the LGM (Figure 5(b)).

How might the ancestral demography of *P. vivipara* revealed by the Bayesian skyline plot be interpreted? Idiosyncrasies of the ecology and life history of contemporary populations of *P. vivipara* may shed light onto the dynamics of effective population size and the climatic-related fluctuations associated with the LGM. *Poecilia vivipara* successfully colonizes rivers and inland habitats varying in salinity from 0 to 36 parts per thousand [22]; the latter figure actually being equal to the salinity of sea water. Populations of *P. vivipara* at either extreme of the salinity gradient differ significantly in traits such as body size and shape, fecundity, and reproductive allotment, which are likely determined genetically [38] [39]. *Poecilia vivipara* also varies markedly in personality traits including boldness, activity, and sociability/shoaling behaviour, which are modulated by environmental factors such as water transparency/visibility, salinity, and dissolved oxygen [40]. Variability in personality traits vanishes, however, under homogenous laboratory conditions, indicating that plasticity underlies the behavioural differences observed in nature [40]. The available data thus suggest that genetics and plasticity play fundamental roles in the distribution of ecological and behavioural traits of *P. vivipara*, which are potentially relevant for the colonization of heterogeneous environments subjected to climatic fluctuations. In particular, plasticity, a major factor in the adaptive process [41], may be a key



**Figure 5.** (a) Palaeoshore line reconstruction of the coastal plain in southeastern Brazil (based on bathymetric data from Bastos & Silva [48]). (b) Current coastline in southeastern Brazil. Figure created using ArcGIS 10.5.0, <https://esri.ca>.

mechanism underlying the increase (decrease) in effective population size associated with habitat expansion (contraction) driven by glacio-eustatic sea level oscillations during the LGM.

The ancestral demography of *P. vivipara* appears strongly correlated with environmental changes driven by the LGM. This is indeed the major task of historical demography, that is, to test for correlation between demographic phenomena, as revealed by variation in DNA sequences and palaeoclimatic episodic events [9]. However, a correlation between variation in effective population size and climatic oscillations does not always emerge. For example, in the Northern hemisphere effective population size increased monotonically in the lizard *Podarcis filfolensis* in the Maltese archipelago [42], whereas effective population size increased and then levelled off in the tree frog *Hylasarda* in the Corsica-Sardinian island system [43]. For these systems, the observed demographic expansion is actually the reverse trend expected under the classic expansion-contraction model, which predicts demographic contraction during glaciations [42]. In the Southern hemisphere, in estuarine habitats of the coastal plains of southeastern Brazil four species of fishes experienced demographic stability, whereas three other species underwent demographic expansion followed by an

asymptote in effective population size [44]. In none of these cases, however, the demographic changes did coincide with the LGM. Global climatic oscillations coupled with local geomorphological features are probably the major exogenous factors contributing to the origin of the observed diversity of demographic regimes. However, as pointed out by Ruzzante *et al.* [45], the life history and ecology of individual species is doubtless a fundamental endogenous determinant of the spatiotemporal demographic dynamics of contemporary populations. In this connection, Ruzzante *et al.* [45] showed that two sympatric species of Patagonian freshwater fishes differ in their historical demography. *Galaxias platei* experienced a bottleneck during the LGM, whereas *Percichthys truch* experienced sustained growth in effective population size for the last 100ka BP. However, the two species differ in physiological traits, habitat preferences, feeding ecology, ability to colonize new habitats, and predation pressure [45], which conceivably explain the observed demographic dynamics of current populations of *G. platei* and *P. truch* [45]. The diversity of factors potentially affecting the demography of populations in a variety of species possibly explain the complexity of outcomes as the cited above, with different factors playing a more important role in each system.

## 5. Conclusion

The emerging complexity and diversity of demographic regimes in terrestrial and aquatic species during the late Pleistocene-Holocene both in the Northern and in Southern hemispheres, in particular regarding the LGM, is remarkable. The pattern of variation in effective population size we uncovered for *P. vivipara*, particularly with respect the role of the LGM, adds to the already known complexity. In this connection, comparative studies of species differing in life history and ecology, but sharing common environments, will be instrumental in uncovering the processes underlying the historical demography of contemporary populations. Such an approach might lead to the discovery of the factors that determine whether the demographic dynamics coincides or not with the episodic climatic events of the Late Pleistocene-Holocene.

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## Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

## References

- [1] Hewitt, G.M. (2004) Genetic Consequences of Climatic Oscillations in the Quaternary. *Philosophical Transactions of the Royal Society, Biological Sciences*, **359**, 183-195. <https://doi.org/10.1098/rstb.2003.1388>
- [2] Liu, J., Guo, X., Chen, D., Li, J., Yue, B. and Zeng, X. (2019) Diversification and Historical Demography of the Rapid Racerunner (*Eremias velox*) in Relation to Geological History and Pleistocene Climatic Oscillations in Arid Central Asia. *Molecular Phylogenetics and Evolution*, **130**, 244-258. <https://doi.org/10.1016/j.ympev.2018.10.029>
- [3] Silveira, M.H.B., Mascarenhas, R., Cardoso, D. and Batalha-Filho, H. (2019) Pleistocene Climatic Instability Drove the Historical Distribution of Forest Islands in the Northeastern Brazilian Atlantic Forest. *Palaeogeography, Palaeoclimatology, Palaeoecology*, **527**, 67-76. <https://doi.org/10.1016/j.palaeo.2019.04.028>
- [4] Margaryan, A., *et al.* (2017) Eight Millenia of Matrilineal Genetic Continuity in the South Caucasus. *Current Biology*, **27**, 2023-2028. <https://doi.org/10.1016/j.cub.2017.05.087>
- [5] Önkamo, S., *et al.* (2017) Identification and Analysis of mtDNA Genomes Attributed to Finns Reveal Long-Stagnant Demographic Trends Obscured in the Total Diversity. *Scientific Reports*, **7**, Article No. 6193. <https://doi.org/10.1038/s41598-017-05673-7>
- [6] Barrick, J.E. and Lenski, R.E. (2013) Genome Dynamics during Experimental Evolution. *Nature Reviews of Genetics*, **149**, 827-839. <https://doi.org/10.1038/nrg3564>
- [7] Bromham, L. (2016) An Introduction to Molecular Evolution and Phylogenetics. Oxford University Press, Oxford.
- [8] Chang, D. and Shapiro, B. (2016) Using Ancient DNA and Coalescent-Based Methods to Infer Extinction. *Biology Letters*, **12**, Article ID: 20150822. <https://doi.org/10.1098/rsbl.2015.0822>
- [9] Ho, S.Y.W. and Shapiro, B. (2011) Skyline-Plot Methods for Estimating Demographic History from Nucleotide Sequences. *Molecular Ecology Resources*, **11**, 423-434. <https://doi.org/10.1111/j.1755-0998.2011.02988.x>
- [10] Shapiro, B., *et al.* (2004) Rise and Fall of the Beringian Steppe Bison. *Science*, **306**, 1561-1565. <https://doi.org/10.1126/science.1101074>
- [11] Drummond, A.J., Rambaut, A., Shapiro, B. and Pybus, O.G. (2005) Bayesian Coalescent Inference of Past Population Dynamics from Molecular Sequence. *Molecular Biology and Evolution*, **22**, 185-1192. <https://doi.org/10.1093/molbev/msi103>
- [12] Froese, D., *et al.* (2016) Fossil and Genomic Evidence Constrains the Timing of Bison Arrival in North America. *Proceedings of the National Academy of Sciences*, **113**, 8057-8063.
- [13] Leonardi, M., *et al.* (2017) Evolutionary Patterns and Processes: Lessons from Ancient DNA. *Systematic Biology*, **66**, e1-e29.
- [14] Lorenzen, E.D., *et al.* (2011) Species-Specific Responses of Late Quaternary Megafauna to Climate and Humans. *Nature*, **479**, 359-364. <https://doi.org/10.1038/nature10574>

- [15] Provan, J. and Bennett, K. (2008) Phylogeographic Insights into Cryptic Glacial Refugia. *Trends in Ecology and Evolution*, **23**, 564-571. <https://doi.org/10.1016/j.tree.2008.06.010>
- [16] Watanabe, L.A., *et al.* (2014) The Past and Present of an Estuarine-Resident Fish, the “Four-Eyed Fish” *Anableps anableps* (Cyprinodontiformes, Anablepidae), Revealed by mtDNA Sequences. *PLoS ONE*, **9**, e101727. <https://doi.org/10.1371/journal.pone.0101727>
- [17] Kowsmann, R.O. and Costa, M.P.A. (1974) Paleolinhas de costa na plataforma continental das regiões sul e norte brasileiras. *Revista Brasileira de Geologia*, **4**, 215-222. <https://doi.org/10.25249/0375-7536.1981215222>
- [18] Correa, I.C. (1996) Les variations du niveau de la mer durant les derniers 17.500 ans BP: l'exemple de la plateforme continentale du Rio Grande do Sul, Brésil. *Marine Geology*, **130**, 163-178. [https://doi.org/10.1016/0025-3227\(95\)00126-3](https://doi.org/10.1016/0025-3227(95)00126-3)
- [19] Cohen, M., *et al.* (2005) A Model of Holocene Mangrove Development and Relative Sea-Level Changes on the Bragança Peninsula (Northern Brazil). *Wetland Ecology and Management*, **13**, 433-443. <https://doi.org/10.1007/s11273-004-0413-2>
- [20] Bassar, R.D., *et al.* (2010) Local Adaptation in Trinidadian Guppies Alters Ecosystem Processes. *Proceedings of the National Academy of Sciences USA*, **107**, 3616-3621. <https://doi.org/10.1073/pnas.0908023107>
- [21] Bassar, R.D., Simon, T., Roberts, W., Travis, J. and Reznick, D.N. (2017) The Evolution of Coexistence: Reciprocal Adaptation Promotes the Assembly of a Simple Community. *Evolution*, **71**, 373-385. <https://doi.org/10.1111/evo.13086>
- [22] Gomes, J.L. and Monteiro, L.R. (2008) Morphological Divergence Patterns among Populations of *Poecilia vivipara* (Teleostei: Poeciliidae): Test of an Ecomorphological Paradigm. *Biological Journal of the Linnean Society*, **93**, 799-812. <https://doi.org/10.1111/j.1095-8312.2007.00945.x>
- [23] Aljanabi, S. and Martinez, I. (1997) Universal and Rapid Salt-Extraction of High Quality Genomic DNA for PCR-Based Techniques. *Nucleic Acids Research*, **25**, 4692-4693. <https://doi.org/10.1093/nar/25.22.4692>
- [24] Schories, S., Meyer, M.K. and Scharlt, M. (2009) Description of *Poecilia (Acanthophaelus) obscura* (Teleostei: Poeciliidae), a New Guppy Species from Western Trinidad, with Remarks on *P. wingei* and the Status of the “Endler’s Guppy”. *Zootaxa*, **2266**, 35-50.
- [25] Tajima, F. (1989) Statistical Method for Testing the Neutral Mutation Hypothesis by DNA Polymorphism. *Genetics*, **123**, 585-595.
- [26] Librado, P. and Rozas, J. (2009) DnaSP v5: A Software for Comprehensive Analysis of DNA Polymorphism Data. *Bioinformatics*, **25**, 1451-1452. <https://doi.org/10.1093/bioinformatics/btp187>
- [27] Templeton, A.R., Crandall, K.A. and Sing, C.F. (1992) A Cladistic Analysis of Phenotypic Associations with Haplotypes Inferred from Restriction Endonuclease Mapping and DNA Sequence Data. III. Cladogram Estimation. *Genetics*, **132**, 619-633.
- [28] Clement, M., Posada, D. and Crandall, K.A. (2000) TCS: A Computer Program to Estimate Gene Genealogies. *Molecular Ecology*, **9**, 1657-1659. <https://doi.org/10.1046/j.1365-294x.2000.01020.x>
- [29] Dupanloup, I., Schneider, S. and Excoffier, L. (2002) A Simulated Annealing Approach to Define the Genetic Structure of Populations. *Molecular Ecology*, **11**, 2571-2581. <https://doi.org/10.1046/j.1365-294X.2002.01650.x>

- [30] Drummond, A.J., Suchard, M.A., Xie, D. and Rambaut, A. (2012) Bayesian Phylogenetics with BEAUti and the BEAST 1.7. *Molecular Biology and Evolution*, **29**, 1969-1973. <https://doi.org/10.1093/molbev/mss075>
- [31] Darriba, D., Taboada, G.L., Doallo, R. and Posada, D. (2012) jModelTest 2: More Models, New Heuristics and Parallel Computing. *Nature Methods*, **9**, 772. <https://doi.org/10.1038/nmeth.2109>
- [32] Rambaut, A., Suchard, M., Xie, D. and Drummond, A. (2014) Tracer v1.6. Computer Program and Documentation Distributed by the Author. <http://beast.bio.ed.ac.uk/Tracer>
- [33] Meredith, R.W., Pires, M.N., Reznick, D.N. and Springer, M.S. (2010) Molecular Phylogenetic Relationships and the Evolution of the Placenta in *Poecilia* (*Micro-poecilia*) (Poeciliidae: Cyprinodontiformes). *Molecular Phylogenetics and Evolution*, **55**, 631-639. <https://doi.org/10.1016/j.ympev.2009.11.006>
- [34] Chikhi, L., Sousa, V.C., Luisi, P., Goossens, B. and Beaumont, M.A. (2010) The Confounding Effects of Population Structure, Genetic Diversity and the Sampling Scheme on the Detection and Quantification of Population Size Changes. *Genetics*, **186**, 983-995. <https://doi.org/10.1534/genetics.110.118661>
- [35] Heller, R., Chikhi, L. and Siegmund, H.R. (2013) The Confounding Effect of Population Structure on Bayesian Skyline Plot Inferences of Demographic History. *PLoS ONE*, **8**, e62992. <https://doi.org/10.1371/journal.pone.0062992>
- [36] Clark, P.U., *et al.* (2009) The Last Glacial Maximum. *Science*, **325**, 710-714. <https://doi.org/10.1126/science.1172873>
- [37] Delrieu-Trottin, E., *et al.* (2017) Population Expansions Dominate Demographic Histories of Endemic and Widespread Pacific Reef Fishes. *Scientific Reports*, **7**, Article No. 40519. <https://doi.org/10.1038/srep40519>
- [38] Araújo, L.G.B.R. and Monteiro, L.R. (2012) Growth Pattern and Survival in Populations of *Poecilia vivipara* (Teleostei: Poeciliidae) Inhabiting an Environmental Gradient: A Common Garden Study. *Environmental Biology Fish*, **96**, 941-951. <https://doi.org/10.1007/s10641-012-0089-z>
- [39] Araújo, M.S., Perez, S.I., Magazoni, M.J.C. and Petry, A.C. (2014) Body Size and Allometric Shape Variation in the Molly *Poecilia vivipara* along a Gradient of Salinity and Predation. *BMC Evolutionary Biology*, **14**, 251. <https://doi.org/10.1186/s12862-014-0251-7>
- [40] Sommer-Trembo, C. (2017) Predation Risk and Abiotic Habitat Parameters Affect Personality Traits in Extremophile Populations of a Neotropical Fish (*Poecilia vivipara*). *Ecology and Evolution*, **7**, 6570-6581. <https://doi.org/10.1002/ece3.3165>
- [41] Berlin, S. (2017) Genetics of Phenotypic Plasticity and Biomass Traits in Hybrid Willows across Contrasting Environments and Years. *Annals of Botany*, **120**, 87-100. <https://doi.org/10.1093/aob/mcx029>
- [42] Salvi, D., Schembri, P.J., Sciberras, A. and Harris, D.J. (2014) Evolutionary History of the Maltese Wall Lizard *Podarcis filfolensis*: Insights on the “Expansion-Contraction” Model of Pleistocene Biogeography. *Molecular Ecology*, **23**, 1167-1187. <https://doi.org/10.1111/mec.12668>
- [43] Bisconti, R., Canestrelli, D., Colangelo, P. and Nascetti, G. (2011) Multiple Lines of Evidence for Demographic and Range Expansion of a Temperate Species (*Hylasarda*) during the Last Glaciation. *Molecular Ecology*, **20**, 5313-5327. <https://doi.org/10.1111/j.1365-294X.2011.05363.x>
- [44] Tschá, M.K., *et al.* (2016) Sea-Level Variations Have Influenced the Demographic History of Estuarine and Freshwater Fishes of the Coastal Plain of Paraná, Brazil.

*Journal of Fish Biology*, **90**, 968-979. <https://doi.org/10.1111/jfb.13211>

- [45] Ruzzante, D.E., *et al.* (2008) Climate Control on Ancestral Population Dynamics: Insight from Patagonian Fish Phylogeography. *Molecular Ecology*, **17**, 2234-2244. <https://doi.org/10.1111/j.1365-294X.2008.03738.x>
- [46] Pillans, B., Chappell, J. and Naish, T. (1998) A Review of the Milankovich Climate Beat: Template for Plio-Pleistocene Sea-Level Changes and Sequence Stratigraphy. *Sedimentary Geology*, **122**, 5-21. [https://doi.org/10.1016/S0037-0738\(98\)00095-5](https://doi.org/10.1016/S0037-0738(98)00095-5)
- [47] Nicol, A. (2011) Landscape History of the Marlborough Sounds, New Zealand. *New Zealand Journal of Geology and Geophysics*, **54**, 195-208. <https://doi.org/10.1080/00288306.2010.523079>
- [48] Bastos, A.C. and Silva, C.G. (2000) Caracterização morfodinâmica do litoral Norte Fluminense, RJ, Brasil. *Revista Brasileira de Oceanografia*, **48**, 41-60. <https://doi.org/10.1590/S1413-77392000000100004>