

Differences in Modified Morgan Phosphorus Levels Determined by Colorimetric and Inductively Coupled Plasma Methods

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ABSTRACT

Phosphorus (P) fertilization is frequently needed for profitable crop production. Modified Morgan P (MMP) is a soil test P used to estimate plant available P in soils. The critical values of MMP for P fertilization and maintenance recommendations are based on the P concentrations measured by a common colorimetric molybdenum blue method although other P quantification methods have also been used for MMP measurements. In this study, we collected 120 surface soil samples of Caribou Sandy loam under potato cultivation or its rotation crops from Maine, USA, and 72 soil samples of Cecil sandy loam with cotton/corn crops under conventional tillage and no-till management with chemical and poultry litter fertilization in Georgia, USA. The MMP levels in all 192 dry samples were greater when they were measured by an inductively coupled plasma (ICP)-based method, compared to the corresponding data produced from colorimetry. Our results show the two sets of data were positively and significantly correlated ($r = 0.93$, $P < 0.001$). In average, the ICP-based MMP level of the 192 samples was $23.3 \text{ mg} \cdot \text{P} \cdot \text{kg}^{-1}$ with standard deviation of 12.9, compared to the average of colorimetric MMP level of $14.9 \text{ mg} \cdot \text{P} \cdot \text{kg}^{-1}$ with standard deviation of 8.8. Based on the observations in this work, both colorimetric and ICP-based methods can be used for P fertilizer recommendation, but a conversion factor should be applied for ICP data as the current recommendation systems are based on colorimetric M & R data.

Keywords: Phosphorus Measurement; Soil Test Phosphorus; Modified Morgan Phosphorus; ICP; Molybdenum Blue Method

1. Introduction

Phosphorus (P) is an essential plant nutrient. P fertilization is frequently needed for profitable crop production. Soil test phosphorus (STP) is an estimate of P in soil that is part of soil P available for plant uptake during a growing season. Modified Morgan P (MMP) is one of many STP methods used in the US. This method has been used to evaluate STP in Maine [1,2] and other northeastern states [3,4]. Traditionally, P concentration in a soil extract is measured by a colorimetric method, which measures the soluble inorganic P [5] or molybdate (Mo)-reactive P [6]. With the advancement of technology, inductively coupled plasma (ICP)-based methods, which mea-

sure total P in a soil extract, are also used in STP measurement [7]. As colorimetric P determination methods are initially used to calibrate STP for P fertilizer recommendations, the difference in STP data determined by colorimetric methods and ICP-based methods need to be examined and documented in order to appropriately interpret ICP-based STP data [7]. Indeed, the difference in Mehlich 3 P (M3P), another STP, measured by Mo-reactive and ICP methods has been rigorously examined by numerous research groups [7-10]. Based on about 6500 soil samples, Pittman *et al.* [7] reported that the M3P values measured by ICP-atomic emission spectrometer (AES) were higher than those determined colorimetrically. These researchers proposed two regression equations. for $P >$ and $< 60 \text{ mg} \cdot \text{kg}^{-1}$ respectively. The first was to convert ICP-based data to colorimetric M3P values to make fertilization recommendations if $STP < 60 \text{ mg} \cdot \text{kg}^{-1}$, and other conversion was for $STP > 60 \text{ mg} \cdot \text{kg}^{-1}$ for other uses.

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The difference in MMP measured by colorimetry and ICP methods has not been extensively investigated so far. In the limited literature noted, the ICP method measured an average of $1.5 \text{ mg}\cdot\text{kg}^{-1}$ more MMP than colorimetry [4]. Some researchers [11] argued that it is a substantial difference since critical soil MMP levels are in the range of $4 - 7 \text{ mg}\cdot\text{kg}^{-1}$. He *et al.* [12] found that MMP concentrations measured by an ICP method were two times greater than those measured by a colorimetric method as the average $\text{MMP}_{\text{Colorimetric}}$ and MMP_{ICP} contents of 10 Maine soil samples were $7.24 \text{ mg}\cdot\text{kg}^{-1}$ and $14.96 \text{ mg}\cdot\text{kg}^{-1}$, respectively. To further explore the difference in MMP measured by a different method, we collected 120 surface soil samples from plots cultivated to potato and its rotation crops in Maine, USA, and determined their MMP levels using different colorimetric and ICP-based methods. The study provides useful reference on more appropriate application of MMP data resulting from different P assay methods.

2. Materials and Methods

2.1. Soil Sample Collection

Soil samples were collected from two sites. The first sample site was the USDA-ARS research field in Presque Isle, Maine, USA (Latitude $46^{\circ}41'N$, Longitude $68^{\circ}2'W$). The soil is classified as Caribou Sandy loam (fine-loamy, isotic, frigid Typic Haplorthods). Potato or rotation crops had been planted with five production systems as 1) Continuous Potato (CP), a non-rotation control; 2) Status Quo (SQ) a typical 2-yr rotation practice in the area: potato (Yr 1) followed by barley (Yr 2); 3) Disease Suppressive (DS): mustard green manure/winter rapeseed (Yr 1)—sorghum sudangrass/winter rye (Yr 2)—potato (Yr 3); 4) Soil Conserving (SC): barley underseeded with timothy (Yr 1)—timothy sod (Yr 2)—potato (Yr 3) with mulch after harvest; and 5) Soil Improving (SI): same as SC, with compost ($20 \text{ Mg}\cdot\text{ha}^{-1}$) added to each crop [13]. Fertilizer was applied at the annual rate of $2240 \text{ kg}\cdot\text{ha}^{-1}$ of commercial $10(N)-10(P_2O_5)-10(K_2O)$ fertilizer (fertilizers forms are ammonium nitrate, diammonium phosphate and potassium muriate) in bands approximately 5 cm to the side and 5 cm below the seed [13]. Extra P input ($39 \text{ kg}\cdot\text{P}\cdot\text{ha}^{-1}\cdot\text{yr}^{-1}$ based on five year average) was added in the SI plots from compost addition. All of the production systems were managed under both rainfed and irrigated conditions and there were five replications of each treatment. Irrigation water (1.25 cm) was applied to all irrigated treatments when 25% of the tensiometers placed in irrigated plots register 50 kPa reading. Soil samples under both irrigated and rainfed managements were collected in May 2010 after the completion of two three-year crop rotations.

The second sample site was the water quality facility at the USDA-ARS, J. Phil Campbell, Sr. Natural Resource Conservation Center, Watkinsville, Georgia, USA ($83^{\circ}24'W$ and $33^{\circ}54'N$) [14,15]. The facility consists of 12 large ($10 \text{ m} \times 30 \text{ m}$) tile-drained plots, located on nearly level ($<2\%$ slope) Cecil sandy loam soil (fine, kaolinitic, thermic Typic Kanhapludults). The facility was managed over ten years for cotton and corn production under combinations of two tillage types, conventional tillage and no-till management, and two fertilizer sources, poultry litter and inorganic (chemical) fertilizer. In the fall of 1997, 2000, and 2005, soil samples (0 to 15 cm) were collected with a tractor mounted hydraulic soil coring device from three locations in each plot. In addition, in Oct 2006, 0- to 2.5, 2.5- to 5, and 5- to 15-cm samples were collected from each plot in the same way.

2.2. Modified Morgan Extraction

For modified Morgan P (MMP), soils (5.0 g dry soil or equivalent) were extracted by 20 mL of $0.62 \text{ M NH}_4\text{OH}/1.25 \text{ M CH}_3\text{COOH}$ (pH 4.8) for 15 mins [12]. MMP was extracted first from a set of 25 wet soil samples. Later MMP was extracted from the whole set of 120 dry soil samples. The extract and soil residues were separated by centrifuging at $14,000 \times g$ for 30 min at 4°C . The supernatant (*i.e.* the extract) was removed and past through a $0.45 \mu\text{m}$ filter. Those supernatants were saved and kept at 4°C prior to P determination. In addition, a set of Maine soil samples was sent to University of Maine Analytical Laboratory for MMP measurements for comparison.

2.3. Phosphorus Determination

Two molybdenum (Mo) blue methods were used in colorimetric MMP determination. The first one was based on He and Honeycutt (H & H) [5] and the second was the Murphy and Riley (M & R) method as modified by Watanabe and Olsen [16]. ICP-AES P was determined using a Spectra Arco Spectrophotometer (Perkin-Elmer, Norwalk, CT) at Oklahoma State University. ICP-AES P analyzed at University of Maine was conducted using a Thermo Jarrell Ash IRIS 1000 Dual-view Spectrometer (Franklin, MA).

3. Results and Discussion

3.1. Difference in P Determined by Two Mo Blue Methods

Figure 1 shows the MMP concentrations in 25 Maine wet soil samples collected from five PP and SQ plots. MMP in most soil samples was greater when measured by M & R than by H & H. The MMP concentration was

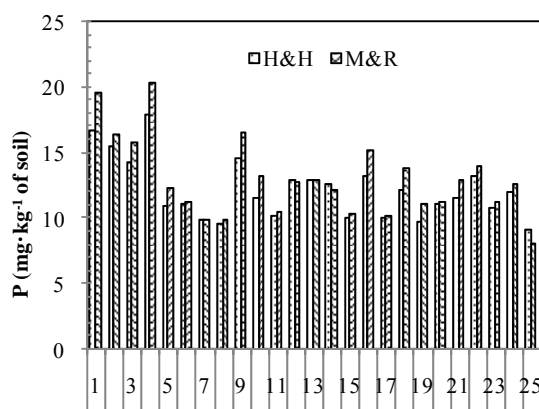


Figure 1. Modified Morgan P concentrations in 25 Maine wet soils from Continuous Potato (PP) and Status Quo (SQ) plots measured by Mo blue methods based He and Honeycutt (H & R) [5] or Murphy and Riley (M & R) [16]. These plots were either rainfed (R) or irrigated (I) with potato (P) or barley (B) grown.

only slightly lower in M & R measurement than by H & H in Sample 12, 14 and 25, apparently due to the fact that the assay errors are greater than the difference between the two methods. The average of MMP of the 25 samples is $12.13 \text{ mg}\cdot\text{kg}^{-1}$ of dry soil with SD of 2.30 measured by H & H, and $12.96 \text{ mg}\cdot\text{kg}^{-1}$ of dry soil with SD of 3.01 measured by M & R. The results were reasonable as H & H method was adapted to measure soluble inorganic orthophosphate only [5,17]. Whereas M & R measures some organic P that may be hydrolyzed during the assay, leading to a greater P measurement than inorganic orthophosphate [17] alone so the P measured by M & R has been alternatively termed “Mo-reactive P” [6,18]. Statistical analysis of data in 1 indicates the two sets of data are positively and significantly correlated ($r^2 = 0.944$, $P < 0.001$). The linear regression is $\text{MMP}_{\text{M \& R}} = 1.271 \text{ MMP}_{\text{H \& H}} + 2.457$ in $\text{mg}\cdot\text{kg}^{-1}$. Our observations imply that whereas M & R method is practical for soil P test measurement, H & H method should be applied for more sophisticated or highly quantitative research involving accurate measurements of inorganic P and organic P, such as in hydrolysable organic P studies [19-22].

3.2. Relationship between Colorimetric M & R MMP and ICP-AES MMP for the Maine Soil

The MMP concentrations of 120 dry soil samples from Maine potato fields measured by the two methods are shown in **Figure 2(a)**. The MMP concentrations of all 120 samples measured by M & R method were less than by ICP-AES. The average of the colorimetric measurements of all samples was $16.9 \text{ mg}\cdot\text{kg}^{-1}$ of soil with SD of 4.5 whereas the average of ICP-AES measurements 27.6

$\text{mg}\cdot\text{kg}^{-1}$ with the SD of 5.9. Specifically, the averages and standard deviations of the colorimetric and ICP-AES measurements were $(15.5 \pm 2.3) \text{ mg}\cdot\text{kg}^{-1}$ and $(24.9 \pm 2.3) \text{ mg}\cdot\text{kg}^{-1}$ for 45 soils from rainfed plots without compost addition, $(14.4 \pm 2.4) \text{ mg}\cdot\text{kg}^{-1}$ and $(24.3 \pm 3.0) \text{ mg}\cdot\text{kg}^{-1}$ for 45 soils from irrigated plots without compost addition, $(23.1 \pm 4.8) \text{ mg}\cdot\text{kg}^{-1}$ and $(34.4 \pm 9.2) \text{ mg}\cdot\text{kg}^{-1}$ for 15 soils from rainfed plots with compost addition, and $(22.2 \pm 4.3) \text{ mg}\cdot\text{kg}^{-1}$ and $(32.1 \pm 6.1) \text{ mg}\cdot\text{kg}^{-1}$ for 15 soils from irrigated plots with compost addition. Compost addition contributed an additional $10 \text{ mg}\cdot\text{kg}^{-1}$ of MMP in these soils. However, irrigation did not significantly impact MMP levels, which is similar to an earlier observation on labile water soluble P in soils from these plots after the first 3-year crop rotation in 2007 [23].

Visual examination of **Figure 2(a)** suggests that the difference in MMP values between colorimetric and ICP-AES methods is not affected by the treatments. The P concentration is a more critical factor as seen in the changing trend above $27 - 30 \text{ mg}\cdot\text{P}\cdot\text{kg}^{-1}$ measured by the colorimetric method. There were 119 soil samples below the value of $30 \text{ mg}\cdot\text{P}\cdot\text{kg}^{-1}$. The linear regression of these data points is $\text{MMP}_{\text{ICP-AES}} = 1.1 \text{ MMP}_{\text{Colorimetric}} + 8.0$ in $\text{mg}\cdot\text{kg}^{-1}$ ($r^2 = 0.92$, $P < 0.001$). The slope near 1 and intercept near 10 imply that the difference between the two measurements was mainly due to a stable (about $10 \text{ mg}\cdot\text{kg}^{-1}$) organic P component in these soils. Our previous work after completing the 3-yr crop rotation also found no significant change of organic P fractions in these soils attributable to crop rotation or irrigation [23]. Previously, a regression of $\text{MMP}_{\text{ICP-AES}} = 0.98 \text{ MMP}_{\text{Colorimetric}} + 1.5$ ($\text{MMP}_{\text{Colorimetric}} < 30 \text{ mg}\cdot\text{P}\cdot\text{kg}^{-1}$) was reported based on 51 soil samples collected from corn fields in 10 Northeastern USA states [4]. The lower constant value suggested a lower level of organic P in these corn fields than that in the potato field we tested. The eight soils with $\text{MMP}_{\text{Colorimetric}} > 27 \text{ mg}\cdot\text{P}\cdot\text{kg}^{-1}$ all were from soils where compost was added. Organic P in these eight soils as determined by the Modified Morgan extraction was greater than for the other soils. These data points can be mathematically expressed by another regression $\text{MMP}_{\text{ICP-AES}} = 6.26 \text{ MMP}_{\text{Colorimetric}} - 134$ in $\text{mg}\cdot\text{kg}^{-1}$ ($r^2 = 0.99$, $P < 0.001$).

The relative difference between the two sets of data is shown in **Figure 2(b)**. The ratio of ICP-AES P/Colorimetric P is 1.3 to 2.2 with the lowest ratios between $20 - 25 \text{ mg MMP}_{\text{Colorimetric}} \text{ kg}^{-1}$ soil. The greater ratio seems to be due to the indigenous organic P at the low MMP levels. The greater ratio shown by the high MMP levels could be attributed to external organic P input from compost. When all data was combined, the ratio of ICP-AES P/Colorimetric P could be expressed by a non-linear equation as shown in **Figure 2(b)**.

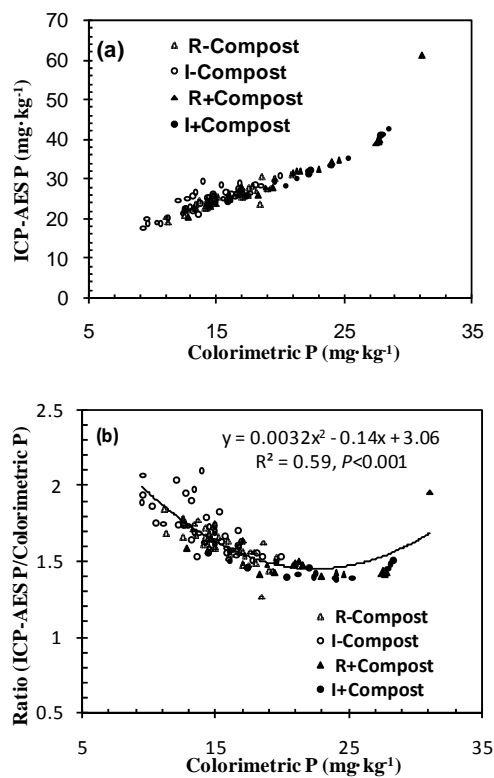


Figure 2. Modified Morgan P concentrations in 120 Maine dry soil samples measured by colorimetric Murphy and Riley method and ICP-AES. (a) In absolute concentrations; (b) in ratio of ICP-AES P/Colorimetric P. All samples were from potato or rotation crop fields under rainfed (R) or irrigated management with (+) or without (-) additional compost application.

3.3. Relationship between Colorimetric M & R MMP and ICP-AES MMP for the Georgia Soil

The MMP concentrations of 72 dry soil samples from Georgia cotton/corn fields measured by the two methods are shown in **Figure 3**. The colorimetric MMP concentrations varied from 0.5 to 41 mg·kg⁻¹ of soil. The ICP-AES MMP concentrations varied from 2.5 to 102 mg·kg⁻¹ of soil. The greater variation of the Georgia soil data than the Maine soil data was apparently due to the different fertilization/soil managements. All Maine soils except those with compost had received the same NPK 10-10-10 fertilizer at the same rate so that the variation of their MMP concentration was relatively small. Those Georgia soils had received different types of fertilizers (chemical fertilizer and poultry litter) with conventional tillage or no-till management. Previous data [14] have shown the no-till effect retained most of P from poultry litter application in the surface soil (0 - 2.5 cm). Indeed, the highest 3 MMP concentrations were observed with the 0 - 2.5 cm soil samples (**Figure 3**). The low MMP concentrations

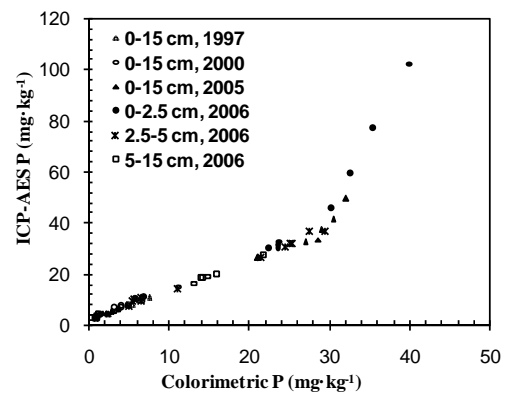


Figure 3. Modified Morgan P concentrations in 72 Georgia dry soil samples measured by colorimetric Murphy and Riley method and ICP-AES. All samples were collected from cotton/corn plots from the indicated soil depths at the specific years.

were with subsurface soil samples or with the samples collected in earlier years as P from poultry litter accumulated in these soils over time [14]. In spite of those differences, the relationship of the colorimetrically and ICP-AES-measured MMP concentrations of these Georgia soils was similar to that of Maine soils as the two-line's pattern was observed with a changing trend started at 27 - 30 mg·P·kg⁻¹ measured by the colorimetric method. The linear regression of the 66 data points with colorimetric MMP < 30 mg·kg⁻¹ is $MMP_{ICP-AES} = 1.2 MMP_{Colorimetric} + 1.9$ in mg·kg⁻¹ ($r^2 = 0.99$, $P < 0.001$). The 11 data points with colorimetric MMP > 27 mg·kg⁻¹ is mathematically expressed by the regression $MMP_{ICP-AES} = 5.57 MMP_{Colorimetric} - 124$ in mg·kg⁻¹ ($r^2 = 0.97$, $P < 0.001$). These two regressions were comparable to those two for Maine soils, indicating the applicability of the observations of these regressions to a wider soil types with different cropping/soil managements. Combined all sample data from both experimental locations, the linear regressions are $MMP_{ICP-AES} = 1.3 MMP_{Colorimetric} + 3.1$ in mg·kg⁻¹ ($r^2 = 0.94$, $P < 0.001$) with colorimetric MMP < 30 mg·kg⁻¹, and $MMP_{ICP-AES} = 5.0 MMP_{Colorimetric} - 102.0$ in mg·kg⁻¹ ($r^2 = 0.91$, $P < 0.001$) with colorimetric MMP > 27 mg·kg⁻¹.

3.4. Effects of Soil Drying and Inter-Laboratory Assay on MMP Measurements

Figure 4 shows the relationship of colorimetric MMP of the 25 field moist soil samples in **Figure 1** with the corresponding dried soils in **Figure 2**. The fact that almost all data points are above the diagonal dash line in **Figure 4** implies that air drying increased the colorimetric MMP level. The difference was generally <3 mg·kg⁻¹ since the average and SD of MMP data were (13.0 ± 3.0) mg·kg⁻¹ and (15.7 ± 2.4) mg·kg⁻¹ for the wet and dry samples,

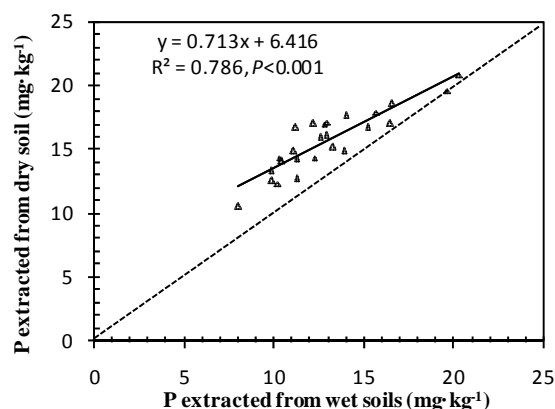


Figure 4. The relationship of MMP in wet and air-dried samples of same Maine soils measured by colorimetric Murphy and Riley method.

respectively. The significant linear regression (**Figure 4**) suggested the regression line was different from zero and that the two sets of data vary. The high value in the dry soil could be attributed to drying-induced P release through microbial cell lysis, organic matter destabilization, P desorption site exposure [24].

To examine the repeatability of ICP-AES MMP measurement from different laboratories, the 120 dry soils were analyzed by the University of Maine Analytic Laboratory. Samples had greater MMP levels, while 109 samples had lower MMP levels when measured at the University of Maine compared to the measurements made at Oklahoma State University (**Figure 5**). The difference between the two sets of data was around $6 \text{ mg}\cdot\text{kg}^{-1}$ and with a SD of $(21.1 \pm 7.0) \text{ mg}\cdot\text{kg}^{-1}$ for the University of Maine data, and $(26.8 \pm 5.9) \text{ mg}\cdot\text{kg}^{-1}$ for the Oklahoma State University data. Statistical analysis indicates that the two sets of data were significantly and highly correlated ($r^2 = 0.90$, $P < 0.001$) (**Figure 5**). Thus, the MMP measurements seem repeatable and the differences between the different labs were within an acceptable range and were similar to the variance found previously for an interlaboratory comparison of Mehlich 3 P [25].

4. Conclusions

Phosphorus (P) fertilization is frequently needed for profitable crop production. Modified Morgan P (MMP) is a soil test P used to estimate plant available P in soils. MMP levels in 120 Caribou Sandy loam soil samples from potato and rotation crops plots in Maine, USA, and 72 Cecil sandy loam soil samples from cotton/corn plots under conventional tillage and no-till managements with chemical or poultry litter fertilization in Georgia, USA, were evaluated by colorimetric Mo blue and ICP-AES

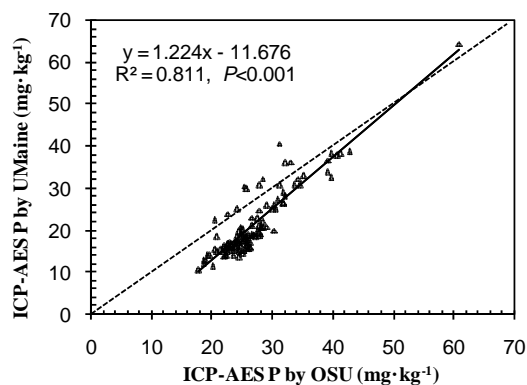


Figure 5. The inter-lab difference in ICP-AES MMP of 120 Maine dry samples measured by Oklahoma state university and university of Maine.

methods. The MMP levels in Maine soils measured by different methods were compared. For the 25 field moist soil samples, the MMP levels measured by M & R method were greater than those measured by a modified Mo blue method, in which the modification improved the specificity on soluble inorganic P measurement. Based on the observations in this work, we recommend the modified Mo blue and ICP methods be used for investigating inorganic P and organic P partition in soil MMP pools.

The data from both Maine and Georgia soil samples show the same pattern in relationships between colorimetrically- and ICP-AES-measured MMP concentrations. That is, there was turning point between 27 - 30 $\text{mg}\cdot\text{P}\cdot\text{kg}^{-1}$ of soil measured by colorimetry. Combined the two sets of data (total 192 soil samples tested), 184 samples were below the value of 30 $\text{mg MMP}_{\text{Colorimetric}} \text{ kg}^{-1}$ measured by Murphy-Riley method. The linear regression of these data points is $\text{MMP}_{\text{ICP-AES}} = 1.3 \text{ MMP}_{\text{Colorimetric}} + 3.1$ in $\text{mg}\cdot\text{kg}^{-1}$. The 19 data points higher than 27 $\text{MMP}_{\text{Colorimetric}}$ can be expressed by a different linear regression equation of $\text{MMP}_{\text{ICP-AES}} = 5.0 \text{ MMP}_{\text{Colorimetric}} - 102.0$ in $\text{mg}\cdot\text{kg}^{-1}$. The highly correlated relationships between the two sets of data imply that both colorimetric and ICP methods can be used for MMP measurements. But a conversion factor should be applied for soil P fertilization recommendation as the current recommendation system is based on the MMP values measured by the colorimetric method. More field samples should be tested for confirming the conversion factor as it could vary with other soil types and cropping management practices due to the presence of different pools of P.

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