

Prediction of the Fragmentation Pathway of Atorvastatin by Using High Resolution Collision Induced Dissociation (HR-MS/MS) Spectral Information

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Abstract

Introduction: A possible fragmentation pathway of atorvastatin was proposed based on rational interpretation of high resolution collision induced dissociation (CID) fragmentation spectral data. **Method**: The mass spectral (MS and MS/MS) data of atorvastatin was obtained by using a flow injection analysis, LC coupled with high resolution mass analyzer system with Q-TOF technology. **Results**: The elemental composition for each major fragment was proposed with a calculated mass error in parts per million (ppm). The mass error found in this study is from 0.3 to 5.7 ppm; strongly supports all the proposed elemental composition of fragments. Based on the fragments, possible fragmentation pathway was proposed. **Conclusion**: The workflow followed for interpretation can also address the structural possibilities of similar type of small organic molecules.

Subject Areas

Analytical Chemistry, Chemistry & Materials Science

Keywords

Atorvastatin, Small Drug Molecules, High Resolution Mass Spectrometry, Fragmentation, Rational Workflow, Interpretation

1. Introduction

A variety of spectroscopic techniques are available for the identification and

characterization of drug molecules. One of the most advanced analytical tools is high resolution mass spectrometry (HRMS) that has become the method of choice in structural characterization of small molecules, drug products, proteins and metabolites in different biological matrix [1] [2] [3] [4]. With advancements in ionization methods and instrumentation, high performance liquid chromatography coupled with high resolution mass spectrometry (LC-HRMS) has become a powerful tool for the research [5]-[11]. Interpretation of the fragmentation spectra and establishment of fragments structures are most time taking and essential tasks. The workflow [12] for the interpretation of the full scan atmospheric pressure ionization mass spectra (MS) and collision induced dissociation fragmentation spectra (MS/MS) of small organic molecules was efficiently applied for structural conformational studies; small nitrogenous organic molecule with hydroxyl and carboxylic acid functional group; atorvastatin was selected for this study.

Atorvastatin [13] [14] [15] is an inhibitor of 3-hydroxy-3-methylglutarylcoenzyme A (HMG-CoA) reductase. This enzyme catalyzes the conversion of HMG-CoA to mevalonate, an early and rate-limiting step in cholesterol biosynthesis. It is used primarily as a lipid-lowering agent and for prevention of events associated with cardiovascular disease especially people with type 2 diabetes, coronary heart disease, or other risk factors. It is an off-white crystalline powder and chemically known as Calcium-(3R,5R)-7-[2-(4-fluorphenyl)-5-isopropyl-3-phenyl-4-(phenylcarbamoyl)-1H-pyrrol-1-yl]-3,5-dihydroxyheptanoathydrat(1:2: 3). The empirical formula of atorvastatin calcium trihydrate is ($C_{66}H_{68}CaF_2$ N_4O_{10})3H₂O and its molecular weight is 1209.4. Atorvastatin free form empirical formula is $C_{33}H_{35}FN_2O_5$; monoisotopic molecular weight is 558.2530 and molecular structure of atorvastatin free form is presented in **Figure 1**.

During this study, high resolution mass spectral data of atorvastatin was generated using electrospray ionization and collision induced dissociation, followed by interpretation using basic interpretation rules and rational workflow [12] approach.



510 515 520 525 530 535 540 545 550 555 560 565 570 575 580 585 590 595 600 605 610 615 620 625 630 635 640 Figure 1. HRMS spectrum of atorvastatin using ESI⁺ ion source.

2. Experiment

2.1. Drug Sample

Atorvastatin was extracted from generic dosage form. A final concentration was about 10 µg/ml in water and methanol for mass spectrometric studies.

2.2. Chemicals and Reagents

The ultrapure water (18.2 $M\Omega$) was obtained using MilliQ apparatus from Millipore (Milford, USA) and the HPLC grade methanol was purchased from J.T. Baker.

2.3. Instrumentation

Shimadzu Prominence 20 AD HPLC (Kyto Japan) was coupled with Triple TOF (AB SCIEX) high resolution mass spectrometer system equipped with dual ionization (electrospray ionization and atmospheric pressure chemical ionization) source was used for this analysis. External calibrant delivery system (CDS) was used to calibrate the TOF mass analyzer with APCI small molecules calibration solution provided by instrument vendor.

2.4. Chromatographic and Mass Spectrometric Conditions

The extracted drug sample of atorvastatin was subjected to high resolution MS and MS/MS analysis via flow injection analysis (FIA) mode; liquid chromatography system was used to introduce the sample to mass spectrometer ion source. Liquid chromatography system set to isocratic flow 0.1 ml/min of mobile phase water and methanol in a ratio of 1:1 and injection volume was 10 μ l. Electrospray ion source was selected to achieve the intense protonated parent ion and high resolution mass analyzer selected to get the accurate mass; which supports to predict the right elemental composition. Spray voltage and collision energy optimized by using direct infusion mode to get optimum quality to spectral data. Experiments were acquired using optimized parameters; spray voltage of +5.0 kV for MS and collision energy (CE) setting of 35 V with a spread of ±15 V applied to all parent ions for CID (collision induced dissociation) for MS/MS.

3. Results and Discussion

The LC coupled with HRMS technique is a modern alternative method for the identification of small molecules and their impurities. The experimental data (MS and MS-MS spectral data) of atorvastatin was generated using a high performance liquid chromatography (HPLC) coupled with high resolution mass spectrometer system (HRMS) with Time of Flight (TOF) mass analyzer via Flow Injection Analysis (FIA) mode and Electro-spray Ionization (ESI⁺) ion source.

The full scan accurate mass and product ion spectrum of atorvastatin was obtained from IDA (information dependent acquisition) experiments. The workflow [12] was applied for the interpretation of high resolution MS and MS/MS spectral data; molecular ion peak was displayed $[M + H]^+$ at m/z 559.2635 Da. The most typical adduct $[M + Na]^+$ at m/z 581.2442 with a less abundant adduct $[M + K]^+$ at 597.2183 in mass spectrum of atorvastatin is further confirming the molecular ion peak at m/z 559.2635 with empirical formula $C_{33}H_{36}FN_2O_5^+$ (Figure 1).

The TOF MS/MS spectrum of atorvastatin also exhibited molecular ion peak at m/z 559.2635 Da as $[M + H]^+$ (calculated for $C_{33}H_{36}FN_2O_5^+$, exact mass 559.2603, difference 5.7 ppm, Figure 2 and Table 1). It was further fragmented in collision cell (Q2) into eleven major fragments at m/z 250.1038, 276.1194, 292.1508, 318.1654, 362.1561, 380.1675, 404.2032, 422.2139, 440.2239, 448.1936 and 466.2046. The elemental composition, exact mass, assigned structures and mass error of these major fragments were illustrated in Figure 2 and Table 1. Predicted fragmentation pathway also presented in Figure 2; the molecular ion peak (m/z 559.2635 Da) was initially fragmented into m/z 466.2046 (calculated formula $C_{27}H_{29}FNO_5^+$, exact mass 466.2024, mass error 4.7 ppm) by the loss of C₆H₇N, fragment ion 466.2046 yielded two ions at *m/z* 448.1936 (calculated formula $C_{27}H_{27}FNO_4^+$, exact mass 448.1919, mass error 3.8 ppm) and 440.2239 (calculated formula $C_{26}H_{31}FNO_4^+$, exact mass 440.2232, mass error 1.6 ppm) by loss of water and by loss of CO, respectively. Fragment m/z 440.2239 was further fragmented into two ions at m/z 422.2139 (calculated formula $C_{26}H_{29}FNO_3^+$, exact mass 422.2126, mass error 3.1 ppm) and 292.1508 (calculated formula $C_{20}H_{19}FN^+$, exact mass 292.1496, mass error 4.1 ppm) by loss of a water molecule and $C_4H_{12}O_4$, respectively. Further, a loss of water and C_3H_6 to the ion with m/z422.2139 lead fragment ions m/z 404.2032 (calculated formula $C_{26}H_{27}FNO_2^+$,

 Table 1. Interpretation of MS/MS spectra of atorvastatin based on rational work flow
 [12].

Measured Mass (<i>m/z</i>)	Electron Paring	Nitrogen Rule	No. of Nitrogen(s)	Proposed Formula	Calculated Mass (<i>m/z</i>)	Error* (ppm)
559.2635	$[M + H]^+$	EN	2	$C_{33}H_{36}FN_2O_5^+$	559.2603	5.7
466.2046	EE	ON	1	$C_{27}H_{29}FNO_5^+$	466.2024	4.7
448.1936	EE	ON	1	$\mathrm{C_{27}H_{27}FNO_4^+}$	448.1919	3.8
440.2239	EE	ON	1	$\mathrm{C}_{26}\mathrm{H}_{31}\mathrm{FNO}_4^+$	440.2232	1.6
422.2139	EE	ON	1	$C_{26}H_{29}FNO_{3}^{+}$	422.2126	3.1
404.2032	EE	ON	1	$\mathrm{C_{26}H_{27}FNO_2^+}$	404.2020	3.0
380.1675	EE	ON	1	$\mathrm{C}_{23}\mathrm{H}_{23}\mathrm{FNO}_3^+$	380.1656	5.0
362.1561	EE	ON	1	$\mathbf{C}_{23}\mathbf{H}_{21}\mathbf{FNO}_2^+$	362.1551	2.8
318.1654	EE	ON	1	$C_{22}H_{21}FN^{+}$	318.1653	0.3
292.1508	EE	ON	1	$C_{20}H_{19}FN^{+}$	292.1496	4.1
276.1194	EE	ON	1	$\mathrm{C_{19}H_{15}FN^{+}}$	276.1183	4.0
250.1038	EE	ON	1	$C_{17}H_{13}FN^{+}$	250.1027	4.4

EE: even electron; EN: even nitrogen; ON: odd nitrogen; *Mass error = difference between measured accurate mass and calculated accurate mass/calculated accurate mass $\times 10^6$.



 Into 180 190 200 210 220 230 240 250 260 270 280 290 300 310 320 330 340 350 360 370 380 390 400 410 420 430 440 450 460 470 480 490 500 510 520 530 540 550 560 570

 Figure 2. CID mass spectra (HR-MS/MS), proposed fragments and probable fragmentation pathway.

exact mass 404.2020, mass error 3.0 ppm) and 380.1675 (calculated formula $C_{23}H_{23}FNO_3^+$, exact mass 380.1656, mass error 5.0 ppm) was observed, respectively. The fragment ion with m/z 362.1561 (calculated formula $C_{23}H_{21}FNO_2^+$, exact mass 362.1551, mass error 2.8 ppm) shows a loss of C_3H_6 to a fragment ion with m/z 404.2032, which was further fragmented at m/z 318.1654 (calculated formula $C_{22}H_{21}FN^+$, exact mass 318.1653, mass error 0.3 ppm) corresponding to the loss of COOH.

The fragment at m/z 292.1508 (calculated formula $C_{20}H_{19}FN^+$, exact mass 292.1496, mass error 4.1 ppm) corresponding to the loss of $C_4H_{12}O_4$ from the ion m/z 440.2239. A subsequent fragment of ion m/z 292.1508 resulting peak at m/z 276.1194 (calculated formula $C_{19}H_{15}FN^+$, exact mass 276.1183, mass error 4.0) by losing methane, which further resulted an ion at m/z 250.1038 (calculated formula $C_{17}H_{13}FN^+$, exact mass 250.1027, mass error 4.4 ppm) corresponding to the loss of C_2H_2 . Fragmentation pathway described above and presented in figure no. 2 is a probable pathway; formation of fragments can be simultaneous because of multiple transitions can occur simultaneously.

4. Conclusion

The molecular ion peak as $[M + H]^+$ in MS/MS spectra of atorvastatin appeared at m/z 559.2635 Da. Further, the CID fragmentation of protonated $[M + H]^+$ ion generated and interpretation was carried followed by the proposal of fragmentation pathway (based on neutral losses, single and multiple bond cleavages). Fragmentation pathway is only a predication. Pathway can be different and there are some other probabilities also considering the occurrence of multiple transitions simultaneously. The mass error found in this study is 0.3 to 5.7 ppm, which strongly supports the fragment ions molecular structures and elemental composition. Nowadays various software tools are available for the interpretation of mass spectrometry data; during the study no software tool was used for interpretation, predication of the fragments structure and pathway of formation. In addition to above, the study also provides the insights about workflow based interpretation of parent and product ion spectra in combination with the basic rules. The workflow [12] applied in this study was found efficient and can be applied for structure verification studies of small organic molecules and for identification of similar type of drug molecules or small organic molecules and their impurities.

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Conflicts of Interest

Declared none.

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Abbreviations

LC:	Liquid chromatography;			
HPLC:	High performance liquid chromatography;			
MS:	Mass spectrometry;			
HRMS:	High resolution mass spectrometer;			
MS/MS:	Tandem mass spectrum;			
<i>m z</i> :	mass-to-charge ratio;			
API:	Atmospheric pressure ionization;			
APCI:	Atmospheric pressure chemical ionization;			
ESI:	Electrospray ionization;			
CID:	Collision induced dissociation;			
Q-TOF: Quadrupole-Time of Flight;				
IDA:	Information dependent acquisition;			
FIA:	Flow Injection Analysis			



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