



Fast and Sustainable Determination of Persistent Organic Pollutants from Organic Fertilizer Using Optimized Microwave-Assisted Extraction Method and Gas Chromatography-Mass Spectrometry

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Abstract

A microwave-assisted extraction was optimized for the evaluation of persistent organic pollutants in liquid fertilizer samples. The extraction process was defined by a 2^3 factorial design. The parameters (temperature and sample and solvent volume) were optimized through the response surface methodology (RSM). The data for the study were based on injection of a mix of organochlorine pesticides. In optimized conditions, the microwave-assisted extraction was carried out at 70°C for 10 minutes using 10 mL of sample and 10 mL of solvent. This study was accomplishment to evaluate the statistical influence of parameters: temperature, solvent volume and sample volume in the microwave-assisted extraction of organochlorine contaminants present in biofertilizer samples obtained from liquid humus and consequently to define the best conditions of process.

Keywords

Microwave-Assisted Extraction, Persistent Organic Pollutants, Organochlorine Pesticides, Biofertilizers

Subject Areas: Analytical Chemistry, Environmental Chemistry

1. Introduction

The liquid humus contains nutrients and organic matter that can bring benefits to the soil and consequently serve as an additive in agriculture, increasing the fertility of the soil or contaminating the environment if it is not con-

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trolled [1]. The production of agricultural fertilizers and industrial organic wastes provides an important means of nutrient recovery. The high content of organic matter present in these wastes contributes to the conditioning of the physicochemical properties of the soil, resulting in an increase in agricultural productivity [2].

The Stockholm Convention, together with the United Nations Environment Programme (UNEP), has defined alternatives to manage, reduce or eliminate persistent organic pollutants (POPs), recognizing that these pollutants have toxic properties. They are resistant to degradation, bioaccumulative and transported by air, water and by migratory species through international boundaries and deposited far from their place of release, where they accumulate in terrestrial and aquatic ecosystems [3].

Organochlorine pesticides (DDT, HCB, HCHs etc.) are classified as POPs and are of global concern with respect to contamination and environmental toxicity [4]. They are one of the most important environmental contaminants; when released in the environment, they may persist or resist to degradation and then remain unchanged for long periods. They are used in effective pest control; its resistance to degradation results in universal contamination of water, soil and foods [5]. They are bioaccumulative, *i.e.*, they are resistant to degradation and their wastes accumulate in animals and persist in the environment, rivers, soils, sediments and other living beings, therefore bioaccumulating up along the food chain [6].

The organochlorine pesticides (OCPs) analysis involves the extraction of these compounds from the matrix, which is difficult by the strong interaction between them and the different components of the matrix, in particular of organic matter. Conventional techniques for removing organic contaminants from complex matrices, such as water, soil and sediment, involve a lot of time in the extraction process, pre-concentration, which is a limiting factor of the entire methodology [7].

Chromatographic techniques are considered the best for the determination of OCPs in a wide variety of matrices, but the extraction and pretreatment methods should be considered before the chromatographic determinations. Conventional extraction methods offer efficiency and accurate results are methods relatively time consuming and hazardous to health due to the use of organic solvents and highly expensive when considering using solvents with high purity [6].

The microwave-assisted extraction (MAE) is a technique using polar solvents such as water, to extract target compounds in solid matrices [8]. The extraction method depends on various physical and chemical parameters, such as temperature, polarity and solvent volume of extraction, irradiation time, stirring and amount of sample in the process [9] etc.

The aim of this study was to optimize a methodology for the extraction of persistent organic pollutants, with emphasis on organochlorine pesticides in biofertilizers obtained from liquid humus using microwave-assisted extraction followed by gas chromatography-mass spectrometry determination.

2. Materials and Methods

2.1. Samples

To the optimization tests, liquid humus samples obtained from the fermentation of agricultural wastes were used. The mixture consisted of 20% wastes (4 L poultry + 4 L of orange peel) with 80% water (32 L), in volume.

2.2. Reagents and Solutions

A mix (2000 $\mu\text{g}\cdot\text{L}^{-1}$) of reference material *p,p'*-DDT, *p,p'*-DDE, *p,p'*-DDD, lindane, aldrin, heptachlor, chlordane, dieldrin and HCB (hexachlorobenzene) in hexane: toluene (1:1, in volume) was used for testing. From the reference material, there was prepared a stock solution of 200 $\mu\text{g}\cdot\text{mL}^{-1}$ in ethyl acetate. Ethyl acetate HPLC grade was supplied by Tedia. Helium (purity: 99.9990%) was supplied by White Martins.

2.3. Chromatographic System

The chromatographic system used was a GC/MS, Shimadzu, GC2010 Plus, equipped with split/splitless injector and a mass spectrometric detector, Shimadzu, MS-QP 2010 Plus. The capillary column used was a BP-5 capillary column, 30 m \times 0.25 mm i.d. and 1.0 μm film thickness (Agilent). Helium at a constant flow rate of 1 $\text{mL}\cdot\text{min}^{-1}$ was used as carrier gas. The injector and detector temperature were 280°C. Other operational conditions are described in **Table 1**. The OCPs were quantified by analytical curve constructed with 6 concentration levels in the range of 10 - 100 $\text{mg}\cdot\text{L}^{-1}$ for each reference material.

Table 1. Gas chromatograph conditions.

Injection	Splitless (1 min)
Injector temperature	250°C
Column temperature	80°C
Flow	He, 1.0 mL·min ⁻¹
Oven temperature program	50°C·min ⁻¹ up to 100°C + 15°C·min ⁻¹ up to 230°C (3:00 min) + 5°C·min ⁻¹ up to 270°C

The spectrometer was operated in full scan mode for determination of retention times of the compounds and in the SIM mode to analyze the samples. The operating conditions of the mass spectrometer are shown in **Table 2**.

2.4. Microwave-Assisted Extraction

The extraction procedure was performed in a microwave digestion system (SPEEDWAVE FOUR[®], with Built-in Non-Contact, Temperature and Pressure Measurement, BERGHOF, Germany) with a capacity of 12 vessels of 60 mL and 1450 W maximum power. To the optimization tests, they were used liquid humus samples obtained from the fermentation of agricultural wastes (20:80, in volume) with the extraction solvent (ethyl acetate). The extraction temperature ranged from 70°C to 110°C, sample and solvent volumes ranged from 5 to 10 mL, the irradiation times (10 min), ramp (5 min) and power (80% - 1160 W) were fixed based on previous experiments.

After the extraction step, the vials were cooled until room temperature and then opened. Then, the material was filtered in 45 µm filter paper, concentrated under vacuum in rotary evaporator (FIZATOM) at 80°C until dryness, reconstituted with 1 mL of ethyl acetate and placed in a vial for the chromatographic determination. For tests with blanks, the above procedures were followed using the sample without addition of the reference material.

2.5. Experimental Design and Optimization of Microwave-Assisted Extraction

The extraction conditions of POPs evaluated in liquid humus samples were optimized based on the analysis of response surface methodology (RSM) of the investigated factors. In this study, the effects of three factors on the response were evaluated using a 2³ factorial design, 16 experiments in duplicate were randomly performed according to the experimental design. The area for each compound was chosen as response-variable in the optimization process.

A two-way interaction linear model was adjusted to the experimental data based on Equation (1):

$$\hat{y} = \beta_0 + \beta_1x_1 + \beta_2x_2 + \beta_3x_3 + \beta_{12}x_1x_2 + \beta_{13}x_1x_3 + \beta_{23}x_2x_3 \quad (1)$$

where \hat{y} is the predicted value (area for each compound), β 's are the model coefficients, and x_1, x_2, x_3 the encoded factors: temperature (x_1 , 70°C - 110°C), sample volume (x_2 , 5 - 10 mL) and solvent volume (x_3 , 5 - 10 mL). The ranges used to evaluate the independent variables were selected based on preliminary experiments.

3. Results and Discussion

3.1. Extraction

Previous experiments in our laboratory, to extract the compound *p,p'*-DDT in sewage sludge samples evaluated the significance of the parameters: temperature, extraction time, and percentage of acetone in the solvent mixture (ethyl acetate-acetone). Based on this, new parameters were selected for the present study. Then, the solvent chosen to extraction was ethyl acetate and the extraction time for all tests was 10 minutes.

The use of microwave energy in the extraction process generates heat of solvent and consequently of the sample which can suffer degradation or conversion of some compounds by different ways, such as exposure to high temperatures and pressure inside the microwave extraction vessel [5]. To achieve the best extraction condi-

Table 2. Operating conditions of mass-spectrometer.

Ion source	220°C	
Interface temperature	250°C	
Operation mode	SIM	
Ionization mode	EL	
OCPs	Retention time (min)	m/z
Hexachlorobenzene	8.83	284, 286
Lindane	9.32	281, 219
Aldrin	11.85	263, 261
Heptachlor	12.98	272, 274, 183, 353
Chlordane	13.77	373, 375
Cis-Chlordane	14.33	373, 375
<i>p,p'</i> -DDE	15.14	
Dieldrin	15.28	79, 277
<i>o,p'</i> -DDD	15.48	
<i>p,p'</i> -DDD	16.05	235, 165, 235, 165
<i>p,p'</i> -DDT	17.18	

tions, a factorial design was used. The actual and encoded levels of the independent variables (temperature, sample volume and solvent volume) for each experiment are shown in **Table 3**.

Table 4 shows the significance of the extraction process. Bold numbers indicates significant factors as identified by the analysis of variance at the 95% confidence level.

From the calculated results it can be seen that the factor 3 (solvent volume) was the most important for the majority of the investigated analytes (strongly positive) except for the compounds heptachlor and *o,p'*-DDD. Only to three compounds, the factor 2 (sample volume) influenced the extraction process, and the major significance of this factor is in the compound hexachlorobenzene. The factor 1 (temperature) was significant for most compounds, but the interaction of this one with the other two factors, *i.e.*, the factor interactions (β_{12} and β_{13}) had low significance in the process, and the factor interactions β_{12} (temperature x sample volume) has significance in five compounds. The factor interactions β_{13} (temperature x solvent volume) was not significant in the extraction process. The interaction between sample volume x solvent volume (β_{23}) is what most influences the process, eight of eleven evaluated compounds were significantly influenced. All calculated regression coefficients (R^2) are in the range 0.99, indicating a good correlation to quantify and interpret the relationship between the experimental data and the factor effects studied. Equation (2) describes an example of the proposed model for the compound Aldrin.

$$\hat{y} = 42.526 + 123.88x_1 + 16.89x_2 + 69.68x_3 + 36.94x_1x_2 - 0.09x_1x_3 + 158.26x_2x_3 \quad (2)$$

3.2. Response Surface Plot Analysis

Response surface graphics are very useful to estimate and to identify the optimal ranges for the two or more factors on the response [10]. In order to determine the optimum ranges of the factors, graphs were plotted using the z-axis (response) against two of the independent variables (temperature, sample volume and solvent volume), maintaining a third one at a fixed value. **Figures 1-3** show an example of the main interactions in the process for the compound Aldrin. Similar plots but not identical were obtained for all the other investigated compounds.

Figure 1 shows the interaction between temperature and the sample volume in response. Increasing the temperature from 70°C to 110°C and increasing the sample volume from 5 to 10 mL, the response value increase. So, best responses are achieved when using 10 mL of sample at a temperature of 110°C, using 10 mL of the extrac-

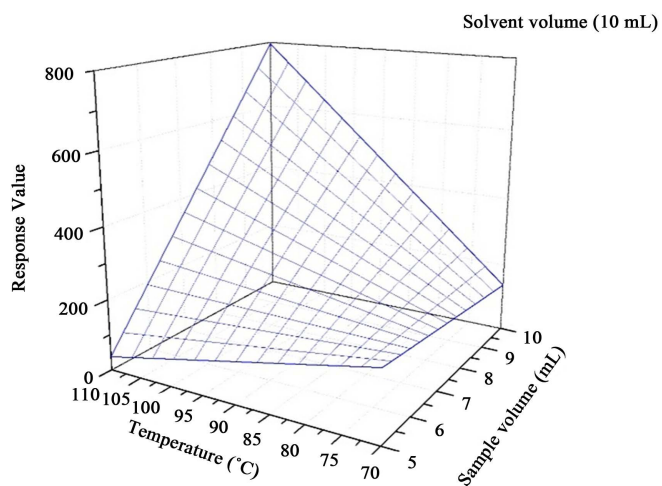


Figure 1. Response surface. Temperature x sample volume.

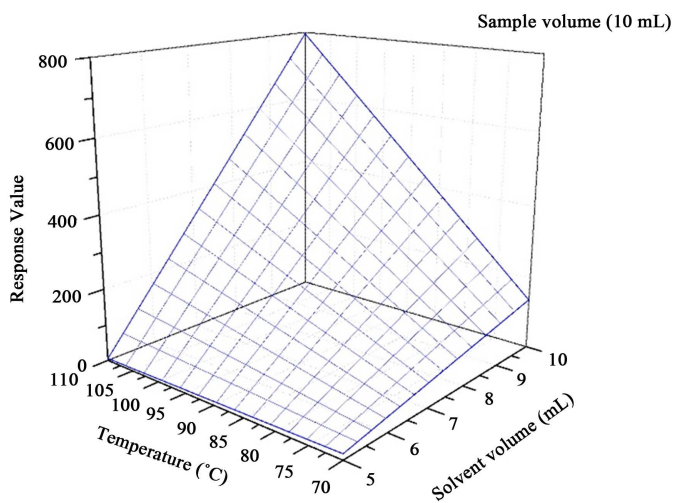


Figure 2. Response surface. Temperature x solvent volume.

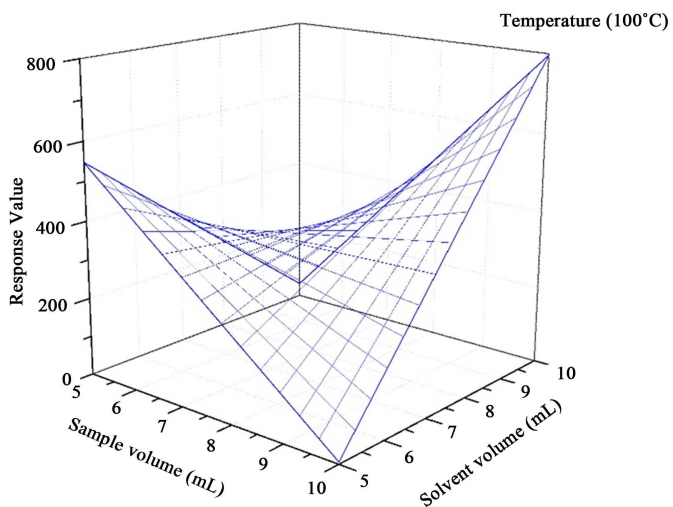


Figure 3. Response surface. Sample volume x solvent volume.

Table 3. Experimental design 2³: actual and encoded levels of the factors.

Experiment	Temperature (°C)	Sample volume (mL)	Solvent volume (mL)
1	70 (-)	5 (-)	5 (-)
2	110 (+)	5 (-)	5 (-)
3	70 (-)	10 (+)	5 (-)
4	110 (+)	10 (+)	5 (-)
5	70 (-)	5 (-)	10 (+)
6	110 (+)	5 (-)	10 (+)
7	70 (-)	10 (+)	10 (+)
8	110 (+)	10 (+)	10 (+)

Table 4. Significance levels of parameters (effects) identified by analysis of variance (ANOVA).

	β_0	β_1	β_2	B_3	B_{12}	β_{13}	B_2	R ²
Hexachlorobenzene	1425.17	-573.39	531.77	1246.99	-491.52	-1256.93	1469.45	0.9996
Lindane	1381.15	-548.96	-1321.4	1820.26	1819.37	-1198.85	-564.00	0.9982
Aldrin	220.96	247.77	33.78	139.36	73.88	-0.1875	316.53	0.9993
Heptachlor	235.23	341.03	-384.0	-351.67	-383.10	-410.48	352.37	0.9997
Chlordane	2356.24	1303.89	-400.54	448.22	-1987.02	-2684.86	3720.20	0.9981
Cis-Chlordane	3550.14	3571.33	-1612.29	186.06	-1494.62	-876.18	5063.64	0.9993
<i>p,p'</i>-DDE	3472.88	1504.30	-1877.10	1595.13	1748.45	-1941.36	1748.54	0.9957
Dieldrin	2211.94	-1642.55	-182.47	864.22	1378.79	-862.27	-1808.21	0.9971
<i>o,p'</i>-DDD	926.99	493.09	-146.09	-659.52	-585.63	-578.72	403.22	0.9991
<i>p,p'</i>-DDD	1172.76	-1199.08	-873.61	720.34	961.31	-1235.10	-1373.56	0.9999
<i>p,p'</i>-DDT	85.33	-51.65	35.04	40.36	-48.53	-28.73	56.83	0.9997

tion solvent. **Figure 2** shows the interaction between solvent volume with the extraction temperature in response. Increasing the solvent volume from 5 to 10 mL and increasing the temperature from 70°C to 110°C the response value increase. Extraction temperature at 110°C combined with a solvent volume of 10 mL provides a better response. **Figure 3** shows that best responses are achieved when the maximum sample volume is equal to the maximum solvent volume. The combination of 10 mL of sample volume, 10 mL of solvent volume (ethyl acetate) at 110°C were considered to provide the best conditions.

Considering the extraction temperature significance in the process, *i.e.*, higher the temperature, higher the yield of the extraction, the chosen temperature was 110°C. Elevated levels of temperature (above 110°C) would increase the internal pressure inside extraction vessels, which could result in loss of analyte. Thus, the maximum levels of factors sample volume and solvent volume, related to recovery levels, achieved the final extraction conditions were defined, without the need of additional experiments.

3.3. Recovery Assay: Yield of Extraction

After defining the best extraction conditions, the recovery of the compounds were evaluated. The recovery assays experiments were performed by adding 125 μL of stock solution of 200 $\mu\text{g}\cdot\text{mL}^{-1}$ to the extraction vessel, the final volume was 20 mL. Thus, inside of vessels, the concentration of POPs was 1.25 $\mu\text{g}\cdot\text{L}^{-1}$. After the extraction procedure, the sample was determined according to the procedure previously described.

Table 5 shows the linear regression, coefficient of determination, average recovery and relative standard deviation of the OCPs studied.

Table 5. Linear regression, coefficient of determination, recoveries and relative standard deviation of the compounds studied.

Compound	Calibration	R ²	Average recovery* (%)	R.S.D* (%)
Hexachlorobenzene	y = 103.1531x + 3335.219	0.9930	90.66 ± 1.79	1.82
Lindane	y = 32.2791x + 1345.398	0.9876	93.07 ± 0.69	1.19
Aldrin	y = 75.41816x + 2372.263	0.9904	91.26 ± 1.07	1.17
Heptachlor	y = 79.14032x + 3423.263	0.9903	105.68 ± 3.25	3.08
Clordane	y = 103.1175x + 2074.485	0.9935	95.40 ± 2.32	2.43
Cis-Clordane	y = 84.81596x + 929.8195	0.9897	93.67 ± 0.90	1.68
<i>p,p'</i> -DDE	y = 86.3497x + 1877.259	0.9931	97.34 ± 1.80	1.83
Dieldrin	y = 71.61183x + 1392.62	0.9943	87.98 ± 1.02	1.45
<i>o,p'</i> -DDD	y = 58.34518x + 8971.62	0.9735	97 ± 2.56	2.27
<i>p,p'</i> -DDD	y = 91.5611x + 4521.11	0.9832	88.1 ± 0.82	1.47
<i>p,p'</i> -DDT	y = 42.1294x - 1180.484	0.9805	100.30 ± 0.81	0.81

*Means ± standard deviation (duplicate) and relative standard deviation (R.S.D).

In the analysis of blank chromatograms, no peaks related to OCPs were observed. The recoveries values (87.98% - 105.68%) showed that the methodology was suitable for determination of organochlorine pesticide in liquid humus samples.

4. Conclusions

Microwave-assisted extraction of organochlorine pesticides from liquid fertilizer samples was satisfactory, considering the recovery percentages obtained (87% to 105%) in the determination of 11 organochlorine pesticides. As could be shown, the extraction solvent and the factor interactions between sample volume and solvent volume were the most important and significant during all the experiments. In general, the factor sample volume was not significant and the factor temperature showed a positive effect on the extraction. After a careful analysis of data and combination of them, considering the ranges defined in the experimental design, the optimal extraction conditions were: 10 mL of sample, 10 mL of solvent (ethyl acetate) and a temperature of 110°C.

The production of liquid humus from organic waste increases the aggregated value of this waste, being a good alternative for soil organic matter replacement. However, the quality of the produced organic matter must be attested. Only then, its use becomes environmentally safe.

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