

The role of catalysis on the formation of an active proto-enzyme in the prebiotic aqueous environment

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ABSTRACT

A model detailing the conditions for the formation of active enzymes from amino acid monomers in the pre-biotic aqueous environment is proposed, making use of only a minimal number of parameters: the intensity of the amino acid source and the experimental rate coefficient for polypeptide hydrolysis. In a closed system with fixed total amino acid concentration, the formation of polypeptides of significant length is precluded by hydrolysis. In the presence of an amino acid source, the behavior of the steady state solutions is dependent on the efficiency of the catalytic activity in the environment. In absence of catalysis the formation of long amino acid chains shows a remarkable similarity to the equilibrium problem, *i.e.* it is equally unlikely that an active protein forms. In the presence of catalysis the concentration of polypeptides of significant length increases dramatically to about the same order of magnitude as the steady state concentration of the monomers. Besides the important notion of the proto-enzyme concentration, the model offers a link between the given environmental conditions (flux of monomers, rates of the peptide bond formation and hydrolysis, size of the amino acid set) and properties of the polypeptide relevant to its catalytic properties, such as its length and conserved number of residues.

Keywords: Prebiotic Chemistry; Enzymes

1. INTRODUCTION

This work aims to discuss, on the basis of known chemistry, if the prebiotic conditions on Earth could allow for the spontaneous formation of catalytic polypeptides without any prior complex RNA-based chemistry

that could code for enzymes with specific amino acid sequences. After the formation of proto-enzymes we may envision the development of a more complex chemistry until a kinetic cycle of reactions involving self-replicating species was reached. We assume a source of amino acids sustained by processes driven by an energy source acting on simple organic compounds and focus on the formation of peptide bonds among amino acid units until the critical length that allows for catalytic properties is attained. It will be shown that the length of the polypeptide depends on how many types of monomeric units are involved in the formation of the chain. There are a number of simulations of the processes taking place in interstellar ices showing how many amino acids may form from simple organic molecules. For example Muñoz Caro *et al.* [1] have shown the formation of 16 amino acids by UV irradiation of ices with components H₂O, CO, CO₂, CH₃OH and NH₃. Maurette [2] suggested that micrometeorites could have acted as chemical reactors forming amino acids from organic precursors upon contact with water and warm temperatures following re-entry. However, given the degree of uncertainty of the actual processes that have formed primordial amino acids, we choose to keep the number of amino acids as a variable and regard direct sources of amino acids as equivalent to sources of their precursors. Since it has been shown that a hydrosphere was interacting with the Earth's crust as early as 4.3 Gyr ago [3], we may also safely assume that the polymerization process took place in the terrestrial aqueous environment, with or without the participation of solid surfaces promoting the peptide bond formation [4,5]. A similar approach was undertaken by Chen and Nowak [6] that investigated the equilibrium configuration of RNA polymers formed from activated monomers and exhibiting self-replication and selection.

It has been well established that micrometeorites, interplanetary dust particles (IDP), and carbonaceous chondrites presently deliver amino acids to the Earth [7]. Larger bodies also exhibit significant inbound fluxes, but

the heating experienced during the atmospheric entry and impact phases is likely to pyrolyze their organic content. The flux of IDPs has been estimated around 3.0×10^7 kg-yr⁻¹ [8] and the content of amino acids in polar micrometeorites has been reported to be 180 - 249 ppm [9] and up to 869 nmol/g [8]. Micrometeorites could thus deliver 2.61×10^4 mol-yr⁻¹ of amino acids, a figure that must be multiplied by a factor of 10^2 to 10^7 based on the lunar cratering record of 3.8 to 4.5 Gyr ago [10]. Wilson [11] estimates a flux of 3.6×10^{10} kg-yr⁻¹ of organic carbon in IDPs that were subjected to mild temperatures and survived re-entry on the early Earth. The oceanic amino acid concentration from electrical discharge and cometary input 4 Gyr ago was estimated by Pierazzo and Chyba [12] to be 2.4×10^{-9} M and 1.5×10^{-11} M for glycine and aspartic acid, respectively.

The extent of the amino acid source is both critical to the formation of long polypeptide chains and to their concentration. This investigation begins by determining the (small) concentration of polypeptides of given lengths in equilibrium conditions, *i.e.* in the absence of amino acid sources. It will proceed to examine the effect of a source and catalysis on the dynamics of the state variables (concentrations of all the polymers of different lengths), determining also the time scale for achieving steady-state conditions. The maximum length of the polypeptides at steady-state will also be determined, a key factor strictly related to their potential catalytic capabilities.

The concentration of the amino acid monomers will be expressed in terms of the intensity of the source or, equivalently, of the incoming flux. Having determined the connection between the concentration of polypeptides of a given length and the steady-state concentration of monomers, we will be able to express the lengths of the polypeptides in terms of the source intensity, the size of the amino acid set, and the parameters governing the reactivity of the peptide bond, *i.e.* bond formation and hydrolysis.

2. RESULTS AND DISCUSSION

An evolutionary process able to build an active enzyme with n amino acid residues from its monomeric components must have produced nearly all the possible polypeptides of length n . It has been determined experimentally that, in order to retain the catalytic activity of an enzyme, a fraction α between 0.1 and 0.2 of its amino acid sequence is constrained [13]. Within this requirement, the number of possible proteins with n amino acid residues chosen from a set of p amino acids is $p^{\alpha n}$. The ratio between the oceanic content of all the polypeptides of length n and the total number of possible structures of the same length is

$$f = x_n V N_A p^{-\alpha n}, \quad (1)$$

where N_A is the Avogadro number and V is the volume of water on the planetary body hosting the prebiotic chemistry. In the assumption that a catalytic protein did form in the prebiotic environment, we must have $f \approx 1$ and **Eq.1** constrains the value of α once we determine x_n .

We begin considering a solution of total content c of amino acid monomers that reaches equilibrium with formation of dipeptides of concentration x_2 , tripeptides of concentration x_3 , and so forth, with x_i indicating the concentration of the polypeptide with i components. The equilibrium constant K is assumed to be independent of i . We have thus

$$K = \frac{x_2}{x_1^2} = \frac{x_3}{x_2 x_1} = \frac{x_i}{x_{i-1} x_1}, \quad (2)$$

that iteratively gives

$$x_i = (K x_1)^{i-1} x_1. \quad (3)$$

The mass balance equation links the concentration x_i to the total amino acid content and the equilibrium constant

$$c = \sum_{i=1}^n i x_i = x_1 \sum_{i=1}^n i (K x_1)^{i-1}. \quad (4)$$

Since the reactants and products have nearly the same free energy [5], we have

$$K' = \frac{\eta}{\eta_h} = \frac{x_2 x_w}{x_1^2} \cong 1, \quad (5)$$

with η being the bimolecular rate coefficient for peptide bond formation and η_h the rate coefficient for hydrolysis. The concentration of water $x_w = 55.55$ M is usually included in the equilibrium constant obtaining $K = K'/x_w \cong x_w^{-1}$, which makes $K x_1$ in **Eq.4** very small (we are considering dilute solutions) and gives, to a very good approximation, $x_1 \cong c$, making the concentration of all the polypeptides given by **Eq.3** virtually negligible

$$x_i = (c/x_w)^{i-1} c. \quad (6)$$

It is thus necessary, to increase the concentration of polypeptides of significant lengths, that the system be endowed with a source of amino acids. This source can be thought of as either external (meteoritic and cometary impacts, IDPs) or internal to the biosphere. High temperatures in hydrothermal vents [14] and electrical discharge in the atmosphere [16] are thought to be viable spontaneous processes that were able to form amino acids from simple compounds and to have provided 10^{10} kg-yr⁻¹ of organics [16] and 10^7 kg-yr⁻¹ of amino acids. In the hypothesis that the system evolved in the presence of a source with the ability to keep the concentration of

monomers x_1 constant, the rate of spontaneous formation of a dipeptide between two aminoacid monomers of molar concentration x_1 is

$$\dot{x}_2 = \eta x_1^2 - x_2 (\eta x_1 + \eta_h x_w) \quad (7)$$

(the formation of x_2 by hydrolysis of x_3 is being neglected). The quantity $\omega_h = \eta_h x_w$ is a constant because of the large excess of water, and its experimental value for the hydrolysis of glycyl-glycine is $6.3 \times 10^{-11} \text{ s}^{-1}$ [17]. It is useful to define the proper frequency of the reactive system as

$$\omega = \eta x_1 + \eta_h x_w, \quad (8)$$

and the ratio ζ of the rates of peptide formation and the overall reaction (peptide formation plus hydrolysis)

$$\zeta = \frac{\omega - \omega_h}{\omega}. \quad (9)$$

Consequently, the ratio between the rates of the catalyzed and uncatalyzed reactions is $(\omega - \omega_h)/\omega_h = \zeta/(1 - \zeta)$. The values taken by the parameter ζ describe all the possible relative rates; for example, $\zeta = 0.5$ means equal rates for peptide bond formation and hydrolysis, while $\zeta = 0.99$ represents a ratio of the rates of 99. With the previous definitions, the solution of Eq.7 is

$$x_2 = \zeta x_1 (1 - e^{-\omega t}). \quad (10)$$

For the formation of peptides with three residues we thus have

$$\dot{x}_3 = \eta x_1 x_2 - \omega x_3 = \eta \zeta x_1^2 (1 - e^{-\omega t}) - \omega x_3 \quad (11)$$

with solution

$$x_3 = \zeta^2 x_1 [1 - e^{-\omega t} (1 + \omega t)]. \quad (12)$$

This procedure can be iterated and one obtains for the formation of the peptide with n residues the equation

$$\dot{x}_n = \eta x_1 x_{n-1} - \omega x_n, \quad (13)$$

with solution

$$\begin{aligned} x_n &= \zeta^{n-1} x_1 \left[1 - e^{-\omega t} \left(1 + \omega t + \frac{1}{2} \omega^2 t^2 + \dots + \frac{1}{(n-2)!} \omega^{n-2} t^{n-2} \right) \right] \\ &= \zeta^{n-1} x_1 \left[1 - e^{-\omega t} \sum_{i=0}^{n-2} \frac{(\omega t)^i}{i!} \right] \end{aligned} \quad (14)$$

Given the definition of $Q(n-1, \omega t)$ as the ratio between the incomplete and complete gamma function of Euler

$$Q(n-1, \omega t) = \frac{\Gamma(n-1, \omega t)}{\Gamma(n-1)}, \quad (15)$$

we may more conveniently express x_n as

$$x_n = \zeta^{n-1} x_1 [1 - Q(n-1, \omega t)]. \quad (16)$$

The inflection point of Q located at $\omega t = n-2$ separates the early from the late stages of polypeptide formation, the latter being close to the stationary state. We now define the number of molecules of the monomer at stationary state $N = x_1 N_A V$ and equate the r.h.s of Eq.16 to x_n from Eq.1, obtaining the evolution equation for n

$$(\zeta p^{-\alpha})^n [1 - Q(n-1, \omega t)] = \frac{\zeta}{N}. \quad (17)$$

Numerical solutions of Eq.17 are shown in Figure 1 for two values of ζ . The curve with $\zeta = 0.5$, corresponding to the already fairly large ratio $\eta/\eta_h = x_w/x_1$, affords polypeptides with lengths limited to 80 amino acid units. The curve defined by $\zeta = 0.99$ shows that the catalytic efficiency necessary to attain polypeptide lengths of 241 units implies $\omega_h/\omega \ll 1$, that is $\eta_c/\eta_h \gg x_w/x_1$.

The elapsed proper time before steady-state is also dependent on the ability of the environment to catalyze peptide bond formation. In fact, since in the absence of catalysis the ratio of the rate coefficients for peptide formation and hydrolysis is about unity (the reaction is nearly thermoneutral), and we may reasonably assume that the concentration of amino acids is much lower than that of water ($x_1 \ll x_w$), we have $\omega \approx \omega_h$ and

$$\zeta \approx \frac{1}{1 + \frac{x_w}{x_1}} \approx \frac{x_1}{x_w}. \quad (18)$$

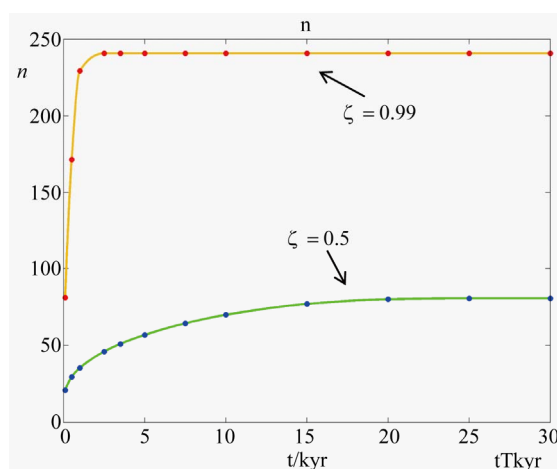


Figure 1. Effect of catalysis on the solutions of Eq.17 ($\zeta = 0.5$, blue points and $\zeta = 0.99$, red points), giving the maximum length of a spontaneously formed polypeptide in the early times of molecular evolution for $\omega = \omega_h/(1 - \zeta)$, $p = 20$, and $\alpha = 0.11$.

On the contrary, in the presence of significant catalysis ($\zeta \approx 1$), the proper frequency is dominated by the polymerization process.

We may now estimate that the uncatalyzed reacting system may attain steady state in the proper time $t_h = (n - 2)\omega_h^{-1} = 3.14 \times 10^{12} \text{ s } (9.96 \times 10^4 \text{ yr})$, while the corresponding time for the catalyzed system would be lowered to $t_c = (1 - \zeta)t_h$, showing that in both cases a steady state concentration of x_n is reached in the very early stages of the chemical evolution. These proper times may be compared to the much longer time (8 - 10 Myr) necessary for the volume of the sea to circulate through the hot areas of the intrusion zones in the sea floor [18, 19] and constituting a sink for the protoenzymes.

We now give two convenient approximate expressions of **Eq.16** valid for small and large t , respectively. For $\omega t \ll n - 2$, we approximate $1 - Q$ as

$$1 - Q(n - 1, \omega t) = 1 - e^{-\omega t} \sum_{i=0}^{n-2} \frac{(\omega t)^i}{i!} \tag{19}$$

$$\cong 1 - (1 - \omega t)(1 + \omega t) = (\omega t)^2$$

and, for $\omega t \gg n - 2$,

$$1 - Q(n - 1, \omega t) \cong 1, \tag{20}$$

that give the following approximation of **Eq.16**

$$x_n = \zeta^{n-1} x_1 \Omega, \tag{21}$$

where $\Omega = (\omega t)^2$ and $\Omega = 1$ for $\omega t \ll n - 2$ and $\omega t \gg n - 2$, respectively. For these two cases, **Eq.17** has the closed-form solution

$$n = \frac{\ln(\zeta / N\Omega)}{\ln(\zeta p^{-\alpha})}. \tag{22}$$

The plots of **Eq.22** given in **Figure 2** for two typical values of the parameter α show that a catalytic protein of significant length could only be formed in a catalyzed medium with $\zeta \approx 1$ and attest that in the absence of catalysis the length of the polypeptides is limited to $n = 4$. Thus the catalytic capability of the spontaneously formed proto-enzymes, directly dependent on their size, strictly depends on the ratio of the rate coefficients for the peptide bond formation and hydrolysis, expressed through the parameter ζ .

Also, the steady state concentrations ($\Omega = 1$) of the polypeptides of length n differ markedly in the uncatalyzed system, where ζ is given by **Eq.18**, and the catalyzed system, where ζ approaches unity. In the uncatalyzed system the solution given by **Eq.21** is very similar to the corresponding solution for the equilibrium problem **Eq.6**, while the effect of catalysis dramatically increases the concentration of long polypeptides by the large factor $(\zeta x_w / x_1)^{n-1}$. Thus high values of ζ in-

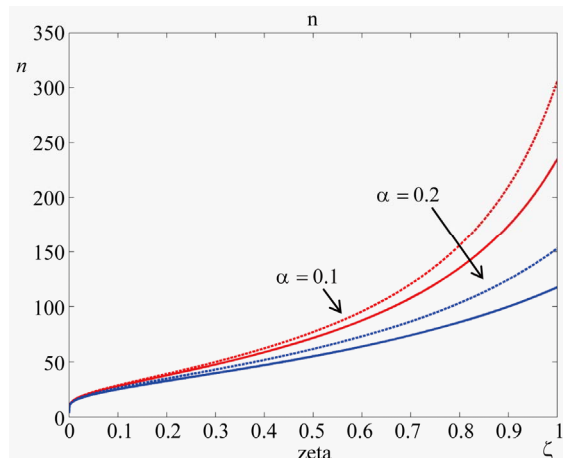


Figure 2. Effect of the catalytic efficiency ζ on the number n of amino acid units constituting an enzyme at steady state according to **Eq.22** for two typical values of the parameter α and the present-day volume of the sea. The effect of different values of the number of available amino acids is also shown: $p = 20$ (solid lines) and $p = 10$ (dashed lines).

crease both the length and the concentration of long polypeptides.

We now turn our attention to determine how the steady state concentration of the amino acid monomers changes with the source. Given a source s of amino acids, the evolution of the concentration x_1 is given by

$$\dot{x}_1 = -\eta x_1^2 - \eta x_1 x_2 - \eta x_1 x_3 - \dots + \eta_h x_w x_2 + \eta_h x_w x_3 + \dots + s. \tag{23}$$

The steady state condition is

$$s = \eta x_1 (x_1 + x_2 + x_3 + \dots) - \eta_h x_w (x_2 + x_3 + \dots) = \eta x_1 \sum_{i=1}^n x_i - \eta_h x_w \sum_{i=2}^n x_i = \eta x_1 \sum_{i=1}^n x_i - \eta_h x_w \sum_{i=1}^n x_i + \eta_h x_w x_1 = (\eta x_1 - \eta_h x_w) \sum_{i=1}^n x_i + \eta_h x_w x_1. \tag{24}$$

For $\omega t \gg n - 2$, **Eq.21** holds with $\Omega = 1$, and we have

$$s = x_1 (\eta x_1 - \eta_h x_w) \sum_{i=1}^n \zeta^{i-1} + \eta_h x_w x_1. \tag{25}$$

We now define $j = i - 1$ and obtain

$$\sum_{i=1}^n \zeta^{i-1} = \sum_{j=0}^{n-1} \zeta^j \cong \sum_{j=0}^{\infty} \zeta^j = \frac{1}{1 - \zeta}, \tag{26}$$

the approximation being valid for large n . Using this approximation and **Eq.9**, **Eq.25** becomes

$$s = \omega_h x_1 \left(\frac{\zeta}{1-\zeta} \right)^2. \quad (27)$$

The flux φ of amino acids on the surface of the early Earth of radius R and the corresponding source are related by

$$sVN_A = 4\pi R^2 \varphi. \quad (28)$$

Thomas [20] estimates the infall rate of organic carbon from interplanetary dust particles between 10^8 and 10^{10} kg·yr⁻¹, with an amino acid content around 3%. Assuming an average content of 10 carbon atoms per amino acid, the estimate made by Thomas translates into an infalling rate between 4.77×10^{23} and 4.77×10^{25} s⁻¹ of amino acids. For the purpose of the following calculations we will make use of the average between these two values and the present-day volume of the sea (today the Earth has a water content of 1.4×10^{21} kg). This choice corresponds to the estimate of Pasek and Lauretta [21] of 10^{-10} M·yr⁻¹ for the flux of carbon during the late heavy bombardment. With this estimate, **Eq.27** gives for the catalyzed system with $\zeta = 0.99$ a concentration of the amino acid monomers of 9.16×10^{-15} M, and we obtain for the stationary-state number of molecules of the monomer $N = x_1 N_A V = 7.72 \times 10^{30}$. **Eq.21** in turns gives $x_{200} = 1.24 \times 10^{-15}$ M.

We now have all the elements that link the fundamental quantities that play a role in the formation of a primordial catalyst. **Eq.1** relates the concentration of a catalytic peptide of length n , the fraction α of conserved residues, and the size of the amino acid set p . **Eqs.21** and **27** connect x_n to the catalytic efficiency ζ for the formation of the peptide bond, the frequency of hydrolysis ω_h , and the source of amino acid monomers. Thus we obtain

$$\left(\zeta p^{-\alpha} \right)^n \frac{(1-\zeta)^2}{\zeta^3} = \frac{\omega_h}{4\pi R^2 \varphi}, \quad (29)$$

that combine the variables ζ, p, α , and n . The significance of **Eq.29** may be discerned by displaying the functional dependence between two quantities while keeping the third constant and regarding the fourth as a parameter. For example, solving **Eq.29** for α versus the polypeptide length n we obtain the plots in **Figure 3**, showing the effect of the catalytic efficiency on the maximum attainable fraction of constrained amino acid units. For $\zeta = 0.5$ **Eq.29** has solutions with positive α only for $n \leq 117$, a fact that we already determined from the results in **Figure 2**. Significant values for both n and α are only attained with $\zeta \geq 0.8$, but, on the other hand, there is a saturation effect around $\zeta \approx 0.99$ that makes very efficient catalysts unnecessary. Conversely, setting $\alpha = 0.11$ and $\zeta = 0.99$, we can calculate the effect of the size of the amino acid set on the attainable length of the sponta-

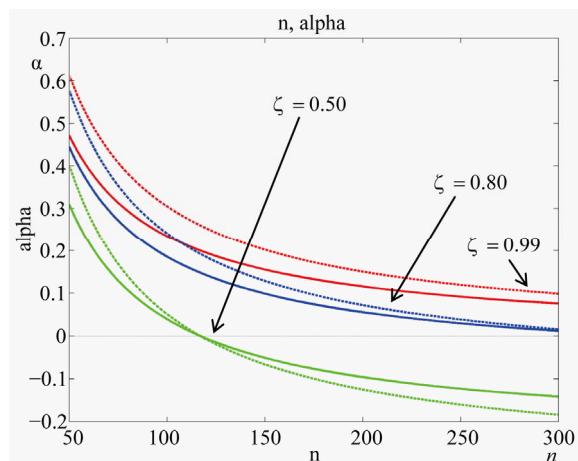


Figure 3. Plots of the attainable fraction α of constrained amino acid units versus the polypeptide length n according to **Eq.29** with $p = 20$ (solid lines) and $p = 10$ (dashed lines) for various catalytic efficiencies: $\zeta = 0.50$ (green), $\zeta = 0.80$ (blue), and $\zeta = 0.99$ (red).

neously forming catalytic polypeptides. The curve in **Figure 4** shows that a set of amino acids of limited size affords considerably longer polypeptides with respect to chains built from a fairly large set of amino acids. Since the catalytic efficiency of an enzyme requires a large structure, the descending curve of **Figure 4** is an indication that the primitive set of amino acids was probably small. The first catalytic polypeptides could have promoted the formation of both a wider range of amino acids and at the same time facilitated peptide bond formation, leading to a self-sustaining cycle in the chemical evolution.

4. CONCLUSIONS

1) The composition of a diluted water mixture of amino acids in equilibrium with the corresponding polypeptides was computed with respect to the polypeptide length. In the case of a system with an initial concentration c of monomers, the equilibrium concentration of a polypeptide of length n is negligibly small, of the order of $(c/x_w)^{n-1} c$.

2) The equation for the time evolution of a system with a source of amino acids was solved both in the uncatalyzed and catalyzed scenarios, affording the steady state concentration of the polypeptides of length n as $x_n = \zeta^{n-1} x_1$. The different values of the parameter ζ in the two cases ($\zeta = x_1/x_w$ for the uncatalyzed case and $\zeta \approx 1$ for the catalyzed case) regulate the concentration.

3) The length of the polypeptides at steady state is also strongly dependent on the value of the parameter ζ . In the absence of catalysis the maximum length is restricted to a few units, while for $\zeta \approx 1$, n may reach a few hundred. The model is able to assess the magnitude of the catalytic efficiency required for the formation of a poly-

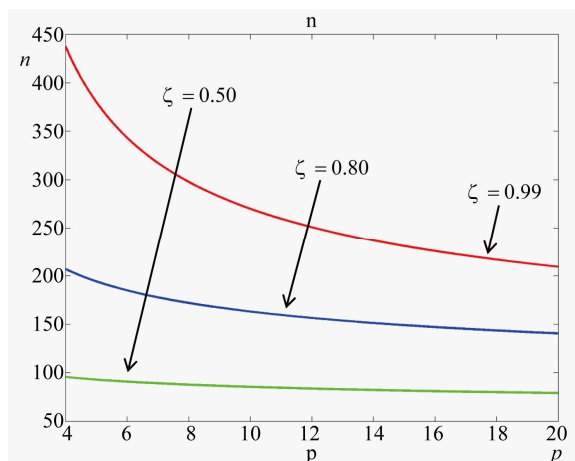


Figure 4. Plots of the number n of amino acid units in an enzyme formed at steady state according to Eq.29 versus the size of the amino acid set for various catalytic efficiencies: $\zeta = 0.50$ (green), $\zeta = 0.80$ (blue), and $\zeta = 0.99$ (red) ($\alpha = 0.11$).

peptide of a given length.

4) From the intensity of the incoming flux of amino acids on the early Earth from interplanetary dust particles, the constant steady state concentration of amino acid monomers is estimated to be 9.2×10^{-15} M for the catalyzed scenario. The concentration of a polypeptide of 200 amino acid units is predicted to be only about one order of magnitude less than the steady state concentration of monomers.

5) The effect of the catalytic efficiency on the maximum attainable fraction of conserved residues is estimated to fade around $\zeta \approx 0.99$. The maximum attainable length of the polypeptides is a decreasing function of the size of the amino acid set.

REFERENCES

- [1] Muñoz Caro, G.M., Meierhenrich, U.J., Schutte, W.A., Barbier, B., Arcones Segovia, A., Rosenbauer, H., Thiemann, W.H.-P., Brack, A. and Greenberg, J.M. (2002) Amino acids from ultraviolet irradiation of interstellar ice analogues. *Nature*, **416**, 403-406. [doi:10.1038/416403a](https://doi.org/10.1038/416403a)
- [2] Maurette, M., Duprat, J., Engranda, C., Gounellea, M., Kurat, G., Matrajta, G. and Toppanid, A. (2000) Accretion of neon, organics, CO₂, nitrogen and water from large interplanetary dust particles on the early Earth. *Planetary and Space Science*, **48**, 1117-1137. [doi:10.1016/S0032-0633\(00\)00086-6](https://doi.org/10.1016/S0032-0633(00)00086-6)
- [3] Mojzsis, S.J., Harrison, T.M. and Pidgeon, R.T. (2001) Oxygen-isotope evidence from ancient zircons for liquid water at the Earth's surface 4,300 Myr ago. *Nature*, **409**, 178-181. [doi:10.1038/35051557](https://doi.org/10.1038/35051557)
- [4] Bujdák, J. and Rode, B.M. (1996) The effect of smectite composition on the catalysis of peptide bond formation. *Journal of Molecular Evolution*, **43**, 326-333. [doi:10.1007/BF02339007](https://doi.org/10.1007/BF02339007)
- [5] Rimola, A., Sodupe, M. and Ugliengo, P. (2007) Aluminosilicate surfaces as promoters for peptide bond formation: An assessment of Bernal's hypothesis by *ab initio* methods. *Journal of the American Chemical Society*, **129**, 8333-8344. [doi:10.1021/ja070451k](https://doi.org/10.1021/ja070451k)
- [6] Chen, I.A. and Nowak, M.A. (2012) From pre-life to life: How chemical kinetics become evolutionary dynamics. *Accounts of Chemical Research*, **45**, 2088-2096. [doi:10.1021/ar2002683](https://doi.org/10.1021/ar2002683)
- [7] Anders, E. (1989) Pre-biotic organic matter from comets and asteroids. *Nature*, **342**, 255-257. [doi:10.1038/342255a0](https://doi.org/10.1038/342255a0)
- [8] Matrajt, G., Pizzarello, S., Taylor, S. and Brownlee, D. (2004) Concentration and variability of the AIB amino acid in polar micrometeorites: Implications for the exogenous delivery of amino acids to the primitive earth. *Meteoritics & Planetary Science*, **11**, 1849-1858. [doi:10.1111/j.1945-5100.2004.tb00080.x](https://doi.org/10.1111/j.1945-5100.2004.tb00080.x)
- [9] Martins, Z., Alexander, C.M. O'D., Orzechowska, G.E., Fogel, M.L. and Ehrenfreund, P. (2007) Indigenous amino acids in primitive CR meteorites. *Meteoritics & Planetary Science*, **12**, 2125-2136. [doi:10.1111/j.1945-5100.2007.tb01013.x](https://doi.org/10.1111/j.1945-5100.2007.tb01013.x)
- [10] Hartmann, W.K., Ryder, G., Dones, L. and Grinspoon, D. (2000) The time-dependent intense bombardment of the primordial Earth/Moon system. *Origin of the Earth and Moon*. University of Arizona Press, Tucson, 493-512.
- [11] Wilson, R. (2009) Organic material in micrometeorites: Processes affecting its delivery to planetary environments. Submitted for the Degree of Doctor of Philosophy, Faculty of Science, The Open University, 180.
- [12] Thomas, P.J., Hicks, R.D., Chyba, C.F. and McKay, C.P. (2006) Comets and the origin and evolution of life. Springer, Berlin, Heidelberg, 148.
- [13] De Ley, J. (1968) Molecular biology and bacterial phylogeny. *Journal of Evolutionary Biology*, **2**, 103-156. [doi:10.1007/978-1-4684-8094-8_3](https://doi.org/10.1007/978-1-4684-8094-8_3)
- [14] Shock, E.L. and Schulte, M.D. (1998) Organic synthesis during fluid mixing in hydrothermal systems. *Journal of Geophysical Research*, **103**, 28513-28527. [doi:10.1029/98JE02142](https://doi.org/10.1029/98JE02142)
- [15] Tian, F., Toon, O.B., Pavlov, A.A. and De Sterck, H. (2005) A hydrogen-rich early Earth atmosphere. *Science*, **308**, 1014-1017. [doi:10.1126/science.1106983](https://doi.org/10.1126/science.1106983)
- [16] Ehrenfreund, P., Irvine, W., Becker, L., Blank, J., Brucato, J.R., Colangeli, L., Derenne, S., Despois, D., Dutrey, A., Fraaije, H., Lazcano, A., Owen, T. and Robert, F. (2002) Astrophysical and astrochemical insights into the origin of life. *Reports on Progress in Physics*, **65**, 1427-1487. [doi:10.1088/0034-4885/65/10/202](https://doi.org/10.1088/0034-4885/65/10/202)
- [17] Radzicka, A. and Wolfenden, R. (1996) Rates of uncatalyzed peptide bond hydrolysis in neutral solution and the transition state affinities of proteases. *Journal of the American Chemical Society*, **118**, 6105-6109. [doi:10.1021/ja954077c](https://doi.org/10.1021/ja954077c)
- [18] Edmond, J.M., Von Damm, K.L., McDuff, R.E. and Measures, C.I. (1982) Chemistry of hot springs on the East

- Pacific Rise and their effluent dispersal. *Nature*, **297**, 187-191. [doi:10.1038/297187a0](https://doi.org/10.1038/297187a0)
- [19] Elderfield, H. and Schultz, A. (1996) Mid-ocean ridge hydrothermal fluxes and the chemical composition of the ocean. *Annual Review of Earth and Planetary Sciences*, **24**, 191-224. [doi:10.1146/annurev.earth.24.1.191](https://doi.org/10.1146/annurev.earth.24.1.191)
- [20] Chyba, C.F., Thomas, P.J., Brookshaw, L. and Sagan, C. (1990) Cometary delivery of organic molecules to the early earth. *Science*, **249**, 366-373. [doi:10.1126/science.11538074](https://doi.org/10.1126/science.11538074)
- [21] Pasek, M. and Lauretta, D. (2008) Extraterrestrial flux of potentially prebiotic C, N, and P to the early earth. *Origins of Life and Evolution of Biospheres*, **38**, 5-21. [doi:10.1007/s11084-007-9110-5](https://doi.org/10.1007/s11084-007-9110-5)