

# Laser induced fluorescence study on the growth of maize plants

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## ABSTRACT

The laser induced fluorescence (LIF) spectra from plants give accurate information on the influences of ultraviolet A (UV-A) and ultraviolet B (UV-B) radiation on the growth of maize plants. In this paper, a thorough LIF study has been done on maize plants grown under controlled conditions. The maize (*Zea mays*) seeds were planted and exposed to UV-A and UV-B radiation from 0 to 10 hours. The decrease in the ratios of peaks of LIF confirm a decrease in the height, diameter of stem, number of leaves in the plants exposed to UV-B and UV-A with the increase in the time of exposure.

**Keywords:** UV-A Radiation; UV-B Radiation; Zea Mays; LIF

## 1. INTRODUCTION

Stratospheric ozone depletion has caused an increase in the amount of ultraviolet-A (UV-A) and ultraviolet-B (UV-B) radiation reaching the earth's surface (Gao *et al.*, 2004). It is suggested that the maximal ozone depletion and peak UV-B levels will occur during the next decade, with a return to pre-1980 levels of stratospheric ozone and UV-B by the middle of this century [1,2]. UV-A radiation has a wavelength range of 320 - 440 nm and UV-B has a wavelength range of 280 - 320 nm. Since UV-B has a shorter wavelength than UV-A, it is the most damaging ultraviolet radiation [2]. The most important parameter for global irradiance at the surface of the earth is solar elevation above the horizon. The spectral alteration in the UV-B range is mainly caused by ozone layer through which solar radiation must pass [3]. Since cloudiness is a very decisive factor for global irradiance, it is difficult to define general criteria for its influence; however the small contribution of transmitted UV-B cannot be neglected since the photochemical and dam-

aging effects of plants increase exponentially with increasing wavelength [3].

Remote sensing techniques are widely used these days to study plants. In remote sensing applications, laser-induced fluorescence (LIF) is a powerful technique, which can be used for early remote sensing of stress conditions in plants [4,5]. LIF is an active sensing technique capable of capturing immediate and specific indications of changes in plant physiology and metabolism as they relate to the concentration and photosynthetic activity of the plant pigment. There are several advantages in the use of pulsed UV lasers as excitation sources. For example, the use of this excitation spectral range is recommended by international eye-safe regulation in remote sensing. Another advantage is that the powerful and reliable laser sources are available in the whole spectral range of UV [5]. The continuous fluorescence can be obtained from a high repetition rate pulsed excitation. Furthermore, the fluorescence emission spectra may have range as wide as possible and hence the increasing information contents in the LIF technique [4,5]. UV LIF emission of green plants has become of great interest with respect to application of remote fluorescence light detection and ranging (LIDAR) techniques for screening the state of health of terrestrial vegetation [4,6]. Green leaves excited by UV light emit red fluorescence between 650 and 800 nm (known as chlorophyll fluorescence) and autofluorescence emission in the blue-green spectral region [5,6].

Plant damage caused by environmental changes can be studied using LIF technique. Mineuchi *et al.* [7] used LIF technique to study the effects of UV-B radiation on peanut leaves. They evaluated the vitality decrease of leaves by measuring the change in a specific fluorescence peak ratio. They found that an early detection and evaluation of UV-B stress could be possible by applying the method of remote sensing. Takeuchi *et al.* [8] investigated leaf samples irradiated by UV-B and found that at two wavelengths, namely 685 and 735 nm, the LIF lifetimes became shorter during UV-B irradiation. They established a

relationship between the lifetimes and plant's activities under UV-B irradiation.

Maize is one of the most important staple foods in the world. Therefore the study of maize seedlings to stimulated UV-A and UV-B light under laboratory conditions is very important for assessment of the responses of maize plants to a stress factor in the fields [9]. The effects of UV-A and UV-B on the growth of maize seedlings have been reported by several researchers [3,9-11]. Several investigators have demonstrated that the effect of UV-B radiation enhancement on plants includes reduction in grain yield, alteration in species competition, susceptibility to diseases and changes in plant structure and pigmentation [2,9]. The enhanced UV radiation can cause deleterious effects to DNA, the photosynthetic system and other target molecules. Some effects on photosynthesis may also include alteration in light penetration into the leaf, changes in chlorophyll and stomata density, and reduction in leaf area [2,12]. The objective of this study was to evaluate the effects of UV-A and UV-B radiation on the growth of maize seedling using laser induced fluorescence (LIF) technique. This may be used to get an overview of what happens when maize plants are growing in UV regions under field conditions, and help find measures on how to protect crops from increasing UV-A and UV-B radiation [9].

## 2. MATERIALS AND METHODS

### 2.1. Growing and Exposure of Maize Seedlings to UV Radiation

Fifteen pots were filled with sandy soil. The depth of the seeds in the soil was kept at 1 cm and the pots were left to stand under normal (white) light during the day and under a dark mode during night time by switching the lights off. Each pot contained 3 seeds. The pots were watered with 100 ml of water every 3rd day for 2 weeks. The seedlings were divided into two groups. One group was exposed to UV-A (Philips, F40T, 12/B1, Actinic) and another group was exposed to UV-B (Philips, TL 40W/12RS, J2) lamb lights. The exposure to UV-A and UV-B radiation was given for up to 10 hours at two hour intervals. Each experiment was done using three replicates. The protocol used in this study was previously reported by Hedimbi *et al.* [9].

### 2.2. Laser Induced Fluorescence Spectrum

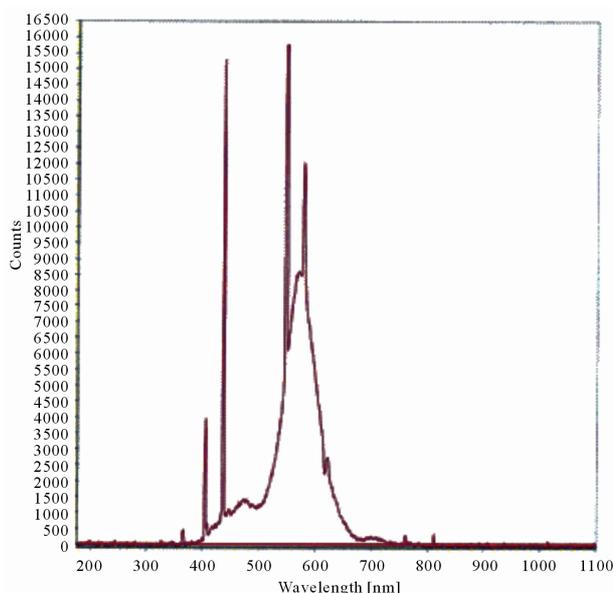
After the exposure of the maize seedlings (every 2 hours), the emission spectrum of the third leaf from the bottom, side of each maize seedling excited by 663 nm was obtained, using Ava Light-D(H)S Deuterium-Halogen light sources which is connected to Ava Spec-2048 Fiber Optic Spectrometer and a computer for displaying

the results in a graph format.

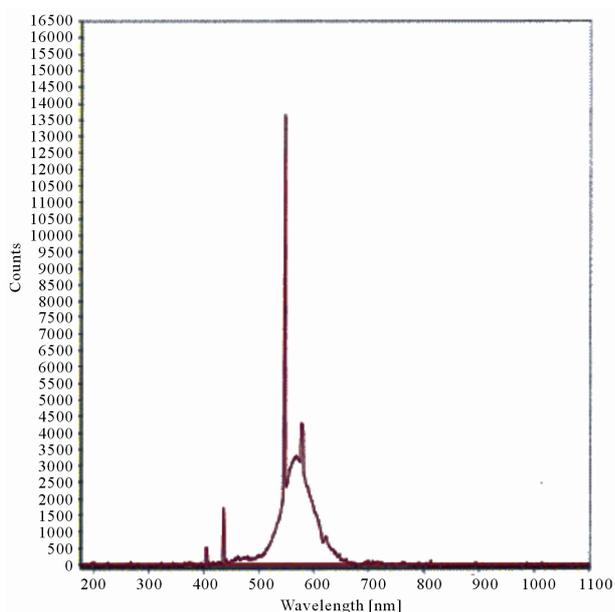
## 3. RESULTS AND DISCUSSIONS

The LIF spectrum was taken from the bottom of third leaf of each plant grown under normal conditions and under stress of UV-A and UV-B exposure and we have shown the spectrum taken for the third leaf in the **Figures 1-10**. For each plant, we have shown the kind of stress given to the plant and time of stress. **Figures 1-5** show LIF spectrum for UV-A stress given to plants for 2 hours (**Figure 1**), 4 hours (**Figure 2**), 6 hours (**Figure 3**), 8 hours (**Figure 4**) and 10 hours (**Figure 5**). **Figures 6-10** shows LIF spectrum of plants exposed to UV-B radiation for 2 hours (**Figure 6**), 4 hours (**Figure 7**), 6 hours (**Figure 8**), 8 hours (**Figure 9**) and 10 hours (**Figure 10**). Each spectrum has four peaks at 410 nm, 450 nm, 560 nm, and 600 nm. The intensity ratios of these peaks as F410/F450, F410/F560, and F560/F600 corresponding to blue to green, blue to red, and chlorophyll fluorescence were calculated and analyzed (**Tables 1 and 2**). It was found that the ratio F560/F600 decreased due to the photosynthesis activity whereas the ratio of F410/F450 was found to have decreased with the increase of UV-A and UV-B radiation but the ratio of F410/F560 was found to be increased with the increase of UV-A and UV-B radiation (**Tables 1 and 2**). Furthermore, it was found that UV-B was more effective than UV-A in reducing photosynthetic activity of maize plants as shown in **Tables 1 and 2**.

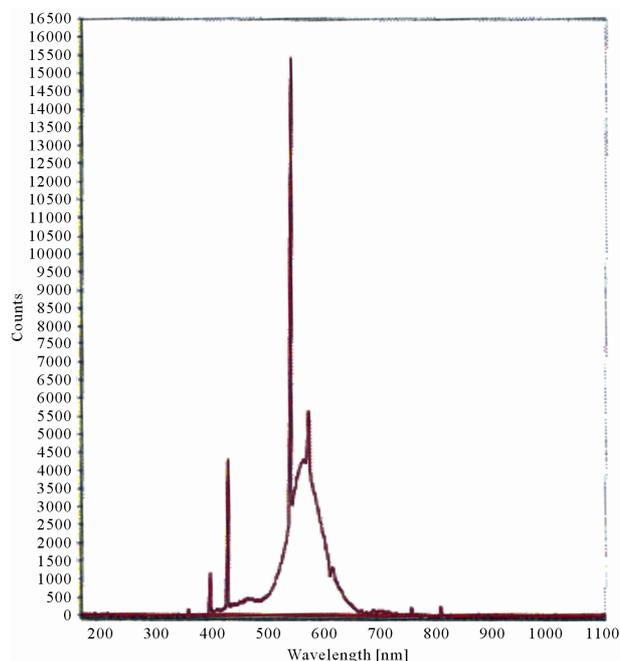
The continuous growth of plants indicates that they can tolerate or adapt to UV radiation and can survive. The epidermis usually protects the inner leaf with high



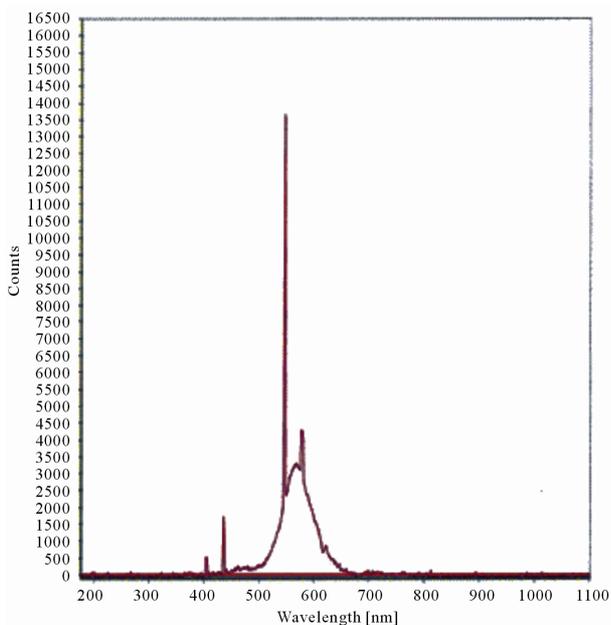
**Figure 1.** LIF spectrum of maize leaves exposed to 0 - 10 hours of UV-A radiation.



**Figure 2.** LIF spectrum of maize leaves exposed to 4 hours of UV-A radiation.

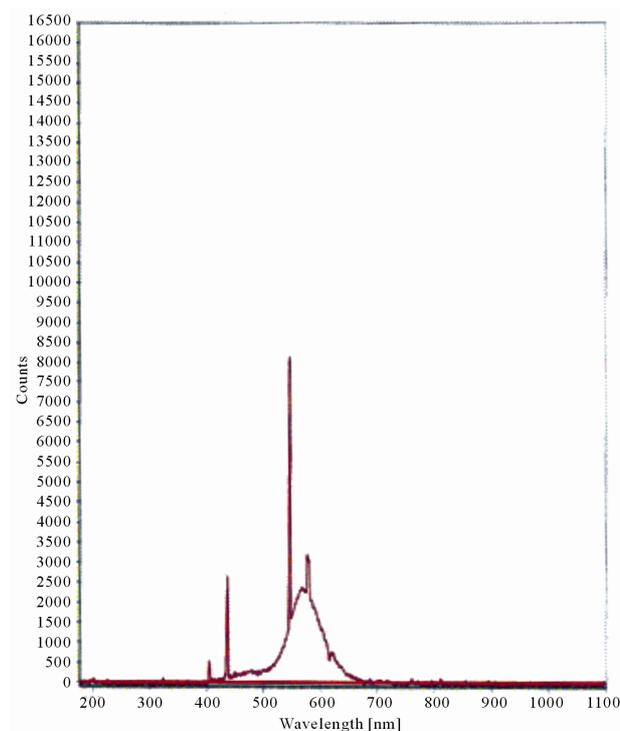


**Figure 4.** LIF spectrum of maize leaves exposed to 8 hours of UV-A radiation.



**Figure 3.** LIF spectrum of maize leaves exposed to 6 hours of UV-A radiation.

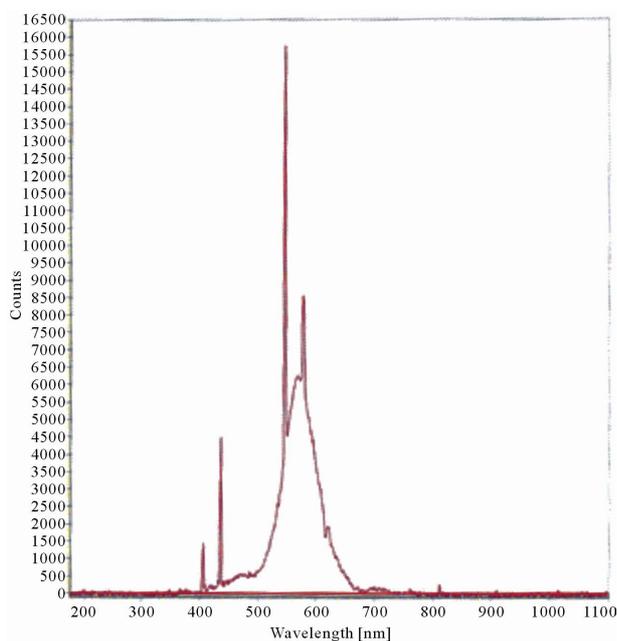
concentrations of flavonoids and anthocyanins, which absorb the UV radiation safely. The UV treatment is affected by photosynthetically active radiation (PAR) levels [13,14]. Whether the presence or absence of activated photomorphogenic, photosynthetic and photoprotective systems during the UV treatment affect plant sensitivity to UV radiation or not has not been investigated so far. For example, the possible role of photosynthetic process and of xanthophyll cycle activated by wavelengths > 500



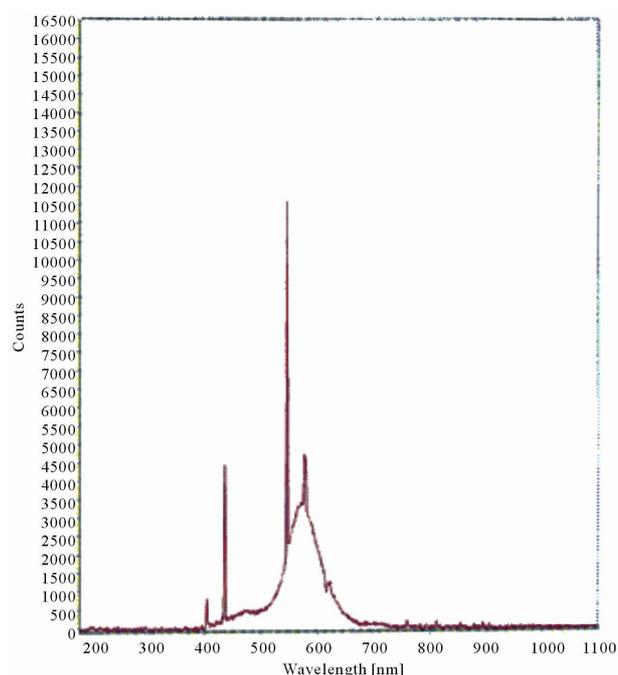
**Figure 5.** LIF spectrum of maize leaves exposed to 10 hours of UV-A radiation.

nm has not been determined because UV irradiation is always been given in the presence of PAR [11].

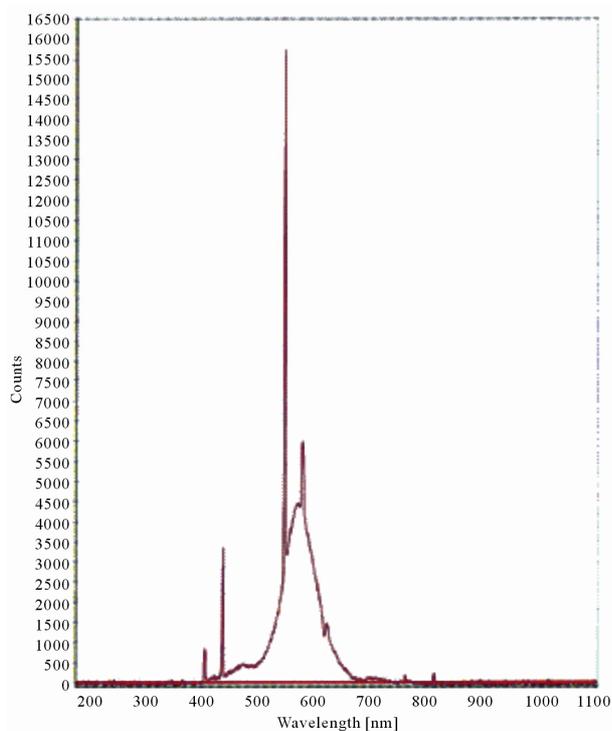
UV-B radiation causes a multitude of physiological and biochemical changes in plants, including inhibition



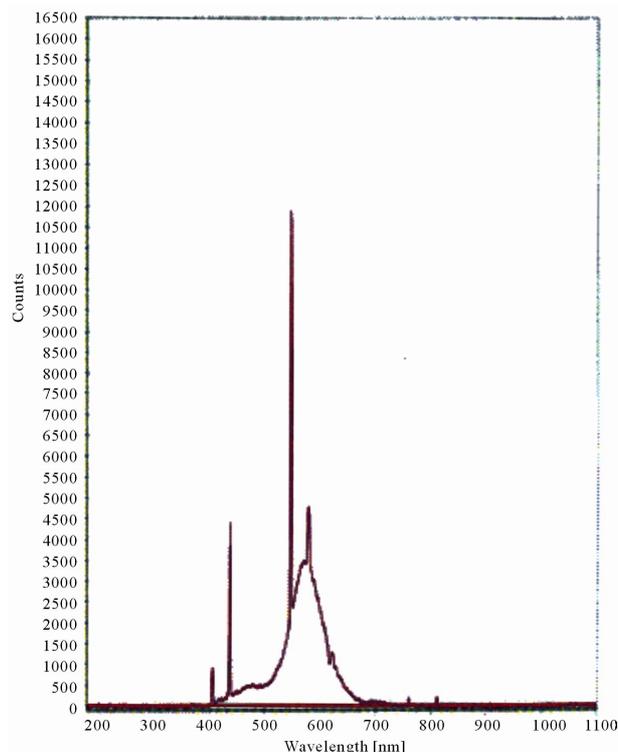
**Figure 6.** LIF spectrum of maize leaves exposed to 2 hours of UV-B radiation.



**Figure 8.** LIF spectrum of maize leaves exposed to 6 hours of UV-B radiation.



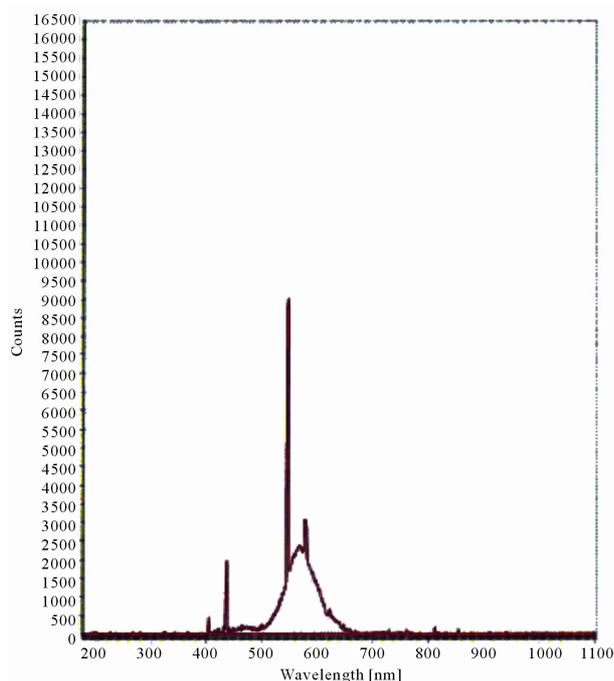
**Figure 7.** LIF spectrum of maize leaves exposed to 4 hours of UV-B radiation.



**Figure 9.** LIF spectrum of maize leaves exposed to 8 hours of UV-B radiation.

of photosynthesis [15,16]. This inhibition is due to reduced levels of chlorophyll (Chl), chloroplast proteins such as Rubisco (ribulose-1,5-bisphosphate carboxylase/oxygenase) and LHCII (light-harvesting chlorophyll a/b-binding protein of photosystem II) [16,17], and pho-

tosynthesis-related gene expression [16,18]. Allen *et al.* [19] reported that loss of Rubisco is a primary factor in UV-B inhibition of photosynthesis in oilseed rape. Further,



**Figure 10.** LIF spectrum of maize leaves exposed to 10 hours of UV-B radiation.

**Table 1.** The fluorescence intensity ratios of maize leaves exposed to UV-A with their respective standard errors of three replicates.

UV-A (hours)	F410/F450	F450/F560	F560/F600
0	0.4 ± 0.0	0.2 ± 0.0	2.9 ± 0.1
2	0.3 ± 0.0	0.2 ± 0.1	2.7 ± 0.1
4	0.3 ± 0.0	0.3 ± 0.0	2.1 ± 0.0
6	0.3 ± 0.0	0.3 ± 0.2	1.7 ± 0.1
8	0.3 ± 0.0	0.6 ± 0.0	1.7 ± 0.1
10	0.2 ± 0.0	0.4 ± 0.1	1.6 ± 0.1

**Table 2.** The fluorescence intensity ratios of maize leaves exposed to UV-B with their respective standard errors of three replicates.

UV-B (hours)	F410/F450	F450/F560	F560/F600
0	0.4 ± 0.0	0.3 ± 0.1	2.5 ± 0.1
2	0.3 ± 0.5	0.3 ± 0.1	2.2 ± 0.3
4	0.3 ± 0.0	0.4 ± 0.8	2.1 ± 0.1
6	0.3 ± 0.0	0.4 ± 0.0	2.0 ± 0.4
8	0.2 ± 0.0	0.4 ± 0.0	1.4 ± 0.2
10	0.2 ± 0.0	0.4 ± 0.1	1.1 ± 0.5

it was apparent that the UV-B induced reduction in Rubisco was greater in UV-sensitive, than in UV-resistant strains [16].

It has been reported that, in chloroplasts under illumination, the large subunit (LSU) of Rubisco is directly fragmented into two polypeptides by reactive oxygen species (ROS) [20,21]. Other authors have demonstrated that UV-B generated ROS induce photodamage to Rubisco [22], and that ROS cause proteolytic degradation of the LSU [23]. Thus, the generation of ROS is thought to be involved in UV-B induced degradation of Rubisco in rice. John *et al.* [24] reported that, in *Arabidopsis*, exposure to UV-B radiation induces expression of senescence-associated genes (SAGs), including SAG12, which encodes a cysteine protease [16,25]. This result suggests that the formation of some kind of protease may also be involved in the enhancement of Rubisco degradation in plants [16].

Singh *et al.* [10] studied the effects of UV-A radiation on the growth of maize plants and their fluorescence spectra. The emission spectra of the second leaf from the bottom side of each maize plant excited by 337 nm were obtained using spex 1680, 0.22 m double monochromator known as spexfluorolog made in USA. The spectra consisted of two peak bands in the blue-green region at 435 nm (F435) and 525 nm (F525) respectively and two peak bands in the red region at 684 nm (F684) and 740 nm (F740) respectively. The ratios of these peaks as (F435/F525), (F435/F684) and (F684/F740) were calculated in each case. The ratio of blue to red (F435/F684) and chlorophyll fluorescence (F684/F740) ratio were found to be decreased. This might be due to the fact that the intensity of red peaks was increased due to reabsorption of light when the plants were treated with UV-A, and hence the ratios (F435/F684) and (F684/F740) were reduced [3,5,10].

LIF obtained from intact leaves under different conditions and their quantitative relation to stress detection has been discussed in detail by Theisen [26]. Fluorescence of plant tissues is considered a serious problem in immunofluorescence microscopy investigation of signified tissues and tissues with vacuolar deposits of phenolic compounds and in some cases it has been eliminated by the use of strong reducing substances [3,5]. Oxidized chlorophylls that emit at 540 nm are released to vacuoles from chloroplasts [3,5]. Broglia [26] found a decrease of laser induced green fluorescence and increase of chlorophyll fluorescence in *Vicia faba* when epidermal tissue had been removed. This shows that the epidermis is both an important protection against UV radiation and a possible topological source of green fluorescence [3,5].

There are two potential primary mechanisms involved in UVB-induced physiological and biochemical damage. DNA lesions, such as cyclobutane pyrimidine dimer

(CPD) and pyrimidine (6-4) pyrimidone photoproducts [(6-4) photoproducts], interfere with DNA replication and transcription [16,27]. The second mechanism is through modification of proteins by photo-oxidation, or by reactive oxygen species (ROS) and free radicals produced during photosensitization [16,22,27]. These modifications include cross-linking, aggregation, denaturation and degradation [16,27].

Gao *et al.* [2] investigated the effects of supplementary UV-B radiation on the growth, yield and seed qualities of maize under field conditions. Increased UV-B radiation caused a significant reduction in dry matter and yield, and affected seed quality as follows: protein, sugar and starch levels decreased, whereas lysine levels increased. Taken together, the results of this study suggest that the enhanced solar UV-B radiation as predicted by atmospheric models will result in reduction of growth and yield of maize crops in the future. This has been confirmed from this study that the enhanced UV-A and UV-B radiation causes a significant reduction in the growth of maize plants and hence in the yield in turn. However, to have proper protection, the excessive exposure of both UV-A and UV-B should be avoided to have better growth of yield. Also, it has been observed that maize can sustain certain level of UV-A, but excessive exposure will be dangerous to plants whereas exposure of UV-B of any level is harmful to plants.

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