

# *In Vivo* Investigation of Zr-Based Bulk Metallic Glasses Sub-Periosteally Implanted on the Bone Surface

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## Abstract

Bulk metallic glasses (BMG) show higher strength and lower Young's modulus than SUS 316L stainless steel and Ti-6Al-4V alloys. This study aimed to investigate the reaction of Zr-based BMG sub-periosteally implanted on the surface of the rat femur, thereby evaluate the possibility of the BMG as biomaterials for osteosynthetic devices.  $Zr_{65}Al_{7.5}Ni_{10}Cu_{17.5}$  BMG ribbons with 10 mm length, 2 mm width and 0.5 mm thickness were implanted sub-periosteally on the femur surface in three male Wistar rats for 6 weeks. Systemic effects were evaluated by measuring Cu and Ni levels in the blood, and local effects were evaluated by the histological observation of the surrounding soft tissues in contact with the BMG. The reaction of the surface of the BMG was examined with scanning electron microscopy. No increase of Cu and Ni levels in the blood was recognized. In the scanning electron microscopy observation, spherical deposits which were considered as sodium chloride crystals were observed. Neither breakage nor pitting corrosion was noted. BMG will be a promising metallic biomaterial for osteosynthetic device that must be removed.

## Keywords

Amorphous Alloy, Bulk Metallic Glasses, Biomaterial, Osteosynthetic Device

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## 1. Introduction

For materials of osteosynthetic devices such as bone plates, intramedullary nails, and screws, crystalline metallic alloys such as SUS 316L stainless steel and Ti-6Al-4V alloys are commonly used. The materials need to possess non-toxicity, anti-corrosiveness, durability, strength, low Young's modulus and biocompatibility.

Bulk metallic glasses (BMG), *i.e.* amorphous alloys, are metallic materials with metastable glassy states. Tensile strength of the Zr-based BMG is 1500 - 1700 MPa [1] and approximately twice higher than that of Ti-6Al-4V alloy and 3 times higher than that of 316L stainless steel. Young's modulus of the Zr-based BMG is 70 to 80 GPa [1], which is closer to that of the bone than those of the conventional biomaterials.

$Zr_{65}Al_{7.5}Ni_{10}Cu_{17.5}$  BMG showed anti-corrosiveness behavior in the physiologically relevant environment [2]-[7] and  $Zr_{52.5}Cu_{17.9}Ni_{14.6}Al_{10.0}Ti_{5.0}$  BMG showed excellent electrochemical properties in the phosphate-buffered saline electrolyte [8]. However, the behaviors of Zr-based BMG have not been investigated so far by

conducting an *in vivo* experiment.

The aim of this study was to investigate the reaction of Zr-based BMG sub-periosteally implanted on the surface of the rat femur, thereby evaluate the possibility of the BMG as biomaterials for osteosynthetic devices.

## 2. Materials and Methods

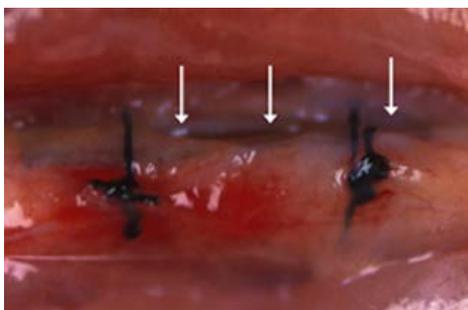
Zr<sub>65</sub>Al<sub>7.5</sub>Ni<sub>10</sub>Cu<sub>17.5</sub> alloy ingots were prepared from a mixture of pure metals; zirconium, aluminum, nickel, and copper, each with a purity of more than 99.9%. Each ingot was prepared by an arc melting method. The alloy was re-melted in a quartz nozzle and rapidly quenched by casting it on the rotating copper roll. BMG ribbon with 10 mm length, 2 mm width and 0.5 mm thickness was obtained. The structure of the formed alloy was analyzed by means of an X-ray diffraction method with Cu K $\alpha$  radiation to ascertain that whole of the structure of the alloy was really amorphous. The free-side surface was polished with 600 grid silicon carbide paper in distilled water, dried for a few days to generate air-formed passive film, and then were sterilized in a high-pressure steam sterilizer (BS-305; TOMY, Tokyo, Japan). It was confirmed that none of these preparation procedures had any influence on the amorphous structure of the alloy.

*In vivo* experiment was approved by the animal committee in our institution based on the result of the study showing excellent anti-corrosiveness in the simulated body fluid [2]-[7]. Fourteen-week-old male Wistar rats were used in the experiment. Each animal was housed in a separated cage (22 cm  $\times$  33 cm  $\times$  13 cm height) and received a standard diet. Six rats weighing 400 to 450 grams were randomly assigned into two groups. The Zr<sub>65</sub>Al<sub>7.5</sub>Ni<sub>10</sub>Cu<sub>17.5</sub> BMG ribbon was implanted sub-periosteally on the femur surface in 3 rats (**Figure 1**). The other 3 rats were assigned as control group.

Under intraperitoneal anesthesia with 75 mg/kg of ketamine hydrochloride and 10 mg/kg of xylazine, the left mid-shaft of femur was exposed between the vastus lateralis and hamstrings and BMG ribbon was implanted sub-periosteally on the surface of the femur of each rat. For preventing migration of the alloy, it was tied up both proximally and distally on the surface of the femur with a 4 - 0 nylon suture string (**Figure 2**). A sham operation was performed on the contra-lateral right femur. The wound was irrigated with normal saline and then was closed by each layer suture. Postoperatively, each animal was allowed free activity in a separate cage.



**Figure 1.** Radiograph of the femur with the BMG ribbon sub-periosteally implanted on the bone surface.



**Figure 2.** Photograph of the femur with the BMG ribbon (arrows) sub-periosteally implanted and tied up both proximally and distally with a 4 - 0 nylon suture string after 6 weeks implantation.

After 6 weeks from implantation, 5 ml of blood samples was obtained from the descending aorta under anesthesia with intra-peritoneal injection of 45 mg/kg of ketamine hydrochloride and 6 mg/kg of xylazine to measure blood level of Cu and Ni. Then, euthanasia with intra-peritoneal injection of 120 mg/kg of ketamine hydrochloride and 16 mg/kg of xylazine, the BMG ribbon was removed. Undecalcified specimens of the femur, periosteum, and surrounding soft tissues that were in contact with the BMG ribbon were prepared and stained with Hematoxylin-Eosin.

The extracted BMG ribbons were lavaged for 20 second with 30 ml of UF detergent in purified water and were fixed by a carbon tape and preserved in an airtight container. The surface of the BMG that had been in contact with the femora were observed by a scanning electron microscopy (XL30FEG, Philips) and constituents of the material formed on the surface of the alloys were identified and semi-quantified by energy dispersive X-ray spectroscopy (DX-4, EDAX Japan, Tokyo).

### 3. Results

#### 3.1. Cu and Ni Blood Levels

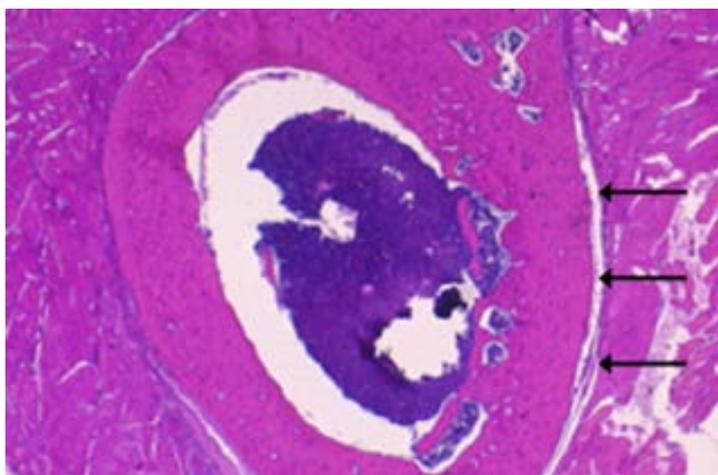
In the investigation on the systemic effects of BMG group, the mean whole-blood Cu concentration was  $101.3 \pm 6.1$   $\mu\text{g/dl}$  and the mean whole-blood Ni concentration was less than  $0.10$   $\mu\text{g/dl}$ . In control group, the mean whole-blood Cu concentration was  $117.3 \pm 11.2$   $\mu\text{g/dl}$  and the mean whole-blood Ni concentration was less than  $0.10$   $\mu\text{g/dl}$ . No increase of the blood levels of Cu and Ni was recognized in BMG group in comparison to those of control group.

#### 3.2. Histological Findings

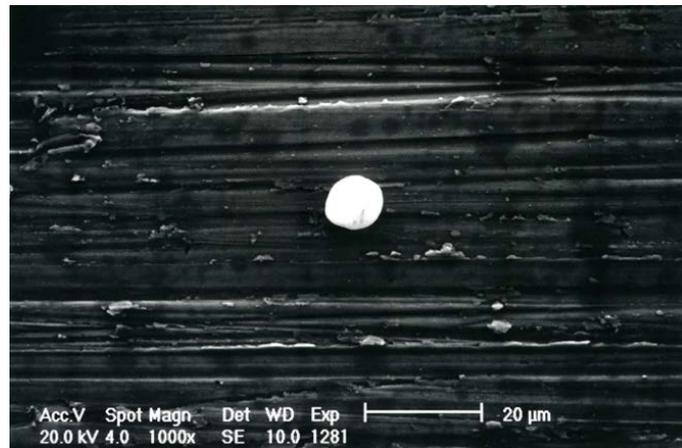
The observation by light microscopy of the H&E stained specimens of the surrounding soft tissues in contact with the BMG disclosed that no findings of the biological effects was recognized such as bone resorption, infiltration of inflammatory cells, cell necrosis or dysplasia, wear debris of the alloy and so on (**Figure 3**).

#### 3.3. Surface Analysis of the Implanted Materials

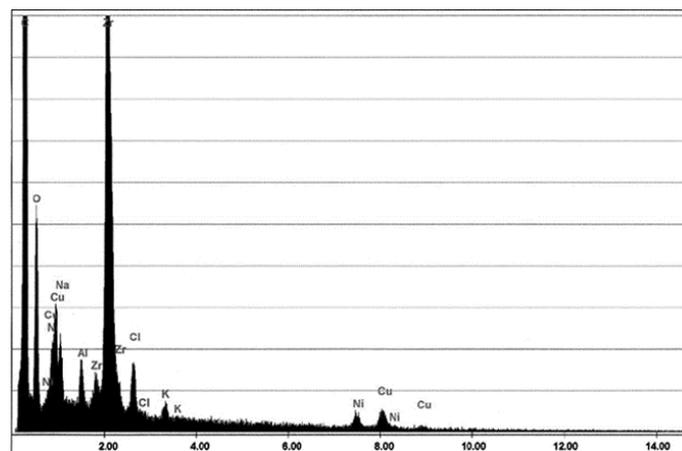
Surface observation by scanning electron microscopy (SEM) revealed spherical deposits (**Figure 4**). Transverse grooves that would be made by polished with 600 grid SiC paper were observed both before implantation and after 6 weeks implantation. The semi-quantitative examination of the constituents of the surface materials formed on the alloy surface by energy dispersive spectroscopy (EDS) disclosed that Na and Cl were recognized (**Figure 5**). Therefore, the spherical deposits were considered as sodium chloride crystals. Neither breakage nor pitting corrosion of the harvested BMG was noted (**Figure 6**).



**Figure 3.** Photomicrograph of the femur and the surrounding soft tissues after 6 weeks implantation and removal of the BMG. The BMG ribbon had been implanted sub-periosteally (arrows). H & E,  $\times 10$ .



**Figure 4.** Surface appearance of the extracted BMG after 6 weeks implantation. Scanning electron microscopy (SEM),  $\times 1000$ .



**Figure 5.** Energy dispersive spectroscopy (EDS) of the surface of the BMG after 6 weeks implantation. The constituents of the surface materials included Na and Cl.



**Figure 6.** The extracted BMG ribbon. This photo shows the side that had been in contact with the femur.

#### 4. Discussion and Conclusion

Current osteosynthetic devices made of SUS 316L stainless steel and Ti-6Al-4V titanium alloy sometimes failed due to their insufficient strength. According to the survey on orthopaedic implant failure by the implant com-

mittee of the Japanese Orthopaedic Association, 2.5% of the osteosynthetic implants failed [9]. To prevent failure, the bone plates have inevitably been made bulky and extensive surgical exposure is necessary to implant such plates. Occasionally it leads difficulty in closing the open wound. Therefore, one of the disadvantages of the current metallic osteosynthetic devices is their insufficient strength. Another disadvantage is their excessively higher Young's modulus. The Young's moduli of currently available metallic devices are 100 GPa ordered and much higher than that of cortical bone, *i.e.*, 20 GPa. This sometimes causes stress shielding and absorption of the bone stabilized with osteosynthetic devices made of these materials [10]-[14].

BMG, *i.e.* amorphous alloys, are materials with no long-range atomic order that are prepared by the solidification of a liquid melt at a sufficient rate to suppress the growth of crystalline phases. Conventional metallic biomaterials such as Ti-6Al-4V alloys and 316L stainless steel are crystalline alloys. Crystalline alloys have grain boundaries, dislocations, and segregations. When the crystalline alloys are loaded, grain boundaries and dislocations easily lead failure. In addition, crystalline alloys have slip plane formed by shearing stress, which results in plastic deformation of the material via translocation of the structure.

On the contrary, BMG have random atom structure and do not have segregations and defects in their structure. In BMG, no slip plane is generated and elastic deformation continues even under considerably large stress because the deformation is caused by massive movement of the atoms. On this account, BMG show higher strength and lower Young's modulus than crystalline alloys. In this way, BMG have some advantages for metallic biomaterials.

In the previous literatures, there had been only two studies that evaluated BMG *in vivo* [15] [16]. There had been another paper that reported metallic intravascular stent with amorphous oxide surface showed excellent corrosion resistance not only *in vitro* but also *in vivo* [17]. Our investigation was the first to implant BMG subperiosteally on the bone surface *in vivo*.

In this study, we used a Zr-based BMG containing nickel that was referred to as carcinogenic. No systemic and local effects were recognized by measuring Cu and Ni levels in the blood and the histological observation of the surrounding soft tissues in contact with the BMG. But nickel free materials might be more appropriate for biomaterials and nickel free Zr-Al-Cu BMG is available now. The future study should investigate nickel free Zr-Al-Cu BMG.

The SEM and EDS results in this study indicated that  $Zr_{65}Al_{7.5}Ni_{10}Cu_{17.5}$  BMG was almost biologically inert. The excellent biocompatibility of titanium and its alloys may result in osseous integration and strong bone-to-metal attachment may cause complications. The detaching test after implantation of the Ti-6Al-4V plates into the tibiae of rabbits indicated that the Ti alloy bonds directly to bone after more than 8 weeks [18]. Therefore, biomaterials with low reactivity *in vivo* like  $Zr_{65}Al_{7.5}Ni_{10}Cu_{17.5}$  BMG might be more appropriate for osteosynthetic device that must be removed.

The problems exist in adopting this material for clinical application. One is that large sized BMG are technically demanding to create. Applying BMG for osteosynthetic devices, even larger sized material should be manufactured. Another problem is that there is no data about non-toxicity, anti-corrosiveness, and durability with long term implantation. In conclusion, BMG will be a promising metallic biomaterial for new osteosynthetic implants but further *in vivo* study should be necessary.

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