

# Tailoring Properties of Biocompatible PEG-DMA Hydrogels with UV Light

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## ABSTRACT

Hydrogels are highly water-absorbent hydrophilic polymer networks, which show potential in many biocompatible applications. In previous work, we demonstrated the feasibility of using poly(ethylene glycol) dimethacrylate (PEG-DMA) gels polymerized with a photoinitiator for encapsulation and stabilization of painted biomimetic membrane arrays for novel separation technologies or biosensor applications. These gels were formed from PEG-DMA monomers suspended in phosphate buffered saline (PBS) solution and gelled by radical polymerization in the presence of the photoinitiator Darocur 1173. In this work, we show that the properties of a PEG-DMA hydrogel formed by photoinitiated polymerization can be tailored by varying the photocrosslinking time. Fourier Transform Infrared Spectroscopy (FTIR) and Raman Spectroscopy (RS) showed that the optimal crosslinking time for the gel was 6 - 10 minutes and that the water content of the gels could be tuned in the range of 50 - 90 wt%. The resistivity was between 0.8 - 3.5  $\Omega$ m, which is comparable to that of PBS. The low resistivity of the gel makes it compatible for encapsulating membranes for (ion channel based) biosensor applications. With FTIR and RS we identified spectral features of the hydrogel, which may serve as a diagnostic tool to monitor changes in the gels due to variation in parameters such as time, pH, temperature, aging or exposure to chemicals or biological material.

**Keywords:** Hydrogel Swelling; UV-Induced Polymerization; FTIR, Raman Spectroscopy; Impedance Spectroscopy

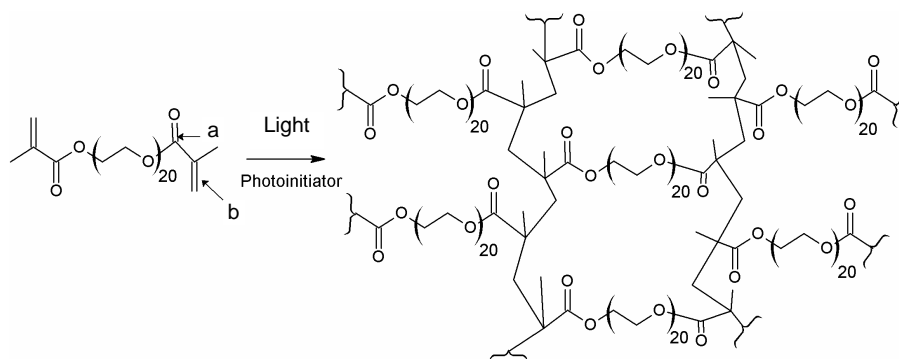
## 1. Introduction

Hydrogels are highly water absorbent hydrophilic polymer networks [1-3]. They are favored as scaffolds for tissue engineering where they may act as supports for cells [4-9] and biomimetic membranes [10,11]. With the advent of methods for producing large scale biomimetic membranes it becomes important to assess material properties of hydrogels as encapsulation materials for technological application of biomimetic membranes [12]. Poly(ethylene oxide) (PEO) synthetic hydrogels are the most commonly used synthetic hydrogel polymers for tissue engineering. Poly(ethylene glycol) (PEG)-based hydrogels are structurally similar to PEO and can be crosslinked by modifying each end with acrylates or methacrylates [7]. These materials are suitable for supporting cells and biomimetic membranes for several reasons: their chemistry and properties can be controlled by varying for example molecular weight, chemical composition, the amount and type of initiator used and the de-

gree of crosslinking, which modifies their mass transport properties, physical properties and biological properties. Also, poly(ethylene glycol)-diacrylate (PEG-DA) hydrogels have been shown to be compatible *in vivo* with porcine islet cells [8] and poly(ethylene glycol)-dimethacrylate (PEG-DMA) hydrogels to be compatible with chondrocytes *in vitro* [9].

Photoactivated PEG-DMA hydrogels are formed from PEG-DMA monomers suspended in aqueous solution and gelate by radical polymerization in the presence of a photoinitiator. The polymerization reaction starts when the solution is exposed to UV light. Each PEG-DMA monomer has two methacrylate groups which can react with up to two other methacrylate groups to make covalent bonds. Thus, each PEG-DMA monomer can covalently link to up to four other PEG-DMA monomers and the resulting polymer forms a covalently crosslinked branched network, see **Figure 1**.

In the design of large scale biomimetic membranes [13-15] stabilization without compromising transport properties



**Figure 1. Chemical structure of PEG-1000-DMA and of the crosslinked PEG-1000-DMA meshwork. Gelation occurs through radical polymerization in which the methacrylate groups participate in an addition reaction to form a branched polymeric network. The crosslinked gel, X1000P, was made from a hydrogel precursor solution containing 65 mM PEG-1000-DMA with 10 mM Darocur photoinitiator in aqueous buffer. The C=O (a) and C=C (b) bonds to which FTIR and RS spectral peaks are assigned are indicated.**

becomes important. In previous work [10,16] we characterized chemically activated and photoactivated PEG-DMA and PEG-DA hydrogels with respect to volumetric stability, water permeability, ionic permeability and porosity and showed successful hydrogel encapsulation of multiple lipid membranes formed across an aperture array.

In this work we address how transport properties of PEG-DMA hydrogels suitable for biomimetic applications can be tailored by varying the photocrosslinking time and thereby the degree of crosslinking. We show that we can change the water absorption capability and the ionic conductivity of the gel, which could be used to control the gel's flux properties. Using Fourier Transform Infrared spectroscopy (FTIR) and Raman spectroscopy (RS) we obtain the spectral fingerprint profile of the gel and find that there is a shift in the C=O peak wavenumber and a decrease in the intensity of the C=C peak due to crosslinking. Furthermore, RS measurements show how the fingerprint is altered with specific spectral peaks receding and other peaks appearing with increased UV irradiation time. The spectral fingerprint of the gel may be applicable as a diagnostic tool to detect changes in the gel in response to variation in parameters, such as temperature, pH, aging and exposure to chemicals.

## 2. Materials and Methods

### 2.1. Materials

Poly(ethylene glycol)-1000-dimethacrylate (PEG-1000-DMA) was from Polysciences (Warrington, PA, USA). 2-Hydroxy-2-methyl-1-phenyl-propan-1-one (Darocur<sup>®</sup> 1173) was from Ciba Specialty Chemicals (Basel, Switzerland). Phosphate buffered saline (PBS) was from Sigma Aldrich Denmark (Brøndby, Denmark).

### 2.2. Preparation of Hydrogels

The hydrogel was prepared as in [10,16]. Briefly, the

hydrogel precursor solution (HPS) consisted of 65 mM PEG-1000-DMA and 10 mM Darocur<sup>®</sup> 1173 (a photoinitiator) in PBS. We will refer to this solution as 1000P. Free-radical polymerisation of 1000P was initiated by 4, 6, 8, 10 or 15 min of photocrosslinking of the HPS by UV light from an EA-140 UV lamp with wavelength  $\lambda = 365$  nm (Spectroline, Westbury, NY, USA) at a lamp distance of 1 cm. The lamp intensity at a distance of 15 cm was  $800 \mu\text{W}\cdot\text{cm}^{-2}$ . We will refer to the crosslinked gel as X1000P.

### 2.3. Swelling Experiments

Equilibrium swelling experiments were used to determine the structural characteristics of the PEG-DMA gels. Irradiated hydrogel samples were prepared from 3 ml of HPS per sample. Excess liquid was poured off and the gels were allowed to equilibrate at ambient temperature ( $\sim 25^\circ\text{C}$ ) for 24 h. Thereafter, the gels were weighed to determine the irradiated gel weight,  $W_{ir}$ . Then they were dried under vacuum for 24 h to remove all water (the dried gel mass corresponded to the expected dry mass of PEG-1000-DMA and Darocur in 3 ml of HPS) and weighed to determine the dry gel weight  $W_d$ , as in [17]. The percentage of water content in the irradiated gels was calculated as  $(W_i - W_d)/W_{ir} \cdot 100\%$  [18]. The dried gels were rehydrated in PBS at ambient temperature ( $\sim 25^\circ\text{C}$ ) for 24 h. Excess liquid was poured off and the gels were allowed to equilibrate at ambient temperature ( $\sim 25^\circ\text{C}$ ) for 24 h. Then the gels were weighed to determine the swollen gel weight,  $W_s$  and percentage of water content determined as  $(W_s - W_d)/W_s \cdot 100\%$ .

### 2.4. Fourier Transform Infrared (FTIR) Spectroscopy

Fourier transform infrared (FTIR) spectroscopy was performed on a Thermo Nicolet Nexus 870 with a Ge  $45^\circ$  Smart Ark attenuated total reflectance (ATR) crystal

from Thermo Scientific, MA, USA. The FTIR spectra were recorded using OMNIC software from Thermo Nicolet Corporation with an average of 32 scans in the range 650 - 4000  $\text{cm}^{-1}$  and at a resolution of 4  $\text{cm}^{-1}$ . Samples were dried in a vacuum desiccator for 24 h prior to analysis in order to reduce the contribution from water in the recorded spectrum.

## 2.5. Raman Spectroscopy

Raman spectra were obtained as in [19] by use of a DILOR-XY 800 mm focal length multichannel spectrometer with horizontal Nd:YVO<sub>4</sub> laser excitation (532.8 nm, ~300 mW, vertically polarized). Rayleigh-scattered light was filtered off with a double pre-monochromator (slit width 500  $\mu\text{m}$ ). The Raman light was collected at 90° scattering, dispersed by use of an 1800 lines/mm grating, and focused onto a charge-coupled device (CCD) detector, cooled to 140 K by liquid nitrogen. The spectral resolution was approximately 4  $\text{cm}^{-1}$ . Hydrogel samples were prepared as described above and were not dried prior to measurements. The recorded spectra were normalized to obtain the same intensity for the spectral peak at 1470  $\text{cm}^{-1}$  (attributed to H-C-H bending). The relative number of C=C bonds in the gels was obtained by comparing the peak height at 1640  $\text{cm}^{-1}$  (attributed to C=C stretching), which was used as a measure of the degree of crosslinking.

## 2.6. Impedance Spectroscopy (IS)

The test cell for impedance spectroscopy measurements consisted of a Teflon support with two Pt electrodes between which a cylindrical crown rubber support (approximately 2.2 mm height and 1.5 cm diameter) filled with gel sample was placed and clamped as previously described [20]. The electrodes were connected to a Frequency Response Analyzer (Solartron 1260, Farnborough, England) and measurements were recorded for 100 data points in the frequency range of 1 Hz - 10 MHz, at a maximum voltage of 0.01 V.

## 3. Results and Discussion

### 3.1. Water Content

The water content of the gel influences the mechanical integrity and strength of the gel as well as the diffusion of nutrients and metabolites through the gel to a cell/biomimetic membrane [7]. Controlling the gel water content is important for the gel's interaction with hydrogel-encapsulated biomimetic membranes, such as those presented in our earlier work [10]. The high water absorbance of hydrogels is one of their most important material properties and therefore, it is crucial to be able to tailor the water content of the gels to suit the application that

they are intended for.

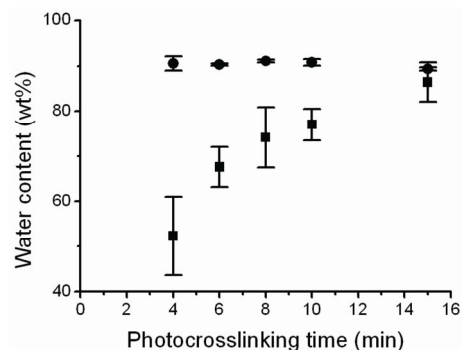
We found that the irradiated and equilibrated PEG-DMA hydrogels contained ~92 wt% water independent of UV irradiation time, see **Figure 2**. However, after the gels were dried and rehydrated overnight, the water uptake ability of the gels increased with an increase in the photocrosslinking time with a water content ranging from 50% for samples irradiated for 4 min to 90 wt% for samples irradiated for 15 min approaching the water content of the non-dried gels, see **Figure 2**.

An explanation for the increased water uptake ability with increased UV irradiation could be that drying the gels causes partial reversible collapse of pores in the gel meshwork as has been observed for similar gels by Wu *et al.* [17], with an increased degree of pore collapse for shorter crosslinking times. Increasing the crosslinking time would increase the crosslinking density and thereby improve the stability of the gel meshwork, leading to less pore collapse. Less pore collapse entails more pores that can be filled with water and thus more water absorption.

Interestingly, Andreopoulos *et al.* [21] report that the water content in a PEG-based gel decreases with increased photocrosslinking time. It is thus possible to tailor the water content of PEG-DMA hydrogels by selecting a suitable UV photocrosslinking time.

### 3.2. Crosslinking Revealed by Fourier Transform Infrared Spectroscopy (FTIR) and Raman Spectroscopy (RS)

In order to analyze whether this trend corresponded to an increase in crosslinking with increased UV irradiation time, we studied samples with FTIR and RS. With FTIR we compared spectral peaks at 1680 - 1780  $\text{cm}^{-1}$  (assigned to C=O bond vibrations and indicated by the label "a" in **Figure 1**) and with RS spectral peaks at 1640  $\text{cm}^{-1}$  (assigned to C=C bonds and indicated by the label "b" in **Figure 1**) [22]. These bands originate from the methacry-



**Figure 2.** Variation in water content of X1000P hydrogel with photocrosslinking time. Gel samples were crosslinked for 4, 6, 8, 10 and 15 min. Irradiated gels (circles) and swollen gels (squares) as a function of photocrosslinking time ( $n = 3$  per time sample).

late groups, which form covalent bonds through an addition reaction during the polymerization. In the addition reaction, the alkene functionality is converted to an alkane functionality. Therefore, the number of C=C bonds can be used as a measure of the degree of crosslinking of the polymer meshwork. In FTIR, the C=C peak overlapped a peak from water, which restricted quantification of the relative number of C=C bonds to dry gels. Therefore we used Raman spectroscopy (RS) to quantify C=C bonds in the wet gel, due to the low Raman scatter from bulk water.

### 3.2.1. FTIR Analysis

The FTIR spectra showed that the C=O peak had shifted when comparing crosslinked PEG-DMA hydrogels with the PEG-DMA monomer, see **Figure 3(a)**. The untreated dry PEG-DMA had a C=O peak at  $1716\text{ cm}^{-1}$ , and with the addition of PBS, the C=O peak was still at  $1716\text{ cm}^{-1}$ . After 15 min UV irradiation the resulting hydrogel sample showed a shift to C=O peak at  $1728\text{ cm}^{-1}$  indicating a change in the chemical environment in the vicinity of the C=O bond. The PEG-DMA monomer contains C=O bonds conjugated with the adjacent C=C bonds. In the crosslinked gel the C=C bond was converted by addition polymerization to a C-C bond and the C=O bonds are no longer conjugated giving rise to the observed shift in agreement with previous observations [23]. Since 15 min of photocrosslinking, shifted the C=O peak cleanly to  $1728\text{ cm}^{-1}$  without any residual peak remaining at  $1716\text{ cm}^{-1}$ , this indicates that virtually all the PEG-DMA monomers are used up in the reaction and effectively no C=O bonds conjugated with C=C bonds remain.

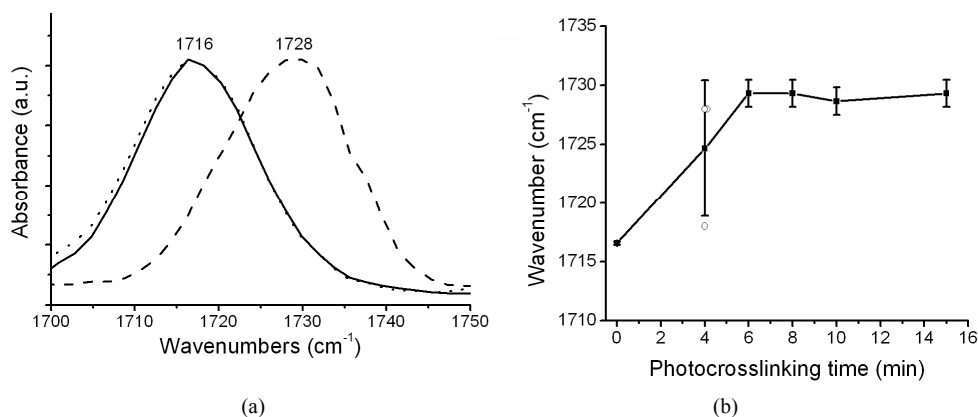
Examining the C=O peak shift as a function of UV irradiation time, we found that for photocrosslinking times

exceeding 6 min the C=O peak is shifted from  $1716\text{ cm}^{-1}$  to  $1728\text{ cm}^{-1}$ , and that the turning point is after 4 min photocrosslinking, where the large standard deviation reflects that some samples have a C=O peak at  $1728\text{ cm}^{-1}$  and others at  $1718\text{ cm}^{-1}$ , see **Figure 3(b)**. This indicates that the polymerization reaction once started occurs rapidly. Either there was no crosslinking and the C=O was at  $1716 - 1718\text{ cm}^{-1}$  or the crosslinking reaction was complete and the C=O peak was at  $1728 - 1730\text{ cm}^{-1}$ . Photocrosslinking times  $< 2$  min were not sufficient to peak cause any crosslinking of the gel (results not shown).

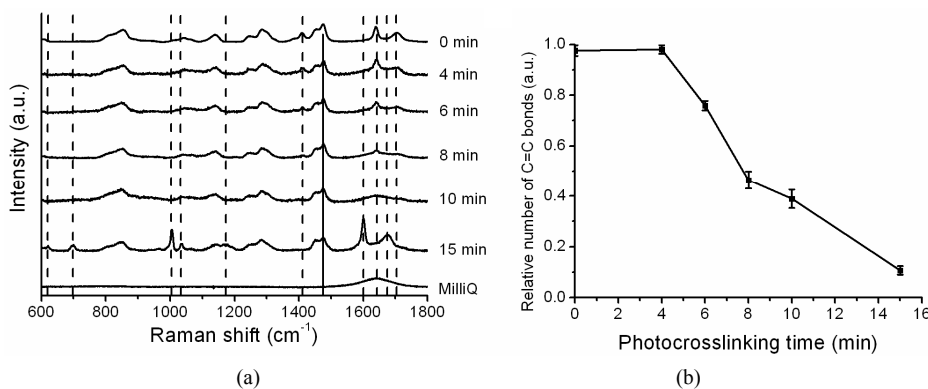
### 3.2.2. RS Analysis

In order to follow the C=C bond formation in the reaction we analyzed how the RS signature of the wet hydrogel changes with photocrosslinking time, see **Figure 4(a)**. Raman peaks at  $1410\text{ cm}^{-1}$ ,  $1640\text{ cm}^{-1}$  (interpreted as stretching of C=C bonds) and  $1705\text{ cm}^{-1}$  decrease in intensity as the photocrosslinking time increases and disappear entirely at a photocrosslinking time of 15 min. Examining the C=C peak with RS, we found that at  $t = 4$  min, virtually all of the original C=C bonds remain, see **Figure 4(b)**. After 6 minutes of photocrosslinking, 25% of the C=C bonds are used up in the crosslinking reaction. The relative number of C=C bonds decreases linearly with increased photocrosslinking time and after 15 minutes, only 10% of the C=C bonds remain. RS measurements showed a linear decrease in the number of C=C bonds, and therefore an increase in crosslinking density, with photocrosslinking time. This correlates with the linear increase in water uptake of the gel, which suggests that the increased water uptake is due to a difference in crosslinking density of the gel.

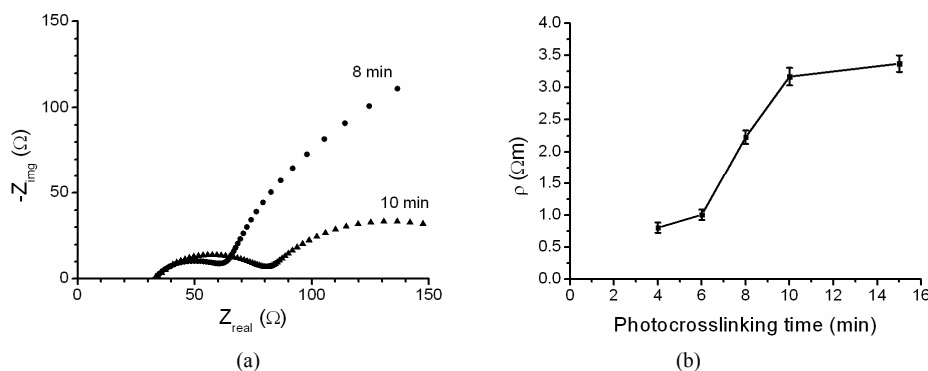
At a photocrosslinking time of 15 min, new Raman



**Figure 3.** Peak shift of the C=O peak due to crosslinking obtained by FTIR. (a) FTIR C=O peaks of PEG-1000-DMA wax (solid), 65 mM PEG-1000-DMA in PBS (dotted) and 65 mM PEG-1000-DMA and 10 mM Darocur in PBS crosslinked with ultraviolet light (X1000P) for 15 min (dashed) are shown. Photocrosslinking the gel results in a peak shift of circa  $12\text{ cm}^{-1}$ ; (b) Effect of photocrosslinking time on FTIR spectra of the X1000P hydrogel. The wavenumber of the C=O peak for X1000P hydrogels with different photocrosslinking times ( $n = 3$ ). Circles indicate the wavenumber of each sample for  $t = 4$  min. Samples were dried in a vacuum dessicator prior to analysis in order to reduce the contribution from water in the recorded spectrum.



**Figure 4.** Spectral fingerprints of X1000P hydrogel as a function of photocrosslinking time obtained by Raman spectroscopy. (a) Raman spectra were normalized to make the peak at 1470 cm<sup>-1</sup> (attributed to H-C-H bending, indicated with a solid line) have equal intensity and then the spectra were offset for clarity. Dashed lines indicate Raman peaks of the gel that change with increased photocrosslinking time; (b) The relative number of C=C bonds in the hydrogel as a function of photocrosslinking time ( $n = 3$ ).



**Figure 5.** Gel impedance (a) and specific electrical resistance,  $\rho$ ; (b) of X1000P hydrogel as a function of the photocrosslinking time ( $n = 3$ ).

peaks appear at 618 cm<sup>-1</sup>, 700 cm<sup>-1</sup>, 1003 cm<sup>-1</sup>, 1035 cm<sup>-1</sup>, 1174 cm<sup>-1</sup>, 1600 cm<sup>-1</sup> and 1676 cm<sup>-1</sup>, see **Figure 4(a)**.

The appearance of Raman peaks at 15 min is likely to be due to reaction end products from the Darocur1173 initiator used in the photocrosslinking reaction. This initiator is split by UV light into a benzoyl radical and an alkyl radical and one of the possible reaction end products is benzoic acid [24]. Benzoic acid Raman peaks that match closely to those observed at 15 min crosslinking are at 618 cm<sup>-1</sup>, 708 cm<sup>-1</sup>, 1035 cm<sup>-1</sup> and 1165 cm<sup>-1</sup> [25] and the Raman peaks at 1003 cm<sup>-1</sup> and 1600 cm<sup>-1</sup> are attributed to C=C aromatic ring stretching from other biproducts of the benzoyl radical. The reaction end products from the benzoyl radical are yellow and those from the alkyl radical generally evaporate during the polymerization process. Although no yellowing of gels was observed, the appearance of additional Raman peaks after 15 min crosslinking indicates that 10 min photocrosslinking is optimal in order to avoid contamination of the gel with reaction end products.

### 3.3. Hydrogel Specific Electrical Resistance

A physical parameter of great interest in determining sample changes is the specific electrical resistance, which can be determined from impedance spectroscopy (IS) measurements using equivalent circuits as models [26, 27]. We collected the impedance plots for the hydrogel at different photocrosslinking times, and clear differences could be observed as shown in **Figure 5(a)**. The fitting of the impedance data by using a non-linear program [28] allows determination of the electrical resistance of the different gel samples ( $R_G$ ), which can be converted to specific electrical resistance,  $\rho$ , using  $\rho = R_G A/l$ , where  $A = 177 \text{ mm}^2$  is the surface area of the gel sample and  $l = 2.2 \text{ mm}$  is the thickness of the gel. Specific electrical resistance values obtained for gel samples are shown in **Figure 5(b)**. The electrical resistance of the gels ranged from 0.7  $\Omega\text{m}$  for 4 min photocrosslinking to 3.5  $\Omega\text{m}$  for 15 min photocrosslinking. For comparison, PBS has a resistivity of 8.4  $\Omega\text{m}$  (Bäckström & Hélix-Nielsen, unpublished results) and deionized water a resistivity of  $1.8 \cdot 10^5 \Omega\text{m}$  [29].

This showed that the gels had a low resistivity (and a high conductivity), making them suitable for use in ion channel-containing hydro gel-encapsulated membranes for biosensor applications. An increase in the specific electrical resistance as a result of an increase in the photocrosslinking time was obtained, which correlated with an increase in the crosslinking density of the gel network. It is remarkable that an increase in photocrosslinking increased the water absorption of the gel, while also increasing its resistivity. We attribute the increased gel resistance to the presence of the hydrophobic initiator Darocur consistent with previous work [16].

#### 4. Conclusion

We show that the properties of a PEG-DMA hydrogel formed by photoinitiated polymerization can be tailored by varying the photocrosslinking time. The water content and the specific electrical resistance of the gel can be controlled between 50 - 90 wt% and 0.7 - 3.5  $\Omega\text{m}$ , respectively within 4 - 15 min crosslinking time. FTIR spectroscopy revealed a change in C=O vibrations from 1716  $\text{cm}^{-1}$  to 1728  $\text{cm}^{-1}$  upon UV illumination consistent with formation of a polymeric network. RS revealed a corresponding decrease in the relative number of C=C bonds indicating the conversion from alkene to alkane bond upon polymerization. Furthermore RS revealed reaction end products from photoinitiator degradation (likely benzoic acid) after prolonged (15 min) UV illumination. Thus both FTIR and RS should be useful as diagnostic tools to study changes in PEG-DMA hydrogels due to e.g. aging, pH, temperature, or chemical exposure.

#### 5. Acknowledgements

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