Effect of Transient Acid Spikes on Developmental Stages of *Lepomis* Fishes

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**Abstract**

The effects of transient acid spikes on development of *Lepomis* sunfish were studied by combining field work with laboratory studies. *Lepomis* sunfish eggs were collected from rocks on nests in two freshwater ponds and tested for their resistance to laboratory induced transient acid spikes. Fertilized eggs on rocks within the same nest were used for each pH transient experiment. Egg masses on one rock from the nest were used as a control while egg masses on another rock were exposed to transient acid spikes. Various stages of development showed different responses to the acidic effects. The major stages studied were fertilized egg, hatched with attached yolk sac, and free swimming with gills. The acid sensitivity followed development stages with the most acid sensitive stage being free swimming with gills. This fieldwork-laboratory data chain supports earlier field studies, providing the final link in an evidentiary chain showing the effects of transient acid spikes on *Lepomis* hatching profiles in freshwater ponds.

**Keywords**

Episodic Acid, pH, *Lepomis*, Sunfishes, Life Stage

**1. Introduction**

There is a developmental sensitivity to pH for most aquatic organisms. Ocean acidification due to increasing amounts of carbon dioxide is a current and future problem for organisms with the gradual lowering of the ocean pH [1]. Carbon dioxide increases also can affect freshwater systems, but episodic acidity causes more problems with organisms in poorly buffered streams, ponds and lakes. Although, there has been a general reduction in the overall extent of acidic precipitation from several decades ago, episodic acidification of aquatic environments still remains a substantial threat to aquatic wildlife [2] [3] [4]. Many studies have
shown that acidified waters are biologically impoverished in some ways [e.g. [5]-[10]]. Effects of sudden anthropogenic inputs producing prolonged acid conditions have been studied [e.g. [11] [12] [13]] as well as field manipulations demonstrating the biological damage caused by acid conditions [e.g., [14] [15] [16] [17] [18]], although it is not clear whether the damage results mainly from the pH changes or mainly from heavy metals (principally aluminum) that are washed from nearby soil by acid rain [19]. However, studies showing population-level effects of ongoing, routine (though likely anthropogenic) acid input are rare [19] [20].

Studies in our lab employing age-analysis of sagittal otoliths and environmental data loggers have linked abnormalities in age structure of Lepomis sunfishes in Massachusetts ponds to poor buffering capacity [21] and to episodic acid spikes [22] [23]. A critical finding in the evidentiary chain developed through these studies was the under-representation of day-classes that experienced a transient acid spike while undergoing “swim-up”, the transition from the yolk-feeding larval form to a juvenile form dependent on gills and a functioning digestive system. Our assumption that the “swim-up” stage of development is especially vulnerable to the effects of acid conditions is therefore critical to our understanding of how episodic acidity impacts Lepomis populations. To verify this key assumption, and to characterize the sensitivity of early life stages of Lepomis sunfishes to acidity, we exposed sunfish eggs, pre-swim-up larvae and post-swim-up fry to acidified pond water in laboratory conditions.

2. Methods

2.1. Study Organism

The small- to-medium-bodied fishes comprising the speciose Lepomis genus in the perch family (Centrarchidae) are found in most freshwater United States ponds and streams east of the great deserts, and are often important forage for larger game fishes [24] [25]. We studied two Lepomis species, Bluegill (L. macrochirus) and Pumpkinseed (L. gibbosus), that are common in ponds in southeastern Massachusetts. Both species are locally variable and the two species sometimes hybridize [26] [27] and although adults of the two species are readily distinguishable, larvae are nearly identical [28]. In addition, individuals do not take up their species-specific dietary niches until the second year of life [29], so bluegills and pumpkinseeds are both morphologically and ecologically indistinguishable in the early stages of life (i.e., they don’t differ as eggs, pre-swim-up larvae, or post-swim-up fry).

Although adult Lepomis sunfishes are hardy and acid-tolerant [30], they build nests in shallow sunlit waters near pond edges where eggs and fry are likely to be exposed to episodic acid spikes caused by run-off during spring rainstorms, as has been documented by Stallsmith et al. [22]. Male sunfishes beat their tails vigorously to clear organic debris from patches of pond bottom, and females lay
adherent eggs on rocks in the centers of these nests.

2.2. Field Collection

To get the early life stage *Lepomis* needed for our episodic acidification lab experiments, we picked up the moderately large rocks (1 - 5 cm diameter) with adhering eggs from the center of sunfish nests in Maquan Pond, Bristol County, Massachusetts. At the 1 m depth of our collections, eggs on the rocks are easy to see because they develop a yellow color soon after fertilization; to standardize egg age we did not collect clear eggs (very young) or darker eggs (older and showing the developing eyes). We identified the species of eggs on the rocks by identifying the species of the male sunfish guarding the nest, and safe handling of the eggs was easy because only the rocks had to be touched and moved. Rocks from the same nest were stored and transported in individual containers.

2.3. Test Procedures

In the laboratory, we used a dissecting microscope to count the number of eggs on each rock, and then allotted rocks into different life-stage/pH/exposure-duration treatments. To examine the effects of different levels of acidity for different durations on three early-life-stages, the individuals associated with a particular nest rock were exposed as eggs or as pre-swim-up larvae or as post-swim-up fry to a particular acidity (pH = 4.0, 4.5, 5.0, 5.9)—with pH = 5.9 used as a control since the typical average acidity of Maquan Pond on days without rainstorms was 5.9 [31]. Exposure to the simulated “acid spike” could last for 0, 3, 12 or 24 hours—which covers the range of duration of transient acid spikes we have observed in Maquan Pond [22]. The leachates from soils that might appear in a pond experiencing a drop in pH were not present in any treatments.

The pH in 500 cc test containers, measured with an Orion 720 pH meter, was established before eggs, larvae, or fry were introduced and then maintained using 0.2% sulfuric acid and 0.2% sodium hydroxide; no additional buffers were used. During the tests, pH was measured at 15 minute intervals and adjusted if necessary. Fluctuations of pH were within 0.02 above or below desired levels.

Following exposure to acid, the number of survivors in each was counted at 6 hour intervals for several times until it reached a constant; survivors in “no acid” control groups, which had very low mortality, were counted once a day. Test organisms were kept in natural Maquan Pond water (pH = 5.9) before and after their periods of exposure to acid, and dead fish were counted and removed as soon as possible to maintain water quality. All experiments were conducted in a laboratory with constant room temperature (18°C) and 12L:12D illumination schedule.

The experimental design is not fully factorial: some treatment combinations are not represented; since we were assigning rocks rather than individuals to different treatments, sample sizes were not equal, but we made sure that a substantial number of rocks from each nest were placed in control treatments (no acid).
2.4. Analysis

All data were first cast into a Microsoft Excel 2007 spreadsheet which was used to calculate basic descriptive statistics (e.g., sample sizes, sample means, and sample standard deviations). With log-transformed survival percentage as the response variable, four-way Analyses of Variance and subsequent paired comparisons in the SYSTAT 11 package were used to determine whether survival was effected by four independent variables: pH, duration of acid exposure, life stage (eggs vs pre-swim-up larvae vs post-swim-up fry), and species (bluegill vs pumpkinseed).

3. Results

Overall, 3163 early-life-stage Lepomis individuals (2459 eggs, 309 pre-swim-up larvae, and 395 Post-Swim-Up fry) taken from seven different nests were exposed to acid conditions in these experiments; another 1592 control individuals from the same nests were grown to the Post-Swim-Up stage in non-acid conditions. Survival was high in the controls (94.0%); survival for groups exposed to acid water ranged from 85.5% to 0% (data pooled for groups with the same pH, duration of acid exposure, and life-stage). Roughly 75 per cent of the individuals used in this study were bluegills (L. macrochirus) and the remainder was pumpkinseeds (L. gibbosus).

Four-way Analysis of Variance to determine the effect on survival (angular transform of per cent that survived) of species, life-stage, acidity (pH), and duration of exposure to acid, showed no difference in survival of bluegills versus pumpkinseeds (p = 0.274). Since survival of these closely related species did not differ, results for pumpkinseeds and bluegills were pooled for subsequent analyses. The other three variables—life-stage, acidity (pH), and duration of exposure—had statistically significant effects (all p < 0.003; Table 1).

Notably, acidity alone—without attendant heavy metal leachate from soils that might occur in a pond—produced substantial and statistically significant reductions in survival (Table 1; Figure 1). Lower pH produces more mortality (p = 0.002), and mortality also increases with longer exposure (p = 0.000). Although acidity reduced survival for all three early life stages, all stages did not suffer

Table 1. ANOVA table for the effects of species (levels = L. gibbosus, L. macrochirus), life-stage (levels = Egg, Pre-Swim-Up, Post-Swim-Up), pH (levels = 4.0, 4.5, 5.0, 5.9), and duration of acid exposure (levels = 3, 6, 12, 24 hrs) on survival of Lepomis in the laboratory.

<table>
<thead>
<tr>
<th>Source</th>
<th>Sum-of-Squares</th>
<th>df</th>
<th>Mean Square</th>
<th>F-ratio</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>SPECIES</td>
<td>0.135</td>
<td>1</td>
<td>0.135</td>
<td>1.220</td>
<td>0.274</td>
</tr>
<tr>
<td>LIFE-STAGE</td>
<td>3.218</td>
<td>2</td>
<td>1.609</td>
<td>14.518</td>
<td>0.000</td>
</tr>
<tr>
<td>PH</td>
<td>1.901</td>
<td>3</td>
<td>0.634</td>
<td>5.719</td>
<td>0.002</td>
</tr>
<tr>
<td>DURATION</td>
<td>2.668</td>
<td>3</td>
<td>0.889</td>
<td>8.024</td>
<td>0.000</td>
</tr>
<tr>
<td>Error</td>
<td>6.539</td>
<td>59</td>
<td>0.111</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>
To examine the relationship between life-stage and vulnerability more closely, we used a 2-way ANOVA to analyze the effects of life-stage (3 levels) and duration of exposure to acid (4 levels) at pH = 4 (**Table 2, Figure 2**). A non-significant interaction term indicates that all three life stages responded to the combining of acid severity and acid duration in the same way (interaction p = 0.533), with higher mortality at longer exposures (p = 0.001); Post-Swim-Up fry seem to experience the highest mortality, while Eggs appear to be least vulnerable. In fact, the vulnerability of the Post-Swim-Up stage to low pH puts this life stage in a category by itself. Pairwise statistical testing, with Bonferroni adjustments for multiple comparisons, show that survival was significantly lower for Post-Swim-Up fry than for Eggs (p = 0.000) or Pre-Swim-Up larvae (p = 0.008), but the survival of Pre-Swim-Up larvae was not significantly lower than survival of Eggs (p = 0.187).

Exposure to various levels of acidity for different lengths of time showed that Post-Swim-Up fry incur high mortality across a wide range of acid conditions. Although 77 per cent of Post-Swim-Up fry survived 6 hours of exposure to water at pH = 5, at pH = 4 less than 40 per cent of Post-Swim-Ups survived six hours equally (p = 0.000).

**Figure 1.** Mean Per cent survival after 6 hours of exposure to prevailing natural water (pH = 5.9) and to acidic water for Lepomis Eggs (pH = 4, pH = 5), Pre-Swim-Up Fry (pH = 4), and Post-Swim-Up Fry (pH = 4, pH = 5). Error bars are lacking because they are too small to be visible on a graph at this scale. Note that the point at the far right is the “prevailing natural pH control” for all three life stages.
Table 2. ANOVA table for the effects of life-stage (Egg, Pre-Swim-Up, Post-Swim-Up), duration of exposure to pH 4 water (3, 6, 12, 24 hrs), and life-stage/duration interaction on survival of Lepomis in the laboratory.

<table>
<thead>
<tr>
<th>Source</th>
<th>Sum-of-Squares</th>
<th>df</th>
<th>Mean Square</th>
<th>F-ratio</th>
<th>P</th>
</tr>
</thead>
<tbody>
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<td>STAGE</td>
<td>3.188</td>
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<td>1.594</td>
<td>13.395</td>
<td>0.000</td>
</tr>
<tr>
<td>DURATION</td>
<td>2.213</td>
<td>3</td>
<td>0.738</td>
<td>6.202</td>
<td>0.001</td>
</tr>
<tr>
<td>STAGE*DURATION</td>
<td>0.611</td>
<td>6</td>
<td>0.102</td>
<td>0.857</td>
<td>0.533</td>
</tr>
<tr>
<td>Error</td>
<td>5.433</td>
<td>46</td>
<td>0.119</td>
<td></td>
<td></td>
</tr>
</tbody>
</table>

Figure 2. Mean percentage survival after exposure to acid water (pH = 4) for varying lengths of time (0, 3, 6, 12, and 24 hrs) of Eggs, Pre-Swim-Up Larvae and Post-Swim-Up Fry. Error bars (standard errors) are lacking when they are too small to be visible on a graph at this scale. Note that the point at the far left is the “prevailing natural pH control” for all three life stages.

of exposure—and less than 10 per cent survived 12 hours (Figure 3). Following heavy rainstorms, we have measured acid events of comparable intensity and duration in the shallow breeding area where the experimental organisms were collected.

4. Discussion

The biological effects of acidic events have been attributed to low pH per se and also to the heavy metals, especially Aluminum, that acid rain washes out from the soil [32]. Malcolm et al. [33] found that the concentration of labile aluminum was the best predictor of the presence of brown trout (Salmo trutta) fry for 22 sites in the UK that had been monitored for 22 years. Evidence from many sources indicates that acid and aluminum are an especially toxic combination. Duis & Oberemm [11] found that long-term exposures to very low pH and high heavy metal concentrations produced very high levels of mortality in pike (Esox lucius), Delonay et al. [34] found that smaller increases in acidity led to significant
mortality of early-life-stage golden trout (*Onchorhyncus aquabonita*) when aluminum was also added, and Ormerod *et al.* [35] found that adding aluminum and acid to a Welsh stream increased the mortality of salmonids by at least a factor of five, compared to acid alone. Palmer *et al.* [30] found in laboratory toxicity tests that adult bluegills (*Lepomis macrochirus*) were also killed after 4 days of exposure—but only with both very high concentrations of aluminum concentration and very low pH. Our study does not resolve the issue of whether the effects of pH versus aluminum are stronger, but it establishes that brief exposure to low pH alone can produce substantial mortality in early life stages of sunfishes.

In addition, our results are consistent with those of Bilk’ko [36], who found that brief exposure to low pH killed eggs of roach (*Rutilus rutilus*) and bream (*Abramis brama*). Jellyman & Harding [37] also found that while 14 days of exposure to pH 4.5 did not affect survival of adults of five New Zealand fishes ((redfin bully *Gobiomorphus huttoni*, *inanga* *Galaxias maculatus*, brown trout *Salmo trutta*, longfin eel *Anguilla dieffenbachii* and koaro *Galaxias brevipinnis*), larval koaro incurred substantial mortality after just 3 days. Malcolm *et al.* [33] also found in brown trout (*S. trutta*) that fry were more sensitive than parr to hydrochemical conditions in headwater streams recovering from acidification. Unlike Geffen [38], who found that reducing pH for two days does not by itself effect mortality of rainbow smelt eggs (*Osmerus mordax*), we found that acid exposure by itself did cause mortality of *Lepomis* eggs, but less mortality than in
the other early life stages. Our laboratory experiments verify that transient acid spikes that are commonly associated with rainfall can produce recruitment failure of sunfishes on a day-class scale.

In a review of the effects of acidification on developing fishes, Sayer et al. [39] noted the heightened vulnerability associated with swim-up: “There is a great increase in vulnerability once dependence on the yolk has come to an end. This period coincides with the movement from the incubation site in the substratum into the open water, where the relevant water chemistry is likely to fluctuate between greater extremes. In waters where acidification is most likely to occur, the early life stages of many freshwater fish species will be close to their survival threshold and only a slight decline in water quality might result in the loss of a complete year class…” By examining sunfishes at several different scales, we have confirmed this prediction. Our finding that post-swim-up fry are significantly more likely to be killed by transient exposure to acidity than other life stages is consistent with the findings of Reader et al. [40] and Serrano et al. [41] that brown trout (Salmo trutta) become much more susceptible to episodic exposure to acid plus aluminum after the yolk-sac has been absorbed.

Most importantly, this finding completes the evidence showing that the episodic pH changes produced by acid rainstorms cause biological damage, by corroborating the conclusion of Stallsmith et al. [22] that day-class gaps in the age distribution of young-of-the-year Lepomis collected at Maquan Pond in Plymouth County, MA resulted from the mortality inflicted on Post-Swim-Up fry by transient acid spikes (pH < 5.3) associated with rainstorms during the years 1990-1992. They found that day-class gaps were significantly associated with the occurrence of acid spikes when the day-class was at Swim-up (Table 3 presents

<table>
<thead>
<tr>
<th>Year</th>
<th>Acid Spike</th>
<th>No Acid Spike</th>
<th>Total</th>
<th>$\chi^2$</th>
<th>P</th>
</tr>
</thead>
<tbody>
<tr>
<td>1990</td>
<td></td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Day-class Gap</td>
<td>7</td>
<td>5</td>
<td>12</td>
<td>4.69</td>
<td>0.018</td>
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<tr>
<td>No Day-Class Gap</td>
<td>3</td>
<td>17</td>
<td>20</td>
<td></td>
<td></td>
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<tr>
<td>1991</td>
<td></td>
<td></td>
<td></td>
<td></td>
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<tr>
<td>Day-class Gap</td>
<td>0</td>
<td>4</td>
<td>4</td>
<td>0.01</td>
<td>&gt;0.50</td>
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<tr>
<td>No Day-class Gap</td>
<td>0</td>
<td>12</td>
<td>12</td>
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<td></td>
</tr>
<tr>
<td>1992</td>
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<td></td>
<td></td>
</tr>
<tr>
<td>Day-class Gap</td>
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<td>17</td>
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<tr>
<td>1990-1992 TOTAL</td>
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<td></td>
<td></td>
<td>11.94</td>
<td>0.000</td>
</tr>
<tr>
<td>Day-class Gap</td>
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<td>11</td>
<td>21</td>
<td></td>
<td></td>
</tr>
<tr>
<td>No Day-class Gap</td>
<td>4</td>
<td>45</td>
<td>49</td>
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</tbody>
</table>
these data as an erratum to a misprinting in the 1996 report).

5. Conclusion

In the *Lepomis* sunfishes of Plymouth County, otolith studies revealed under-represented year-classes in poorly buffered ponds [21]. Otolith studies on young-of-the-year sunfishes revealed poor recruitment of day-classes associated with transient acid spikes during Swim-Up [22], and the present study links the recruitment failures of day-classes to the high mortality of Swim-Up individuals exposed to acid in controlled conditions. Based on year-to-year associations between acidic inputs and year-class abundances, Hudd [19] has also inferred that episodic acidification produces reduced recruitment of estuarine fishes (burbot *{Lota lota}*, perch *{Perca fluviatilis}*, and smelt *{Osmerus eperlanus}*). For our case of pond-living sunfishes, a complete inferential chain of causation has been established—from transient acid spikes to population abnormalities in age structure indicating recruitment failure—before the biological effects of acid conditions seriously reduced or eliminated the populations.

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References


