

# Disposal and Treatment Methods for Pesticide Containing Wastewaters: Critical Review and Comparative Analysis

Mariam T. Al Hattab, Abdel E. Ghaly

Department of Process Engineering and Applied Sciences, Faculty of Engineering, Dalhousie University, Halifax, Canada.  
Email: [abdel.ghaly@dal.ca](mailto:abdel.ghaly@dal.ca)

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## ABSTRACT

Pesticides provide the primary means for controlling organisms that compete with man for food and fibre or cause injury to man, livestock and crops. They played a vital role in the economic production of wide ranges of vegetable, fruit, cereal, forage, fibre and oil crops which now constitute a large part of successful agricultural industry in many countries. After application to the target areas, pesticide residues are removed from applicators by rinsing with water which results in the formation of a toxic wastewater that represents a disposal problem for many farmers. Pesticides can adversely affect people, pets, livestock and wildlife in addition to the pests they are intended to destroy. The phenomenon of bio-magnification of some pesticides has resulted in reproductive failure of some fish species and egg shell thinning of birds such as peregrine falcons, sparrow hawk and eagle owls. Pesticide toxicity to humans include skin and eye irritation and skin cancer. Therefore, care must be exercised in the application, disposal and treatment of pesticides. Currently, disposal of pesticide wastewater is carried out by: 1) land cultivation, 2) dumping in soil pits, plastic pits and concrete pits or on land and in extreme cases in streams near the rinsing operation, 3) use of evaporation beds and 4) land filling. These methods of disposal are unsafe as the surface run off will reach streams, rivers and lakes and the infiltration of the wastewater into the local soil will eventually reach ground water. The treatment methods currently used for pesticide wastewater include: 1) incineration (incinerators and open burning), 2) chemical treatments ( $O_3/UV$ , hydrolysis, Fenton oxidation and KPEG), 3) physical treatments (inorganic, organic absorbents and activated carbon) and 4) biological treatments (composting, bioaugmentation and phytoremediation). Therefore, the choice of safe, on farm disposal techniques for agricultural pesticides is very important. A comparative analysis was performed on 18 methods of pesticide disposal/treatment using six criteria: containment, detoxification ability, cost, time, suitability for on farm use, size and evaporation efficiency. The results indicated that of the 18 methods evaluated, 9 scored above 80/100 and can be used on farm. They were organic absorbents (97), composting (94), bioaugmentation (92), inorganic absorbents (90), Fenton oxidation (86),  $O_3/UV$  (83), activated carbon (82), hydrolysis (82), and land cultivation (80). The other methods are not suitable for on farm use as they suffered from containment problems, high cost and variability of effectiveness.

**Keywords:** Pesticides; Health; Soil; Water; Disposal; Treatment; Land Cultivation; Disposal Pits; Evaporation Ponds; Landfills; Incineration;  $O_3/UV$ ; Fenton Oxidation; Hydrolysis; Bioremediation; Bioaugmentation

## 1. Introduction

Pesticides provide the primary means for controlling organisms that compete with man for food and fibre or cause injury to man, livestock and crops. They are classified based on the pest they control as shown in **Table 1**. Pesticide expenditures account for 13% - 22% of total costs of production per hectare. The worldwide pesticide consumption and expenditures in 2007 (**Table 2**) were 2.37 billion kg and 39.4 billion dollars, respectively [1]. **Table 3** illustrates the top 10 countries applying higher rates (kg/hectare) of pesticides [2]. Ten companies (**Table 4**) account for 87.2% of the total sales [3].

Pesticides played a vital role in the economic produc-

tion of wide ranges of vegetable, fruit, cereal, forage, fibre and oil crops which now constitute a large part of successful agricultural industry in many countries. They lower crop losses, increase revenue to farmers from the additional marketable yield obtained with their use and thus lower the cost of production per unit output [4]. Other benefits include: 1) reduced uncertainty of crop loss from pests, 2) increased profit to farm input suppliers (machinery, fertilizer, chemicals and seed companies) from increased sale, 3) benefit to consumers through decreased price of raw foods or improved quality of food products and 4) benefit to society as whole (farmers, consumers, farm suppliers, food processors) from increased employment opportunities and expanded export

**Table 1. Common types of pesticides.**

Pesticide	Pest to be controlled
Insecticide	Insects
Herbicide	Undesirable plants
Rodenticide	Rats, mice and other rodents
Nematicide	Nematodes
Fungicide	Fungal diseases
Acasicide	Mites and spiders
Bactericide	Bacteria

**Table 2. 2007 worldwide consumption and expenditures of pesticide active ingredients [1].**

Pesticide	Consumption (10 <sup>6</sup> kg)	Expenditures (10 <sup>6</sup> \$)
Herbicides	955	15,512
Insecticide	405	11,158
Fungicide	235	9216
Other*	775	3557
<b>Total</b>	<b>2370</b>	<b>39,443</b>

\*Other includes mematicide, fumigants and other miscellaneous.

**Table 3. Top 10 countries applying pesticide at higher rates in 2000 [2].**

Country	Application Rate (kg/ha)
Costa Rica	51.2
Colombia	16.2
Japan	12.0
Netherlands	9.4
Korea	6.6
Ecuador	6.0
Portugal	5.3
France	4.6
Greece	2.8
Uruguay	2.7

**Table 4. Top 10 pesticide companies in 2007 [3].**

Company	Country	Sales (10 <sup>6</sup> \$)	Market Share (%)
Bayer	Germany	7458	19.0
Syngenta	Switzerland	7285	18.5
BASF	Germany	4297	10.9
Dow Agro Science	USA	3779	9.6
Monsanto	USA	3599	9.1
Du Pont	USA	2369	6.0
Makhteshim Agan	Israel	1895	4.8
Nufarm	Australia	1470	3.7
Sumitomo Chemical	Japan	1209	3.0
Aystra Life Science	Japan	1035	2.6
<b>Total</b>	<b>N/A</b>	<b>34,396</b>	<b>87.2</b>

Sales are in millions of dollars; Total Worldwide Sales = 39443.

of food products [5,6]. The benefit/cost ratio vary from 4 to 33 (for every dollar spent on pesticide farmers receive an additional \$4 - 33 in revenue) depending upon crop rotation and year [4,7].

After pesticides are applied to the target areas, pesticide residues remain in containers and application equipment. These residues are removed from applicators by rinsing with water which results in the formation of a toxic wastewater that can adversely affect people, pets, livestock and wildlife [8-10]. The resulting ecological impact of unsafe disposal of pesticides can be severe depending on the type of pesticide and the amount contained in the wastewater. The phenomenon of biomagnification of some pesticides has resulted in reproductive failure of some fish species [11,12] and egg shell thinning of birds such as peregrine falcons, sparrow hawk and eagle owls [13]. Pesticide toxicity to humans include skin and eye irritation and skin cancer [14]. Therefore, care must be exercised in the application, disposal and treatment of pesticides. Currently, disposal of pesticide wastewater is carried out by several methods including: 1) land cultivation, 2) dumping in soil pits, in ditches, in lagoons, on land, and in extreme cases in sewers and streams near the rinsing operation, 3) use of evaporation pond and 4) land filling. These methods of disposal are totally unsafe. The surface run off will reach streams, rivers and lakes and the infiltration of the wastewater into the local soil will eventually end up in the ground water. The treatment methods currently used for pesticide containing wastewater include: 1) incineration, 2) chemical treatment, 3) physical treatment and 4) biological treatment. These treatment methods either require land or are expensive and suffer from variability of effectiveness [15]. Thus, the development and selection of safe, on farm disposal/treatment technique for agricultural pesticides is paramount.

The aim of this study was to review the current methods of disposal and treatment of pesticides and to perform a comparative analysis to determine the most appropriate method for on farm use.

## 2. Pesticide Disposal Methods

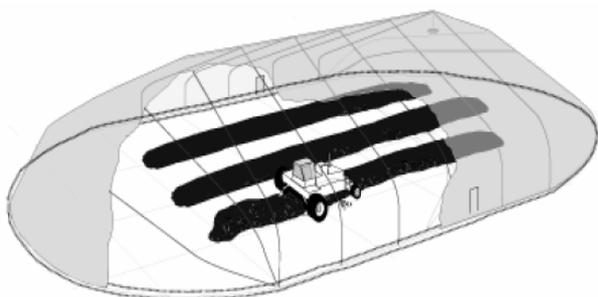
The methods for the disposal of low level pesticides (Table 5) include: land cultivation, disposal pits, evaporation ponds and landfills [16-20]. There are three types of disposal pits: soil pit, plastic pit and concrete pit.

### 2.1. Land Cultivation

In this method, excavated contaminated soil is spread out in a thin layer on uncontaminated soil (Figure 1) in order to allow for natural chemical and biological processes to transform and degrade the contaminants. Soil contains microbes (fungi, algae and bacteria) capable of

**Table 5. Disposal methods of pesticide containing wastewater.**

Method	Description	Advantages	Disadvantages
Land Cultivation	Place liquid wastes in plow zone of soil for subsequent weathering	On-site use Simple technology	Land requirements Possible runoff and leaching Slow and variable decomposition Restricted vegetation
Disposal Pits	Place liquid wastes in pits containing soil and open to air for subsequent weathering	On-site use Simple technology Secure containment	Slow decomposition Limited lifetime of pit Effectiveness varies with climate
Evaporation Ponds	Place liquid wastes in lined ponds open to air for subsequent weathering	On-site use Simple technology Secure containment	Slow decomposition Limited lifetime of pond effectiveness Varies with climate
Landfills	Burial of wastes in soil	Generally available Complete removal	Land requirements High transportation costs Possible runoff and leaching

**Figure 1. Land cultivation of contaminated soil [21].**

metabolizing pesticides [22,23]. The ability of bacteria to metabolize pesticides has been well documented by several researchers. Bhadhade *et al.* [24] reported that soil bacteria was capable of degrading 83% - 93% of the organo-phosphorous pesticide monocrotophos. Ohshiro *et al.* [25] reported a 96% reduction in isoxathion from the organophosphorous pesticide by bacteria isolated from turf green soil. Kearney *et al.* [26] reported that soil microbes were capable of degrading 90% of the alachlor pesticide within 30 - 40 days. Tang and You [27] reported that the triazophos bacteria was capable of degrading 33.1% - 95.8% of pesticides in soil.

Racke and Coats [28] reported that after soil has been treated with a pesticide a few times its microorganisms build up a need for that pesticide which results in fairly rapid degradation of any additional applications. Schoen and Winterlin [29] stated that natural soil degradation is effective when low concentrations of the pesticide are present, but with high concentrations of pesticide it becomes much more difficult to degrade. Felsot [30] stated that land cultivation is only effective for compounds that can be biotransformed or biomineralized by soil microbes. Somasundaram *et al.* [31] reported that the ability of soil microbes to degrade certain pesticides is affected by pesticide toxicity to soil microbes that are responsible for the degradation. Felsot *et al.* [22] stated that land cultivation

is effective if the pesticide is degraded at the same or faster rate than it is applied to the field. Felsot *et al.* [32] noted that land cultivation can be enhanced by the addition of organic amendments such as sewage sludge.

## 2.2. Soil Pit

A primary method for disposing of liquid pesticide waste is by dumping it in an unlined soil evaporation pit (**Figure 2**), usually  $15 \times 15 \times 1$  m [33]. Schoen and Winterlin [29] reported that factors such as chemical structure and concentration of pesticide play a major role in the degradation of pesticides in soil pits. Gan and Koskinen [34] stated that the dissipation of the pesticide decreases as the concentrations of pesticide increases. Dzantor and Felsot [35] and Gan *et al.* [36] noted that high pesticide concentrations may cause microbial toxicity which would inhibit the degradation of the pesticide.

Several researchers [22,34,36,37] noted that the prolonged dissipation of pesticides opens a window for runoff and leaching, especially at higher pesticide concentrations. Gan *et al.* [36] reported that 50% dissipation of the alachlor pesticide in soil, at concentrations of 4 and 300 mg/kg took approximately 2 and 52 weeks, respectively. Gan *et al.* [38] noted that atrazine pesticide took approximately 4 and 24 weeks to be dissipated to half the concentration of 7 and 6400 mg/kg in soil, respectively. Schoen and Winterlin [29] noted that captan, trifluralin and diazinon at concentrations of 100 mg/kg took 1 - 2, 116 - 189 and 77 - 160 weeks to dissipate to half the concentration while captan, trifluralin and diazinon at concentrations of 1000 mg/kg took 30 - 48, 168 - 544 and 77 - 160 weeks to reach 50% disappearance in soil, respectively.

## 2.3. Plastic Lined Pit

This method for disposal of pesticide waste requires proper selection of the site to avoid leaching and runoff.

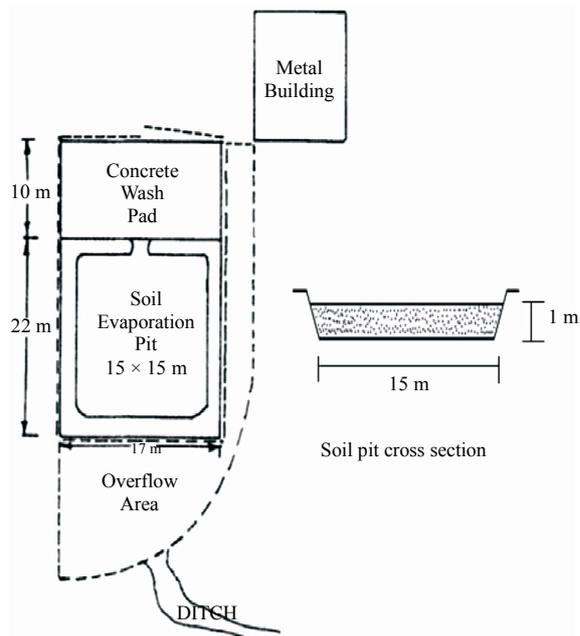


Figure 2. A soil pit for disposal of pesticide water [33].

The site should be in an area where there is no danger of contaminating dwellings groundwater sources and surface water used for crop and livestock production. The pit should be on a levelled ground with a depth of 0.5 - 1 m covered with a plastic liner and a layer of soil is laid on top of the liner (Figure 3). The pit should be open to the atmosphere in order to allow for water evaporation into the atmosphere. A roof cover will prevent the water level from raising due to rain or snow. The wastewater is pumped into the pit for pesticide biodegradation by soil microbes [22]. Hall *et al.* [20] reported that the presence of microbes in the soil water mixture in plastic lined pits was responsible for the degradation of pesticide and no accumulation of pesticide was noted in the pits. Junk and Richard [39] evaluated the effectiveness of 90,000 L polyethylene lined disposal pit with over 150 kg of 25 different types of pesticides for over 2 years and concluded that this method was in fact effective for disposal of pesticide waste with insignificant release to air and water surroundings.

Figure 4 illustrates a cross section view of a simple small scale plastic pit used to dispose of pesticide waste. It consists of a plastic drum with a length and width of 75 × 55 cm, respectively. Inside the drum is a mixture of 15 kg of soil and 60 L of water, that was used to treat pesticide waste which was introduced into the system through the inlet. Junk *et al.* [19] used 56 plastic containers filled with 15 kg of soil and 60 L of water to test the degradation of alachlor, atrazine, trifluralin, 2,4-D ester, carbaryl and parathion and found this system not suitable for atrazine but was effective and very rapid for 2,4-D and carbaryl. They concluded that: 1) the plastic container

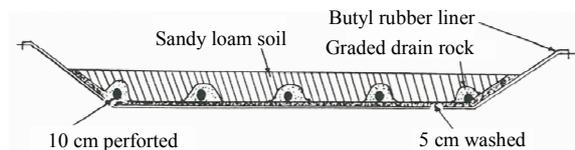


Figure 3. A plastic lined evaporation pit for disposal of pesticide wastewater [16].

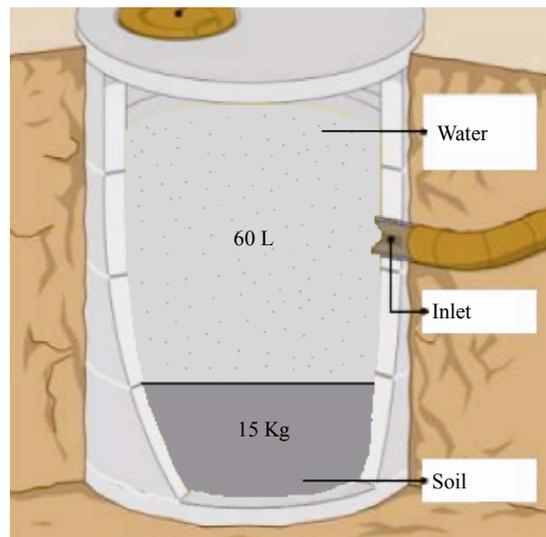


Figure 4. A cross section of a plastic pit for disposal of pesticide wastewater [40].

provided satisfactory containment for most common pesticides, 2) soil was a satisfactory source for microorganisms, 3) aeration and buffers had questionable value, 4) half life concept for degradation was not applicable and 5) sampling from small disposal sites was a problem.

## 2.4. Concrete Pit

Similar to the plastic lined pit, the concrete pit should be on levelled ground with a depth of 0.5 to 1 m, a length of 8 - 10 m and a width of 3.5 m and reinforced with 0.20 m thick concrete walls. The pit consists of a top and bottom layer of gravel that is 4 cm in diameter with the middle layer consisting of topsoil (Figure 5). The pit should also have a cover to prevent rise in water level from rain or snow but remain open to the atmosphere in order to allow for water evaporation.

Johnson and Hartman [41] tested the microbiological activity in a concrete pit and concluded that the degradation process in the pit was effective and no long-term accumulation of pesticide was present. Junk and Richard [39] tested the effectiveness of 30,000 L concrete disposal pit with over 50 kg of 40 different types of pesticides for 8 years and concluded that this method was in fact effective for disposal of pesticide waste with insignificant release to air and water surroundings. Hall [42] tested the effectiveness of an open concrete disposal pit

for the degradation of 45 pesticides over five months and concluded that the biodegradation of the pesticides was successfully accomplished and the pit did not leak or pollute the air, but the system was too large and complicated for most farms.

## 2.5. Evaporation Beds

Lined evaporation beds (**Figure 6**) are used for the disposal of pesticide wastewater. Leach lines underneath the soil surface supply the beds with the pesticide residues from washing equipment. The pesticides rise to the beds surface where they are degraded through photochemical, chemical and biological actions and are distributed via air vapour. Some of the beds have hydrated lime incorporated into the soil in order to aid in the degradation of certain pesticides. A medium scale disposal system of this type costs up to \$50,000 to construct [16,22].

Hodapp and Winterlin [44] reported a reduction in the diazinon pesticide of 62.54% using lined evaporation bed without lime and a degradation of 77.75% with lime. They also reported an ethyl parathion reduction of 69.83% using lime treatment in the beds and a reduction of 45.45% without the use of lime. Winterlin *et al.* [16] tested ten (6 × 12 × 1 m) lined (with a butyl rubbermembrane liner and 36 cm of sandy loam soil) evaporation beds to determine their pesticide decay effectiveness. Pesticide rainsate was introduced through subsurface tiles in limited amounts and the effects of geography, climate and lime application were examined. The method appeared to be beneficial for disposal of some pesticides but not all. Over 100 pesticides were tested, but only 46 were actually detected.

This method for the disposal of pesticide containing wastewaters is advantageous because the beds are economical, little maintenance is required, do not build up high levels of pesticides and are effective in degrading as well as containing the pesticides without excessive exposure through air vapour [16,44]. It is considered an

economical, on-site method of disposal which requires only annual monitoring. The disadvantages appear to be the development of a high concentration of residue in the top layer of the soil and the difficulty in acquiring a representative sample [33].

## 2.6. Land Filling

Landfills are sites that dispose of waste by burial into the soil where microorganisms are used to change the composition of the toxic elements. Landfills for pesticides (**Figure 7**) are equipped with drying pits (**Figure 8**) containing soil to provide the microbes needed to break down the pesticide components into non harmful elements. A nearby sump for the propose of draining and rinsing the containers that have not been fully emptied or rinsed [47,48].

Munnecke [49] reported that soil bacteria were capable of hydrolyzing ethyl parathion found in pesticide container residues within 16 h. Johnson and Lavy [50] reported that carbofuran, thiobencarb and triclopyr buried in degrading containers dissipated to 50% of the initial concentration with the first 94 days or less, while benomyl took 179 - 1020 d before 50% dissipation. They also noted that the rates of dissipation decreased with an increase in soil depth.

Yasuhara *et al.* [51] detected 190 compounds in landfill leachates in Japan. Williams *et al.* [52] reported that the pesticide mecoprop is found in landfill leachate because it is resistant to anaerobic degradation. Christensen *et al.* [53] noted the presence of the pesticide bentazon, N,N-Diethyltoluamide and mecoprop in landfill leachate because of their persistence to anaerobic landfill conditions. Alloway and Ayres [54] noted the presence of the pesticides atrazine and simazine in landfill leachate.

## 3. Pesticide Treatment Methods

The pesticides treatment methods (**Table 6**) include: 1) thermal treatment, 2) chemical treatments, 3) physical

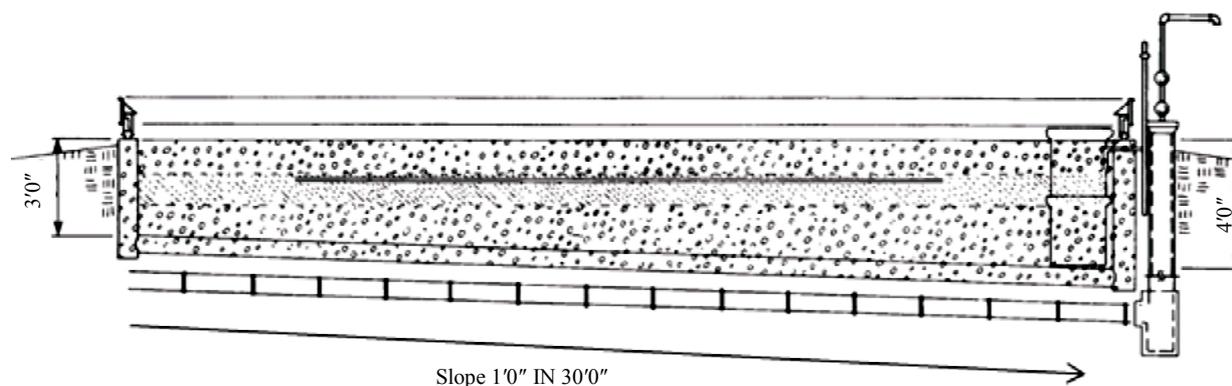


Figure 5. A cross section of concrete pit for disposal of pesticide wastewater [20].



Figure 6. Evaporation beds for disposal of pesticide wastewater [43].



Figure 7. A landfill for disposal of pesticide wastes [45].



Figure 8. The landfill leachate collection pit [46].

treatments and 4) biological treatments. Thermal treatments include incineration and open burning. Chemical treatments include ozonation/UV radiation, Fentonoxidation, hydrolysis and KPEG. Physical treatments are based on absorption using activated carbon, inorganic and organic materials. Biological treatments include composting, phytoremediation and bioaugmentation.

### 3.1. Incineration

Pesticide Incineration is a high temperature oxidation process where the pesticide is converted into inorganic gases (water vapour, CO<sub>2</sub>, volatile acids, particles and metal oxides) and ash [22,54]. Incineration of pesticide should be operated at temperatures higher than 1000°C so that the pesticide can be treated within the first 2 seconds. At such temperatures, smoke production is nil and the generated combustion gases are similar to those generated by wood burning [22]. Temperatures lower than 1000°C can also be used as long as the incineration time of the pesticide does not exceed 2 seconds [23]. However, lower temperatures tend to produce toxic intermediate products [56].

Kennedy *et al.* [57] noted change in the combustion efficiency over the temperature range of 600°C - 1000°C. Ferguson and Wilkinson [55] reported that incineration has 99.99% destruction efficiency at temperatures of 1000°C and a retention time of 2 s in the combustion zone. Steverson [58] reported a 99% destruction efficiency for 16 currently used insecticides and herbicides at temperatures ranging from 200°C to 700°C. Linak *et al.* [59] reported an incineration efficiency of greater than 99.99% for dinoseb. Ahling and Wiberger [60] noted that the incineration of fenitrothion and malathion at temperatures lower than 600°C gave emissions of 1% - 2% of the pesticide amount added and temperatures above 700°C would be required to achieve safe destruction and emission levels.

Table 6. Current treatment methods of pesticide containing wastewater.

Method	Description	Advantages	Disadvantages
Thermal	Controlled combustion of either liquid waste or concentrated residue	Destructive Rapid No by-products	High costs Complex Not useful for some chemical
Chemical	Chemical destruction through use of oxidative, reductive, hydrolytic or catalytic reagents	Destructive Rapid	High costs Complex Variable effectiveness
Physical	Removal of chemicals from wastewater by adsorption and/settling	Rapid Possible on-site use	No destruction involved By-products for disposal
Biological	Use of micro-organisms to destroy chemicals	Destructive	High costs Susceptible to shock Relatively slow Variable effectiveness

Incinerators (**Figure 9**) capable of achieving high levels of destruction are equipped with a combustion chamber, an afterburner, scrubbers and electrostatic filters. Ferguson and Wilkinson [55] reported the following performance standards for incinerating hazardous wastes: 1) the incinerator must achieve a destruction and removal efficiency greater than 99.99% for each of the chemicals present in the waste feed, 2) HCl emissions must not exceed 1.8 kg/h or 1% of the HCl in the stack gas prior to entering any pollution control equipment and 3) the particulate matter emitted must not exceed 180 mg/DSCM when corrected to 7.0% O<sub>2</sub>. The advantages of incineration include: 1) effectiveness in degrading chlorinated organics, 2) destruction efficiency of 99.99% and 3) setup at locations next to plants generating the waste. The disadvantages of incineration technology include: 1) need for sophisticated equipment 2) production of cyanide in the off gas during the incineration of organonitrogen pesticides, 3) too costly and complex, 4) it is intended for centralized large scale disposal and 5) not recommended for inorganic pesticides [22].

### 3.2. Open Burning

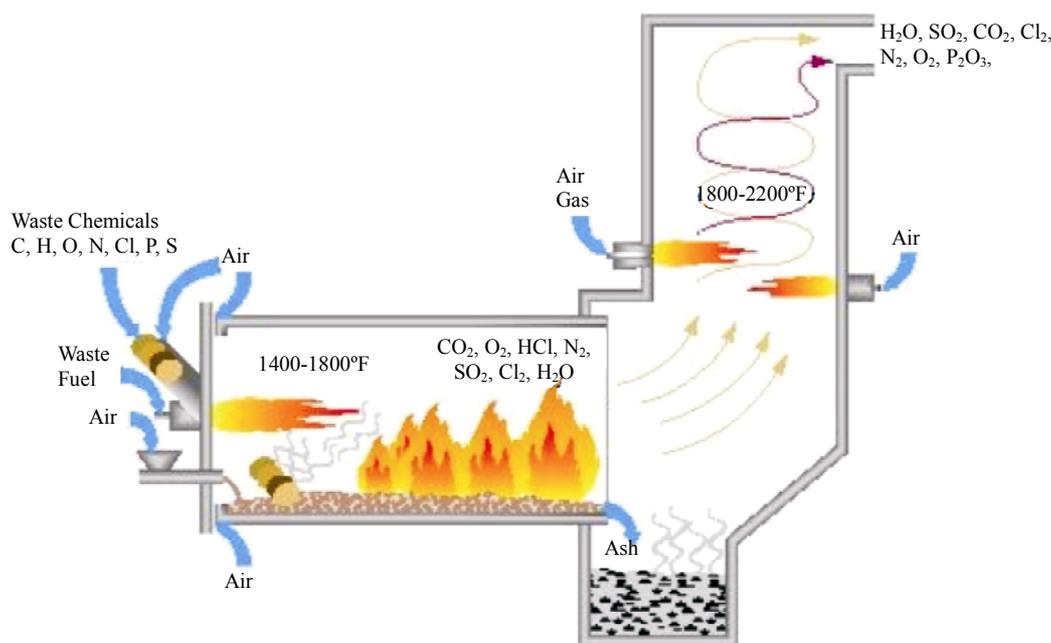
This method combusts pesticides and pesticide waste containers by piling up empty paper and plastic containers and setting them on fire (**Figure 10**). Although this method is inexpensive and convenient, it is hazardous to workers, plants and animals. It is prohibited in some cases by the Regional Air Quality regulations in the US [1]. It emits gases, smoke and fumes into the atmosphere as well as toxic residues that are left in the containers.

Adebona *et al.* [63] noted several products of incomplete combustion, polyaromatic hydrocarbons and low levels of dioxins in open burning tests on 22.7 kg insecticide bags. Oberacker *et al.* [64] noted that after burning bags containing phorate, 2% of the phorate was released into the air and 0.5% remained in the solid residues. Felsot *et al.* [22] reported that bags containing atrazine released 13% of the remaining product into the air while 25% remained as residue. Such results indicate that the temperatures for complete combustion were not reached or were not maintained long enough in order to obtain destruction efficiencies of 99.99% or greater [22].

### 3.3. Ozonation/UV Radiation

The use of ozone and UV radiation to enhance the oxidation of aromatic compounds was investigated by several researchers [17,28,65]. Ozonation is more effective treatment method in the presence of UV light because it can form hydrogen radicals which are very effective oxidizing agents [66,67]. The benefits of this process are its mobility, ease of operation and rapid effects. The disadvantages are its high energy consumption and initial equipment cost [18].

Kuo [68] used a UV/ozonation system (**Figure 11**) consisting of a medium-pressure mercury vapor lamp with a water cooling jacket and an ozone generator. The lamp power consumption was 150 W and was capable of 14.3 W output (at 3.0 mW/cm<sup>2</sup> at a distance of 9 cm). The O<sub>3</sub> was pumped at a rate of 400 mg O<sub>3</sub>/hr/L solution. A solution of 2% KI was used for absorbing the residual ozone from the reactor.

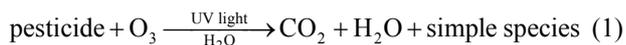


**Figure 9.** An incinerator for pesticide wastes [61].



Figure 10. Open burning of pesticide wastes [62].

Somlich *et al.* [17] noted that irradiation of the alachlor pesticide achieved de-chlorination of the compound, while ozonation works to oxidize the compound into several intermediate products. Under Ultraviolet irradiation, the photon absorption by the carbonyl present in the compound is then followed by the loss of the chlorine. The pesticide degradation reactions that take place under UV/ozonation are as follow [69]:

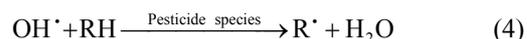


Kearney *et al.* [18] monitored the degradation of alachlor using a UV/O<sub>3</sub> system by measuring the concentration of the 14 CO<sub>2</sub> released. Under UV radiation, the

alachlor pesticide was completely depleted from the water with the presence of oxygen within 25 minutes while it and took 50 minutes before it was fully depleted with ozone alone.

### 3.4. Fenton Oxidation

The Fenton process can be used as part of an oxidative system (Figure 12) to treat and degrade pesticides. It consists of hydrogen peroxide (H<sub>2</sub>O<sub>2</sub>) and iron salts at low pHs [71]. The iron salts act as a catalyst, increasing the effectiveness of the H<sub>2</sub>O<sub>2</sub> by forming highly reductive hydroxyl radicals. The radicals are capable of oxidizing other species that are present in the solution as follows [72].



Hydroxyl radicals are very powerful oxidizing agents with a 2.33 V oxidative potential [72]. The rate of degradation of organic pollutants is strongly accelerated by UV irradiation. The photolysis of the Fe<sup>3+</sup> complexes allows the regeneration of Fe<sup>2+</sup> thus allowing the reaction to proceed much quicker in the presence of H<sub>2</sub>O<sub>2</sub> [73]. The advantages of this method for pesticide treatment are: low cost, ease of operation, simplicity and the wide range of temperature that can be used [72].

Fallmann *et al.* [70] noted a 72% reduction in 100 ppm

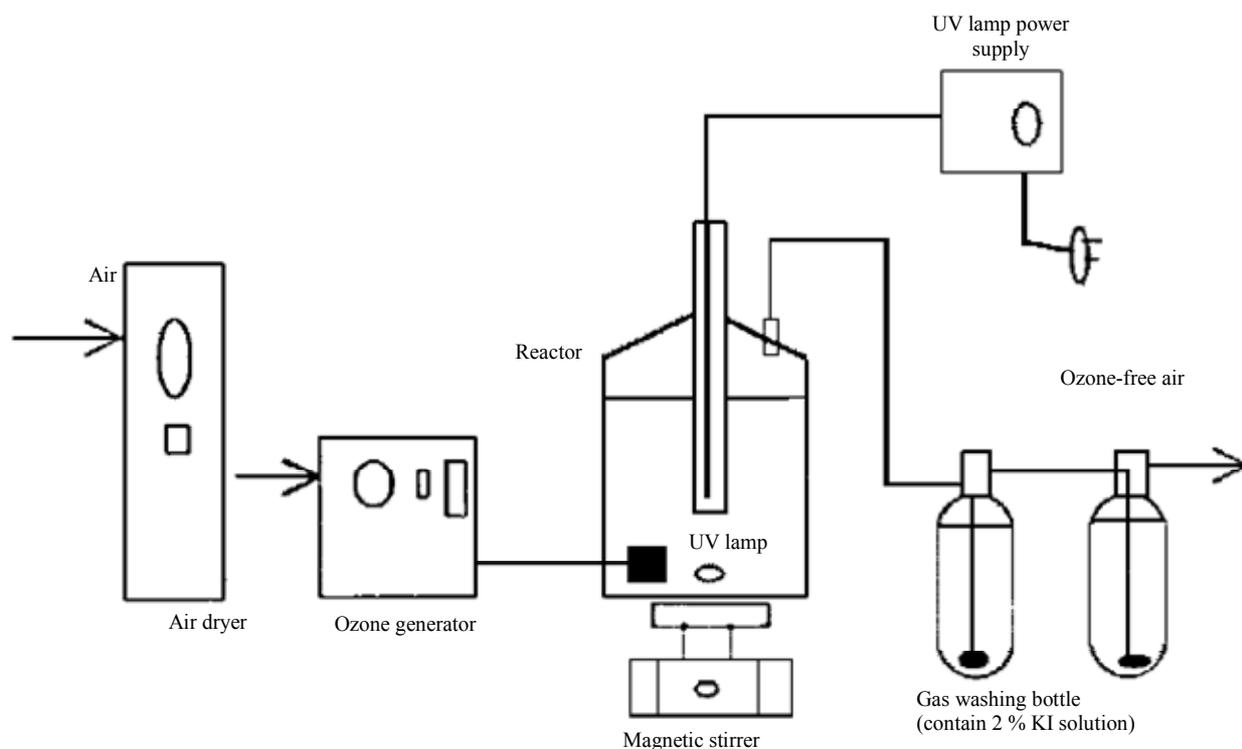
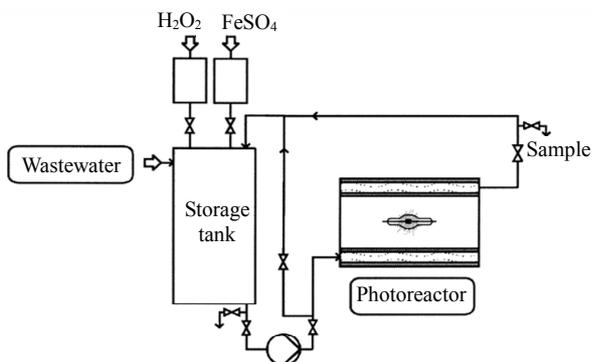


Figure 11. A UV/O<sub>3</sub> system for the treatment of pesticide wastewater [68].



**Figure 12.** A photo assisted Fenton system for treatment of pesticide wastewater [70].

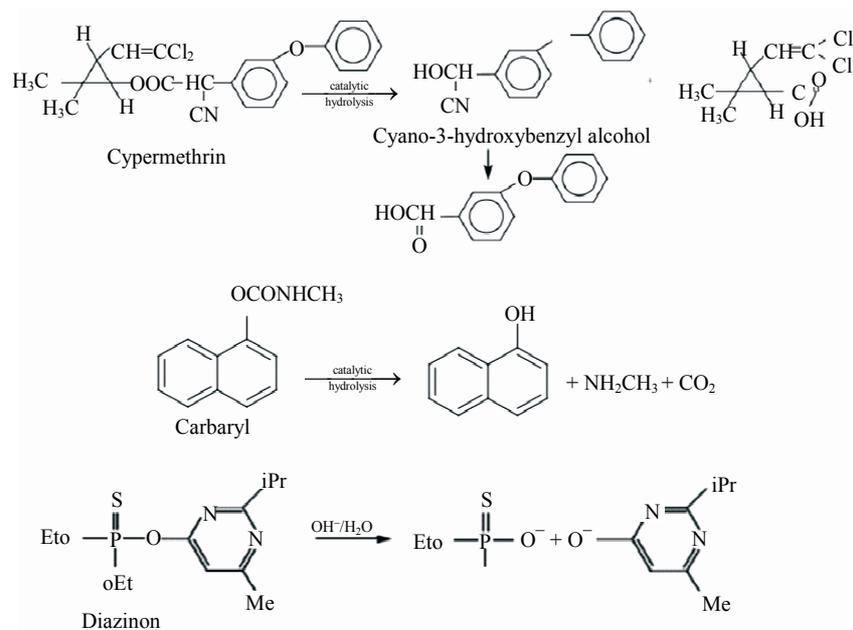
total organic carbon solution using 23 mL of hydrogen peroxide and a reaction time of 124 minutes in a photo assisted Fenton process. Larson *et al.* [74] reported that in the presence of ferric perchlorate and a mercury lamp, the atrazine pesticide had a half-life of less than 2 minutes compared to 1500 minutes when iron salt was not present. Huston and Pignatello [75] noted a half-life of less than 10 minutes for the captan pesticide using UV assisted Fenton reagent at a pH of 2.8. Pignatello and Sun [76] reported a half-life of 2 minutes for methyl parathion using UV assisted Fenton reagent. Doong and Chang [77] reported a half-life of less than 10 minutes for alachlor pesticide under photo assisted Fenton reagent at a pH of 2.8.

### 3.5. Hydrolysis

This method for pesticide treatment works by hydrolyz-

ing the ester linkages found in pesticide compounds, including pyrethroids, carbamates, organophosphates and acetanilides. These compounds can be hydrolyzed in solutions with high pH levels [22]. Desmarchelier [78] used calcium hydroxide for ester hydrolysis and found it to be a safer alternative to sodium and potassium hydroxides for the hydrolysis of fenitrothion pesticide. Lee *et al.* [79] noted that under basic conditions, sodium perborate was more effective in the hydrolysis of organophosphorus than sodium hydroxide, because the peroxide anion released from sodium perborate is much more reactive to organophosphorus than the hydroxyl ion. Qian *et al.* [80] noted an enhancement in the hydrolysis process of mevinphos, diazinon, methyl parathion, malathion and parathion in lake water (10 mg/L) using sodium perborate at pH of 9.88. However, with the presence of soil, the reaction was noted to be significantly slower and the concentration of perborate had to be increased by four folds.

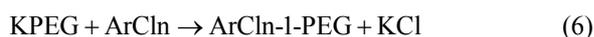
Metal oxide and divalent metal ions have been noted for their ability to catalyze the hydrolysis of organophosphorus insecticides. Smolen and Stone [81] reported that the phosphorothionate insecticides (chlorpyrifos-methyl, zinophos, diazinon, parathion-methyl and runnel) and phosphorooxonates (chlorpyrifos-methyl oxon and paraoxon) were most effectively catalyzed by Copper (II). The downside of catalysis using metal oxides is the formation of products with significant toxicity [22]. Badawi and Ahmed [82] noted that the hydrolysis of the pesticides diazinon, cypermethrin and carbaryl (Figure 13) was effective and accelerated by the addition of a copper (II) ion complex.



**Figure 13.** Hydrolysis of cypermethrin, carbaryl and diazinon pesticides [82].

### 3.6. KPEG

Potassium polyethylene glycol ether (KPEG) is capable of destroying chlorinated pesticides. Chlorinated hydrocarbons and cyclodienes are resistant to degradation by hydrolysis. Dechlorinating these pesticides with KPEG would then enable their biodegradation through land treatment processes. KPEG was found to be capable in dechlorinating polychlorinated biphenyls (PCBs) in soil and solvents [83,84]. In older formulations of phenoxy herbicide, KPEG was found to be capable of degrading dioxins and dibenzofurans [85]. The reaction that takes place consists of a nucleophilic substitution and a phase transfer at the carbon-halogen bond as illustrated by the following equations [22].



where:

PEG = polyethylene glycol monomethyl ether

Ar = aromatic nucleus

Taylor *et al.* [86] reported that the vessel for the KPEG reaction consists of a 55-gal drum (surrounded with heat tape capable of maintaining the temperature at 70°C - 85°C) and an electric motor with a mixer (**Figure 14**). With the reagents KOH and PEG, this vessel was capable of degrading 98% of phenoxy herbicide waste. The generated waste contained in the drums can remain there, eliminating the need for transfer into another container. Vapor emitted from the reaction drums are condensed in a water drum, the remaining condensables are trapped in the scrubber containing sodium hypochlorite solution. Vapors are then passed through an activated carbon adsorbent and as well as a molecular sieve [86].

The materials and chemicals needed for the KPEG process are easy to find [22]. The disadvantage of the KPEG process are 1) high clay content, acidity and high natural organic matter interferes with KPEG reaction and

2) its not recommended for large waste volumes with concentrations above 5% for chlorinated contaminants. If necessary, emissions can be controlled by construction of a vent system with scrubber and absorbent.

### 3.7. Inorganic Absorbents

Pesticide adsorption can be performed using anionic clays (layered double hydroxides) which are simple to prepare [87], hydrotalcite, which occurs in nature may also be used as a layered double hydroxide (LDH). A variety of compounds can be formed by changing the cation metal. In order for a material to be considered as a good adsorbent it must possess the following properties: 1) a granular structure, 2) insoluble in water, 3) chemical stability and 4) have a high mechanical strength [88]. **Figure 15** is multi-functional gravity filter which can be used for various water treatment methods by employing various adsorbent media [89].

Niwas *et al.* [90] reported that styrene supported zirconium (IV) tungstophosphates was successful in adsorbing the pesticide phosphamidan. Inacio *et al.* [86] noted that the inorganic adsorbent  $\text{Mg}_3\text{AlCl}$  was capable of adsorbing the MCPA herbicide within 30 - 45 minutes at room temperature. Boussahel *et al.* [91] noted a removal efficiency in ayanaz in and atrazine of 85% - 90% using  $\text{CaCl}_2$  or  $\text{CaSO}_4$ . Bojemueller *et al.* [92] reported that the pesticide metolachlor can be adsorbed by bentonites and the adsorption efficiency can be doubled by increasing the temperature. Li *et al.* [93] noted that the pesticide glyphosate was adsorbed on the external surface of  $\text{MgAl-LDH}$  at low concentrations, while at high glyphosate concentrations an inter layer ion exchange occurred.

### 3.8. Organic Absorbents

Various organic materials can be used as good adsorbents for pesticide removal. Ahmaruzzaman and Gupta [88] reported that rice husk is insoluble in water, possesses an

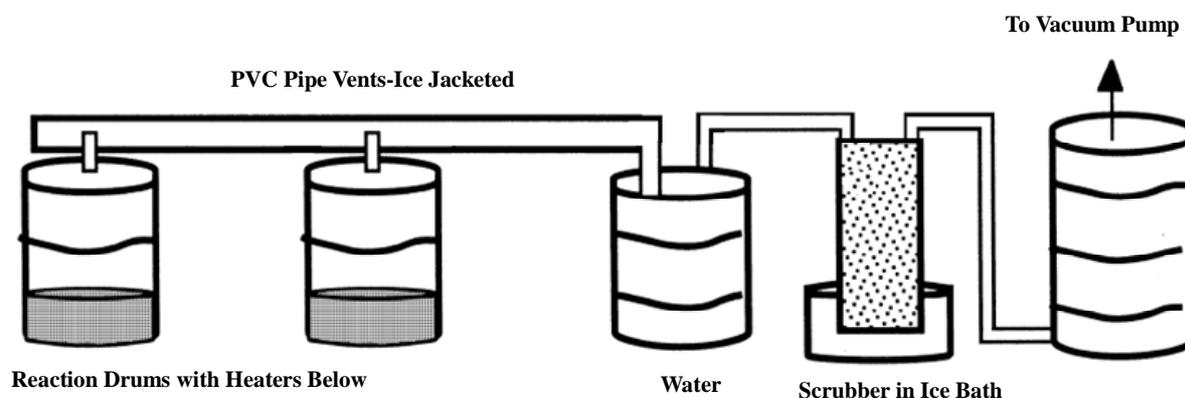


Figure 14. A KPEG process with vent system [86].



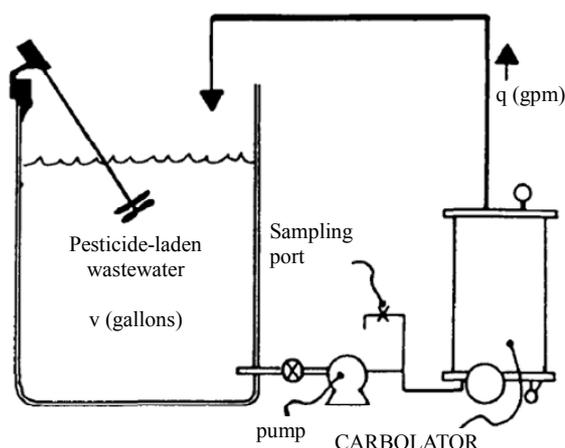
**Figure 15.** Filters used to treat pesticide wastewater through adsorption [89].

irregular granular structure and has a high mechanical strength and chemical stability that make it a good adsorbent. Chowdhury *et al.* [94] noted that treated rice husk was capable of removing 89% - 97% of malachite green pesticide. Akhtar *et al.* [95] investigated the adsorption potential of selected agricultural waste materials (rice, barn, bagasse fly ash from sugarcane and rice husk) for the pesticide removal of methyl parathion from wastewater and reported pesticide removal efficiencies in the range of 70% - 90% within 90 minutes. Memon *et al.* [96] reported that thermally treated watermelon peels were capable of removing 99% of the methyl parathion pesticide. Al hattab and Ghaly [97] reported a captan removal efficiency of 99.2% and 98.5% using hay and soybean plant residues, respectively.

### 3.9. Activated Carbon

Carbon adsorption treatment method for pesticide containing wastewater is used in the pesticide manufacturing industry as well as in pesticide cleanup [98,99]. The activated carbon system consists of a prefilter made up of sand or an alum flocculation chamber with a carbon filter [100-102]. Dennis and Kobylinski [100] reported on a Carbolator (**Figure 16**) system which uses a suspended bed of carbon packed in bags of floating porous polyethylene in order to avoid clogging. The water was continuously recirculated through the carbon filters by directing it back into the waste holding tank.

Felsot *et al.* [22] reported that rinsewater containing malathion, propoxur, chlorpyrifos, diazinon and dimethoate were all removed to nondetectable levels using the Carbolator. This process reduced the amount of waste generated by several magnitudes through efficiently absorbing pesticides from the water. Kobylinski *et al.* [99] used a Carbolator 35B to remove baygon, dimethoate, diazinon, runnel, malathion, dursban and 2,4-D and found that the higher the molecular weight of the compound the more favourable the effect of adsorption by activated carbon. Similar findings were also reported by other researchers [104,105].



**Figure 16.** Recirculation through activated carbon [103].

Honeycutt *et al.* [69] reported that a waste stream containing 100 ppm chlorophenols was reduced to 1 ppm using activated carbon. Giusti *et al.* [106] reported a carbon activated adsorption of 3.6% and 98.5% for methanol (molecular weight of 32 g/mol) and 2-ethyl hexanol (molecular weight of 130.2 g/mol), respectively. Sarkar *et al.* [107] reported an adsorbent efficiency of 98% - 99% for the removal of the isoproturon pesticide using powdered activated charcoal. Gupta *et al.* [108] reported an adsorption efficiency of 70% - 80% using activated charcoal for removing pesticides. Word and Getzen [109] reported that a decrease in pH increased the adsorption of aromatic acid compounds due to enhancement of carbon surface properties.

The activated carbon is very effective in removing pesticides and it does not require extensive monitoring. The disadvantages include: 1) the need for a skilled chemist for field testing, 2) the high cost and 3) this process is only capable of adsorbing solutions with concentrations of less than 1000 ppm [69].

### 3.10. Composting

This treatment method relies primarily on microbial activity and aeration efficiency. Microorganisms that are naturally occurring in the materials increase significantly in numbers and begin to decompose biodegradable compounds which results in the release of carbon dioxide as well as the production of metabolic heat, causing the temperature of the compost to rise to 60°C - 70°C. As the compost temperature increases, three succession of microbes occur: psychrophilic, mesophilic and thermophilic [22].

Racke and Frink [110] reported a complete degradation of carbaryl during the composting of sewage sludge. Petruska *et al.* [111] achieved a complete degradation of diazinon pesticide using dairy manure compost. Rose and Mercer [112] reported a 100% degradation of parathion

insecticide in cannery wastes. Singh [113] reported a degradation efficiency of 96.03% for the endosulfan pesticide after 4 weeks, using composted soil with a moisture level of 38%. Al hattab and Ghaly [97] achieved a captan removal efficiency of 92.4% in the first four days using hay compost.

Several researchers stated that polyhalogenated hydrocarbons, used in pesticides, can be metabolized under anaerobic conditions [114-117]. However, other researchers noted that pesticides may largely persist unchanged during the composting process [110,111,118]. Muller and Korte [118] noted little to no degradation of the aldrin, dieldrin and monolinuron during the composting of sewage waste sludge. Strom [119] noted the presence of chlordane in finished compost from various US municipalities.

### 3.11. Phytoremediation

In this method plants are used to contain and remove harmful environmental contaminants as shown in **Figure 17** [120]. Kruger *et al.* [121] reported a degradation efficiency in atrazine of 65% after 9 weeks in soil where *Kochi* sp. was planted. Coats and Anderson [122] reported that degradation of atrazine, metolachlor and trifluralin was enhanced in soils where the *Kochi* sp. plant grows. Olette *et al.* [123] reported that the aquatic plants *L. minor*, *C. aquatic* and *E. Canadensis* were capable of removing 2.5% - 50% of dimethomorph and flazasulfuron present in the water. Buyanovsky *et al.* [124] noted that the fungi rhizosphere was capable of degrading carbofuran by using it as its carbon source. Gordon *et al.* [125] noted that 95% of trichloroethylene was removed from wastewater by hybrid polar trees during growing season. Stearman *et al.* [126] noted that in constructed wetlands, cells with plants were capable of removing 77.1% and 82.4% of simazine and metolachlor, respectively, while cells without plants were only capable of removing 64.3% and 63.2%, respectively. Wang *et al.* [127] reported that in the first 20 days of plant growth, oilseed rape seedlings were capable of removing 20% of

chlorpyrifos pesticide.

### 3.12. Bioaugmentation

This method uses isolated microbes for the degradation of pesticides. The pesticides are quickly metabolized and converted to products with a lower toxicity under aerobic and anaerobic conditions [22]. Some pesticides are capable of being degraded by certain bacterial strains as their sole nutrient or carbon source. Such microbial reactions are known as mineralizing because of the large amount of carbon dioxide released during metabolism [22].

Dichlorodiphenyltrichloroethane, normally resistant to biodegradation, has been reported to have been metabolized in an anaerobic microbial culture, followed by an aerobic one [128]. Bhadhade *et al.* [24] reported that bacteria isolated from soil was capable of degrading 83% - 93% of the organophosphorous pesticide, monocrotophos. Ohshiro *et al.* [25] reported a 96% reduction in isoxathion from the organophosphorous pesticide by bacteria isolated from turf green soil. Tang and You, [27] reported that the triazophos bacteria is capable of degrading 33.1% to 95.8% of pesticides.

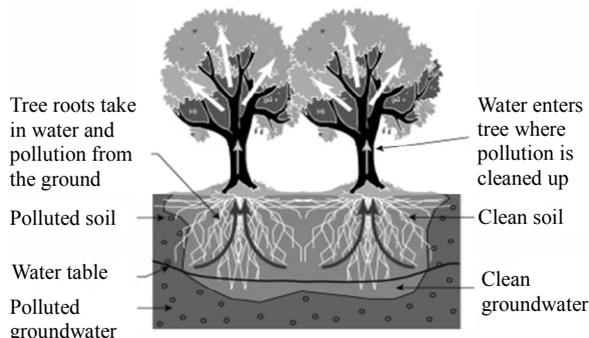
The down side of such a method is the establishment of the microbes in the presence of other microbial populations present in the contaminated soil [22]. Acea *et al.* [129] noted that the population of the introduced bacteria may be reduced due to susceptibility to predation or starvation.

## 4. Comparative Analysis

Seven criteria (**Table 7**) were used for the evaluation of pesticide disposal and treatment methods: 1) containment ability, 2) detoxification ability, 3) time 4) cost, 5) suitable for on farm use, 6) size and 7) water evaporation. Each criterion was then assigned a number from 1 to 20 which was determined by the degree of importance of the criterion. These values were then used to determine the ability of each method to meet a given criterion as shown in **Tables 8-25**. The sum of the scores obtained from each method were then used to determine its overall performance (**Table 26**). The results indicated that of the 18 methods of pesticide disposal and treatment evaluated, 9 scored above 80/100 and can be used on farm. These were organic absorbents (97), composting (94), bioaugmentation (92), inorganic absorbents (90), Fenton oxidation (86) O<sub>3</sub>/UV (83), activated carbon (82), hydrolysis (82) and land cultivation (80). The other methods are not suitable for on farm use as they suffered from containment problems, high cost and variability in detoxifying effectiveness.

## 5. Conclusion

Pesticides provide the primary means for controlling



**Figure 17. Phytoremediation to clean up pesticide contaminated grounds [1].**

**Table 7. Criteria used in the comparative analysis.**

Criteria	Importance	Description
Containment	20	The system should be able to contain the low level pesticide wastewater with no leakage or overflow so that it does not create pollution problems
Detoxification ability	20	The system should reduce the traces of pesticides by breaking the molecules down into harmless compounds
Cost	15	The cost should be appealing to farmers
Time	15	The rate of degradation should be faster than the rate of application
Suitability	12	The system should take into consideration local climate and type of pesticides used
Size	10	An on-site method is the goal since transport is costly and requires special equipment and precautions
Evaporation	8	The system should allow water to easily evaporate but not the pesticide so that clean water is returned to the ecosystem

**Table 8. Evaluation criteria for land cultivation.**

Criteria	Description	Score
Containment (20)	Runoff will contain pesticide residues Possible contamination of ground water from heavy rain or snow melt	14
Detoxification ability (20)	Dependent on the type and concentration of pesticides Relatively effective in degrading some pesticides	16
Cost (15)	Minimum energy costs required for operation and maintenance Land is radially available and economical	15
Time (15)	Dependent on the type and concentration of pesticides	10
Suitability (12)	Effectiveness is pesticide dependant System will be subjected to freeze/thaw cycles in Nova Scotia	9
Size (10)	Depends on amount of pesticide being disposed	8
Evaporation (8)	Water is absorbed by soil to provide the moisture needed for biodegradation	8

**Table 9. Evaluation criteria for soil pit.**

Criteria	Description	Score
Containment (20)	Runoff will contain pesticide residues Possible contamination of ground water after heavy rain or snow melt The area needs to be fenced to prevent accidents	12
Detoxification Ability (20)	Dependent on the type and concentration of pesticides High pesticide concentration may inhibit degradation	14
Cost (15)	Minimum energy cost required for operation and maintenance Space and materials are available on farm	15
Time (15)	Dependent on the type and concentration of pesticides	9
Suitability (12)	Effectiveness is pesticide dependent System will be subject to freeze/thaw cycles in Nova Scotia	8
Size (10)	Depends on amount of pesticide being disposed	8
Evaporation (8)	Effective water evaporation	6

**Table 10. Evaluation criteria for plastic pit.**

Criteria	Description	Score
Containment (20)	Noted to contain pesticides satisfactorily The area needs to be fenced to prevent accidents	18
Detoxification Ability (20)	Dependent on the type and concentration of pesticides Reported to be very effective for 2,4-D and Carbaryl but had little effect on Atrazine	14
Cost (15)	Costs of liner for large size pit is high Commercial garbage containers are readily available and very economical	12
Time (15)	Dependent on the type and concentration of pesticides	10
Suitability (12)	Effectiveness is pesticide dependent This system will be subjected to freeze/thaw cycles in Nova Scotia and may crack	6
Size (10)	Depends on amount of pesticide being disposed Commercial garbage container can be used for small amounts	9
Evaporation (8)	Effective water evaporation	6

**Table 11. Evaluation criteria for concrete pit.**

Criteria	Description	Score
Containment (20)	Is more durable and will contain pesticides The area needs to be fenced to prevent accidents	18
Detoxification Ability (20)	Reported to be very successful with most pesticides	18
Cost (15)	The cost of materials for this system is high	9
Time (15)	Dependent on the type and concentration of pesticides	10
Suitability (12)	Costly but little maintenance is required The system is subjected to freeze/thaw cycles in Nova Scotia	8
Size (10)	Depends on amount of pesticide being disposed	8
Evaporation (8)	Effective water evaporation	6

**Table 12. Evaluation criteria for evaporative beds.**

Criteria	Description	Score
Containment (20)	Will contain pesticides The area needs to be fenced to prevent accidents	18
Detoxification Ability (20)	Dependent on the type and concentration of pesticides	16
Cost (15)	The materials and construction are expensive	9
Time (15)	Dependent on the type and concentration of pesticides	10
Suitability (12)	Costly but little maintenance is required The system is subjected to freeze/thaw cycles in Nova Scotia	8
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Effective water evaporation	6

**Table 13. Evaluation criteria for landfilling.**

Criteria	Description	Score
Containment (20)	With proper leachate collection system this method would be adequate	18
Detoxification Ability (20)	Dependent on pesticides ability to degrade under anaerobic conditions	12
Cost (15)	Properly constructed landfills are costly	8
Time (15)	Dependent on the pesticide type and concentration	10
Suitability (12)	Suitable for all pesticides	12
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	No evaporation (used mostly for solid waste)	8

**Table 14. Evaluation criteria for incineration.**

Criteria	Description	Score
Containment (20)	Intermediate dangerous products can be produced and emitted into the atmosphere Scrubber and electrofilters can reduce emissions	14
Detoxification ability (20)	Very effective at removing the original pesticide May produce other harmful products	14
Cost (15)	Very high because of the sophisticated equipment and construction involved High energy consumption	3
Time (15)	Rapid	15
Suitability (12)	Unsuitable for on farm use because of cost and complexity	3
Size (10)	Lacks standard specification for pesticides	2
Evaporation (8)	Water released to atmosphere as vapor may contain some pesticide	4

**Table 15. Evaluation criteria for open burning.**

Criteria	Description	Score
Containment (20)	Intermediate toxic products can be produced and emitted into the atmosphere Must be isolated and kept away from residential areas Direction of wind should be considered when choosing location	10
Detoxification ability (20)	Temperatures for complete combustion are not reached Pesticide residues still remain in the pit	10
Cost (15)	Economical	13
Time (15)	Relatively fast	13
Suitability (12)	Unsuitable for on farm use because of toxic emissions	5
Size (10)	Depends on the amount of pesticides to be disposed	8
Evaporation (8)	Water released into the atmosphere as vapour may contain pesticides	8

**Table 16. Evaluation criteria for O<sub>3</sub>/UV.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Dependent on the type and concentration of pesticides Works well on aromatic compounds Other harmful compounds maybe produced	12
Cost (15)	High equipment cost High energy consumption	8
Time (15)	Rapid	15
Suitability (12)	Can be used on the farm quite easily	12
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Purified water can be recycled	8

**Table 17. Evaluation criteria for Fenton oxidation.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Effective pesticide destruction only under acidic conditions and absence of other substances that can complex with the iron ions	14
Cost (15)	Low cost	11
Time (15)	Rapid if equipped with UV radiation	13
Suitability (12)	Ease of operation and low costs make it suitable for small scale operation by farmers	12
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Purified water can be recycled	8

**Table 18. Evaluation criteria for hydrolysis.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Effective in treating pesticides with an ester linkage Unable to treat chlorinated pesticides	14
Cost (15)	Materials are readily available and economical	14
Time (15)	Can be accelerated by addition of Cu(II) complex	13
Suitability (12)	Release of toxic products makes it unsuitable for on farm use	5
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Purified water can be recycled	8

**Table 19. Evaluation criteria for KEPG.**

Criteria	Description	Score
Containment (20)	Effective if properly equipped with a vent system consisting of a scrubber and adsorbent to control emissions	16
Detoxification ability (20)	Dependent on the type and concentration of pesticides Not suited for chlorinated pesticides with concentrations above 5%	10
Cost (15)	Materials are readily available and economical Energy is required	12
Time (15)	Biodegradation after dechlorination requires time	10
Suitability (12)	The ease of obtaining the chemicals required makes it suitable for farmers	12
Size (10)	Cannot be used as a remedy for large scale treatment	5
Evaporation (8)	Purified water can be recycled	8

**Table 20. Evaluation criteria for inorganic absorbents.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Dependent on the type and concentration of pesticides Effective in destroying chlorinated pesticides	18
Cost (15)	Relatively economical	12
Time (15)	Faster with increased temperature	12
Suitability (12)	Materials are readily available and easy to prepare	12
Size (10)	Depends on amount of pesticides in wastewater	8
Evaporation (8)	Purified water can be recycled	8

**Table 21. Evaluation criteria for organic absorbents.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Effective pesticide removal ability of various types	20
Cost (15)	Economical Materials are readily available	15
Time (15)	Relatively fast	14
Suitability(12)	Ease of operation makes it suitable for farmers	12
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Purified water can be recycled	8

**Table 22. Evaluation criteria for activated carbon.**

Criteria	Description	Score
Containment (20)	Effective containment of pesticides	20
Detoxification ability (20)	Effective in treating wastewaters with pesticide concentrations less than 1000 ppm	18
Cost (15)	High cost Energy is required	9
Time (15)	Relatively fast	13
Suitability (12)	Not suitable for farmers because a skilled chemist is required for field testing	6
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Purified water can be recycled	8

**Table 23. Evaluation criteria for composting.**

Criteria	Description	Score
Containment (20)	Effective containment if a bioreactor is used	18
Detoxification ability (20)	Effective in biodegrading pesticides	20
Cost (15)	Economical	15
Time (15)	Dependent on the pesticide type and concentration	13
Suitability (12)	The ease of operation and limited care required make this treatment option suitable for farmers	12
Size (10)	Depends on amount of pesticide being disposed	8
Evaporation (8)	Water is absorbed by the organic material to provide moisture for biodegradation	8

**Table 24. Evaluation criteria for bioaugmentation.**

Criteria	Description	Score
Containment (20)	Effective containment if a bioreactor is used	18
Detoxification ability (20)	Effective in biodegrading pesticides	20
Cost (15)	Economical	13
Time (15)	Dependent on the type and concentration of pesticides	13
Suitability (12)	Can be easily used on farm sites	12
Size (10)	Depends on the amount of pesticide being disposed	8
Evaporation (8)	Water is absorbed by organic material to provide moisture for biodegradation	8

**Table 25. Evaluation criteria for phytoremediation.**

Criteria	Description	Score
Containment (20)	Runoff will contain pesticide residues The area should be fenced to avoid accidents Possible contamination of ground water	12
Detoxification ability (20)	Dependent on pesticide type and plant type	12
Cost (15)	Materials can be easily found Relatively economical	13
Time (15)	Long time is required for pesticide degradation	8
Suitability (12)	The ease of operation and low cost make this method suitable for on farm use	12
Size (10)	Depends on amount of pesticide being disposed	8
Evaporation (8)	Effective evaporation and transeaporation	8

**Table 26. Comparative analysis.**

Criteria	LC	SP	PP	CP	EB	LF	I	OB	O <sub>3</sub> /UV	FO	H	KP	IA	OA	AC	C	BA	PR
Containment (20)	14	12	18	18	18	18	14	10	20	20	20	16	20	20	20	18	18	12
Detoxification ability (20)	16	14	14	18	16	12	14	10	12	14	14	10	18	20	18	20	20	12
Cost (15)	15	15	12	9	9	8	3	13	8	11	14	12	12	15	9	15	13	13
Time (15)	10	10	10	10	10	10	15	13	15	13	13	10	12	14	13	13	13	8
Suitability (12)	9	8	6	8	8	12	3	5	12	12	5	12	12	12	6	12	12	12
Size (10)	8	8	9	8	8	8	2	8	8	8	8	5	8	8	8	8	8	8
Evaporation (8)	8	6	6	6	6	8	4	8	8	8	8	8	8	8	8	8	8	8
<b>Total (100)</b>	<b>80</b>	<b>73</b>	<b>75</b>	<b>77</b>	<b>75</b>	<b>76</b>	<b>55</b>	<b>67</b>	<b>83</b>	<b>86</b>	<b>82</b>	<b>73</b>	<b>90</b>	<b>97</b>	<b>82</b>	<b>94</b>	<b>92</b>	<b>73</b>

LC = land cultivation; FO = Fenton oxidation; SP = soil pit; H = hydrolysis; PP = plastic pit; KP = (KPEG) potassium polyethylene glycol ether; CP = concrete pit; IA = inorganic adsorbents; EB = evaporation bed; OA = Organic adsorbents; LF = landfilling; AC = activated carbon; I = incineration; C = composting; OB = open burning; BA = bioaugmentation; O<sub>3</sub>/UV = Ozone/Ultraviolet radiation; PR = phytoremediation.

organisms that compete with man for food and fibre or cause injury to man, livestock and crops. They played a vital role in the economic production of wide ranges of vegetable, fruit, cereal, forage, fibre and oil crops which now constitute a large part of successful agricultural industry in many countries. After pesticides are applied to the target areas, pesticide residues are removed from applicators by rinsing with water which results in the formation of a toxic wastewater that represents a disposal problem for many farmers. Pesticides can adversely affect people, pets, livestock and wildlife in addition to the pests they are intended to destroy. The phenomenon of biomagnification of some pesticides has resulted in reproductive failure of some fish species and egg shell thinning of birds such as peregrine falcons, sparrow hawk and eagle owls. Pesticide toxicity to humans include skin and eye irritation and skin cancer. Currently, disposal of pesticide wastewater is carried out by: 1) land cultivation, 2) dumping in soil pits, plastic pits and concrete pits or on land, and in extreme cases in streams near the rinsing operation, 3) use of evaporation beds and 4) land filling. These methods of disposal are totally unsafe as the surface run off will reach streams, rivers and lakes and the infiltration of the wastewater into the local soil will eventually end up in the ground water. The treatment methods currently used for pesticide containing wastewater include 1) incineration (incinerators and open burning), 2) chemical treatments (O<sub>3</sub>/UV, hydrolysis, Fenton oxidation and KPEG), 3) physical treatments (inorganic adsorbents, organic adsorbents and activated carbon) and 4) biological treatments (composting, bio-augmentation and phytoremediation). Some of these methods either require land or are expensive and suffer from variability of effectiveness. Therefore, care must be exercised in the application, disposal and treatment of pesticides. A comparative analysis was performed on 18 methods of pesticide disposal and treatment using six

criteria: containment, detoxification ability, cost, time, suitability for on farm use, size and evaporation efficiency. The results indicated that of the 18 methods evaluated, 9 scored above 80/100 and can be used on farm. These were organic adsorbents (97), composting (94), bioaugmentation (89), inorganic adsorbents (90), Fenton oxidation (86), O<sub>3</sub>/UV (83), activated carbon (82), hydrolysis (82) and land cultivation (80). The other methods are not suitable for on farm use as they suffered from containment problems, high cost and variability in detoxifying effectiveness.

## 6. Acknowledgements

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## REFERENCES

- [1] US EPA, "Pesticides and Pesticide Containers, Regulation for Acceptance and Recommended Procedures for Disposal and Treatment," US Environmental Protection Agency, Municipal Environmental Research Laboratory, Cincinnati, 2011. <http://www.epa.ca>
- [2] WRI, "Pesticide Use (Most Recent) by Country," World Resources Institute, Woolwich, 2011. <http://www.nationmaster.com>
- [3] AWCN, "The World's Top 10 Pesticide Firms," Agrow World Crop Protection News, London, 2008. <http://www.gmwatch.org>
- [4] J. K. Horowitz and E. Lichtenberg, "Insurance, Moral Hazard, and Chemical Use in Agriculture," *American Journal of Agricultural Economics*, Vol. 75, No. 4, 1993, pp. 926-935. [doi:10.2307/1243980](https://doi.org/10.2307/1243980)
- [5] E. C. Oerke and H. W. Dehne, "Safeguarding Production-Losses in Major Crops and the Role of Crop Protection," *Crop Protection*, Vol. 23, No. 4, 2004, pp. 275-285. [doi:10.1016/j.cropro.2003.10.001](https://doi.org/10.1016/j.cropro.2003.10.001)
- [6] J. Cooper and H. Dobson, "The Benefits of Pesticides to

- Mankind and the Environment,” *Crop Protection*, Vol. 26 No. 9, 2007, pp. 1337-1348. [doi:10.1016/j.cropro.2007.03.022](https://doi.org/10.1016/j.cropro.2007.03.022)
- [7] C. Osteen and M. Livingston, “Pest Management Practices,” Agricultural Resources and Environmental Indicators, United States Department of Agriculture, Washington DC, 2006. <http://www.ers.usda.gov>
- [8] T. J. Centner, “Unwanted Agricultural Pesticides: State Disposal Programs,” *Journal of Environmental Quality*, Vol. 27, No. 4, 1998, pp. 736-742. [doi:10.2134/jeq1998.00472425002700040002x](https://doi.org/10.2134/jeq1998.00472425002700040002x)
- [9] C. Wilson and C. Tisdell, “Why Farmers Continue to Use Pesticides Despite Environmental Health and Sustainability Costs,” *Ecological Economics*, Vol. 39, No. 3, 2001, pp. 449-462. [doi:10.1016/S0921-8009\(01\)00238-5](https://doi.org/10.1016/S0921-8009(01)00238-5)
- [10] R. L. Ridgway, J. C. Tinney, J. T. MacGregor and N. J. Starler, “Pesticide Use in Agriculture,” *Environmental Health Perspectives*, Vol. 27, 1978, pp. 103-112. [doi:10.1289/ehp.7827103](https://doi.org/10.1289/ehp.7827103)
- [11] D. W. Connell, “Bioaccumulation Behaviour of Persistent Organic Chemicals with Aquatic Organisms,” *Review of Environmental Contamination and Toxicology*, Vol. 101, 1988, pp. 117-154. [doi:10.1007/978-1-4612-3810-2\\_3](https://doi.org/10.1007/978-1-4612-3810-2_3)
- [12] K. Verschuere, “Handbook of Environmental Data on Organic Chemicals,” Van Nostrand Reinhold, New York, 1977.
- [13] C. E. Lundholm, “DDE-Induced Eggshell Thinning in Birds: Effects of p,p'-DDE on the Calcium and Prostaglandin Metabolism of the Eggshell Gland,” *Comparative Biochemistry and Physiology Part C: Pharmacology Toxicology Endocrinology*, Vol. 118, No. 2, 1997, pp. 113-128. [doi:10.1016/S0742-8413\(97\)00105-9](https://doi.org/10.1016/S0742-8413(97)00105-9)
- [14] R. Spiewak, “Pesticides as a Cause of Occupational Skin Diseases in Farmers,” *Annals of Agricultural and Environmental Medicine*, Vol. 8, No. 1, 2001, pp. 1-5.
- [15] N. E. Kowal and H. R. Pahren, “Health Effects Associated with Wastewater Treatment and Disposal,” *Journal (Water Pollution Control Federation)*, Vol. 54, No. 6, 1982, pp. 677-687.
- [16] W. L. Winterlin, S. R. Schoen and C. R. Mourer, “Disposal of Pesticide Wastes in Lined Evaporation Beds,” *Treatment and Disposal of Pesticide Wastes*, Vol. 259, 1984, pp. 97-116. [doi:10.1021/bk-1984-0259.ch006](https://doi.org/10.1021/bk-1984-0259.ch006)
- [17] C. J. Somlich, M. T. Kearney and S. Elsasser, “Enhanced Soil Degradation of Alachlor by Treatment with Ultraviolet Light and Ozone,” *Journal of Agricultural Food Chemistry*, Vol. 36, 1988, pp. 1322-1326. [doi:10.1021/jf00084a049](https://doi.org/10.1021/jf00084a049)
- [18] P. C. Kearney, Q. Zeng and J. M. Ruth, “A Large Scale UV-Ozonation Degradation Unit-Field Trials on Soil Pesticide Waste Disposal,” *ACS Symposium Series*, Vol. 259, 1984, pp. 195-209.
- [19] G. A. Junk, J. J. Richard and P. A. Dahm, “Degradation of Pesticides in Controlled Water-Soil Systems,” *Treatment of Pesticide Wastes, ACS Symposium Series*, Vol. 259, 1984, pp. 125-154.
- [20] C. V. Hall, J. Baker, P. Dahm, L. Freiburger and G. Gorder, “Safe Disposal Methods for Agricultural Pesticide Wastes,” US Environmental Protection Agency, Municipal Environmental Research Laboratory, Cincinnati, 1981. <http://www.epa.gov/>
- [21] FRTR, “Composting (*ex Situ* Biological Treatment),” Federal Remediation Technologies Roundtable, Center for Environmental Research Information, Cincinnati, 2008. <http://www.frtr.gov/matrix2/section4/4-12.html>
- [22] A. S. Felsot, K. D. Racke and D. J. Hamilton, “Disposal and Degradation of Pesticide Waste,” *Reviews of Environmental Contamination and Toxicology*, Vol. 177, 2003, pp. 123-200. [doi:10.1007/0-387-21725-8\\_3](https://doi.org/10.1007/0-387-21725-8_3)
- [23] R. F. Krueger and D. J. Severn, “Regulation of Pesticide Disposal,” *Treatment and Disposal of Pesticide Wastes, ACS Symposium Series*, Vol. 259, 1984, pp. 3-15. [doi:10.1021/bk-1984-0259.ch001](https://doi.org/10.1021/bk-1984-0259.ch001)
- [24] B. J. Bhadhade, S. S. Sarnaik and P. P. Kanekar, “Bioremediation of an Industrial Effluent Containing Monocrotophos,” *Current Microbiology*, Vol. 45, No. 5, 2002, pp. 346-349. [doi:10.1007/s00284-002-3681-1](https://doi.org/10.1007/s00284-002-3681-1)
- [25] K. Ohshiro, T. Kakuta, T. Sakai, H. Hirota, T. Hoshino and T. Uchiyama, “Biodegradation of Organophosphorus Insecticides by Bacteria Isolated from Turf Green Soil,” *Journal of Fermentation and Bioengineering*, Vol. 82, No. 3, 1996, pp. 299-305. [doi:10.1016/0922-338X\(96\)88823-4](https://doi.org/10.1016/0922-338X(96)88823-4)
- [26] P. C. Kearney, R. G. Nash and C. S. Helling, “Pesticide Degradation Properties,” *Proceedings, National Workshop on Pesticide Waste Disposal*, US Environmental Protection Agency, Water Engineering Research Laboratory, Cincinnati, 1985, pp. 35-42.
- [27] M. Tang and M. You, “Isolation, Identification and Characterization of a Novel Triazophos-Degrading *Bacillus* sp. (TAP-1),” *Microbiological Research*, Vol. 167, No. 5, pp. 299-305. [doi:10.1016/j.micres.2011.10.004](https://doi.org/10.1016/j.micres.2011.10.004)
- [28] K. D. Racke and J. R. Coats, “Enhanced Biodegradation of Pesticides in the Environment,” *American Chemical Society*, No. 426, Washington DC, 1990.
- [29] R. S. Schoen and W. L. Winterlin, “The Effects of Various Soil Factors and Amendments on the Degradation of Pesticide Mixtures,” *Journal of Environmental Science and Health Part B*, Vol. 22, No. 3, 1987, pp. 347-377. [doi:10.1080/03601238709372561](https://doi.org/10.1080/03601238709372561)
- [30] A. S. Felsot, “Landfarming Pesticide-Contaminated Soils,” *Pesticide Remediation in Soils and Water*, Wiley, New York, 1998, pp. 129-160.
- [31] L. Somasundaram, J. R. Coats, D. K. Racke and H. M. Stahr, “Application of the Microtox System to Assess the Toxicity of Pesticides and Their Hydrolysis Metabolites,” *Bulletin of Environmental Contamination Toxicology*, Vol. 44, No. 2, 1990, pp. 254-259. [doi:10.1007/BF01700144](https://doi.org/10.1007/BF01700144)
- [32] A. S. Felsot, J. K. Mitchell and E. K. Dzantor, “Remediation of Herbicide Contaminated Soil by Combinations of Landfarming and Biostimulation,” *SSSA Special Publication*, Vol. 43, 1995, pp. 237-257.
- [33] W. Winterlin, J. N. Seiber, A. Craigmill, T. Baier, J. Woodrow and G. Walker, “Degradation of Pesticide Waste Taken from a Highly Contaminated Soil Evaporation Pit in California,” *Archives of Environmental Contamination*

- and Toxicology, Vol. 18, 1989, pp. 734-747.  
[doi:10.1007/BF01225011](https://doi.org/10.1007/BF01225011)
- [34] J. Y. Gan and W. C. Koskinen, "Pesticide Fate and Behavior in Soil at Elevated Concentrations," In: P. C. Kearney and T. R. Roberts, Eds., *Pesticide Remediation in Soils and Water*, Wiley, Chichester, 1998, pp. 59-84.
- [35] E. K. Dzantor and A. S. Felsot, "Microbial Responses to Large Concentrations of Herbicides in Soil," *Environmental Toxicology and Chemistry*, Vol. 10, No. 5, 1991, pp. 649-655. [doi:10.1002/etc.5620100511](https://doi.org/10.1002/etc.5620100511)
- [36] J. Gan, W. C. Koskinen, R. L. Becker and D. D. Buhler, "Effect of Concentration on Persistence of Alachlor in Soil," *Journal of Environmental Quality*, Vol. 24, No. 6, 1995, pp. 1162-1169.  
[doi:10.2134/jeq1995.00472425002400060016x](https://doi.org/10.2134/jeq1995.00472425002400060016x)
- [37] J. M. Davidson, P. S. Rao, L. T. Ou, W. B. Wheeler and D. F. Rothwell, "Adsorption, Movement and Biological Degradation of Large Concentrations of Selected Pesticides in Soil," US Environmental Protection Agency, Municipal Environmental Research Laboratory, Cincinnati, 1980.
- [38] J. Gan, R. L. Becker, W. C. Koskinen and D. D. Buhler, "Degradation of Atrazine in Two Soils as a Function of Concentration," *Journal of Environmental Quality*, Vol. 25, No. 5, 1996, pp. 1064-1072.  
[doi:10.2134/jeq1996.00472425002500050019x](https://doi.org/10.2134/jeq1996.00472425002500050019x)
- [39] G. A. Junk and J. J. Richard, "Pesticide Disposal Sites: Sampling and Analysis," *ACS Symposium Series*, Vol. 259, 1984, pp. 69-95.
- [40] B. Kinnear, "Wastewater Technology: Engineering a Healthier Society," University of Southern California, Los Angeles, 2011. <http://www.illumina.usc.edu>
- [41] L. M. Johnson and P. A. Hartman, "Microbiology of a Pesticide Disposal Pit," *Bulletin of Environmental Contamination and Toxicology*, Vol. 25, No. 3, 1980, pp. 448-455. [doi:10.1007/BF01985553](https://doi.org/10.1007/BF01985553)
- [42] C. V. Hall, "Pesticide Waste Disposal in Agriculture," *Treatment and Disposal of Pesticide Wastes*, Vol. 3, 1984, pp. 27-36. [doi:10.1021/bk-1984-0259.ch003](https://doi.org/10.1021/bk-1984-0259.ch003)
- [43] LANL, "Aerial Tour of Los Alamos National Laboratory," Los Alamos Study Group, Albuquerque, 2004. <http://www.lasg.org>
- [44] D. M. Hodapp and W. Winterlin, "Pesticide Degradation in Model Soil Evaporation Beds," *Bulletin of Environmental Contamination and Toxicology*, Vol. 43, No. 1, 1989, pp. 36-44. [doi:10.1007/BF01702235](https://doi.org/10.1007/BF01702235)
- [45] B. Plumer, "The Cost of Superfund Neglect," The New Republic, Washington DC, 2006. <http://www.plumer.blogspot.com>
- [46] KPBSWD, "Central Peninsula Landfill," Solid Waste Department, Kenai Peninsula Borough Solid Waste Department, Soldotna, 2011. <http://www2.borough.kenai.ak.us/solidwaste/>
- [47] M. Ghassami, S. C. Quinlivan and H. R. Day, "Landfills for Pesticide Waste Disposal," *Environmental Science and Technology*, Vol. 10, No. 13, 1976, pp. 1209-1214.  
[doi:10.1021/es60123a005](https://doi.org/10.1021/es60123a005)
- [48] P. Truong and B. Hart, "Vetiver System for Wastewater Treatment," Pacific Rim Vetiver Network Technical Bulletin, San Antonio, 2000. <http://www.vetiver.org>
- [49] D. M. Munnecke, "Enzymic Detoxification of Waste Organophosphate Pesticides," *Journal of Agricultural and Food Chemistry*, Vol. 28, No. 1, 1980, pp. 105-111.  
[doi:10.1021/jf60227a025](https://doi.org/10.1021/jf60227a025)
- [50] W. G. Johnson and T. L. Lavy, "In-Situ Dissipation of Benmethyl, Carbofuran, Thiobencarb and Triclopyr at Three Soil Depths," *Journal of Environmental Quality*, Vol. 23, No. 3, 1994, pp. 556-562.  
[doi:10.2134/jeq1994.00472425002300030022x](https://doi.org/10.2134/jeq1994.00472425002300030022x)
- [51] A. Yasuhara, H. Shiraishi, M. Nishikawa, T. Yamamoto, T. Uehiro, O. Nakasugi, T. Okumura, K. Kenmotsu, H. Fukui, M. Nagase, Y. Ono, Y. Kawagoshi, K. Baba and Y. Noma, "Determination of Organic Components in Leachates from Hazardous Waste Disposal Sites in Japan by Gas Chromatography-Mass Spectrometry," *Journal of Chromatography A*, Vol. 774, No. 1, 1997, pp. 321-332.
- [52] G. M. Williams, I. Harrison, C. A. Carlick and O. Crowley, "Changes in Anantipmeric Fraction as Evidence of Natural Attenuation of Mecoprop in a Limestone Aquifer," *Journal of Contaminant Hydrology*, Vol. 64, No. 3-4, 2003, pp. 253-267. [doi:10.1016/S0169-7722\(02\)00206-1](https://doi.org/10.1016/S0169-7722(02)00206-1)
- [53] T. H. Christensen, P. Kjeldsen, P. L. Bjerg, D. L. Jensen, J. B. Christensen, A. Baun, H. J. Albrechtsen and G. Heron, "Biogeochemistry of Landfill Leachate Plumes," *Applied Geochemistry*, Vol. 16, No. 7, 2001, pp. 659-718.  
[doi:10.1016/S0883-2927\(00\)00082-2](https://doi.org/10.1016/S0883-2927(00)00082-2)
- [54] B. J. Alloway and D. C. Ayres, "Chemical Principles of Environmental Pollution," *Water, Air and Soil Pollution*, Vol. 102, No. 3, 1997, pp. 216-218.
- [55] T. L. Ferguson and R. R. Wilkinson, "Incineration of Pesticide Wastes," *Treatment and Disposal of Pesticide Wastes*, ACS Symposium Series, Vol. 259, 1984, pp. 181-191.
- [56] D. A. Oberacker, "Incineration Options for Disposal of Waste Pesticides," In: J. S. Bridges and C. R. Dempsey, Eds., *Pesticide Waste Disposal Technology*, US Environmental Protection Agency, Noyes Data Corporation, 1988.
- [57] M. V. Kennedy, B. J. Stojanovic and F. L. Shuman, "Chemical and Thermal Methods for Disposal of Pesticides," *Residue Reviews*, Vol. 29, 1969, pp. 89-104.
- [58] E. M. Steverson, "Incineration as a Pesticide Remediation Method," *Pesticide Remediation in Soils and Water*, Wiley, New York, 1998, pp. 85-103.
- [59] W. P. Linak, J. A. Mulholland, J. A. McSorley, R. E. Hall, R. K. Srivastava, J. V. Ryan, M. G. Nishioka, J. Lewtas and D. M. DeMarini, "Application of Staged Combustion and Reburning to the Co-Firing of Nitrogen-Containing Wastes," *Hazardous Waste and Hazardous Materials*, Vol. 8, No. 1, 1991, pp. 1-15. [doi:10.1089/hwm.1991.8.1](https://doi.org/10.1089/hwm.1991.8.1)
- [60] B. Ahling and K. Wiberger, "Incineration of Pesticides Containing Phosphorus," *Journal of Environmental Quality*, Vol. 8, No. 1, 1979, pp. 12-13.  
[doi:10.2134/jeq1979.00472425000800010003x](https://doi.org/10.2134/jeq1979.00472425000800010003x)
- [61] J. J. Santoleri, "Incineration," *Pollution Issues*, Advameg, Flossmoor, 2011.

- <http://www.pollutionissues.com/Ho-Li/Incineration.html>
- [62] CART, "Allowable Open Burning," Country Acres Resource Team, Adams County Extension, Brighton, 2008. <http://www.coopext.colostate.edu/adams/sa/openburning.htm>
- [63] B. Adebona, A. Shafagati, E. J. Martin and R. C. Chawla, "Laboratory Evaluation of Products of Incomplete Combustion Formed from Burning of Agricultural Product Bags," *Waste Management, ACS Symposium Series*, Vol. 510, 1992, pp. 63-77.
- [64] D. A. Oberacker, P. C. Lin, G. M. Shaul, D. T. Ferguson, V. S. Engleman, T. W. Jackson, J. S. Chapman, J. D. Evans, R. J. Martrano and L. L. Every, "Characterization of Emissions Formed from Open Burning of Pesticide Bags," *Pesticide Waste Management, ACS Symposium Series*, Vol. 510, 1992, pp. 78-94.
- [65] A. Mokrini, D. Oussi and S. Esplugas, "Oxidation of Aromatic Compounds with UV Radiation/Ozone/Hydrogen Peroxide," *Water Science and Technology*, Vol. 35, No. 4, 1991, pp. 95-102. [doi:10.1016/S0273-1223\(97\)00014-0](https://doi.org/10.1016/S0273-1223(97)00014-0)
- [66] W. H. Glaze, "Drinking-Water Treatment with Ozone," *Environmental Science and Technology*, Vol. 21, No. 3, 1987, pp. 224-230. [doi:10.1021/es00157a001](https://doi.org/10.1021/es00157a001)
- [67] G. R. Peyton and W. H. Glaze, "Mechanism of Photolytic Ozonation," *Photochemistry of Environmental Aquatic Systems, ACS Symposium Series*, Vol. 327, 1987, pp. 76-88.
- [68] W. S. Kuo, "Photocatalytic Oxidation of Pesticide Residue," *Journal of Environmental Science and Health, Part B: Pesticides, Food Contaminants, and Agricultural Wastes*, Vol. 37, No. 1, 2002, pp. 65-74. [doi:10.1081/PFC-120002898](https://doi.org/10.1081/PFC-120002898)
- [69] R. D. H. Paulson, H. LeBaron, G. Rolofson and C. Ganz, "Chemical Treatment Options for Pesticide Wastes Disposal," In: J. S. Bridges and C. R. Dempsey, Eds., *Pesticide Waste Disposal Technology*, Park Ridge, 1988, pp. 73-86.
- [70] H. Fallmann, T. Krutzler, R. Bauer, S. Malato and J. Blanco, "Applicability of the Photo-Fenton Method for Treating Water Containing Pesticides," *Catalysis Today*, Vol. 54, No. 2-3, 1999, pp. 309-319. [doi:10.1016/S0920-5861\(99\)00192-3](https://doi.org/10.1016/S0920-5861(99)00192-3)
- [71] Y. Sun and J. J. Pignatello, "Chemical Treatment of Pesticide Wastes, Evaluation of Fe(III) Chelates for Catalytic Hydrogen Peroxide Oxidation of 2,4-D at Circumneutral pH," *Journal of Agricultural Food Chemistry*, Vol. 40, 1992, pp. 322-327. [doi:10.1021/jf00014a031](https://doi.org/10.1021/jf00014a031)
- [72] C. Ozdemir, S. Sahinkaya and M. Onucyildiz, "Treatment of Pesticide Wastewater by Physicochemical and Fenton Processes," *Asian Journal of Chemistry*, Vol. 20, No. 5, 2008, pp. 3795-3804.
- [73] S. Malato, J. Blanco, A. Vidal, P. Fernandez, J. Caceres, P. Trincado, J. C. Oliveira and M. Vincent, "New Large Solar Photocatalytic Plant: Set-Up and Preliminary Results," *Chemosphere*, Vol. 47, No. 3, 2002, pp. 235-240. [doi:10.1016/S0045-6535\(01\)00220-X](https://doi.org/10.1016/S0045-6535(01)00220-X)
- [74] R. A. Larson, M. B. Schlauch and K. A. Marley, "Ferric Ion Promoted Photodecomposition of Trizines," *Journal of Agricultural Food Chemistry*, Vol. 39, No. 11, 1991, pp. 2057-2062. [doi:10.1021/jf00011a035](https://doi.org/10.1021/jf00011a035)
- [75] P. L. Huston and J. J. Pignatello, "Degradation of Selected Pesticide Active Ingredients and Commercial Formulations in Water by the Photo-Assisted Fenton Reaction," *Water Research*, Vol. 33, No. 5, 1999, pp. 1238-1246. [doi:10.1016/S0043-1354\(98\)00330-3](https://doi.org/10.1016/S0043-1354(98)00330-3)
- [76] J. J. Pignatello and Y. Sun, "Complete Oxidation of Metolachlor and Methyl Parathion in Water by the Photoassisted Fenton Reaction," *Water Research*, Vol. 29, 1995, pp. 1837-1844. [doi:10.1016/0043-1354\(94\)00352-8](https://doi.org/10.1016/0043-1354(94)00352-8)
- [77] R. Doong and W. Chang, "Photoassisted Iron Compound Catalytic Degradation of Organophosphorus Pesticide with Hydrogen Peroxide," *Chemosphere*, Vol. 37, No. 13, 1998, pp. 2563-2572. [doi:10.1016/S0045-6535\(98\)00038-1](https://doi.org/10.1016/S0045-6535(98)00038-1)
- [78] J. M. Desmarchelier, "Kinetics of Alkaline and Peroxide-Catalyzed Hydrolysis of Fenitrothion," *Journal of Environmental Science and Health, Part B*, Vol. 22, No. 4, 1987, pp. 403-411.
- [79] G. Lee, R. A. Kenley and J. S. Winterle, "Reaction of Sodium Perborate with Organophosphorus Esters," *Treatment and Disposal of Pesticide Wastes, ACS Symposium Series*, Vol. 259, 1984, pp. 211-219.
- [80] C. Qian, P. G. Sanders and J. N. Seiber, "Accelerated Degradation of Organophosphorus Pesticides with Sodium Perborate," *Bulletin of Environmental Contamination and Toxicology*, Vol. 35, No. 1, 1985, pp. 682-688. [doi:10.1007/BF01636573](https://doi.org/10.1007/BF01636573)
- [81] J. M. Smolen and A. T. Stone, "Divalent Metal Ion-Catalyzed Hydrolysis of Phosphorothionate Ester Pesticides and Their Corresponding Oxonated," *Environmental Science and Technology*, Vol. 31, No. 6, 1997, pp. 1664-1673. [doi:10.1021/es960499q](https://doi.org/10.1021/es960499q)
- [82] A. M. Badawi and S. M. Ahmed, "Hydrolysis of Pesticides in Wastewater Catalyzed by Cu(II) Complexes of Silyl Based Cationic Micelles," *Journal of Dispersion Science and Technology*, Vol. 31, No. 4, 2010, pp. 577-582. [doi:10.1080/01932690903192804](https://doi.org/10.1080/01932690903192804)
- [83] D. J. Brunelle and D. A. Singleton, "Chemical Reaction of Polychlorinated Biphenyls on Soils with Poly(Ethylene glycol)/KOH," *Chemosphere*, Vol. 14, No. 2, 1985, pp. 173-181. [doi:10.1016/0045-6535\(85\)90096-7](https://doi.org/10.1016/0045-6535(85)90096-7)
- [84] A. Kornel and C. J. Rogers, "PCB Destruction: A Novel Dehalogenation Reagent," *Journal of Hazardous Materials*, Vol. 12, No. 2, 1985, pp. 161-176. [doi:10.1016/0304-3894\(85\)85004-4](https://doi.org/10.1016/0304-3894(85)85004-4)
- [85] T. O. Tiernan, D. J. Wagel, J. H. Garrett, G. F. VanNess, J.G. Solch and L. A. Harden, "Laboratory and Field Tests to Demonstrate the Efficacy of KPEG Reagent for Detoxification of Hazardous Wastes Containing Polychlorinated Dibenzo-p-Dioxins (PCDD) and Dibenzofurans (PCDF) and Soils Contaminated with Such Chemical Wastes," *Chemosphere*, Vol. 18, 1989, pp. 835-841. [doi:10.1016/0045-6535\(89\)90205-1](https://doi.org/10.1016/0045-6535(89)90205-1)
- [86] M. L. Taylor, J. A. Wentz, M. A. Dosani, W. Gallagher and J. S. Greber, "Treating Chlorinated Wastes with KPEG Process," US Environmental Protection Agency, Engineering Laboratory, Cincinnati, 1990.

- [87] J. Inacio, C. Taviot-Gueho, C. Forano and J. P. Besse, "Adsorption of MCPA Pesticide by MgAl-Layered Double Hydroxides," *Applied Clay Science*, Vol. 18, No. 5-6, 2001, pp. 255-264. [doi:10.1016/S0169-1317\(01\)00029-1](https://doi.org/10.1016/S0169-1317(01)00029-1)
- [88] M. Ahmaruzzaman and V. K. Gupta, "Rice Husk and Its Ash as Low-Cost Adsorbents in Water and Wastewater Treatment," *Industrial and Engineering Chemistry Research*, Vol. 50, No. 24, 2011, pp. 13589-13613.
- [89] VWST, "Gravity Filter," Veolia Water Solutions and Technology, Saint Maurice, 2011. <http://www.veoliawaterst.com/filtraflo/en/>
- [90] R. Niwas, U. Gupta, A. A. Khan and K. G. Varshney, "The Adsorption of Phosphamidon on the Surface of Styrene Supported Zirconium (IV) Tungstophosphate: A Thermodynamic Study," *Colloids and Surfaces, A: Physicochemical and Engineering Aspects*, Vol. 164, 2000, pp. 115-119. [doi:10.1016/S0927-7757\(99\)00247-2](https://doi.org/10.1016/S0927-7757(99)00247-2)
- [91] R. Boussahel, A. Montiel and M. Baudu, "Effects of Organic and Inorganic Matter on Pesticide Rejection by Nanofiltration," *Desalination*, Vol. 145, No. 1, 2002, pp. 109-114. [doi:10.1016/S0011-9164\(02\)00394-6](https://doi.org/10.1016/S0011-9164(02)00394-6)
- [92] E. Bojemueller, A. Nennemann and G. Lagaly, "Enhanced Pesticide Adsorption by Thermally Modified Bentonites," *Applied Clay Science*, Vol. 18, No. 5-6, 2001, pp. 277-284. [doi:10.1016/S0169-1317\(01\)00027-8](https://doi.org/10.1016/S0169-1317(01)00027-8)
- [93] F. Li, Y. Wang, Q. Yang, D. G. Evans, C. Forano and X. Duan, "Study on Adsorption of Glyphosate (N-Phosphonomethyl Glycine) Pesticide on MgAl-Layered Double Hydroxides in Aqueous Solution," *Journal of Hazardous Materials*, Vol. 125, No. 1-3, 2005, pp. 89-95. [doi:10.1016/j.jhazmat.2005.04.037](https://doi.org/10.1016/j.jhazmat.2005.04.037)
- [94] S. Chowdhury, R. Mishra, P. Saha and P. Kushwaha, "Adsorption Thermodynamics, Kinetics and Isothermic Heat of Adsorption of Malachite Green onto Chemically Modified Rice Husk," *Desalination*, Vol. 265, No. 1, 2011, pp. 159-168. [doi:10.1016/j.desal.2010.07.047](https://doi.org/10.1016/j.desal.2010.07.047)
- [95] M. Akhtar, S. M. Hasany, M. I. Bhangar and S. Iqbal, "Low Cost Sorbents for the Removal of Methyl Parathion Pesticide from Aqueous Solutions," *Chemosphere*, Vol. 66, No. 10, 2007, pp. 1829-1838. [doi:10.1016/j.chemosphere.2006.09.006](https://doi.org/10.1016/j.chemosphere.2006.09.006)
- [96] G. Z. Memon, M. I. Bhangar, M. Akhtar, F. N. Talpur and J. R. Memon, "Adsorption of Methyl Parathion Pesticide from Water Using Watermelon Peels as a Low Cost Adsorbent," *Chemical Engineering Journal*, Vol. 138, No. 1-3, 2008, pp. 616-621. [doi:10.1016/j.cej.2007.09.027](https://doi.org/10.1016/j.cej.2007.09.027)
- [97] M. Al hattab and A. E. Ghaly, "Sequential Remediation Processes for Low Level Pesticide Wastewater," *Journal of Environmental Protection*, Vol. 3, No. 2, 2012, pp. 150-163. [doi:10.4236/jep.2012.32019](https://doi.org/10.4236/jep.2012.32019)
- [98] P. R. Atkins, "The Pesticide Manufacturing Industry-Current Waste Treatment and Disposal Practices," *Water Pollution Control Research Series*, US Environmental Protection Agency, Washington DC, 1972.
- [99] E. A. Kobylinski, W. H. Dennis and A. B. Rosencrance, "Treatment of Pesticide-Laden Waste Water by Recirculation through Activated Carbon," *Treatment of Pesticide Wastes, ACS Symposium Series*, Vol. 259, 1984, pp. 125-154.
- [100] W. H. Dennis and E. A. Kobylinski, "Pesticide-Laden Wastewater Treatment for Small Waste Generators," *Journal of Environmental Science and Health, Part B: Pesticides, Food Contaminants, and Agricultural Wastes*, Vol. 18, No. 3, 1983, pp. 317-331. [doi:10.1080/03601238309372372](https://doi.org/10.1080/03601238309372372)
- [101] J. C. Nye, "Treating Pesticide-Contaminated Wastewater, Development and Evaluation of a System," *Treatment and Disposal of Pesticide Wastes, ACS Symposium Series*, Vol. 259, 1984, pp. 153-160.
- [102] S. E. Dwinell, "Treatment of Pesticide Wastes: Regulatory and Operational Requirements for Successful Treatment Systems," *Pesticide Waste Management, ACS Symposium Series*, Vol. 510, 1992, pp. 113-126. [doi:10.1021/bk-1992-0510.ch009](https://doi.org/10.1021/bk-1992-0510.ch009)
- [103] W. H. Dennis, "A Practical System to Treat Pesticide-Laden Wastewater," *Proceedings: National Workshop on Pesticide Waste Disposal*, US Environmental Protection Agency, Denver, 1985.
- [104] J. C. Morris and W. J. Weber Jr., "Adsorption of Biochemically Resistant Materials form Solution," *Advanced Waste Treatment Research Program*, US Public Health Service, Cincinnati, 1964, AWTR-9 Report 999-WP-11
- [105] J. C. Morris and J. Weber Jr., "Adsorption of Biochemically Resistant Materials Form Solution," *Part II, US Public Health Service, Advanced Waste Treatment Research Program*, Cincinnati, 1966, WTR-16, Report 999-P-33.
- [106] D. M. Giusti, R. A. Conway and C. T. Lawson, "Activated Carbon Adsorption of Petrochemicals," *Journal-Water Pollution Control Federation*, Vol. 46, No. 5, pp. 1974-1979.
- [107] B. Sarkar, N. Venkateswralu, R. N. Rao and C. Bhattacharjee, "Treatment of Pesticide Contaminated Surface Water for Production of Potable Water by a Coagulation-Adsorption-Nanofiltration Approach," *Desalination*, Vol. 212, No. 1-3, 2007, pp. 129-140. [doi:10.1016/j.desal.2006.09.021](https://doi.org/10.1016/j.desal.2006.09.021)
- [108] V. K. Gupta, I. Ali Suhas and V. K. Saini, "Adsorption of 2,4-D and Carbofuran Pesticides Using Fertilizer and Steel Industry Wastes," *Journal of Colloid and Interface Science*, Vol. 299, No. 2, 2006, pp. 556-563. [doi:10.1016/j.jcis.2006.02.017](https://doi.org/10.1016/j.jcis.2006.02.017)
- [109] T. M. Word and F. W. Getzen, "Influence of pH on the Adsorption of Aromatic Acids on Activated Carbon," *Environmental Science and Technology*, Vol. 4, No. 1, 1970, pp. 64-67. [doi:10.1021/es60036a006](https://doi.org/10.1021/es60036a006)
- [110] K. D. Racke and C. R. Frink, "Fate of Organic Contaminants during Sewage Sludge Composting," *Bulletin of Environmental Contamination and Toxicology*, Vol. 42, No. 4, 1989, pp. 526-533. [doi:10.1007/BF01700232](https://doi.org/10.1007/BF01700232)
- [111] J. A. Petruska, D. E. Mullins, R. W. Young and E. R. Collins, "A Benchtop System for Evaluation of Pesticide Disposal by Composting," *Nuclear and Chemical Waste Management*, Vol. 5, No. 3, 1985, pp. 177-182. [doi:10.1016/0191-815X\(85\)90076-2](https://doi.org/10.1016/0191-815X(85)90076-2)
- [112] W. W. Rose and W. A. Mercer, "Fate of Insecticides in Composting Agricultural Wastes. Fate of Pesticides in Composted Agricultural Wastes," *National Canners As-*

sociation, Washington DC, 1968.

- [113] D. K. Singh, "Biodegradation and Bioremediation of Pesticide in Soil: Concept, Method and Recent Developments," *Biodegradation*, Vol. 48, 2008, pp. 35-50.
- [114] T. F. Castro and T. Yoshida, "Effect of Organic Matter on the Biodegradation of Some Organochlorine Insecticides in Submerged Soils," *Soil Science and Plant Nutrition*, Vol. 20, No. 4, 1974, pp. 363-370. doi:10.1080/00380768.1974.10432607
- [115] J. F. Brown, R. E. Wagner, H. Feng, D. L. Bedard, M. J. Brennan, J. C. Carnahan and R. J. May, "Environmental Dechlorination of PCBs," *Environmental Toxicology and Chemistry*, Vol. 6, No. 8, 1987, pp. 579-593. doi:10.1002/etc.5620060802
- [116] P. Adriaens, Q. Fu and D. Grbic-Galic, "Bioavailability and Transformation of Highly Chlorinated Dibenzo-*p*-Dioxins and Dibenzofurans in Anaerobic Soils and Sediments," *Environmental Science and Technology*, Vol. 29, No. 9, 1995, pp. 2252-2260. doi:10.1021/es00009a015
- [117] M. J. Zwiernik, J. F. Quensen and S. A. Boyd, "FeSO<sub>4</sub> Amendments Stimulate Extensive Anaerobic PCB Dechlorination," *Environmental Science and Technology*, Vol. 32, No. 21, 1998, pp. 3360-3365. doi:10.1021/es9801689
- [118] W. P. Muller and F. Korte, "Ecological Chemical Evaluation of Waste Treatment Procedures," *Environmental Quality and Safety*, Vol. 5, 1976, pp. 215-236.
- [119] P. F. Strom, "Pesticides in Yard Waste Compost," *Compost Science and Utilization*, Vol. 8, No. 1, 2000, pp. 54-60.
- [120] S. D. Cunningham, T. A. Anderson, P. A. Schwab and F. C. Hsu, "Phytoremediation of Soils Contaminated with Organic Pollutants," *Advances in Agronomy*, Vol. 56, 1996, pp. 55-114. doi:10.1016/S0065-2113(08)60179-0
- [121] E. L. Kruger, T. A. Anderson and J. R. Coats, "Phytoremediation of Soil and Water Contaminants," *ACS Symposium Series*, Vol. 664, 1997, pp. 1-4.
- [122] J. R. Coats and T. A. Anderson, "The Use of Vegetation to Enhance Bioremediation of Surface Soils Contaminated with Pesticide Wastes," US Environmental Protection Agency, Office of Research and Development, Washington DC, 1997. <http://www.epa.gov/>
- [123] R. Olette, M. Couderchet, S. Biagianni and P. Eullaffroy, "Toxicity and Removal of Pesticides by Selected Aquatic Plants," *Chemosphere*, Vol. 70, No. 8, 2008, pp. 1414-1421. doi:10.1016/j.chemosphere.2007.09.016
- [124] G. A. Buyanovsky, R. J. Kremer, A. M. Gajda and H. V. Kazemi, "Effect of corn Plants And Rhizosphere Populations on Pesticide Degradation," *Bulletin of Environmental Contamination and Toxicology*, Vol. 55, 1995, pp. 689-696. doi:10.1007/BF00203754
- [125] M. Gordon, N. Choe, J. Duffy, G. Ekuan, P. Heilman, I. Muiznieks, M. Ruszaj, B. B. Shurtleff, S. Strand, J. Wilmoth and L. A. Newman, "Phytoremediation of Trichloroethylene with Hybrid Poplars," *Environmental Health Perspectives Supplements*, Vol. 106, No. 4, 1998, pp. 1001-1004. doi:10.1289/ehp.98106s41001
- [126] K. Stearman, D. George, K. Carlson and S. Lansford, "Pesticide Removal from Container Nursery Runoff in Constructed Wetland Cells," *Journal of Environmental Quality*, Vol. 32, No. 4, 2003, pp. 1548-1556. doi:10.2134/jeq2003.1548
- [127] L. Wang, X. Jiang, D. Yan, J. Wu, Y. Bian and F. Wang, "Behavior and Fate of Chlorpyrifos Introduced into Soil-Crop Systems by Irrigation," *Chemosphere*, Vol. 66, No. 3, 2007, pp. 391-396. doi:10.1016/j.chemosphere.2006.06.038
- [128] F. K. Pfaender and M. Alexander, "Extensive Microbial Degradation of DDT *in Vitro* and DDT Metabolism by Natural Communities," *Journal of Agricultural Food Chemistry*, Vol. 20, No. 4, 1972, pp. 842-846. doi:10.1021/jf60182a045
- [129] M. J. Acea, C. R. Moore and M. Alexander, "Survival and Growth of Bacteria Introduced into Soil," *Soil Biology and Biochemistry*, Vol. 20, No. 4, 1988, pp. 509-515. doi:10.1016/0038-0717(88)90066-1