

Determination of 7 Indicator Polychlorinated Biphenyls (PCBs) Residues in Porphyra by Ultrasonic Extraction and Gas Chromatography (GC)

Liangliang Tian¹, Feng Han¹, Youqiong Cai¹, Cong Kong¹, Yongfu Shi¹, Yuan Wang¹, Guangxin Yang¹, Qianyun Zhan^{1,2}, Dongmei Huang^{1*}

¹Aquatic Product Quality Inspection and Test Center (Shanghai), Ministry of Agriculture of China, East China Sea Fisheries Research Institute, Shanghai, China

²College of Food Science and Engineering, Ocean University of China, Qingdao, China

Email: liangliangjia@126.com, hdm2001@126.com

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Abstract

A method was developed for determination of 7 indicators the polychlorinated biphenyls (PCBs) residues in porphyra by gas chromatography (GC) with electron capture detector (ECD). The PCBs were extracted with hexane/methylene chloride (1:1, v/v) by ultrasonic extraction and the samples were cleaned up by concentrated sulfuric acid and Alumina-N solid phase extraction cartridge. The analytes were quantified by an internal standard method. Under optimal experimental conditions, good linearity was observed in the range of 5 - 200 ng/mL and the correlation coefficients were 0.9994 - 0.9998. The limit of quantitation (LOQ) for target analytes ranged from 6.0 to 10.0 µg/kg. At the spiked levels of 10, 50, 100 µg/kg, the average recoveries ranged from 90.9% to 102% with the relative standard deviations 2.12% - 6.32%. The result showed that the proposed method was rapid, and could be used for the determination of the PCBs in porphyra.

Keywords

Polychlorinated Biphenyls (PCBs), Porphyra, Ultrasonic Extraction, Gas Chromatography (GC)

1. Introduction

The PCBs (Polychlorinated biphenyls) which were widely used in industry from 1929 to 1979 in world have caused global environmental pollution problems because of their their persistence and multiple dimensions of toxicity. Many offshore waters have been contaminated by the PCBs in our country, the concentration is more than 30 ng/L standards that was promulgated by the US EPA [1]. The PCBs may damage the immune system, reproductive system, nervous system with bioaccumulation and carcinogenicity in human body, and that were

*Corresponding author.

listed to be one of the 12 kinds of persistent organic pollutants to priority control in the Stockholm convention.

Porphyra contains rich vitamin, iodine, protein, inorganic salt, etc and can make food after drying. China accounts for the world first in porphyra output [2]. Porphyra is one of the main economic seaweed in our country. Porphyra lives in the high-water line of the shallow intertidal zone and grows best in the nitrogen-rich water (such as the sewage outfall), so it is easy to enrich organic pollutants and contaminated by the PCBs. In order to monitor the safe state of the porphyra, there is a need to establish a detection method.

Accurate quantification of PCBs requires efficient extraction and detection methods to minimize the interferences contributed by the matrix. The detection methods are mainly gas chromatography (GC) and gas chromatography mass spectrometry (GC/MS). The PCB recovery has been performed by Soxhlet extract [3], sonication techniques [4] and other newer techniques such as microwave extraction [5], supercritical fluid extraction [6], and accelerated solvent extraction (ASE) [7], which are time-consuming or require the corresponding instrument, and the sample types are shellfishes, seaweed, blue crab, treebark, fish, etc.. The porphyra contains chlorophyll, carotenoids, lutein, phycoerythrin, blue protein, etc. and almost no fat, that matrix is very different with any other organisms. Accordingly, it is needed to establish a method to detect the PCBs in the porphyra because there is no an effective method suitable for porphyra. So this paper describes a quick, effective, simple and accurate method to monitor the PCBs that the extraction method is soaking-ultrasonic and the detection is method GC.

2. Materials and Method

2.1. Materials and Instrument

HPLC grade solvents purchased from Mallinckrodt Baker, Inc were used in this study. PCB contaminants were purchased from Dr. Ehrenstorfer Company. Porphyra samplings were purchased from supermarket. Neutral alumina solid phase extraction cartridge (Alumina-N SPE, 500 mg/6 mL) were purchased from Agela Company.

The GC system consisted of an Agilent 7890A GC (Agilent Technologies, Shanghai, China), equipped with an electron capture detector (ECD). The GC was fitted with a J & W DB-5MS fused silica capillary column (5% phenyl polysiloxane non-polar stationary phase, 30 m, 0.25 mm i.d. and 0.25 μm film thickness) from Agilent (J&W Scientific, Folsom, CA, USA). Nitrogen with a purity of 99.999% was used as the carrier gas.

Stock solutions were prepared for all standard substances at 1.0 $\mu\text{g/mL}$ in n-hexane. Spiking and calibration mixtures at various concentration levels were obtained by combining aliquots of stock solutions and internal standard with n-hexane and stored at -20°C . The concentration of internal standard in all the calibration mixtures and final sample solutions was 40 ng/mL.

2.2. Method

About 2.5 g of porphyra sample were weighed in a 50 mL centrifuge tube and spiked with 40 ng PCB 198 as the internal standard. The extraction was made with two 25 mL aliquots of a mixture of n-hexane/dichloromethane (1:1 v:v) [8] for 2 h of soaking and 20 min of ultrasonic. Then each mixture was centrifuged at 8000 r/min for 8 min. The two extracts were joined together and evaporated in a rotary evaporator to dry at a low temperature and weak vacuum. The residue was dissolved by 8 mL n-hexane twice and then the dissolved solution was subjected to cleanup.

The dissolved solution was added 3 mL concentrated sulfuric acid and fully shocked for 2 min, Then the mixture was centrifuged at 2000 r/min for 5 min. The supernatant was normally passed through an Alumina-NSPE cartridge conditioning with 5 mL n-hexane without any pressure. The eluate was collected and another 3 mL n-hexane was used to rinse the analytes. The eluate was dried under a gentle nitrogen stream at 40°C . The residue was reconstituted with 1 mL n-hexane and mixed in a vortex stirrer.

2.3. GC Analysis

The final samples were detected on an Agilent 7890A GC, coupled to an electron capture detector (ECD). The column temperature program was as below: 150°C hold for 0 min, ramp at $15^\circ\text{C min}^{-1}$ to 200°C , hold for 0 min, ramp at 2°C min^{-1} to 270°C , hold for 2 min, ramp at $10^\circ\text{C min}^{-1}$ to 280°C , and hold for 1 min. The carrier gas (nitrogen) flow rate was 1.2 mL min^{-1} , injection port temperature was 280°C , and injection volume was 1 μL . The injection was made in the splitless mode with purge on after 0.75 min.

3. Results and Discussion

3.1. Extraction Method

The extraction methods of the PCBs widely used are Soxhlet extraction, accelerated solvent extraction (ASE) and the ultrasonic extraction at present. Soxhlet extraction time is so long that needs 24 h. ASE that runs at high temperature and high pressure conditions requires specialized equipment and cost high. Ultrasonic extraction can shorten the extraction time relative to the Soxhlet extraction and has no special requirements for conditions of use relative to the ASE, so that meets the requirements of improving the efficiency of extracting and easy to use. In this study, the result showed that the extracting effect of ultrasonic after soaking was better than ultrasonic without soaking. The recovery of the method of Soaking for 2 h and ultrasonic for 20min could reach 90%, so this extract method was used.

3.2. Optimization of Sample Purification

In order to determine the trace level of the PCBs, it is advantageous to eliminate any possible interference from samples. The composition of the impurities in Porphyra is very complex. There was some matrix to interfere the PCB 28 and PCB 153 to detect when only concentrated sulphuric acid was used to purify the sample, and it could not reduce interference by increasing the amount of sulfuric acid. With alumina-NSPE only, the solution after purification was not clear which proved that the impurities could not be completely removed. Combined with concentrated sulfuric acid and alumina-NSPE, the target analytes could be accurately analyzed. **Figure 1** presented the chromatograms of 7 chemicals at 10 $\mu\text{g}/\text{kg}$ of spiked level.

3.3. Method Performance

The calibration curves for detection of the target compounds were obtained by performing a linear regression analysis on standard solution using the ratio of standard area to internal standard area (PCB 198) against analyte concentrations ranging from 5 to 200 ng/mL containing 40 ng/mL internal standard. Good linearity was obtained for all analytes, with correlation coefficients of R^2 ranged from 0.9994 to 0.9998. The limit of detection (LOD), defined as the concentration which yield a signal-to-noise (S/N) equal to 3, ranged from 2 $\mu\text{g}/\text{kg}$ to 4 $\mu\text{g}/\text{kg}$ (**Table 1**). The limit of quantification (LOQ), defined as the concentration which yield an S/N equal to 10, ranged from 6 $\mu\text{g}/\text{kg}$ to 10 $\mu\text{g}/\text{kg}$ (**Table 1**). The analytes recovery of this procedure was evaluated by spiking 10, 50 and 100 $\mu\text{g}/\text{kg}$ of each standard analyte and 40 ng internal standard to sample at three levels in replicates of six. The results are listed in **Table 2**; the average recoveries of each compound ranged from 90.9% to 102%. The reproducibility of this method was represented by percent relative standard deviation (R.S.D.) at each level for each compound and these values are also summarized in **Table 2**. The results show that the method's precision was within 10%, which is very satisfactory.

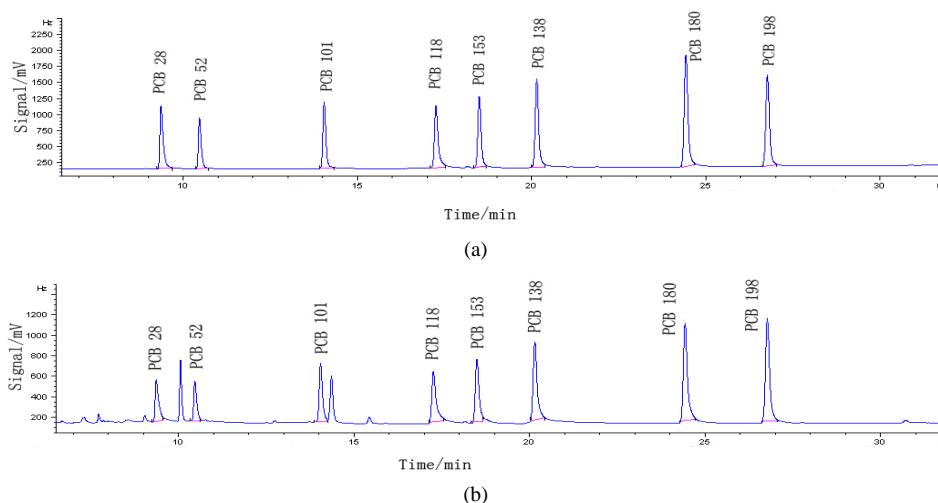


Figure 1. GC chromatograms of PCBs standard (a) and real sample (b).

Table 1. Linearity, the limit of detection (LOD) and the limit of quantification(LOQ) for 7 indicator PCBs.

compounds	Calibration equation	R ²	LOD (µg/kg)	LOQ (µg/kg)
PCB28	$Y = 0.5226X - 0.02819$	0.9994	4.0	10.0
PCB52	$Y = 0.3430X - 0.00171$	0.9997	4.0	10.0
PCB101	$Y = 0.4976X - 0.01042$	0.9997	3.0	9.0
PCB118	$Y = 0.6187X - 0.02979$	0.9996	3.0	9.0
PCB153	$Y = 0.5947X - 0.01647$	0.9997	3.0	9.0
PCB138	$Y = 0.8151X - 0.03589$	0.9994	3.0	9.0
PCB180	$Y = 1.0978X - 0.04450$	0.9998	2.0	6.0

Table 2. Recoveries and R.S.D. of spiked porphyra (n = 6).

compounds	Spiked level (10 µg/kg)		Spiked level (50 µg/kg)		Spiked level (100 µg/kg)	
	Recovery (%)	R.S.D. (%)	Recovery (%)	R.S.D. (%)	Recovery (%)	R.S.D. (%)
PCB 28	95.2	4.21	101	3.78	93.2	3.26
PCB 52	92.8	3.58	92.8	3.97	98.3	2.68
PCB 101	96.5	5.06	95.2	4.66	95.1	4.01
PCB 118	98.1	4.32	98.4	6.32	90.9	3.82
PCB 153	94.6	3.77	97.8	4.21	102	3.21
PCB 138	93.4	2.58	96.2	3.35	99.3	2.12
PCB 180	98.9	3.05	96.6	3.12	98.5	2.44

4. Conclusion

In this investigation, a comprehensive analytical method was developed for simultaneous extraction and determination of seven indicator PCBs in porphyra. The method clearly demonstrated good linearity, accuracy, and precision. Good recoveries ranging from 90.9% to 102% were reached. This new method may be suitable for the surveillance of the PCBs compounds in porphyra contaminated by the sea environment.

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