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## ABSTRACT

Partial Differential Equations (PDEs) have been already widely used to simulate various complex phenomena in porous media. This paper is one of the first attempts to apply PDEs for simulating in real 3D structures. We apply this scheme to the specific case study of the microbial decomposition of organic matter in soil pore space. We got a 3D geometrical representation of the pore space relating to a network of volume primitives. A mesh of the pore space is then created by using the network. PDEs system is solved by free finite elements solver Freefem3d in the particular mesh. We validate our PDEs model to experimental data with 3D Computed Tomography (CT) images of soil samples. Regarding the current state of art on soil organic matter decay models, our approach allows taking into account precise 3D spatialization of the decomposition process by a pore space geometry description.

Keywords: Partial Differential Equations; Soil; Microbial Decomposition; Pore Space; 3D Geometry Modelling; Computed Tomography

## 1. Introduction

It is a fact that soil structure is a heterogeneous media and organic matter decomposition in soil is one of the most complex ecological processes. The nature of organic matter, the dynamic of microorganisms, the environmental conditions etc. are the major factors which influence the decomposition of organic matter. Despite that microbial dynamics plays an important role in soil organic matter decomposition, the majority of organic matter models make the assumption that carbon limitation is controlled only by the intrinsic degradability of organic matter [1]. There are few models that take into account the rule of exoenzymes produced by microorganisms to convert complex substrates into available compounds [2]. Moreover, most of organic matter models do not consider the physical heterogeneity that controls partly the availability of organic matter to microorganisms [3].

In the recent contributions [4,5], the authors tried to simulate the diffusion of soluble carbon substrate in soil in 2D or 1D. Some attempts were also made to simulate enzyme diffusion in artificial structured environments [6,7]. But none of these models consider explicitly the real 3D soil structure. It is obvious that soil organic matter modeling needs to be in 3D soil structure because 1D or 2D representations of soil structure [8,9] can only supply partial information on the nature of overlapping [10]. Yet the improved performance of 3D X-ray computed tomography sensors now makes it possible to obtain very high resolution images of soil sample volume.



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Indeed, such non-destructive imaging technologies (X-ray tomography or CAT, laser scanning confocal microscopy or LSCM) can generate digital gray scale images that visualize 3D soil structure at aggregate [11] and also smaller intra-aggregate scale (16  $\mu$ m in [12]; 3  $\mu$ m in [13]).

Recently, the new model of Monga *et al.* in [14] simulated the decomposition of organic matter at a microscale using computer tomography images of real 3D soil but it does not take into account the diffusion process of dissolved organic matters. The main reason is that it costs a lot of time for computing diffusion process in 3D soil structure even with a very strong computer.

In this study we present an organic matter model that focuses on both transformation and diffusion processes in a 3D soil structure. We use Partial Differential Equations (PDEs) in order to simulate biological activity. In fact, models available in literature that simulates the biological activity in porous media use different techniques like cellular automata to simulate biomass growth in biofilms [15], Partial Differential Equations (PDEs) to simulate biomass recycling in fungal colonies [16] and multiagent system to simulate the impact of earthworms on soil structure [17]. In this work, we focus on the PDE method because 1) PDEs have been widely used to simulate various complex phenomena, particularly including transformation and diffusion processes in porous media but few studies have used this approach to biological soil systems; 2) the recently increasing performances of computer PDE solvers make it possible to simulate more and more complex systems.

This paper dealt with the application of PDEs to simulate organic matter degradation in real micro 3D structures and also took the diffusion into account. We tested our method using high resolution 3D CT of real soil images. Our work faces the problem of solving a nonlinear PDE system (reaction-diffusion) in a non-regular 3D mesh.

The paper is organized as follows. Section 2 presents the biological model. Section 3 presents the mathematical model of soil organic matter decay using PDE system. The PDE system is formed by reaction diffusion equations. The approximation of the model weak solutions is found by using a finite element method and a Newton algorithm. In Section 4, we implement the resolution algorithm by using Freefem3d software [18,19]. In Section 5, we present a validation of our PDE model to a experimental result. Section 6 presents a conclusion and some perspectives. Detailed Freefem3d code is given in **Appendix**.

## 2. Biological Model

Our aim is to simulate microbial decomposition in a nonregular 3D geometric volume shape defined by pore space. In this case pore space is described by a set of volume primitives [14,20-22]. Therefore, we take as input the following data:

- Geometrical representation of pore space using a network (attributed relational valuated graph) volume primitives [14,20-22].
- Parameters describing the initial spatial distribution of elements involved in biological activity: microbial biomass (MB), fructose (F), solid microbial wastes (SW), decomposed soluble microbial metabolites (DSM), un-decomposed soluble microbial metabolites (UDSM) and inorganic organic matter (CO<sub>2</sub>).

Thus, we assume that the microbial decomposition process involves six biological elements noted MB, F, SW, DSM, UDSM and CO<sub>2</sub>. We denote F the amount of fructose which is added into the environment. And we assume that it diffuses through water path to be consumed by microbial biomass (MB) for their maintenance and growth. It is supposed that MB does not move, so we assume that their diffusion coefficient is very small comparing with others. We further assume that the dead MB is transformed into three kinds of organic matters: solid microbial wastes (SWs) which cannot be consumed by MB and cannot diffuse, un-decomposed soluble microbial metabolites (UDSMs) which can diffuse but cannot be consumed by MB, decomposed soluble microbial metabolites (DSMs) which can be consumed again by MB and can diffuse. MB breathes to produce inorganic carbon (CO<sub>2</sub>) [2]. Figure 1 summarizes the biological process inside one spatial unit. The output of the simulation system for each step time is the precise distribution of biological activity parameters, i.e.: MB density, F density, SW density, DSM density, UDSM density and CO<sub>2</sub> density.

Therefore, we provide a kind of animated film showing spatially the evolution of biological characteristics. From these information, we can easily compute the classical global evolution curves:  $CO_2$  content, UDSM content, MB content...

## 3. Presentation of the Model Using PDE

Let  $\Omega \subset \mathbb{R}^3$  be the domain representing the soil pore space. Let  $t \ge 0$  be a given time and

 $x = (x_1, x_2, x_3)^T \in \Omega$  be a point of the pore space. We denote b(x,t), f(x,t),  $m_1(x,t)$ ,  $m_2(x,t)$ , n(x,t) and c(x,t) the densities of microbial (MB), of fructose (F), of solid microbial wastes (SW), of un-decomposed soluble microbial metabolites (UDSM), of decomposed soluble microbial metabolites (DSM) and of inorganic organize matter (CO<sub>2</sub>) at position  $x \in \Omega$  and at time t, respectively.

In the next subsections, we presented replicator equations for the variations of quantities of the six biological elements.



Figure 1. Biological processes inside one spatial unit (geometrical primitive).

## 3.1. Microbial Biomass (MB)

Let  $\mathcal{V}$  be an elementary volume in  $\Omega$ . The variation of the quantity of MB in  $\mathcal{V}$  is due to: microbial diffusion, microbial growth, microbial mortality and microbial breathing. Thus during the breathing, microbial decomposers lose a part of their masses. The following equation summarizes the process:

variation of 
$$b$$
=diffusion of  $b$ +growth of  $b$ -mortality and breathing of  $b$ 

We assume that the MB growth depends on the quantities of fructose (F) and of decomposed soluble microbial metabolites (DSMs). We also assume that microbial consumes F and DSMs according to the Monod equation, proposed in J.D. Murray's book [23], as follows:

$$\frac{\partial b}{\partial t} = \frac{kf}{K_b + f}b + \frac{kn}{K_b + n}b,\tag{1}$$

where the variables are set as follows. *b* is the concentration of microbial decomposers, *k* is the maximal growth rate,  $K_b$  is the half-saturation constant, *f* is the concentration of fructose, *n* is the concentration of decomposed soluble microbial metabolites.

The above equation describes microbial decomposers growth rate. Thus, the variation of b is expressed by the following equation:

$$\frac{\partial b}{\partial t} = D_b \Delta b + \left(\frac{kf}{K_b + f} + \frac{kn}{K_b + n} - \mu - r\right)b, \qquad (2)$$

where  $\Delta$  is the diffusion operator,  $D_b$  represents the microbial decomposers diffusion coefficient,  $\mu$  is set to the mortality rate, r is set to the breathing rate.

#### 3.2. Fructose (F)

F density variation comes from: F diffusion and consumption of F by microbial decomposers. Thus, the

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following equation summarizes F density variation process:

variation of 
$$n$$
  
= diffusion of  $n$  – growth of  $b$  (consumption)

F variation is ruled by the following equation:

$$\frac{\partial f}{\partial t} = D_f \Delta f - \frac{kf}{K_b + f} b, \qquad (3)$$

where  $\Delta$  is the diffusion operator,  $D_f$  represents the F diffusion coefficient.

#### 3.3. Solid Microbial Wastes (SWs)

SW quantity variation comes from the transformation of a part of MB mortality.

variation of 
$$m_1$$
 = a part of mortality of  $b$ .

Thus SW quantity variation is expressed by the following equation:

$$\frac{\partial m_1}{\partial t} = \alpha \mu b, \tag{4}$$

where  $\alpha$  ( $\alpha \in [0,1)$ ) is the rate of dead MB transformed into SW.

## 3.4. Undecomposed Soluble Microbial Metabolites (UDSM)

UDSM quantity variation is caused by the transformation of a part of MB mortality and the diffusion process as follows:

variation of 
$$m_2$$
  
= diffusion of  $m_2$  + a part of mortality of b.

UDSM quantity variation is expressed by the following equation:

$$\frac{\partial m_2}{\partial t} = D_{m_2} \Delta m_2 + (1 - \alpha - \beta) \mu b, \qquad (5)$$

where  $\Delta$  is the diffusion operator,  $1 - \alpha - \beta (1 - \alpha - \beta \in [0, 1))$  is the rate of dead MB transformed into UDSM.

## 3.5. Decomposed Soluble Microbial Metabolites (DSM)

DSM density variation comes from: DSM diffusion, a part of dead MB and consumption of DSM by microbial decomposers. Thus, the following equation summarizes DSM density variation process:

variation of 
$$n$$
 = diffusion of  $n$   
+ a part of mortality of  $b$   
- growth of  $b$  (consumption)

DSM variation is ruled by the following equation:

$$\frac{\partial n}{\partial t} = D_n \Delta n + \beta \mu b - \frac{kn}{K_b + n} b, \tag{6}$$

where  $\Delta$  is the diffusion operator,  $D_n$  represents DSM diffusion coefficient and  $\beta$  is the rate of dead MB transformed into DSM.

### 3.6. Inorganic Carbon (CO<sub>2</sub>)

Inorganic carbon  $(CO_2)$  variation is due to its diffusion and to its production by microbial decomposers (MB) during breathing.

variation of 
$$c$$
 = diffusion of  $c$  + production of  $c$ 

The CO<sub>2</sub> dynamics equation reads as follows:

$$\frac{\partial c}{\partial t} = D_c \Delta c + rb, \tag{7}$$

where  $\Delta$  is the diffusion operator,  $D_c$  is set to the CO<sub>2</sub> diffusion coefficient.

#### 3.7. Boundary Conditions

We assume that biological activities act only insde  $\Omega$ and there is not any outside forces that effect on the dynamics inside  $\Omega$ . It means that flow is null on  $\partial\Omega$ for all variables. Therefore, on the border of  $\Omega$  noted  $\partial\Omega$ , we use the Neumann boundary conditions.

#### **3.8.** The Complete PDE System

In this section we use the above equations describing variable variations to set a global PDE system modeling biological dynamics. Let T > 0 be a fixed time and let's define

$$\Omega_T = \Omega \times [0, T]. \tag{8}$$

Therefore, the whole system of partial differential equations governing the biological model becomes in  $\Omega_T$ :

$$\frac{\partial b}{\partial t} = D_b \Delta b + \left(\frac{kf}{K_b + f} + \frac{kn}{K_b + n} - \mu - r\right)b, \quad (9)$$

$$\frac{\partial f}{\partial t} = D_f \Delta f - \frac{kf}{K_b + f} b, \qquad (10)$$

$$\frac{\partial m_1}{\partial t} = \alpha \mu b, \tag{11}$$

$$\frac{\partial m_2}{\partial t} = D_{m_2} \Delta m_2 + (1 - \alpha - \beta) \mu b, \qquad (12)$$

$$\frac{\partial n}{\partial t} = D_n \Delta n + \beta \mu b - \frac{kn}{K_b + n} b, \qquad (13)$$

$$\frac{\partial c}{\partial t} = D_c \Delta c + rb. \tag{14}$$

We use Neumann homogeneous boundary conditions and the following initial conditions in  $\Omega$ :  $b(x,0) = b_0(x)$ for MB,  $f(x,0) = f_0(x)$  for F,  $m_1(x,0) = m_{10}(x)$  for SW,  $m_2(x,0) = m_{20}(x)$  for UDSM,  $n(x,0) = n_0(x)$ for DSM and  $c(x,0) = c_0(x)$  for CO<sub>2</sub>. The initial conditions are given by the functions of position x, *i.e.* amount of the biological elements at the beginning are known at any position in  $\Omega$ .

Therefore, the above PDE system describes precisely microbial decomposition of organic matter in soil. In the following section, we show how to solve this PDE system which can practically simulate soil biological activity.

## 4. Numerical Solution of the Complete PDE Model

#### **4.1. PDE Vectorial System Formulation**

We simplify the system writing by transforming it into a vector form. Let's define vectors in the following way:

$$u = (u_1, u_2, u_3, u_4, u_5, u_6)^{1},$$
  
=  $(b, f, m_1, m_2, n, c)^{t}$  (15)

$$u_0 = \left(b_0, f_0, m_{10}, m_{20}, n_0, c_0\right)^{\mathrm{T}}.$$
 (16)

The diffusion coefficients matrix  $\underline{D}$  is defined as follows:

The reaction terms of equations are represented by functions  $F_{i}$ ,  $i = 1, 2, \dots, 6$  defined as follows:

$$F_1(u) = \left(\frac{ku_2}{K_s + u_2} + \frac{ku_5}{K_s + u_5} - \mu - r\right)u_1, \quad (18)$$

$$F_2(u) = -\frac{ku_2}{K_a + u_2} u_1,$$
 (19)

$$F_3\left(u\right) = \alpha \mu u_1,\tag{20}$$

$$F_4(u) = (1 - \alpha - \beta) \mu u_1, \qquad (21)$$

$$F_{5}(u) = \beta \mu u_{1} - \frac{k u_{5}}{K_{s} + u_{5}} u_{1}, \qquad (22)$$

$$F_6(u) = ru_1. \tag{23}$$

Let's define the vector function F such that

$$F(u) = (F_{1}(u), F_{2}(u), F_{3}(u), F_{4}(u), F_{5}(u), F_{6}(u))^{\mathrm{T}}.$$
(24)

The vector form of the system is

$$\begin{cases} \partial_t u = div(\underline{D}\nabla u) + F(u) & \text{in } \Omega_T, \\ \frac{\partial u}{\partial n} = 0 & \text{on } \partial\Omega \times [0, T], \\ u(t=0) = u_0 & \text{in } \Omega. \end{cases}$$
(25)

#### 4.2. System Variational Formulation

Let's introduce the following Sobolev space

$$V = \left\{ v \in \left( H^1(\Omega) \right)^6 : \frac{\partial u}{\partial n} = 0 \text{ in } \partial \Omega \right\}.$$
 (26)

Assuming that data are sufficiently regular, the variational formulation consists in finding a function  $u(t) \in V$  such that:

$$\int_{\Omega} \frac{\partial u}{\partial t} v dx + \int_{\Omega} \underline{D} \nabla u \nabla v dx = \int_{\Omega} F(u) v dx \quad \forall v \in V.$$
(27)

#### 4.3. Resolution Scheme of the Variational System

#### 4.3.1. Building a Mesh $\Omega_h$ of Domain $\Omega$

A mesh of pore space  $(\Omega)$  can be built from a geometrical representation of pore space [14,20-22]. The following algorithm can then be use to build a mesh:

 Computing the geometrical primitives network representing pore space

We follow the approach described in [14,20-22]. In detail, we compute an adjacency graph representing pore space using 3D computed tomography image of a soil sample. Geometrical primitives can be chose, for instance, the balls as in [12]. Therefore, we obtain an adjacency graph of balls describing pore space where

each node is attached to a ball representing a pore within common sense [14,20-22].

• Building a mesh by using all centers of the balls

From all centers of the balls, we build a tetrahedra mesh for  $\Omega$ . In order to do that, we propose using the Delaunay triangulation program developed at INRIA by the GAMMA project [24,25].

Building the mesh following this way is exactly the same idea as in [14,22] where the transformation process acts inside each ball and the diffusion process acts between the connected balls. Instead of the geometrical modelling approach using updating graph in [14,22], here we consider in this work the mathamatical modelling approach. We are trying to use the benefits of the approach in order to better model biological activities.

#### • Pruning the mesh

We do pruning the mesh in order to neglect edges between two centers of balls that do not connect. Now we get a mesh  $\Omega_h$  of domain  $\Omega$ .

After building a mesh  $\Omega_h$  of domain  $\Omega$ , we solve the variational formulation (27) in the following discrete space:

$$V_{h} = \left\{ v \in \left( C\left(\overline{\Omega}\right) \right)^{6} : \forall K \in \Omega_{h} \ \left( v_{|K} \in P_{1} \right)^{6} \right\}.$$
(28)

#### 4.3.2. Numerical Scheme

The numerical resolution of the problem is divided into three steps:

 Step 1: We discretize the problem via finite element method in the finite dimensional space V<sub>h</sub>. The problem consists in solving the following system

$$\frac{\partial U}{\partial t} + BU = F(U), \qquad (29)$$

where

$$B_{i,j} = \int_{\Omega} \underline{D} \nabla \phi_i \nabla \phi_j \mathrm{d}x \quad \forall i, j = 1, 2, \cdots, Ndof, \qquad (30)$$

and

$$F(U)_{i} = \int_{\Omega} F_{i}(U) \phi_{i} dx \quad \forall i = 1, 2, \cdots, Ndof, \qquad (31)$$

where Ndof is the number of freedom degrees. The sequence  $(\phi_i)_{1 \le i \le Ndof}$  defines the basis functions of the space  $V_h$ .

• Step 2: We use implicit scheme to discretize time.

Let  $N_t$  be a positive integer denoting the number of time steps.

We call the step time  $\delta t = \frac{T}{N_t}$ .

The numerical scheme is:

$$\frac{U^n - U^{n-1}}{\delta t} + BU^n = F(U^n), \quad n \in \mathbb{N}^*,$$
(32)

which can be as follows:

$$(I + \delta tB)U^n = U^{n-1} + \delta tF(U^n), \quad n \in \mathbb{N}^*,$$
 (33)

where I is the identity matrix,  $U^n$  is the solution at time  $n * \delta t$ .

• Step 3: We linearize the function  $F(U^n)$ . We ignore  $U^n$  at the time  $n * \delta t$ . Therefore, we note  $U^*$  the approximation of  $U^n$  and  $\delta U$  the correction term. We obtain:

$$U^n = U^* + \delta U, \tag{34}$$

so the numerical scheme becomes:

$$(I+\delta tB)\delta U = U^{n-1} - U^* + \delta t \Big(F\Big(U^n\Big) - BU^*\Big).$$
(35)

We consider the following approximation:

$$F(U^n) \simeq F(U^*). \tag{36}$$

The numerical scheme becomes:

$$(I+\delta tB)\delta U = U^{n-1} - U^* + \delta t \left(F\left(U^*\right) - BU^*\right).$$
(37)

#### 4.3.3. Resolution Algorithm

From the initial conditions, we know  $U^0$ . Then in order to find solution  $U^n$  knowing  $U^{n-1}$ , we initialize  $U^*$ to  $U^{n-1}$ . Afterwards, we repeat the numerical scheme by replacing the term  $U^*$  by  $U^* \leftarrow U^* + \delta U$  after each iteration. Two stopping criteria are defined:

- we define a maximal number of iterations called *N* max;
- we define a very small positive real  $\epsilon_a$  and we stop the loop if the norm of  $\delta U$  is smaller than  $\epsilon_a$ .

When one of the stopping criteria is satisfied,  $U^*$  represents the searched solution. Thus, the resolution algorithm can be described as follows:

1) N max and  $\epsilon_a$  are fixed.

2) 
$$n \ge 1$$
,  $U^{n-1}$  is known

a)  $U^* \leftarrow U^{n-1}$ ,

b) the beginning of the loop for linearization. i) Solve

$$(I+\delta tB)\delta U = U^{n-1} - U^* + \delta t \left(F\left(U^*\right) - BU^*\right), \quad (38)$$

ii) Update

$$U^* \leftarrow U^* + \delta U, \tag{39}$$

iii) If the condition on  $\epsilon_a$  or N max is satisfied, the loop is stopped.

c)  $U^n \leftarrow U^*$ .

In **Appendix**, we give the Freefem3d code we implement for solving the system.

#### 5. Experimental Results on Real Data

We have validated our PDE model on real data of sand. The soil is sampled in the surface layer (0 - 20 cm) of an agricultural field at the INRA experimental site of Feucherolles (FEU) (50 km west of Paris) in France. The soil columns were scanned by means of a high resolution micro-CT machine (SIMCT Equipment: SIMBIOS, University of Abertay Dundee, Scotland) operating at 90 KeV and a current of 112 mA.

Using X-ray tomography we compute a 3D Computed Tomography (CT) image of a sand sample. The sizes of the original image are: 1650  $\mu$ m × 1650  $\mu$ m × 1745  $\mu$ m. The voxel is cubic of resolution: 5  $\mu$ m × 5  $\mu$ m × 5  $\mu$ m. **Figure 2** shows slices (1650  $\mu$ m × 1650  $\mu$ m) of the 3D volume image. We extract from the (1650  $\mu$ m × 1650  $\mu$ m × 1745  $\mu$ m) image a (400  $\mu$ m × 400  $\mu$ m × 400  $\mu$ m) image for memory requirements and computing time reasons. **Figure 3** shows successive slices (400  $\mu$ m × 400  $\mu$ m) of the (400  $\mu$ m × 400  $\mu$ m × 400  $\mu$ m) 3D image extracted.

The second stage provides a set of voxels corresponding to pore space. The threshold is computed using the expected porosity of the sample. **Figure 4** shows a cross section representing pore space (white color), the porosity is 35%. Using the geometric model for the 3D representation of the pore space described in [14,20-22], we calculate the network of balls.

Afterwards the balls are drained for the following values of water pressure  $\psi = -0.01$  MPa and  $\psi = -0.001$  Mpa corresponding respectively to situations that are relatively wet (high water pressure) and dry (low water pressure), respectively. For water pressure  $\psi = -0.01$  MPa we get 649776 balls and for water pressure  $\psi = -0.001$  MPa we get 569175 balls. Figure 5 shows perspective views of the ball based pore space representation. Figure 6 shows the tetrahedral mesh building from all the centers of the balls.

We built the model for two different values of water pressures (high water pressure, wet and low water pressure, dry). Initial microbial biomass (40 g) is put on the mesh such that probability of microbial being in a node is proportional to the radius of the bal which corresponds to the node. At the beginning, 300 g fructose is uniformly distribute in the mesh. Diffusion coefficients of fructose, CO<sub>2</sub> and DSM are equal to  $2 \times 10^6 \,\mu m^2/h$ [26] while diffusion coefficient of microbe is equal to  $10 \,\mu m^2/h$  which is very small in comparison with diffusion coefficient of the others since we assume that microbial does not move. We compare experimental carbon flow curves as well as dissolved organic matter (DOC) amount with the ones provided by our PDE model. We test our PDE model for five different types of microbial species: Arthrobacter sp 3R, 7R and 9R, Rhodococcus sp 6L and 5L. Parameter values for simulation are taken from the investigation of Coucheney in [27] which are given in Table 1.

Figure 7 shows an example of the simulation results obtained from the PDE model. The four classical global evolution curves ( $CO_2$ , DOC, microbial biomass and



Figure 2. Successive slices (1650  $\mu m$   $\times$  1650  $\mu m)$  of CT image of a sand soil sample.



Figure 3. Successive slices (400  $\mu$ m × 400  $\mu$ m) of the (400  $\mu$ m × 400  $\mu$ m × 400  $\mu$ m) 3D image extracted.



Figure 4. Cross section representing pore space (white color), the porosity is 35%.



Figure 5. Perspective views of the ball based pore space representation. We display only the balls whose radius is at least 10.



Figure 6. A tetrahedral mesh building from the centers of 1000 balls.

Table 1. Parameter values are used in the simulation where  $k(\text{day}^{-1})$ ,  $K_b(\text{gCg}^{-1})$ ,  $\mu(\text{day}^{-1})$  and  $r(\text{day}^{1})$ .

Species	k	$K_b$	μ	r	β	α
3R	17	0.0005	1.5	0.2	0.72	0.2
9R	9.6	0.001	0.5	0.2	0.55	0.2
7R	8	0.00014	0.2	0.3	0.20	0.2
6L	9	0.0005	0.22	0.45	0.20	0.2
5L	8.16	0.0007	0.4	0.25	0.55	0.2

fructose) are presented for *Rhodococcus* sp 6L. One can see in the figure that fructose was rapidly decomposed by microbial for both cases (high and low water pressures). It was completely decomposed around the second day. The microbial biomass is therefore grows fast in the first two days. It is then decreases the days after. Since growth of  $CO_2$  is proportional to the microbial biomass in our model. Hence, the amount of  $CO_2$  grows fast at the second day then it grows slower the days after. This result is similar to what we observe from the data of  $CO_2$  for the both cases with high and slow water pressures.

In **Figure 8**, we show a comparison between experimental data and our PDE model about  $CO_2$  and DOC of of *Arthrobacter* sp 3R, 7R and 9R. The left hand side is about wet condition (high water pressure) and the right hand side is about dry condition (low water pressure). Black solid and black dash are about  $CO_2$  and DOC predicted from the PDE model. Grey solid with square marks and tri-angle marks are about  $CO_2$  and DOC from



Figure 7. Simulation results of four biological elements of *Rhodococcus* sp 6L are presented. The above figure is about simulation with high water pressure and the below figure is about simulation with low water pressure. In the above figure: grey solid with square marks is about experimental  $CO_2$ , grey solid with tri-angle marks is about experimental DOC. In the below figure: grey solid with circle marks is about experimental  $CO_2$ , grey solid with tetra-angle marks is about experimental DOC. In the below figure: grey solid with circle marks is about experimental  $CO_2$ , grey solid with tetra-angle marks is about experimental DOC. Both: black solid is about  $CO_2$  from the model, black dash is about DOC from the model, grey dark solid is about microbial biomass, grey dark dots is about fructose.



Figure 8. Comparison between model and experimental data for *Arthrobacter* sp 3R, 7R and 9R. On the left hand side is about high water pressure, on the right hand side is about low water pressure. Solid line is about  $CO_2$  from the PDE mode, dash line is about DOC from the model, square is about experimental  $CO_2$ , circle is about experimental DOC.



Figure 9. Comparison between model and experimental data forfor *Rhodococcus* sp 6L and 5L. On the left hand side is about high water pressure, on the right hand side is about low water pressure. Solid line is about  $CO_2$  from the PDE mode, dash line is about DOC from the model, square is about experimental  $CO_2$ , circle is about experimental DOC.

experimental data in the wet condition (left), while grey solid with circle marks and tetra-angle marks are about  $CO_2$  and DOC from experimental data in dry condition (right). Similar comparison can be seen in **Figure 9** for *Rhodococcus* sp 6L and 5L. It is noticed that we assumed that the sum of decomposable and undecomposable bacteria metabolites (DMM + UDMM) is simulated at the end of the experiment in order to measure the bound value of the DOC. Therefore, the DOC-model is expected greater than DOC-data. One can see in **Figures 8** and 9 that DOC-model is greater than DOC-data in most cases except the case Arthrobacter 3R and 9R with high water pressure. It can be seen that  $CO_2$  fits very well to  $CO_2$ -data for all cases. It is sligtly different between the two with Arthrobacter 7R in low water pressure.

### 6. Conclusion and Perspectives

In this study we have proposed a new model of organic matter decomposition in the 3D soil pores. The novelties of our approach are to consider the real 3D microstructures of soil, to take into account diffusion and transformation processes. To our knowledge, this is the first attempt to describe diffusion process in 3D soil structure thanks to the strength of PDEs. We applied our PDE model to real CT images of soil in which we add realistic amount of organic matter and microorganisms. We validated our model to experimental results. In particular, we compare experimental carbon flow curves as well as DOC amount with the ones provided by our PDE model at two different water pressures. The simulation results show that our PDE model fits reasonably well to the experimental results. In this current contribution, we just consider the dynamics of single microbial species. In future studies, simulations with real experimental data will be carried out with our model in order to analyze microbial competition of degradation under different water contents.

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# Appendix. Algorithm Code Using Freefem3D

The freefem3d code corresponding to the algorithm is the following:

Step 1. Getting the mesh from the source file.

meshM = read(medit, "tetranuage.mesh")

**Step 2.** Defining the data. Functions  $f_i$ ,  $i = 1, 2, \dots, 6$  of type *femfunction* defined on the mesh M are used to represent reaction terms of the system. To solve the nonlinearity of the reaction functions, we use the same type to define approximate functions eb, ef,  $em_1$ ,  $em_2$ , en, and ec. The same function type is used to build terms of correction db, df,  $dm_1$ ,  $dm_2$ , dn, and dc. The parameters such as diffusion coefficients, growth and death rate of MB are declared using *double* type.

**Step 3.** Defining and initializing the functions representing variable densities. Initial functions are declared with the *femfunction* type defined on M. The *ofstream* function permits to create a file to save solutions. The quantities of the variables are obtained by integrating their density in the pore space represented by the mesh M. Thus, we use the following Freefem3d code to implement the numerical scheme:

// opening of a file named "outputpathname" to save results

of stream out put = of stream("outputpathname");

// saving of the time and initial quantities

$$output << 0 << " << int [M](b) << " << int [M](f) << " << int [M](m_1) << " << int [M](m_2) << " << int [M](n) << " << int [H](c) << "" << int [M](b + f + m_1 + m_2 + n + c) << endl;$$

// defining the time loop, nt is the number of time steps for  $(doublet = 1; t \le nt; t + +)$ 

// we initialize the iteration counter to solve the nonlinearity

k = 0;// we initialize approximate functions eb = b; ef = f;  $em_1 = m_1;$   $em_2 = m_2;$  en = n; ec = c;// loop for linearizing the system do {

//we redefine the reaction terms as functions of approximate terms.

// solving the system to find correction terms. // Db, Df, Dn, Dc are diffusion coefficients.

 $solve(db, df, dm_1, dm_2, dn, dc)inM$  $test(v_1, v_2, v_3, v_4, v_5, v_6)$  $int(db * v_1) + int(dt * Db * grad(db) * grad(v_1)) +$  $int(df * v_2) + int(dt * Df * grad(dn) * grad(v_2)) +$  $int(dm_1 * v_2) +$  $int(dm_2 * v_4) +$  $int(dn * v_5) + int(dt * Dn * grad(de) * grad(v_5)) +$  $int(dc * v_6) + int(dt * Dc * grad(dc) * grad(v_6))$  $int((b-eb+dt*f1)*v_1)$  $-int(dt * Db * grad(eb) * grad(v_1)) +$  $int((f-ef+dt*f2)*v_2)$  $-int(dt * Df * grad(ef) * grad(v_2)) +$  $int((m_1 - em_1 + dt * f3) * v_3) +$  $int((m_2 - em_2 + dt * f 4) * v_4) +$  $int((n-en+dt*f5)*v_5)$  $-int(dt * Dn * grad(en) * grad(v_5)) +$  $int((c-ec+dt*f6)*v_6)$  $-int(dt * Dc * grad(ec) * grad(v_6));$ } // updating of approximate functions eb = eb + db; ef = ef + df;  $em_1 = em_1 + dm_1;$  $em_2 = em_2 + dm_2 ;$ en = en + dn; ec = ec + dc; // incrementing of the counter k k = k + 1;// we stop the loop if one of the criteria is satisfied. } while  $((int [M](db+df+dm_1+dm_2+dn+dc) > \varepsilon_a))$  $\&(k \leq kmax))$ // updating the solutions b = eb; f = ef; $m_1 = em_1;$  $m_2 = em_2$ ; n = en; c = ec; // updating time counter T = T + dt;// We save solutions.

 $\begin{aligned} &output << T << "<< int [M](b) << " << int [M](f) << " \\ &< int [M](m_1) << " << int [M](m_2) << " \\ &< int [M](n) << " << int [M](c) << " \\ &< int [M](b + f + m_1 + m_2 + n + c) << endl; \\ &// We save the isovalues. \\ &save(medit, "b.", t, b, M); \\ &save(medit, "f.", t, f, M); \end{aligned}$ 

save(medit,"m<sub>1</sub> / m<sub>1</sub>."t,m<sub>1</sub>,M); save(medit,"m<sub>2</sub> / m<sub>2</sub>."t,m<sub>2</sub>,M); save(medit,"n / n."t,n,M); save(medit,"c / c."t,c,M); } //End of progam