

“Qi-Invigorating” Chinese Tonic Herbs (*Shens*) Stimulate Mitochondrial ATP Generation Capacity in H9c2 Cardiomyocytes *in Situ* and Rat Hearts *ex Vivo*

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ABSTRACT

In the study, using ethanol extracts of *Renshen* (*Panax ginseng* C A Meyer), *Xiyangshen* (*Panax quinquefolius* L.) and *Dangshen* (*Codonopsis pilosulae*), we investigated the effect of these “Qi-invigorating” Chinese tonic herbs on mitochondrial ATP generation capacity (ATP-GC) in H9c2 cardiomyocytes *in situ* and rat hearts *ex vivo*. All three types of “*Shens*” stimulated mitochondrial ATP-GC, with *Renshen* being most potent. While a parallel enhancement in mitochondrial ATP-GC was observed in *Renshen*- and *Xiyangshen*-pretreated rats, *Dangshen* treatment did not produce detectable effect *ex vivo*. The discrepancy between *in situ* and *ex vivo* assays for *Dangshen* may be attributed by its limited oral-bioavailability to the heart. The tissue specific activity of *Shens* on mitochondrial ATP-GC may be explained by the “Meridian Theory” in traditional Chinese medicine.

Keywords: Qi; Chinese Medicine; ATP; Mitochondria; Cardiomyocytes

1. Introduction

“*Renshen*”, a Chinese “*pinyin*” for Radix Ginseng (*Panax ginseng* C A Meyer), has a long history of use in Chinese medicine for promoting health. Early pharmacological investigations have demonstrated the adaptogenic effect of *Renshen*, which can enable the body to cope with various stress conditions [1]. Recent experimental studies have shown that *Renshen* or its ginsenosides produces anti-aging, anti-carcinogenic, anti-inflammatory and antioxidant actions [2-6]. According to the theory of Chinese medicine, Chinese tonic herbs can be classified into four functional categories, namely, “Yang-invigorating”, “Yin-nourishing”, “Qi-invigorating” and “Blood-enriching”, with various “*Shens*” being categorized in the “Qi” family. Due to the popularity and therapeutic values of “Qi-invigorating” herbs, the investigation of biological activities and the underlying mechanisms in relation to “Qi-invigoration” is of great pharmacological interest. In this regard, a recent study has demonstrated the relationship between “Qi-invigorating” action and energy level in skeletal muscle isolated from the thigh of “Qi-invigorating” herb-treated rats [7]. However, the pharmacological basis of “Qi-invigorating” action has yet to be established.

Previous experimental findings in our laboratory have

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demonstrated that all “Yang-invigorating” herbs are capable of enhancing mitochondrial ATP-GC in both cell and animal studies [8,9]. As a subcategory of “Yang-invigorating” herbs, “Qi-invigorating” herbs may also stimulate mitochondrial ATP generation capacity in various tissues. In the present study, using the ethanol extracts of *Renshen*, *Xiyangshen* (*Panax quinquefolius* L.) and *Dangshen* (*Codonopsis pilosulae*), we investigated the effect of “Qi-invigorating” Chinese tonic herbs on mitochondrial ATP-GC using *in situ* and *ex vivo* assay systems.

2. Materials and Methods

2.1. Herbal Materials

Renshen, *Xiyangshen* and *Dangshen* were purchased from a local herbal dealer (Lee Hoong Kee). The herbs were authenticated by the supplier and voucher specimens were deposited in the Division of Life Science, Hong Kong University of Science and Technology (HKUST). Individual herb was extracted by 95% ethanol under reflux for 2 h, as previously described [8], and obtained the ethanol extract. All the extracts were dried by evaporating the solvent under reduced pressure and the dried extracts at varied yields (see **Table 1**) were stored at 4°C until use.

Table 1. Yields of herbal extractions.

	Renshen	Xiyangshen	Dangshen
Yield (%)	10.7	16.5	41.5

All herbal extracts were obtained by ethanol extraction under reflux for 2 h.

2.2. Cell Culture

H9c2 cell, which is a subclone of the original clonal cell line derived from embryonic BD1X rat heart tissue, was purchased from American Tissue Culture Centre (ATCC). The cells were cultured as mono-layers in Dulbecco's Modified Eagle's Medium (DMEM) (Gibco BRL Life Technologies, Grand Island, NY, US), supplemented with 10% (v/v) fetal bovine serum (FBS), 100 IU/mL of penicillin (Sigma, St. Louis, MO), 100 µg/mL of streptomycin and 17 mM NaHCO₃. All cells were grown under an atmosphere of 5% (v/v) CO₂ in air at 37°C.

2.3. Measurement of ATP-GC *in Situ*

H9c2 cardiomyocytes were seeded at a density of 2.5×10^4 cells/well into 24-well microtiter plates. After the cell attachment, cells were incubated with herbal extracts for 4 h at 37°C. Control group was given vehicle (DMSO) only. After the incubation, the ATP-GC assay was performed as previously described [8]. In brief, the removal of herbal extract-containing medium was followed by cell membrane permeabilization with digitonin (50 µg/mL) in an incubation buffer (120 mM KCl, 5 mM KH₂PO₄, 2 mM EGTA, 10 mM HEPES, 0.1 mM MgCl₂, 0.5% BSA, pH 7.4). Pyruvate (15 mM), malate (5 mM) and ADP (10 µM) were added for mitochondrial ATP generation at 37°C, which was monitored at increasing time intervals ranging from 0 to 15 min. The reaction was terminated by the addition of 60 µL of perchloric acid (30%, w/v), and the reaction mixtures were then centrifuged at $540 \times g$ for 10 min at 4°C, and the supernatant were measured for ATP content by luciferase assay (ATPlite, PerkinElmer Inc., MA).

The mitochondrial ATP-GC of untreated control was estimated by computing the area under the curve of the graph (AUC₁) plotting ATP generated (nmol/mg protein) against time (0, 7.5 and 15 min) and expressed in arbitrary unit. For herbal extract-treated samples, AUC₁ values of increasing incubation times were normalized to a respective mean control value from untreated cells and expressed as percent control. The area under the curve (AUC₂) of the graph plotting percent control values against incubation time (7.5 and 15 min) was computed and expressed in arbitrary unit. Data of herbal extract-treated groups were expressed as percent control. The two-step data processing aims at minimizing the inter-animal and inter-assay variabilities under the present experimental conditions [10].

2.4. Animal Care

Female adult Sprague-Dawley rats (8-week old; 200 - 250 g) were maintained under a 12 h dark/light cycle at about 22°C and allowed water and food *ad libitum*. Experimental protocols were approved by the Research Practice Committee at HKUST. Animals were randomly divided into groups, with five animals in each. In the experiment, rats were administered intragastrically with herbal extracts at daily doses of 0.5 to 3 g/kg for 3 consecutive days and were killed by decapitation 24 hours after the last dosing. Control animals were administered with the vehicle (water) only. Hearts were excised from control or herbal extract-pretreated rats, mitochondrial fractions were then isolated and subjected to the measurement of mitochondrial ATP generation capacity (ATP-GC) *in vitro*.

2.5. Preparation of Mitochondrial Fractions

Myocardial mitochondrial fractions were prepared from heart tissue homogenates by differential centrifugation at 4°C. In brief, heart ventricular tissue samples were excised and rinsed with ice-cold heart sucrose buffer (320 mM sucrose, 1 mM EDTA, 50 mM Tris-base, pH 7.4). A 10% (w/v) heart homogenate was prepared by homogenizing the minced ventricular tissue with a teflon-glass homogenizer at 4000 rpm for 30 min. The homogenate was centrifuged at $600 \times g$ for 10 min at 4°C to remove nuclei and cell debris. The supernatant was then centrifuged at $9200 \times g$ for 30 min at 4°C to sediment the mitochondria. The pellets were resuspended in heart sucrose buffer and constituted the mitochondrial fractions.

2.6. Destruction of Contaminating ATP in Commercial ADP Preparation

The contaminating ATP was removed by an enzymatic procedure. To briefly describe, ATP was converted to ADP by reacting it with glucose at 25°C during a 18 h incubation in the reaction mixture (34 mM commercial ADP, 100 mM Tris-HCl buffer (pH 8.1), 2 mM MgCl₂, 3.4 mM glucose, 1 mM NADH, 5.6 U hexokinase/mL, and 6 U glucose-6-phosphate dehydrogenase/mL). To destroy the formed NADPH, the solution was acidified with 0.1 volume of 1 M HCl and allowed to stand at 25°C for 10 min. The solution was then alkalized by adding 0.017 volume of 6 M NaOH, and all enzymes were inactivated by heating the reaction mixture to 95°C for 15 min [11].

2.7. Measurement of ATP-GC

Aliquots (50 µL) of mitochondrial fractions (1 mg protein/mL) were mixed with 50 µL of substrate solution (3 mM pyruvate and 3 mM malate) and 25 µL of pretreated

ADP (30 mM) solution, and the reaction mixtures were incubated for increasing period of time (0, 10 and 20 min) at 37°C to allow mitochondrial ATP generation. The ATP content was then measured as described before.

2.8. Protein Assay

Protein concentration was determined by Bio-Rad protein assay kit (Bio-Rad, Hercules, CA), using bovine serum albumin as standard.

2.9. Statistical Analysis

All data were expressed as mean \pm standard error of the mean (SEM), unless otherwise specified. Data were analyzed by one-way analysis of variance (one-way ANOVA) and inter-group difference was detected by Least Significant Difference (LSD) when $p < 0.05$.

3. Results

Figure 1 shows that pretreatment with *Renshen* at concentrations ranging from 50 to 300 $\mu\text{g/mL}$ significantly increased the mitochondrial ATP-GC in a concentration dependent manner in H9c2 cardiomyocytes, with the extent of stimulation being 42% at 300 $\mu\text{g/mL}$. *Xiyangshen* and *Dangshen* extracts significantly stimulated mitochondrial ATP-GC by 26% and 25%, respectively, at the concentration of 300 $\mu\text{g/mL}$ (**Figures 2 and 3**).

As shown in **Figure 4**, pretreatment with *Renshen* extract at daily doses of 0.5 and 1 g/kg significantly enhanced the ATP-GC in mitochondria isolated from rat hearts in a dose-dependent manner, with the extent of

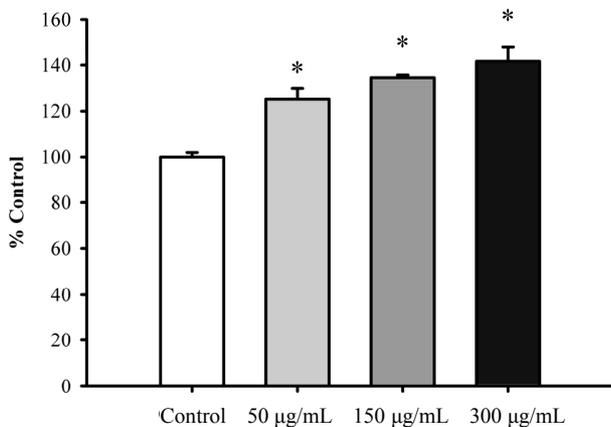


Figure 1. The effect of *Renshen* extract on mitochondrial ATP-GC in H9c2 cardiomyocytes. Cells were pre-incubated with *Renshen* extract at concentrations of 50 to 300 $\mu\text{g/mL}$ for 4 h. Upon the removal of herbal extract-containing medium, the cells were subjected to the measurement of ATP-GC *in situ*. Data were expressed in percent control with respect to the untreated control. Values given are means \pm SEM, with $n = 3$. *Significantly different from the control group.

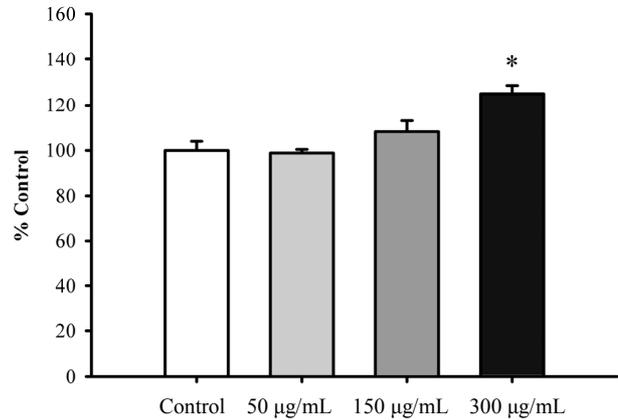


Figure 2. The effect of *Xiyangshen* extract on mitochondrial ATP-GC in H9c2 cardiomyocytes. Cells were pre-incubated with *Xiyangshen* extract at concentrations of 50 to 300 $\mu\text{g/mL}$ for 4 h. Mitochondrial ATP-GC was measured as described in **Figure 1**. Data were expressed in percent control with respect to the untreated control. Values given are means \pm SEM, with $n = 3$. *Significantly different from the control group.

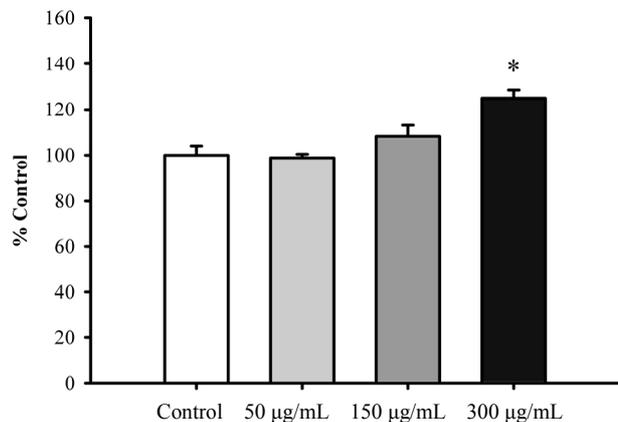


Figure 3. The effect of *Dangshen* extract on mitochondrial ATP-GC in H9c2 cardiomyocytes. Cells were pre-incubated with *Dangshen* extract at concentrations of 50 to 300 $\mu\text{g/mL}$ for 4 h. Mitochondrial ATP-GC was measured as described previously. Data were expressed in percent control with respect to the untreated control. Values given are means \pm SEM, with $n = 3$. *Significantly different from the control group.

stimulation being 9.3% and 22.9%, respectively. **Figure 5** shows that pretreatment with *Xiyangshen* extract at daily doses of 2 and 3 g/kg significantly increased mitochondrial ATP-GC, with the extent of stimulation being 13% and leveled at the high dose. However, *Dangshen* pretreatment at daily doses up to 3 g/kg did not produce any detectable effect on ATP generation in mitochondria isolated from rat hearts (**Figure 6**).

4. Discussion

In the present study, we demonstrated that the ethanol

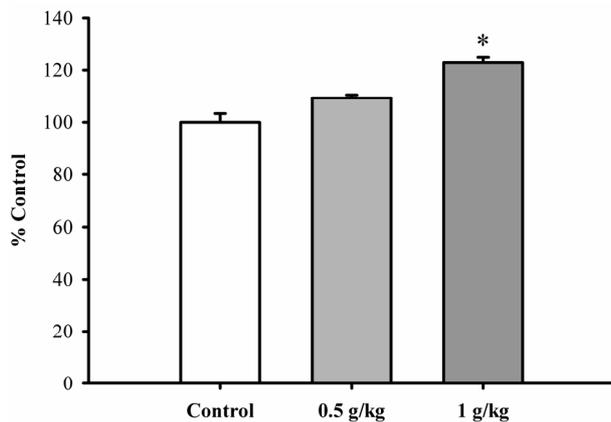


Figure 4. The effect of *Renshen* extract on ATP-GC in mitochondria isolated from rat hearts. Rats were orally administered with *Renshen* extract at daily doses of 0.5 and 1 g/kg for 3 consecutive days. Heart mitochondrial fractions were isolated and subjected to the measurement of ATP-GC *ex vivo*. Data were expressed in percent control with respect to the untreated control animals. Values given are means \pm SEM, with $n = 5$. *Significantly different from the control group.

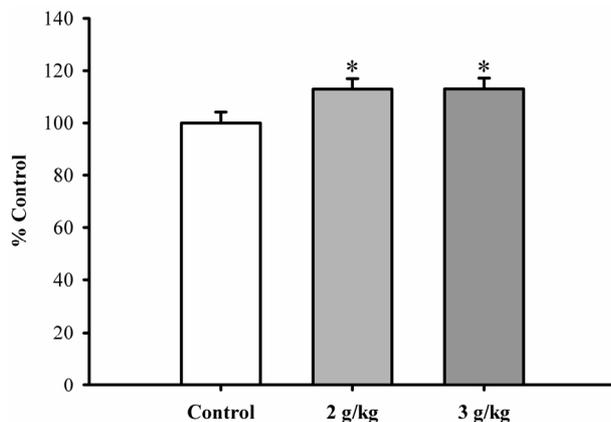


Figure 5. The effect of *Xiyangshen* extract on ATP-GC in mitochondria isolated from rat hearts. Rats were orally administered with *Xiyangshen* extract at daily doses of 2 and 3 g/kg for 3 consecutive days. ATP-GC of heart mitochondria was measured as described in Figure 4. Data were expressed in percent control with respect to the untreated control animals. Values given are means \pm SEM, with $n = 5$. *Significantly different from the control group.

extracts of all three types of “Shens” (namely, *Renshen*, *Xiyangshen* and *Dangshen*) increased mitochondrial ATP-GC in H9c2 cardiomyocytes, with *Renshen* being most potent among the tested “Shens”. This finding, which is consistent with the observation in “Yang-invigorating” herbs [9], suggested that “Qi-invigorating” herbs can also stimulate mitochondrial ATP-GC in H9c2 cardiomyocytes. The stimulation of ATP-GC by *Renshen* and *Xiyangshen* in the cell-based *in situ* assay was associated with a parallel enhancement of ATP-GC in mito-

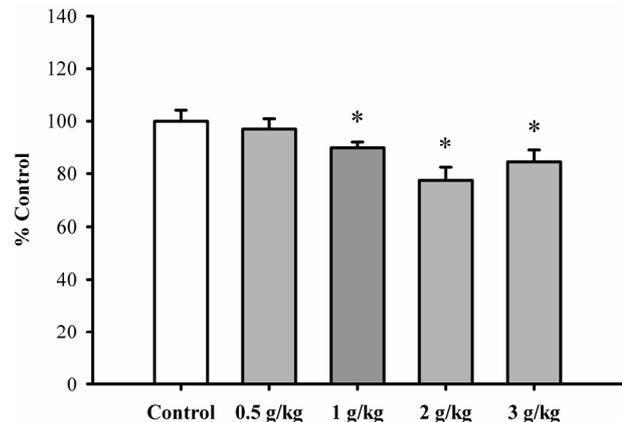


Figure 6. The effect of *Dangshen* extract on ATP-GC in mitochondria isolated from rat hearts. Rats were orally administered with *Dangshen* extract at daily doses ranging from 0.5 to 3 g/kg for 3 consecutive days. ATP-GC of heart mitochondria was measured as described previously. Data were expressed in percent control with respect to the untreated control animals. Values given are means \pm SEM, with $n = 5$. *Significantly different from the control group.

chondria isolated from *Renshen*- or *Xiyangshen*-pretreated rats, with the extent of stimulation by *Renshen* being larger than that of *Xiyangshen* at the same crude herb equivalent dose. However, *Dangshen* treatment did not cause any detectable change in ATP-GC in rat heart mitochondria. The discrepant observation between *in situ* and *ex vivo* assays for *Dangshen* on mitochondrial ATP-GC may be related to its limited oral bioavailability to the heart.

Dangshen has been used as a milder and more economical substitute for *Renshen*, and sometimes they are interchangeably used in many Chinese herbal formulations. However, a pharmacopoeia published in the Qing Dynasty differentiated *Dangshen* from the *Renshen* family, in which *Dangshen* belongs to the family of Campanulaceae while *Renshen* and *Xiyangshen* are grouped under the family of Aralaceae [12]. Although *Dangshen* shares similar biological activities with *Renhsen* and *Xiyangshen*, the tissue specificity varies in these two groups of herbs. It has been demonstrated that majority of the metabolites of *Renshen* and *Xiyangshen* could be found in the liver, heart and small intestine [1], while the action of *Dangshen* seemed to be limited to digestive, immune and respiratory systems [13-15]. Consistent with this, *Dangshen*, as observed in the present study, produced stimulatory effect on ATP-GC in H9c2 cardiomyocytes *in situ* but not in rat hearts *ex vivo*. In the context of Chinese medicine, this finding may be explained by the “Meridian Theory” (*Jingluo*), in which “Meridian” is the path that “Qi” (life energy) flows. According to the literature on *materia medica*, *Renshen* and *Xiyangshen* enter the “Meridians” of heart, spleen and lung

after oral administration and then bring about the beneficial effects to the respective organs. *Dangshen* only enters the “Meridians” of spleen and lung, but not heart, which may be attributed to its inability to stimulate the ATP-GC in rat heart mitochondria.

In conclusion, the results obtained from the present study showed that the three tested “Qi-invigorating” *Shens* in Chinese medicine stimulated the ATP-GC *in situ* in the cell-based assay system. Interestingly, the “Qi-invigorating” *Shens* herbs may produce the beneficial action in a tissue specific manner. Further investigations should examine whether “Yang-invigorating” and “Qi-invigorating” Chinese tonic herbs can stimulate mitochondrial ATP-GC *ex vivo* in a tissue specific manner according to the “Meridian Theory” in Chinese medicine.

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