## An Algorithm for Classification of Algal Blooms Using MODIS-Aqua Data in Oceanic Waters around India

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## ABSTRACT

Increasing incidences and severity of algal blooms are of major concern in coastal waters around India. In this work an automatic algorithm has been developed and applied to a series of MODIS-Aqua ocean color data to classify and monitor four major algal blooms in these waters (*i.e., Trichodesmium erythareum, Noctiluca scintillans/miliaris* (green/brown), and *Cochlodinium polykrikoides* (red)). The algorithm is based on unique spectral signatures of these blooms previously reported by various field sampling programs. An examination of the algorithm results revealed that classified blooms agree very well with in-situ data in most oceanic waters around India. Accuracy assessment based on overall, user's and producer's accuracy and Kappa accuracy further revealed that the producer's/user's accuracy of the four algal blooms were 100%/100%, 79.16%/79.16%, 100%/80%, 100%/86.95%, respectively. The Kappa coefficient was 1.01. These results suggest that the new algorithm has the potential to classify and monitor these major algal blooms and such information is highly desired by fishermen, fish farmers and public health officials in this region. It should be noted that coefficients with the new algorithm may be fine-tuned based on more in-situ data sets and the optical properties of these algal blooms in oceanic waters around India.

Keywords: Algal Blooms; Arabian Sea; MODIS; Automated Algorithm; India.

## **1. Introduction**

Green, red and brown algal blooms have been reported to increase spatially and temporally in many coastal and offshore waters around the world. Increased incidences and severity of such algal blooms could be due to nutriaent enrichment of these waters supplied from anthropogenic or natural sources, hydrographic changes, or climate change impacts [1-9]. The appearance, persistence and epidemic of some of these blooms have also been reported to cause fish mortality, shellfish poisoning, physiological impairment, and numerous ecological and health impacts [1,5] and references therein.

Over the past decades, an increase in the frequency of algal blooms has also been reported in coastal and offshore waters around India [10-17]. Major algal blooms dominating in this region are *Trichodesmium erythareum*, *Trichodesmium thiebautii*, *Noctiluca scintillans*, *Noctiluca milaris*, and *Cocholodinium ploykrikoides* [11,12,14, 15]. *Trichodesmium* is a typical genus of pelagic bluegreen algae, which has two species *T. thiebautii* and *T. erythareum*—both occurring mostly in tropical and subtropical seas. These species are known as nitrogen fertilizer because of their N<sub>2</sub> fixing action. They are distinguishable by their occurrence on the surface waters, and

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are a common feature from February to May when large areas of the sea (particularly Arabian Sea and coastal waters around India) are covered with clumps of sawdust colored algae features [17].

Until the late 1990s, N. miliaris, a large dinoflagellate, was a minor component of phytoplankton populations in the Arabian Sea, appearing sporadically in bloom form in coastal regions during the summer southwest monsoon (June-September). Recently, N. miliaris blooms have begun to appear with increased frequency and intensity following the winter northeast monsoon (November-February) [14]. Several cruises conducted by the National Institute of Oceanography (India) have also documented the appearance of extensive blooms of N. miliaris in the late winter to early spring for several consecutive years. N. scintillans is also published as N. miliaris which turns the water green or red depending on its pigment variations and associated constituents. Blooms of N. scintillans have been observed in coastal waters of Oman and India (northeastern Arabian Sea, Orissa coast, Kochi coast, and Gulf of Mannar) almost every year with variations in their abundances [10,12,15]). On the other hand, C. polykrikoides is a common ichthyotoxic "red water" bloom species associated with extensive fish kills and great economic loss in Japanese and Korean waters [5,7].



This species has become a persistent HAB (harmful algal bloom) problem in the Arabian Gulf and the Gulf of Oman and its recurrence may be related to increased nutrient enrichment of coastal waters from domestic and industrial inputs, natural meteorological and oceano-graphic forcings, and the recent introduction of this species through ballast water discharge [18].

Phytoplankton serves as the base of the aquatic food web, providing an essential ecological function for all aquatic life. However, there are algal blooms that have negative economic and health impacts. Therefore, it is very essential to identify the type of algal bloom that occurs in this region. Traditionally, algal blooms are recognized and monitored using ship-borne water sampling techniques at discrete locations, but such techniques cannot provide sufficient spatial and temporal coverage to monitor algal blooms. Satellite remote sensing has become a complementary tool for monitoring algal blooms with its synoptic coverage, frequent revisit, high observing efficiency and relatively low cost.

Many algorithms have been developed to estimate chlorophyll (Chl-a) concentrations (used as an index of phytoplankton) using satellite ocean color data (e.g., the SeaWiFS Ocean Chlorophyll four-band algorithm OC4v4; MODIS-Aqua Ocean Chlorophyll three-band algorithm OC3) [19,20]. While this approach has been valid in oceanic waters where a change in the concentration of Chl-a mainly causes a shift in the blue to green ratios of upwelling light fields [21], the ratios used in these algorithms can vary in response to factors besides Chl-a concentration and therefore introduce large errors in pigment retrievals from satellite data in coastal waters with high colored dissolved organic matter (CDOM) and suspended sediment (SS) contents [17,22,23]. A number of other op- tical methods have been developed and used with satelli- te data to provide a valuable tool for detection and mapp- ing of algal blooms ([24-32]). More recently, an improved algorithm (ABI) has been developed and thor- oughly validated by [9]. This algorithm appears to de- termine Chl-a concentrations helping to differentiate blooms from other constituents in complex waters. However, such a pigment algorithm is not intended for classifying the different types of algal blooms.

Our objective is to develop an automated algorithm that can be used for operational classification and monitoring of algal blooms in coastal and offshore waters around India. A sensitivity analysis is performed to evaluate the new algorithm using satellite data and field observations. Finally, the accuracy of classification algorithm is assessed and the results are discussed.

## 2. Data and Methods

#### 2.1. Satellite Data

The data used in this study were MODIS-Aqua Level 1A

(L1A) data of the Arabian Sea, Indian Ocean and Bay of Bengal of different periods (7 & 8 Oct. 2008, 5 April 2005, 24 Jan. 2006, 9 Feb. 2008, 23 Nov. 2008, 3 June 2009, 17 April 2009) obtained from the NASA Goddard Space Flight Centre (http://oceancolor.gsfc.nasa.gov/). These data were processed to obtain remote sensing reflectance Rrs (for the bands 412, 443, 488, 531, 547, 667, 678, 748, and 869 nm) using SeaDAS integrated with the CCS (Coastal Correction Scheme) scheme, [33]. These data were further processed using CAAS algorithm and the atmospheric correction problems were eliminated. Chlorophyll concentration was estimated using ABI algorithm [9]. The remote sensing reflectance data were further processed using a new algorithm to classify different types of algal blooms in oceanic waters around India.

#### 2.2. Field Data

Many field sampling programs have previously recorded and documented the different types of algal blooms in coastal waters around India (Table 1). Because all the photosynthetic phytoplankton contain Chl-a, the measurement of Chl-a is a routine work for monitoring phytoplankton blooms and its observations during different bloom periods is presented in Table 1. It should be recognized that Chl-a is an imprecise indicator of phytoplankton biomass so that even accurate determinations of Chl-a bear uncertain relationships to the abundance and species composition of phytoplankton [10]. Thus, consideration must be given to the various physical, chemical and biological properties associated with algal blooms. During 7 and 8 Oct. 2008, an intense bloom of N. scintillans was recorded in coastal waters of the Gulf of Mannar where it turned the waters dark green because of high cell concentrations (around  $13.5 \times 10^5$  cells L<sup>-1</sup>). The presence of N. scintillans was revealed by the microscopic examination. The size ranged from 400 to 1200 microns [15] and associated water temperature, salinity, dissolved oxygen and nutrients measured were characteristically different from surrounding waters (water temperature 29.5°C, salinity 34.2 psu, dissolved oxygen 4.86 ml·L<sup>-1</sup>, phosphate 8.28  $\mu$ g·L<sup>-1</sup> and ammonia 85  $\mu$ g·L<sup>-1</sup>) [15]. During 24 Jan 2006 and other occasions in winter, blooms of N. millaris were sampled and their associated physical, chemical and biological conditions (pH, nutrient, dissolved oxygen, Chl and cell concentrations) were recoded in the northern Arabian Sea [14,17]). In-situ studies [12] reported an intense bloom of C. polykrikoides on 23 Nov. 2008, which caused the fish kill in the Gulf of Oman and significantly diminished the dissolved oxygen in bloomed waters. On several occasions in 2008 and 2009, blooms of T. erythareum were observed with abnormal salinity and Chl-a levels along the southeast coast of India (Tamil

SNo	Name of Bloom	Date	Location	Lat/Long/Chl-a	Reference
1	T. erythareum	6 <sup>th</sup> to 20 <sup>th</sup> May 2005	From Mangalore to Quilon on the southwest coast of India	12°59'N and 74°31'E	Anoop et al. (2006)
2	N. scintillans	2 to 12 Oct 2008	Gulf of Mannar	09°16'47.6"N/79°11'17.1"E to 09°14'28.5"N/78°54'21.6"E/ 0.116 mg·m <sup>-3</sup>	Gopakumar <i>et al.</i> (2009)
3	<i>Noctiluca</i> Blooms	July to December 2008	South east coast of India	Mandapam & Keelakari coast	Anantharaman <i>et al.</i> (2010)
4	N. scintillans	5 April 2005	Orissa coast	Chl-a high	Mohanty <i>et al.</i> (2007)
5	T. erythareum	16th March 2007.	Kalpakkam waters on the southeast coast of India	12°33'N/80°11'E/Chl-a increased 20 times than the pre-bloom values.	Satpathy et al. (2007)
6	N. miliaris	3 <sup>rd</sup> -19 <sup>th</sup> Jan 2003, 27 <sup>th</sup> Feb 5 <sup>th</sup> Mar 2003)/24 Jan 2006	Arabian sea	Chl-a 0.63 mg $\cdot$ m <sup>-3</sup>	Gomes et al. (2008)
7	N. miliaris	17 Sept 2004.	Kerala coast	High Chl-a	Joseph et al. (2008)
8	C. polykrikoides	13 Jan 2009	Gulf of Oman		http://www.riob.fr/I MG/pdf/Muthanna_ Alomar.pdf
9	C. polykrikoides	23 Nov 2008	Gulf of Oman		Gheilani (2009)
10	T. erythareum	19 Feb 2008	Kalpakkam waters on the southeast coast of India	Chl-a 42.15 mg·m <sup>-3</sup>	Mohanty <i>et al.</i> (2010)
11	T.erythareum	29 May to 12 June 2009	Kollam and Kochi coast	(08°59.492'N, 75°59.334'E) (09°56.183'N, 75°54.948'E)	Padmakumar <i>et al.</i> (2010)

# Table 1. Different types of algal blooms recorded and reported by various studies in coastal and offshore waters around India.

Nadu coast), southwest coast of India (from Mangalore to Quilion and Kollam to Kochi), west coast of India (Goa and Gujarat) [11-13,17,34]. Similar observations

made by others reported these blooms to consistently occur in the same locations and spread to the offshore waters (**Table 1**).

## 3. Background of the New Algorithm

Remote sensing reflectance spectra were obtained from MODIS-Aqua data (7 & 8 Oct. 2008, 5 April 2005, 24 Jan. 2006, 23 Nov. 2008, 3 June 2009, 17 April 2009 and 9 Feb 2008) for four algal blooms (reported/known) in various coastal waters around India. These blooms are previously well-documented by various field sampling programs, namely *T. erythareum*, *N. scintillans*, *N. millaris*, and *C. polykrikoides*. It was found that the remote sensing reflectance spectra and their derivatives of each algal bloom have the unique signatures forming the basis of developing a new classification algorithm (**Figure 1**).

## 3.1. Spectral Analysis

The features of the reflectance curves are important as they give insight into the spectral characteristics of different algal blooms. Figure 2 displays the MODIS-Aqua reflectance spectra of four types of algal blooms that were sampled by many field programs in different coastal areas [11-13,15,17,34,35]. It is observed that the reflectance spectra of different algae are distinct because of their unique absorption and reflectance characteristics. All four algal blooms have a minimal value in the blue region which is due to the combined absorption by phytoplankton pigments, CDOM and non-algal particles (NAP); a maximal value in the green region due to the minimal values of total absorption; a minimal value in the red region (around 667 nm) due to chlorophyll absorption and a lower value in the near-infrared (> 750 nm) due to increased absorption by sea-water itself. Chlorophyll fluorescence is observed with its peak position at around 678nm, although it shifts towards the longer wavelengths owing to the species composition and concentration [36-39]. Trichodesmium contains both the Chl-a and bilin pigments (phycoerythrin and phyco- cyanin) which have characteristic absorption spectra. As an outcome of this, T. erythatreum shows a smooth variation in the magnitude from  $R_{rs}(531)$  to  $R_{rs}(443)$ . The reflectance values are high at 531 and 547 nm, which imply high pigment concentrations with high backscattering coefficients at these wavelengths. The peak at 678 nm is due to Chl-a and the peaks at 531 and 547 nm are due to bilin pigments and perhaps other pigments [40]. It was reported that during the bloom period ammonia was significantly high (about 392.80  $\mu$ ·mol·L<sup>-1</sup>), especially on the day of high cell density. This could be due to the diazotropic nature of Trichodesmium. Earlier reports showed a two fold increase of ammonia concentration in bloomed waters of Trichodesmium at the same locality [12].

The *N. miliaris* bloom shows a reflectance peak at  $R_{rs}(748)$ , which may be due to the combined effect of Chl-a absorption, shifted fluorescence and particulate backscattering. This peak could be caused by floating of

these algae in the water [28]. The N. scintillans bloom shows a decrease in reflectance (due to absorption) at  $R_{rs}(443)$ , a steep rise from  $R_{rs}(488)$  to  $R_{rs}(531)$ , and a linear line from  $R_{rs}(531)$  to  $R_{rs}(547)$ . A smooth rise towards  $R_{rs}(412)$  may be the result of atmospheric correction or algal matter itself. By contrast, the reflectance spectra of C. polykrikoides collected at different times (in coastal waters of Oman) are consistently the same .The spectral reflectance curves showed an overall increment in magnitude which is due to the increase in cell abundance and Chl-a concentration. The steepness in the range of 443 to 531 nm; a prominent peak at 547 nm; a distinct trough at 667 nm and a peak at 678 nm were also observed. Low  $R_{rs}(\lambda)$  values observed in the blue/green domain are due to the outcome of the interacting absorption characteristics of both chlorophyll and carotenoid pigments. Overall, the noticeable increment in  $R_{rs}(\lambda)$  that peaked at 547 nm is due to the outcome that the absorption by chlorophylls and carotenoids were minimal and in consequence backscattering by cells remained the main factor governing  $R_{rs}(\lambda)$  [36,39]. The position of this peak around 547 nm is considered a distinctive feature of chlorophyll containing algae and is regarded as an indicator of their presence in natural waters [36].

Different algal blooms absorb or reflect energy from different wavelengths in unique way; this provides the ability to identify the presence or absence of different algal blooms. An investigation of the distinctive spectral reflectance characteristics of different algal blooms allowed determining certain band ratios and reflectance differences and using these in the classification algorithm to classify four types of algal blooms from satellite data sets in oceanic waters around India. Since the classification algorithm is based on the specific reflectance signatures of different phytoplankton, the contribution of CDOM or SS to these reflectance features are expected minimal or their impacts are expected negligible in the classification outputs (mainly clustering of pixels rather than determining concentrations like Chl-a).

#### **3.2. Derivative Analysis**

Derivatives of second order or higher are relatively less sensitive to variations in illumination intensity, as well as spectral variations of sunlight and skylight (Tsai and Philpot, 1998). The derivative analysis, which amplifies spectral inflections and enhances detection of small spectral variations, can be used to closely examine the spectral reflectance patterns. This technique provides information regarding the convexity and concavity of a given reflectance spectrum. In this classification technique, we examined the second derivative of  $R_{rs}(\lambda)$  ( $d\lambda^2 R_{rs}$ ) to indicate the centre of the secondary peak and trough of  $R_{rs}(\lambda)$ .  $d\lambda^2 R_{rs}$  is derived using.

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Figure 1. Flowchart (referred as "neqn") describing the algal bloom classification algorithm.



Figure 2. Remote sensing reflectance spectra obtained from.

$$\frac{d^2 R_{rs}}{d\lambda^2} = \frac{(R_{rs}(\lambda_{i+1}) - 2R_{rs}(\lambda_i) + R_{rs}(\lambda_{i-1}))}{\Delta\lambda^2}$$
(1)

where the finite band resolution  $\Delta \lambda = (\lambda_i - \lambda_{i+1})$ . It was found that each bloom has distinct derivative spectra as shown in **Figure 3**. The derivative spectra have a negative maximum at 667 nm and a negative minimum value at 531 nm for the *T. erythareum* bloom. *N. scintillans* has two negative troughs at 531nm and 667nm of the derivative spectra. In the derivative spectra of *N. miliaris* a positive maximum peak is observed at 667 nm and a minimum peak at 531 nm. Two negative troughs at 531 nm and 667nm are observed in the derivative spectra of *C. polykrikoides*. In these derivative spectra the prominent peak at 667 nm reflects the absorption maxima of Chl-a, whereas the peaks appearing at 488 nm and 531 nm reflect the absorption feature of carotenoids [40].

In this analysis, two sets of reflectance difference/ratios along with a difference in derivative spectra were used to differentiate the four types of algal blooms, as follows:

$$\left[\frac{(R_{rs}488)}{(R_{rs}547 - R_{rs}488)}\right] * \left[d^2 R_{rs}488 - d^2 R_{rs}443\right]$$
  
for *T. erythareum* (2)

$$\left[\frac{(R_{rs}488)}{R_{rs}547 - R_{rs}488}\right] - \left[\frac{R_{rs}443}{R_{rs}547}\right] * \left[d^2 R_{rs}488 - d^2 R_{rs}443\right]$$

#### for N. scintillans, N. miliaris and C. polykrikoides (3).

A scatterplot of the output from Equations (2) and (3) (Figure 1) versus reflectance difference (547 - 443) is shown in Figure 4 where all four types of algal blooms

are well clustered and distinguished. The means of each cluster of the algal blooms were calculated and for each algal bloom type the distances toward class means were calculated (**Figure 4**).

Then, the classification was performed as follows:

The shortest distance to a class mean was found

• If the shortest distance to a class mean was nearest to the user-defined threshold, then this class name is assigned to the output pixel.

• Else the undefined value was assigned to the nearest algal bloom type.

#### 4. Accuracy Assessment

The error matrix can be used for a series of descriptive and analytical statistical techniques. Perhaps the simplest descriptive statistic is overall accuracy which is computed by dividing the total correct (*i.e.*, the sum of the major diago being correctly classified and is really a measure of nal) by the total number of pixels in the error matrix. In addition, accuracies of individual categories can be computed in a similar manner. However, this case is a little more complex in that one has a choice of dividing the number of correct pixels in that category by either the total number of pixels in the corresponding row or the corresponding column. Traditionally, the total number of correct pixels in a category is divided by the total number of pixels of that category as derived from the reference data (i.e., the column total). This accuracy measure indicates the probability of a reference pixel omission error. This accuracy measure is often called "producer's accuracy" because the producer of the classification is interested



neqn

Figure 4. Scatterplot of neqn (Figure 1) vs  $R_{rs}$  difference at 547 and 443 nm.

in how well a certain area can be classified. If the total number of correct pixels in a category is divided by the total number of pixels that were classified in that category, then this result is a measure of commission error. This measure, called "user's accuracy" or reliability, is indicative of the probability that a pixel classified on the image actually represents that category on the ground [41-43].

Thus, an error matrix was generated for the classification technique. Each row of the table was reserved for one of the remote sensing class used by the classification algorithm. Each column displays the corresponding ground truth classes in an identical order. This table was used to properly analyze the validity of each class as well as the classification as a whole. In this way one can evaluate in more detail the efficacy of the classification

Overall accuracy = 
$$\left(\frac{D}{N}\right) \times 100\%$$
 (4)

D: Total number of correct classifications,

N: Total number of classifications.

The overall accuracy of the classification technique was calculated. But just because 90% classifications were accurate overall, does not mean that each category was successfully classified at that rate. So the accuracy of each class type was also performed. The user's accuracy and the producers accuracy was calculated;

User's Accuracy = 
$$\left(\frac{D}{R}\right) \times 100\%$$
 (5)

*R*: Number in row total.

The User's Accuracy was computed for each row;

Producer's Accuracy = 
$$\left(\frac{D}{C}\right) \times 100\%$$
 (6)

*C*: Number in column total.

The Producer's Accuracy was computed for each column. The user's accuracy and producer's accuracy for the four types of algal blooms (*T. erythareum*, *N. scintillans*, *N. millaris*, and *C. polykrikoides*) were calculated.

Another measure of map accuracy is the Kappa coefficient, which is a measure of the proportional (or percentage) improvement by the classifier over a purely random assignment to classes. For an error matrix with *r*-rows, and hence the same number of columns, where A = the sum of r diagonal elements,

 $B = \text{sum of the r products (row total \times \text{column total)}}.$ 

Kappa Coefficient, 
$$\hat{K} = \frac{NA - B}{N^2 - B}$$
 (7)

where N is the number of pixels in the error matrix (the sum of all r individual cell values). If the Kappa coefficient is 1, then the classification can be said to be 100% correct.

#### 5. Results

The new algorithm was applied to a series of MODIS-Aqua data to classify different types of algal blooms in coastal oceanic waters around India (Arabian Sea, Indian Ocean and Bay of Bengal). The scattering particles that cause the water to be turbid can be composed of many things, including sediments and phytoplankton. In addi-

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tion to the spectral criteria for each bloom, thresholds were thus applied to exclude extremely turbid waters in coastal regions (e.g., the Gulf of Kutch and Cambay) where the concentration of suspended sediments is consistently high through out the year and dominates phytoplankton populations. Of several examples, an intense bloom of *N. scintillans* was observed in coastal waters of the Gulf of Mannar where it turned the waters dark green (field data on the right of **Figure 5**) because of high cell concentrations (around  $13.5 \times 10^5$  cells·L<sup>-1</sup>) [15]. The classified bloom was consistent with the field data, ABI\_Chl-a and color composite images and appeared to spread along the coast of Gulf of Mannar during 2 to 13 Oct. 2008; no cloud-free scenes were available except 8 Oct. 2008 (**Figure 5**).

It was noted that the occurrence and persistence of this bloom at the narrow strip of coastal area from Kilakarai to Pamban were due to the favorable environmental conditions, i.e., high air temperature, high sea surface temperature and salinity, low pH, absence of water currents, high concentration of nutrients, absence of rain and the favorable wind along the shore. All these factors influenced the sustenance of the bloom around the Islands and coastal waters, where the cell density ranged from 5.1  $\times$  $10^5$  cells·L<sup>-1</sup> to  $13.5 \times 10^5$  cells·L<sup>-1</sup> [15]. They also observed high suspended sediments and Chl-a (115.89  $\mu g \cdot L^{-1}$ ) during the bloom event. The environmental parameters during the waning phase of this bloom were notably at increased levels; e.g., surface water temperature 29.5°C, salinity 34.2 psu, dissolved oxygen 4.86 ml L<sup>-1</sup>, phosphate 8.28  $\mu$ g L<sup>-1</sup> and ammonia 85  $\mu$ g L<sup>-1</sup>[15].

During 5 April 2005, an intense *N. scintillans* bloom was detected by MODIS-Aqua data in coastal waters off the Rushikulya River mouth (**Figure 6**). The color composite image showed the linear spread of the bloom and it can be confirmed by the ABI\_Chl-a image (**Figure 6**, bottom panels). An *in-situ* study observed a prominent discoloration of the surface water in this region caused by dense and red-colored patches of this bloom of approximately 16 square kilometres (Lat.  $19^{\circ}22$ 'N and Long.  $85^{\circ}02$ 'E) [12]. A relatively low-moderate temperature (26.7°C to 30.6°C) was reported to trigger the appearance of N. scintillans bloom in this area.

**Figure 7** shows an example of N. milaris bloom with consistent spatial patterns (in the ABI\_Chl-a and color composite images) in the Gulf of Oman during 24 Jan. 2006. Field observations indicate that the bloom covered a wide area and turned the water dark green with streaks of red patches. It persisted for a fortnight in the Gulf and accounted for almost 69% of the phytoplankton population at off station on 24 Jan. 2006.

The MODIS-Aqua image on 23 Nov. 2008 showed an intense bloom of *C. polykrikoides* covering a wide area from coastal waters of Oman to the Gulf of Aden (**Figure 8**).



Figure 5. Algal bloom map of the Gulf of Mannar generated from MODIS-Aqua data (8 Oct. 2008) using the new algorithm (for detecting *N. scintillans* bloom). Right image is the field photograph of this bloom which was sampled by Gopakumar *et al.* (2009) during this period. Location: Lat. 09'14'28.5''N, Long. 78'54'21.6''E. Note that the color scale used for ABI\_Chla is kept same for other images in Figures 6-11. In the color composite image, white color represents sediment dominated waters and red color represents bloom waters.

This bloom was responsible for a massive fish kill in these waters, where dissolved oxygen dropped from 5  $\text{mg}\cdot\text{L}^{-1}$  to 0.1  $\text{mg}\cdot\text{L}^{-1}$  within a day, [35].

An example of *T. erythareum* bloom detected from MODIS-Aqua data in the southeastern Arabian Sea during the onset of the southwest monsoon (3 June 2009) is shown in **Figure 9**. Field observations confirmed that the bloom developed off Kollam (08°59.492'N, 5°59.334'E), with a pale brown to pinkish red surface water discoloration, spreading over an area of approximately 10 km<sup>2</sup> on 3 June 2009. Pale brown indicated healthy algae at the peak of its photosynthetic activity, while pinkish red indicated the presence of photosynthetically less active filaments. The bloom area was very fertile with copious quantities of dissolved oxygen (6.85 ml·L<sup>-1</sup>), PO<sub>4</sub>-P (0.108  $\mu$ mol·L<sup>-1</sup>) and SiO<sub>4</sub> (1.29  $\mu$ mol·L<sup>-1</sup>). Lower NO3-N (0.028  $\mu$ mol·L<sup>-1</sup>) values in the bloom area did

not appear to affect *T. erythareum* growth from molecular nitrogen fixation. However, lower NO3-N values altered the normal phytoplankton composition of this area [17]. *T. erythareum* bloom is a very common phenomenon in coastal waters of the eastern Arabian Sea (Mangalore to Bombay), and is well-documented by scientists at the National Institute of Oceanography (**Figure 10**) [44]. It is interesting to note that a filament pattern of *T. erythareum* bloom and their spatial distribution is clearly captured in the ABI\_Chl-a and color composite images.

The spatial distribution of *T. erythareum* bloom is shown in **Figure 11** for Kalpakkam coastal waters of the Bay of Bengal (19 Feb 2008). An *in-situ* study indicated that this bloom appeared during the relatively high temperature and salinity (> 31 psu) conditions, low nitrogen and high phosphate and total phosphorus conditions (Mohanty, *et al.* [12]). As an effect, the contribution of



Figure 6. Algal bloom map of the Orissa coast (Rushikulya River mouth) on the Bay of Bengal generated from MODIS-Aqua data (5 April 2005) using the new algorithm (for detecting *N. scintillans* bloom). Right image is the field photograph of this bloom which was sampled by Mohanty *et al.* (2007) during this period. In the color composite image, white color represents sediment dominated waters and red color represents bloom waters.



Figure 7. Algal bloom map of the Gulf of Oman generated from MODIS-Aqua data (24 Jan. 2006) using the new algorithm (for detecting *N. milaris* bloom). Right image is the field photograph of this bloom from Gomes *et al.* (2008) during this period.



Figure 8. Algal bloom map of the Gulf of Aden and Oman generated from MODIS-Aqua data (23 Nov. 2008) using the new algorithm (for detecting *C. polykrikoides* bloom). Right image is the field photograph of this bloom which was sampled by Gheilani (2009) during this period.



Figure 9. Algal bloom map of the eastern Arabian Sea generated from MODIS-Aqua data (3 June 2009) using the new algorithm (for detecting Trichodesmium A bloom). Right image is the field photograph of this bloom which was sampled by Padmakumar, *et al.* (2010) during this period. In the color composite image, white color represents sediment dominated waters and green color represents bloom waters.

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*T. erythareum* to phytoplankton density ranged from 7.79% to 97.01% and concentrations of chlorophyll and phaeophytin increased abnormally (42.15 mg·m<sup>-3</sup> and 46.23 mg·m<sup>-3</sup> respectively) during the bloom period [12].

To validate the efficacy of the new algorithm, accuracy assessment was performed for all four types of algal blooms. The overall accuracy was found to be 92.3%. But just because 93.2% classifications were accurate overall, does not mean that each category was successfully classified at that rate. Thus, the accuracy of each bloom type was also calculated. The user's accuracy and the producer's accuracy were 100% and 100% for C. polykrikoides, 79.16% and 79.16% for N. scintillans, 100% and 80% for N. millaris, 100% and 86.95% for T. erythareum (Table 2). The Kappa coefficient was 1.01. These statistics demonstrate that the new algorithm is very effective in terms of detecting and classifying all five algal bloom types, making MODIS-Aqua data offer unrivalled utility in monitoring the algal blooms in coastal and offshore waters around India.

Overall Accuracy =  $((13+19+8+20)/65) \times 100 = 92.3\%$ Producer's Accuracy, User's Accuracy

 $CP = (13/13) \times 100 = 100\%, CP = (13/13) \times 100 = 100\%,$ 

NS =  $(19/24) \times 100 = 79.16\%$ , NS =  $(19/24) \times 100 = 79.16\%$ ,

 $NM = (8/10) \times 100 = 80\%$ ,  $NM = (8/8) \times 100 = 100\%$ ,

TE =  $(20/20) \times 100 = 100\%$ , TE =  $(20/23) \times 100 = 86.95\%$ ,

Kappa Coefficient = NA-B/N<sup>2</sup>-B, N = 65: A = (13+19+8+20) = 60, B =  $((13 \times 3) + (19 \times 9) + (8 \times 8) + (20 \times 20)) = 674$ , Kappa Coefficient = 1.01.

## 6. Discussion

Recently, a new bio-optical algorithm was developed to provide quantitative assessments of (chlorophyll) of algal blooms in complex waters around India [9]. This algorithm seems reasonable in discriminating pigment patches from other water constituents, but is not intended to classify the different types of algal blooms in these waters. In this study, an automated algorithm involving certain band ratios and band differences has been developed to classify and monitor four dominant algal blooms (T. erythareum, N. scintillans, N. milaris, and C. polykrikoides) in coastal and offshore waters around India. The band ratios/differences and criteria used in this algorithm are based on the unique spectral signatures of each bloom type. It should be noted that atmospherically distorted and improbable negative values at  $R_{rs}(412)$  and  $R_{rs}(678)$  caused by the standard atmospheric correction were neglected or flagged out. The grey color in the classified image and black color in the ABI-Chl image represent cloud/no data in the Figures 5-11. The performance of this algorithm was tested on several MODIS-Aqua imageries and the classified blooms were validated with the *in-situ* data, chlorophyll image and color composite images. Comparison with previous studies showed that the spectral shape of the reflectance spectra of C. polykrikoides from this study were similar to those of the same species documented in coastal waters of Korea [30] and at Bahía Fosforescente [22]. Its absorption and back scattering properties were already described by Kutser 26. However, the reflectance spectra of other species/bloom types were quite different from those reported in the previous studies from other regions. It is important to note that the difference in reflectance spectra could be attributed to many factors, such as pigment concentration, atmospheric correction, bloom patchiness and sub-pixel variability (MODIS-Aqua pixel resolution of 1.1 km may include phytoplankton, suspended sediments and clear waters). It is therefore difficult to compare satellite pixel measurements of reflectance with the in-situ point measurements. The algal bloom classified as T. erythareum (in Figure 9) by the new algorithm was consistent with field measurements, which showed that the surface water discoloration was caused by the accumulation of T. ervthraeum and that water column also contained a colony of T. thiebautii [17]. The surface water color in the bloomed region varied from pale brown to pinkish red. Pale brown indicated healthy algae at the peak of its photosynthetic activity, while pinkish red indicated the presence of photosynthetically less active filaments. The T. erythareum bloom was observed off Kollam (08° 59.492'N, 75°59.334'E), with a pale brown surface water discoloration, spreading over an area of approximately 10 km<sup>2</sup> on 3 June 2009. Qualitative and quantitative analyses of bloom samples revealed that in bloomed waters Trichodesmium erythraeum contributed 90% of the surface phytoplankton population. The remaining 10% was predominantly composed of diatoms and dinoflagellates. Cell density was  $1.14 \times 10^6$  filaments L<sup>-1</sup>,  $1.968 \times$  $10^6$  filaments L<sup>-1</sup> and  $1.51 \times 10^6$  filaments L<sup>-1</sup> at blooms sites off Kollam. [17]. The MODIS-Aqua cholorphyll image and the color composite image also revealed the presence of T. erythareum bloom along the Kollam coast.

T. erythareum (Figure 10), which was classified along

Table 2. Accuracy assessment of the new algorithm for different types of algal blooms in coastal and offshore waters around India.

	СР	NS	NM	TE	Total
СР	13	0	0	0	13
NS	0	19	0	0	19
NM	0	2	8	0	10
TE	0	3	0	20	23
Total	13	24	8	20	65

CP: C. polykrikoides; NS: N. scintillans; NM: N. milaris; TE: T. erythareum.



Figure 10. Algal bloom map of the eastern Arabian Sea generated from MODIS-Aqua data (17 April 2009) using the new algorithm (for detecting Trichodesmium A bloom). Right image is the field photograph of this bloom which was sampled by scientists at NIO (NIO report, 2009) during this period. In the color composite image, white color represents sediment dominated waters and green color represents bloom waters.



Figure 11. Algal bloom map of Kalpakkam coastal waters generated from MODIS-Aqua data (19 Feb. 2008) using the new algorithm (for detecting Trichodesmium B bloom). Left bottom image is the field photograph of this bloom which was sampled by Mohanty, *et al.* (2010) during this period. In the color composite image, white color represents sediment dominated waters and green color represents bloom waters.

the Mangalore coast on 17 April 2009, was previously sampled and reported by the NIO researchers [10]. It was observed during their cruises that the diazotrophic algal blooms of Trichodesmium dominated the entire coastal region from Mangalore in the south to Ratnagiri in the north. The blooms were visible on the surface of waters with their characteristic brown color, similar to sawdust sprinkled over water and long winding streaks. During the decay phase of this bloom, Noctiluca populations and slaps were also found in these waters. These findings conclude that the T. erythareum bloom could trigger other species during its decay phase. The color composite image showed the linear spread of the bloom and it can be confirmed by the Chl-a image (Figure 10). The new algorithm also captured the T. erythareum bloom on 19 Feb 2008 in coastal waters of Kalpakkam (south-eastern part of India), where an *in-situ* study reported a prominent discoloration of the surface water caused by dense and vellowish green colored streaks of about 4 to 5 m width and 10 m - 20 m long patches (at point measurement scale). The phytoplankton responsible for discoloration was confirmed to be Trichodesimum (specific species not reported) and T. erythareum bloom had the similar reflectance signature of similar blooms observed on 3 June 2009. The color composite and Chl-a images also confirmed the existence of these blooms.

There have been two different spectral signatures recognized from the MODIS-Aqua data for Noctiluca blooms, which are often reported as N. scintillans and N. milaris. However, some studies have recorded N. milaris as N. scintillans. The difference in the spectral reflectance signatures of these blooms may be due to their cell size, shape and patchiness and perhaps their associated constituents in water. Noctiluca is a heterotrophic dinoflagellate and some have photosynthetic symbiont and also feeds on other plankton which might influence their optical characteristics [45]. These blooms appear green or red color on different occasions. A prominent discoloration of the surface water off the Rushikulya River mouth on 5 April 2005 (Figure 6) is an example of the noticeably dense and red-colored blooms of N. scintillans covering a wide area of approximately 16 sq.km (field measurement scale) [12]. Their qualitative and quantitative analyses of phytoplankton revealed that the density of *N. scintillans* was  $2.38 \times 10^5$  cell L<sup>-1</sup> against the total cell count of  $3.01 \times 10^5$  cell·L<sup>-1</sup>, sharing almost 80% of the total phytoplankton standing crop. This bloom was reported to be associated with 29 other species of phytoplankton, which included nine species of dinoflagellates, 19 species of diatoms and one species of cyanobacteria (Trichodesmium erythraeum) [12]. From their analyses it is evident that the classification algorithm accurately classified the different types of blooms. Though the N. scintillans bloom appeared many times along this coast,

its appearance on 5 April 2005 was unique because of the reported cell density being relatively higher than the usual cell densities and exhibiting visible changes in physical and chemical properties of seawater off the Rushikulya River mouth and along the Orissa coast. The large coverage of the blooms of *N. scintillans* was also evident in Arabian Sea waters (**Figure 5**, **Figure 6**, and **Figure 8**) and these blooms had the spectral signatures similar to those of *N. scintillans* blooms previously confirmed by the field measurements on 5 April 2005.

Intricate patches of the *C. polykrikoides* bloom were distinguished and classified from other algal blooms with its distinct spectral signatures. The negative values at  $R_{rs}(412)$  of *C. polykrikoides* spectra were the result of the atmospheric correction problem in productive waters [46-48] and therefore not used in the present analysis. Satellite imagery showed two intense blooms of *C. polykrikoides* and *N. scintillans* in coastal and offshore waters of Oman on 23 Nov. 2008 (**Figure 8**). While the former one was confirmed by the field measurements, [34], the latter one had the reflectance signatures similar to the *N. scintillans* bloom signatures observed on 5 April 2005. The color composite and Chl-a images indicated the patterns of these blooms off Oman.

Many of the previous studies have focused in detecting a particular bloom type or its special features/characteristics (e.g., [26,28,30,49,50]). On the contrary, the new algorithm simultaneously classified four major algal blooms in oceanic waters around India. The use of the band ratio/differences along with the necessary criteria resulted in good discrimination between these algal bloom types, however the coefficients and thresholds used in our algorithm may require being fine-tuned based on more in-situ data sets and when an accurate atmospheric correction algorithm is developed for complex waters. However, the criteria used to identify turbid water pixels work very well in many coastal regions around India, and therefore no further changes are warranted at this stage.

#### 7. Conclusions

The new algorithm has proved useful in classifying the four types of algal blooms in coastal and offshore waters around India. These algal blooms are characterized by different spectral signatures. The peaks and troughs observed in the reflectance spectra are related to different phytoplankton properties, e.g., chlorophyll-a and other pigment absorption, cell density/bloom intensity and associated optical properties. This algorithm takes advantages of these specific reflectance features and utilizes the combination of reflectance band difference/ratios and derivative signatures for bloom classification. The classified blooms are closely consistent with field measurements data, ABI\_Chla and color composite images; thus its classification accuracy is high for all the classified blooms. These results suggest that the new algorithm has the potential to classify and monitor the investigated algal blooms. The new algorithm will become increaseingly valuable as further improvements are made based on more *in-situ* data sets for different types of algal blooms. The algorithm coefficients may require being fine-tuned when an accurate atmospheric correction algorithm is developed for retrieval of water-leaving radiance in complex waters around India. Our further work will therefore focus on improving these parts and providing a detailed validation with large *in-situ* data sets.

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