

# **Comparison of Different Sampling Techniques** for the Identification of Aromatic **Hydrocarbons from Fire Effluents**

Abdulrhman M. Dhabbah<sup>1\*</sup>, A. Yacine Badjah-Hadj-Ahmed<sup>2</sup>, Anna A. Stec<sup>3</sup>, T. Richard Hull<sup>3</sup>

<sup>1</sup>King Fahd Security College, Riyadh, Kingdom of Saudi Arabia

<sup>2</sup>Chemistry Department, College of Science, King Saud University, Riyadh, Kingdom of Saudi Arabia

<sup>3</sup>Centre for Fire Hazards and Science, School of Physical Sciences and Computing, University of Central Lancashire, Preston, UK Email: \*a\_dhabbah@hotmail.com

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### Abstract

Huge amounts of various polymers are being used in many fields with numerous benefits. However, their great ability to ignition and rapid flame spreading make these materials dangerous for human life and properties due to the release of highly toxic combustion products. The present work aims to investigate several methods of sampling and identification of aromatic hydrocarbons produced by controlled burning of low-density polyethylene (LDPE) using a toxicity tube furnace. Five different sampling methods were used: solid phase micro extraction (SPME), syringe, tedlar bags, sorption tubes, and gas-solution absorbers (midget impingers). The produced hydrocarbons were analysed by gas chromatography coupled to mass spectrometry with and without pyrolysis. Among the tested techniques, the most convenient sampling method was using syringe with a glass vessel which allowed detection of the highest amount of aromatic hydrocarbons at both 800°C and 600°C, and then followed by SPME. On the other hand, the use of gas-solution absorber (midget impinger) showed poorer results. Regarding the use of tedlar bags and sorption tubes, they did not give satisfactory results. Several carcinogenic or possibly carcinogenic compounds were identified in the combustion products, such as benzene, naphthalene, anthracene and pyrene.

## **Keywords**

Polyethylene, Combustion, Fire Effluents, Sampling, Polycyclic Aromatic Hydrocarbons, GC-MS

## **1. Introduction**

For several decades, increasing quantities of synthetic polymeric materials have

been used in various fields, such as construction, transport, electrical and electronic equipment, furniture etc. Unfortunately, the benefits and wide range of application of these materials are undermined by their great liability for ignition and flame spreading, which are accompanied by the release of toxic combustion products [1]. In fire atmospheres, fire effluents (toxic gases, visible smoke, and heat) play a crucial role negatively affecting the environment and life safety of many creatures [2].

Exposure to fire effluents and smoke may cause acute and chronic toxicities. The acute effects can occur immediately whereas the chronic manifestations appear over an extended period of time [1] [3] [4] [5]. This usually happens either after inhalation of particulate matters which are sufficiently small to penetrate and accumulate in the respiratory tract or inspiration of the generated smoke, leading to impaired vision, or breathing difficulties with severe pain in multiple organs including eyes, nose, throat and chest [3] [6].

Acute toxicity may occur after a single exposure to a toxic substance following inhalation of a single dose or more than one within 24 h [1] [7]. On the other hand, chronic exposure depends on the accumulated doses occurring after continuous or repeated exposure to a toxin over an extended period ranging from months to years, causing irreversible toxic effects [8]. The chronic toxicants may exhibit a range of morbidities due to respiratory sensitizers (such as isocyanates) or teratogenic and mutagenic effects, such as those caused by halogenated dibenzo-p-dioxins and dibenzofurans. Other chronic toxicants, which are considered as carcinogens, include aromatic hydrocarbons compounds. They are formed and released in fires, mainly due to incomplete combustion, and can be divided into two categories: volatile organic compounds (VOCs) such as benzene, and semi-volatile organic compounds (SOVCs) including polycyclic aromatic hydrocarbons (PAHs) such as naphthalene, benzo(a)pyrene and 2-nitrophenol [1] [9].

A proper fire effluent sampling system should give a representative and realistic sample of the test atmosphere, without affecting this sample. This depends on a number of factors including nature of the species, the temperature, the length and type of material used for sampling. Therefore, it is necessary to take these factors into account when sampling and analysing a fire effluent to ensure accurate identification and quantification of fire effluent products [10] [11] [12].

For generating aromatic hydrocarbons in fire, certain specific instruments should be used to simulate different fire conditions: non-flaming, well-ventilated flaming and under-ventilated flaming conditions.

As an example, at  $600^{\circ}$ C the rate of burning is almost steady and the process is well ventilated, while at  $800^{\circ}$ C the pyrolysis rate is higher, and it can be regarded as under-ventilated [13].

As an example, it is possible to produce aromatic hydrocarbons under a wide range of fire conditions using a toxicity tube furnace (Model NFX70-100, developed by Concept Equipment, UK) [14]. On the other hand, different methods can be used for trapping and sampling individual aromatic hydrocarbons, such as: gas-solution absorbers (midget impingers), solid phase micro-extraction (SPME), sorption tubes, tedlar bags and syringe [12].

Gas solution absorbers (also known as midget impingers) are one of the most common methods which are designed as bubble tubes used for collecting gaseous chemicals into a liquid solution. In this sampling technique, a known volume of air is bubbled through the impinger containing a specific solvent which will chemically react with or physically dissolve the chemical of interest [15]. Solid-phase microextraction (SPME) which was developed by Pawliszyn in 1989, may be also used to collect gases from fire effluent. It is a simple, fast and solventless extraction technique and proved to be a good alternative to traditional extraction methods. SPME uses a special syringe containing a fused silica fiber coated with a suitable polymer [16] [17]. When immersed in a liquid sample or exposed to volatile components emitted from a liquid or solid matrix, the fiber adsorbs and concentrates the volatile compounds. Solid sorption tube is another method for sampling combustion gases including aromatic hydrocarbons. This technique is based on adsorption of volatile and semi-volatile analytes on a solid sorbent packed in a tube, followed by their desorption step using heat and a flow of inert gas. It proved to be a sensitive sampling technique which can be used instead of traditional concentration methods such as desorption by solvent [18].

After sampling aromatic hydrocarbons by one of the previously cited methods, they are analyzed by gas chromatography coupled with mass spectrometry (GC-MS). In the case of effluent gas collected on sorption tube, it can be analyzed by insertion of it in a pyrolysis instrument connected to a gas chromatograph coupled with a mass spectrometer (Py-GC-MS). GC-MS is a versatile and highly sensitive analytical technique used in many fields. GC-MS combines the advantages of high-resolution capillary columns with the excellent properties of mass spectrometers. It is very useful for elucidation of complex samples with high sensitivity, selectivity and specificity [19].

As mentioned earlier, pyrolysis can be coupled to GC-MS for investigation of thermal degradation of various organic samples. In fire effluent detection, this method can be used to thermally degrade the sample to produce smaller fragments that are separated on a GC column then detected in a mass spectrometer. Py-GC/MS is particularly useful for examination of combustion products resulting from burning polymeric materials that are difficult to dissolve in a solvent [20].

The present study aims to investigate the combustion products resulting from burning low-density polyethylene (LDPE), which is widely used in many fields such as industry and household furniture, by using five different sampling techniques in order to compare them. These methods were: gas-solution absorbers (midget impingers), solid phase micro-extraction (SPME), syringe, tedlar bags and sorption tubes, while the analysis was carried out by gas chromatography coupled to mass spectrometry and focused on aromatic hydrocarbons. The experimental conditions used for each technique were selected according to previous studies [21] [22] [23] [24] [25].

#### 2. Experimental

Producing aromatic hydrocarbons under a wide range of fire conditions was achieved using a toxicity tube furnace (Model NFX70-100, developed by Concept Equipment, UK) (**Figure 1**) [14]. This instrument was developed to produce and estimate pyrolysis or combustion effluents under specific laboratory conditions. Thus, it is a small scale static tube furnace in which the test specimen (typically 1 g, or 0.1 g for low density materials), is pushed in a crucible, into the middle of the furnace tube and then heated to temperatures of 600°C or 800°C, with flowing air at 2 litres per min.

The controlled combustion of low-density polyethylene (LDPE) was carried out by burning 1.00 g in toxicity tube furnace (Model NF X70-100) at 800°C and 600°C. The combustion products were then trapped and sampled using gas-solution absorbers (midget impingers), solid phase micro extraction (SPME), syringe, sorption tubes and tedlar bags [12]. A picture of each sampling device is shown in **Figure 2**.

The detailed experimental parameters used in sampling and analysis of fire effluents are given in **Table 1**. **Table 2** and **Table 3** show the conditions used for collecting the fire effluents produced at 800°C and 600°C. The collection procedures based on SPME and syringe, used in the present work, were described in Bulletin 922 from Supelco [21]. In the case of gas-solution absorption, the sampling was carried out by methods described in EPA 8270D and EPA 8275 [22] [23]. All these sampling methods were followed by gas chromatography hyphenated to mass spectrometry (GC-MS), except experiments based on sorption tubes in which the effluents were analyzed by using pyrolysis-gas chromatography-mass spectrometry (Py-GCMS) according to methods described in EPA TO-15 and EPA TO-17 [24] [25]. Below is a brief summary to describe the different methods used.

#### 2.1. Sampling Using Gas-Solution Absorbers

In this method, collection of fire effluents for absorption of gases in solution was







Figure 2. Pictures showing the different sampling devices used in the present work.

Fable 1. Experimental	parameters for analysis	ng LDPE combustion	products by C	GC-MS and Py-GCMS
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<table-container>SamplesLDEIPge of sorptiontesNaxTexa AGVolume of infectionN/AN/AN/AIPge of SPME File0/monodement (Social Social Sociel Social Social Social Social Social Social Social Social</table-container>	Method	SPME and syringe (Supelco bulletin 922)	EPA 8270	EPA 8275	Sorption tube (EPA TO-15 and TO-17)				
NoneTen A GVolume of injectionN/A1.00 µLN/AType of SPME Fibe100 µn polytimethylsloxanNoneNoneType of syringeNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsNone(GASTIGHT) O µLNoneSolventsSolvents(GASTIGHT) O µLNoneSolventsSolvents(GASTIGHT) O µL(GASTIGHT) O µLSolventsSolvents(GASTIGHT) O µL(GASTIGHT) O µLSolventsSolvents(GASTIGHT) O µL(GASTIGHT) O µLSolventsSolvents(GASTIGHT) O µL(GASTIGHT) O µLSolventsSolvents(GASTIGHT) O µL(GASTIGHT) O µLSolventsSolventsSolvents(GASTIGHT) O µL </th <th>Samples</th> <th></th> <th>LD</th> <th>PE</th> <th></th>	Samples		LD	PE					
Volume of injectionN/A1.0 μLN/AType of SPME Fibre00 μn polydimethylsiloxanNoneNoneType of syringNone(GASTIGHT) 50 μLNoneSolventsNonedeionised water, out acctonitiveNoneSolventsCG-MSP-GC-MSInstrument typeThermo Circle Circ	Type of sorption tube		None		Tenax AG				
Type of SPME File100 µn polydimethylsiloxanNoneNoneType of syringeNone(GASTIGHT > 500 µLNoneSolventsNoneNoneNoneSolventsOnePer Ge-MSPer Ge-MSAnalytical instrumentGe-MSPer Ge-MSNoneColumnThermo TG-SQG30 meter, 0/257 µLPer Autosynther GA with built-in AutosyntherMass range41 - 650 anu35 - 650 µL41 - 650 anuMass range41 - 650 anu0.94 sec/scan0.95 sec/scan0.59 sec/scanObtectorMsEtatosyntherFIDIndiction methodManualSplitProgramables pilit/splitles ingector/SSS1Infiedom20°C20°C20°CSplitles closed 3 min20°C 440°C 4 (2 min.) (2 min.) (2 min.) (2 min.) (2 min.) 	Volume of injection	N/A	1.00	μL	N/A				
Type of syringeNoneNoneSolventsNoneNoneAnalytical instrumentNoneNoneInstrument typeColumnThermo TG-SQG3 meter, U.S.2 mm ID, 0.25 mm ID, 0.2	Type of SPME Fiber	100 µm polydimethylsiloxane	Nor	ne	None				
SolventsNonedeionised water, toluer and constructionNoneAnalytical instrumentGC - MSPropage And constructionPropage And constructionTolument representationThermo-Ticre - Scient and constructionAll - 650 and constructionAll - 650 and constructionMass range41 - 650 and35 - 650 arm41 - 650 and constructionAll - 650 and constructionMass range41 - 650 and0.94 sec/scan0.95 sec/scan0.59 sec/scanSolution0.95 sec/scan0.94 sec/scan0.95 sec/scan0.59 sec/scanDetectorManualSelitFitoInjectorSplites closed 3 minSplitProgrammabilities representationInitial Setpoint20° C20° C20° CSolution and construction35° C2 min/100° di solution40° C4 di construction40° C4 di constructionSolution and construction20° C20° C20° C20° CSolution and construction20° C120° C120° C120° C1Solution and construction20° C120° C120°	Type of syringe	None	(GASTIGH	T) 500 μL	None				
Analytical instrument Instrument typeGC-MSPy-GC-MSInstrument typeThermo-Scientific Trace 1300Pic-Mass can ge AutosamplerColumnThermo-TG-SQC30 meters.JSC JUNCA1 - 650 anu AutosampleMass range41 - 650 anu A5 - 6500.95 sec/scanMass range41 - 650 anu A5 - 6500.95 sec/scanScan time0.59 sec/scan0.95 sec/scanOctor0.94 sec/scan0.95 sec/scanDetectorManulScan timeInjectorManulSplitSplites (closed 3 min.)SplitProgramale splits (signer)Initial Setpoint220°C280°CSpreaceSS <sup>5</sup> C (2 min.)/100°, signer, 40°C (4 or C4 meter)Splits (signer, 40°C (4 or C4 meter)Anstrument typeSS <sup>5</sup> C (2 min.)/100°, signer, 40°C (4 or C4 meter)Splits (signer, 40°C (4 or C4 meter)SpreaceSS <sup>5</sup> C (2 min.)/100°, signer, 30°C (2 min.)Splits (signer, 30°C (2 min.)	Solvents	None	deionised water, tolue	ene and acetonitrile	None				
Instrument type       Thermo Scientific Trace 1300       PE Autosystem GC with building Autosympter         Column       Thermo Scientific Trace 1300       Pe Autosympter       Autosympter         Mass range       41 - 650 anu       35 - 650 amu       41 - 650 anu       41 - 650 anu         Mass range       41 - 650 anu       0.94 sec/scan       0.95 sec/scan       0.59 sec/scan         Scan time       0.59 sec/scan       0.94 sec/scan       0.95 sec/scan       0.59 sec/scan         Detector       Ms       FID       500       FID         Injecton       Manual       Split       Porgramable splits second in	Analytical instrument		GC-MS		Py-GC-MS				
ColumnThermo TG-SQC30 meter, J.25 mm ID, 0.25 µmMass range41 - 650 amu35 - 650 µm41 - 650 amuScan time0.59 sec/scan0.94 sec/scan0.95 sec/scan0.59 sec/scanCarrierManualFIDDetectorManualSplites (closed 3 min.)Splites (closed 3 min.) <th c<="" th=""><th>Instrument type</th><th>Thern</th><th>PE Autosystem GC with built-in Autosampler</th></th>	<th>Instrument type</th> <th>Thern</th> <th>PE Autosystem GC with built-in Autosampler</th>	Instrument type	Thern	PE Autosystem GC with built-in Autosampler					
Mass range41 - 650 amu35 - 650 am.41 - 650 am.Scan time0.59 sec/scan0.94 sec/scan0.95 sec/scan0.59 sec/scanCarrierHe Im/JDetectorMSFIDInjectorManualAutosamplerFogrammable split/splitless injector (PSSI)Initial Setpoint220°C280°C280°COven temperature35°C (2 min.)/10.0°40°C (4 d°C (4 min.)0° cyan.)/10.0° -min <sup>-1</sup> /200°C (2 min.)	Column	Thermo TG-SQC30 meter, 0.25 mm ID, 0.25 μm df							
Scan time0.59 sec/scan0.94 sec/scan0.95 sec/scan0.59 sec/scanCarrierHe ImL/min.DetectorMSFIDInjector methodManualAutosamplerInjectorSplitless (closed 3 min.)SplitProgrammable split/splitless injector (PSSI)Initial Setpoint220°C280°C280°CSome memperature35°C (2 min.)/10.0°40°C (4 min <sup>-1</sup> /200°C (2 min.)/10.0° min <sup>-1</sup> /200°C (2 mi	Mass range	41 - 650 amu	) amu	41 - 650 amu					
Carrier       He Im/min.         Detector       MS       FID         Injection method       Manual       Autosampler         Injector       Splitless (closed 3 min.)       Split       Programmable splitless (injector (PSSI) injector (PSSI)         Initial Setpoint       220°C       280°C       280°C         Oven temperature       35°C (2 min.)/10.0° + min <sup>-1</sup> /20°C (2 min.)       0'C amin.)	Scan time	0.59 sec/scan	0.94 sec/scan 0.95 sec/scan		0.59 sec/scan				
DetectorMSFIDInjection methodManualAutosamplerInjectorSplitless (closed 3 min.)SplitProgrammable split/splitless (injector (PSSI)Initial Setpoint220°C280°C280°COven temperature35°C (2 min.)/10.0°40°C (440°C (40°C (4)Sum temperature35°C (2 min.)/30.0°40°C (440°C (4)0°C (4)0°C (4)Oven temperature35°C (2 min.)/30.0°(2 min.)/10.0° min <sup>-1</sup> /20°C (2 min <sup></sup>	Carrier		He 1m	L/min.					
Injection methodManualAutosamplerInjectorSplitless (closed 3 min.)SplitProgrammable split/splitless (njector (PSSI) injector (PSSI)Initial Setpoint220°C280°COven temperature35°C (2 min.)/10.0°40°C (440°C (4)Nin <sup>-1</sup> /220°C (2 min.)/30.0°40°C (4)40°C (4)00°(10 min.)(2 min.)C (2 min.)00°	Detector		MS		FID				
Injector         Splitless (closed 3 min.)         Split         Programmable split/splitless (njector (PSSI))           Initial Setpoint         220°C         280°C           Sobre temperature         35°C (2 min.)/10.0°.         40°C (4         40°C (4           Min <sup>-1</sup> /220°C (2 min.)/30.0°.         min.)/10.0°.min <sup>-1</sup> /270°C min.)/10.0°.min <sup>-1</sup> /320°C         50°C (2 min.)/10.0°.min <sup>-1</sup> /230°C           Min <sup>-1</sup> /300.0°C (5 min.)         (2 min.)         C (2 min.)	Injection method	Manual		Autosampler					
Initial Setpoint         220° C         280° C           Oven temperature         35° C (2 min.)/10.0°.         40° C (4         40° C (4           min <sup>-1</sup> /20° C (2 min.)/30.0°.         min.)/10.0°.min <sup>-1</sup> /270° C min.)/10.0°.min <sup>-1</sup> /320°         50° C (2 min.)/10.0°.min <sup>-1</sup> /230° C (2 min.)/10.0°.min <sup>-1</sup> /230° C (10 min.)	Injector	Splitless (closed 3 min.)	sed 3 min.) Split		Splitless (closed 3 min.) Split		Programmable split/splitless injector (PSSI)		
Oven temperature $35^{\circ}C(2 \text{ min.})/10.0^{\circ}\cdot$ $40^{\circ}C(4)$ $40^{\circ}C(4)$ $50^{\circ}C(2 \text{ min.})/10.0^{\circ} \cdot \text{min}^{-1}/230^{\circ}C$ $\min^{-1}/220^{\circ}C(2 \text{ min.})/30.0^{\circ}\cdot$ $\min)/10.0^{\circ} \cdot \text{min}^{-1}/270^{\circ}C \text{ min.})/10.0^{\circ} \cdot \text{min}^{-1}/320^{\circ}C$ $50^{\circ}C(2 \text{ min.})/10.0^{\circ} \cdot \text{min}^{-1}/230^{\circ}C$ $\min^{-1}/300.0^{\circ}C(5 \text{ min.})$ $(2 \text{ min.})$ $C(2 \text{ min.})$ $(10 \text{ min.})$	Initial Setpoint		220°C		280°C				
	Oven temperature	35°C (2 min.)/10.0°· min <sup>-1</sup> /220°C (2 min.)/30.0°· min <sup>-1</sup> /300.0°C (5 min.)	40°C (4 min.)/10.0°⋅min <sup>-1</sup> /270°C (2 min.)	40°C (4 min.)/10.0°·min <sup>-1</sup> /320° C (2 min.)	50°C (2 min.)/10.0°⋅min <sup>-1</sup> /230°C (10 min.)				
Total run time         25.00 min.         29.00 min.         34.00 min.         30.00 min.	Total run time	25.00 min.	29.00 min.	34.00 min.	30.00 min.				

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		SPME			Syrin	ge	Gas-			
Methods	Direct	Tedlar bag*	Glass sampling vessel**	Direct	Tedlar bag*	Glass sampling vessel**	Toluene	Acetonitrile	Deionised water	tube***
Ignition time					35	5 sec				
Extinction time					2:00	) min.				
Experiment time					0:00 to	4:00 min.				
Flow rate					1 L	/min.				
Sampled volume of fire effluent		None			200 µ	L		10 mL		None
Sampling time	<ol> <li>Direct out betwee ignition t</li> <li>In case done for</li> <li>In case was for 5</li> </ol>	SPME sampli een 1:20 and 1 time e of tedlar bag 10 sec. after en e of glass vesse :00 min. after	ng was carried :30 min. after sampling was nd of experiment l sampling time heating step	<ol> <li>Direct</li> <li>Direct</li> <li>Direct</li> <li>In cat</li> <li>carried</li> <li>experint</li> <li>In cat</li> <li>perform</li> <li>step</li> </ol>	ct syringe sam in. after ignition ise of tedlar ba out immediat ment ise of glass ves med immediat	npling was done on time ag sampling was ely after end of ssel, sampling was ely after heating	For all s was fro	olvents, samp om 35 sec to 3	ling time :30 min.	Sampling time was from 35 sec to 3:30 min.

#### Table 2. Experimental conditions for sampling of aromatic hydrocarbon compounds at furnace temperature 800°C.

\*Tedlar bags were plugged in all cases after 10 seconds from beginning of experiments; \*\*Glass sampling vessel was heated in oven before sampling for 20 min. at 40°C; \*\*\*Sorption tube experiments were carried out, but unfortunately the results were not reproducible.

		SPME	Syringe						
	direct	glass vessel	direct	glass					
Ignition time		85 se	ec						
Extinction time		2:45 m	iin.						
Experiment time		0:00 to 4:30 min.							
Flow rate	1 L/min.								
Sampled volume of fire effluent		None	200 µL						
Sampling time	<ol> <li>Direct SPME out between 2:20 ignition time</li> <li>In case of glas was for 5:00 min</li> </ol>	sampling was carried ) and 2:30 min. after s vessel sampling time . after heating step	<ol> <li>Direct syrin done 2:20 min.</li> <li>In case of gl was performed heating step</li> </ol>	ge sampling was . after ignition time ass vessel, sampling l immediately after					

**Table 3.** Experimental conditions for sampling of aromatic hydrocarbons compounds at furnace temperature of 600°C.

done using midget impingers (**Figure 2(F)**) [12]. The test atmosphere was drawn or pushed through the absorbing media at a measured rate for a specified period of time. In order to extract a wider range of compounds, three solvents with increasing polarities were used, namely toluene, acetonitrile and deionised water [10]. The three solvents were separately injected as blank samples in the same chromatographic conditions, and no impurities were detected. The experimental conditions corresponding to LDPE combustion are reported in **Table 2**, while the method used for sampling fire effluents products at 800°C are reported in **Table 4**.

 Table 4. Methods used for collecting fire effluents.

Sample	Condition		Methods											
Polymer	Air flow/ temperature	gas ab (El	-solut sorbe PA 82	rion ers 70)	gas ab (El	-solut sorbe PA 82	ion rs 75)	SPME (direct)	SPME (gas sampling bulb 250 mL)	SPME (tedlar bag)	Syringe (direct)	Syringe (gas sampling bulb 250 mL)	Syringe (tedlar bag)	Sorption tube
LDPE (NFX)	2 L·min <sup>-1</sup> / 800°C	Toluene	cessful cetonitrile	cessful test	scessful test	cessful cetonitrile	ccessful test	Successful test	Successful test	No detectable peaks	Successful test	Successful test	No detectable peaks	No detectable peaks
LDPE (NFX)	2 L·min <sup>-1</sup> / 600°C	1	₹ No tes	ons t	Suc	∢ No tes	ons t	Successful test	Successful test	No test	Successful test	Successful test	No test	No test

#### 2.2. Sampling Using Solid Phase Micro Extraction (SPME)

Sampling with SPME (Figure 2(C)) is widely used for many analytical applications. It is an easy and convenient technique applied as an extraction and concentration step prior to GC-MS. The test atmosphere is pumped at a constant rate then sampled with SPME device either directly or after being collected using a tedlar bag (Figure 2(B)) [12] [21]. In addition, we have also developed a procedure for collecting the fire effluents into a glass vessel equipped with two valves before sampling with SPME. In this method, the volatile species were flowed through a 250 mL glass sampling vessel. This glass bulb (shown in Figure 2(A)) was then heated at 40°C for 20 minutes; after that the aromatic hydrocarbons were sampled by SPME for 5 minutes. In case of using the tedlar bag, it is necessary to filter the gas effluent to remove particulates and avoid inlet plugging; for this purpose glass wool filters, have been found effective [12].

## 2.3. Sampling Using Syringe

We have developed a simpler and more efficient sampling method of fire effluents using a gas syringe (Figure 2(E)). It proved to be a good alternative to SPME for collecting combustion products; it is easier and faster, allowing a rapid injection into GC-MS. Furthermore, it can save both cost and time of analysis. The combustion products are taken either directly using a gas syringe or after being collected in a 250 mL glass sampling vessel as mentioned earlier. After collecting these volatile species in a glass bulb, it was heated in an oven at 40°C for 20 minutes; after that the aromatic hydrocarbons were directly sampled using a 200  $\mu$ L gas syringe [12].

#### 2.4. Sampling Using Solid Sorption Tubes

The sampling using solid sorption tubes (Figure 2(D)) was carried out with a pyrolysis unit connected to a GC-MS instrument (Py-GC-MS). The sorption tube was packed with Tenax AG sorbent. The main problem we encountered when using this technique was that inlet plugging can occur due to soot accu-

mulation. In this case, a decrease in sample flow rate was observed [9] [12] [13].

#### 2.5. Sampling Using Tedlar Bags

Tedlar bag (**Figure 2(B**)) is made of a polyvinyl fluoride film (PVF) and equipped with a polytetrafluoroethylene (Teflon, PTFE) valve. It is used to collect various gas samples such as volatile organic compounds (VOCs). Tedlar film (PVF) is a polymer resistant to high temperature; it can be used up to 170°C. Moreover, it is chemically inert, resistant to corrosion with low absorption and permeability of gases [12]. However, in our experiments tedlar bags were plugged in all cases after only 10 seconds from beginning of experiment.

#### 2.6. Analysis of LDPE Combustion Products

Two instruments were used for analysing aromatic hydrocarbons after sampling them by different methods: GC-MS model Thermo Scientific (USA) Trace 1300 and Py-GC-MS model PE Autosystem GC (Perkin Elmer, USA) with built-in Autosampler. On the other hand, all the experimental parameters were set according to previous studies [12] [19] [20].

#### 3. Results and Discussion

All experiments at 800°C were done in duplicate except those using sorption tubes and tedlar bags which did not show detectable combustion products compared with other methods. It appeared that the best results were obtained by using SPME and syringe, either directly or with glass vessel, while the other sampling techniques were unfruitful. Thus, the investigations using SPME and syringe which gave the best results were repeated with a furnace temperature of 600°C, as shown in Table 4.

The LDPE samples were burnt at 800°C and 600°C using a toxicity tube furnace NF X70-100. The results reported in **Table 2** and **Table 3**, show that the burning process was much faster at 800°C than at 600°C; the ignition time which corresponds to the beginning of burning was 35 s. at 800°C instead of 85 s. at 600°C. Similarly the extinction time corresponding to the end of LDPE combustion was 2:00 and 2:45 min. at 800°C and 600°C, respectively. For this reason, the selected sampling conditions were adapted according to the combustion rate at each temperature as indicated in both tables. As an example, the sampling time when using both SPME and syringe was 1:20 and 2:20 min., at 800°C and 600°C, respectively. The collection of aromatic hydrocarbons produced by LDPE burning was performed by different methods of sampling such as SPME, syringe, gas-solution absorbers and sorption tubes. The results obtained in fire effluent analysis are shown in **Figure 3** and **Figure 4**, and in **Table 5** and **Table 6**.

#### 3.1. Aromatic Hydrocarbons Produced at 800°C

Burning LDPE at 800°C can be considered as under-ventilated process; at this



Figure 3. Chromatograms of aroatic hydrocarbons extracted by SPME and syringe at 800°C.

temperature, three solvents were used for sampling with gas-solution absorbers (midget impingers) according to procedures EPA 8270 and 8275 at 800°C. These solvents were: deionized water, toluene and acetonitrile. No hydrocarbon compounds were detected in the deionized water extract, due to the low polarity of hydrocarbons compared to water; whereas acetonitrile and toluene which are less polar, allowed collection of few aromatic hydrocarbons. Nevertheless, all the results obtained with gas-solution absorbers according to whether EPA 8270 or EPA 8275 procedures were not satisfactory. **Figure 3** shows the results obtained by collection with SPME and syringe, either directly or using glass sampling vessel at 800°C.

In case of direct sampling both techniques showed that intensity of volatile compounds was higher than that of the heavy products, whereas using glass sampling vessel gave higher intensity for the heavier hydrocarbons. On the other hand, as mentioned in Table 2 and Table 4, the results obtained with sorption



**Figure 4.** Chromatograms of aromatic hydrocarbons extracted by SPME and syringe at 600°C.

**Table 5.** Summary of aromatic hydrocarbons detected after burning LDPE at 800°C and sampled with SPME and syringe methods.

PAHs	MW	Structure	Retention time (min.)	SPME direct	SPME glass	Syringe direct	Syringe glass
1,4-cyclohexadiene	80		2.07	$\checkmark$	Х	$\checkmark$	$\checkmark$
Benzene	78		2.27	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Toluene	92		3.70	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Styrene	104		5.91		$\checkmark$	$\checkmark$	$\checkmark$
Ethyl benzene	106		7.63	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Indene	116		8.64		$\checkmark$	$\checkmark$	$\checkmark$
Benzene, propyl	120		8.68	$\checkmark$	$\checkmark$	Х	$\checkmark$

Continued						
Indane	118	9.34	Х	$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene, 1,2dihydro	130	10.36	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene	128	10.95	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene, 1-methyl	142	12.59	$\checkmark$		$\checkmark$	$\checkmark$
Biphenyl	154	13.74	Х	$\checkmark$	Х	$\checkmark$
Biphenylene	152	14.72		$\checkmark$	$\checkmark$	$\checkmark$
Anthracene	178	18.64	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Pyrene	202	22.45	$\checkmark$		$\checkmark$	$\checkmark$

**Table 6.** Summary of aromatic hydrocarbons detected after burning LDPE at 600°C and sampled with SPME and syringe methods.

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PAHs	MW	Structure	Retention time (min.)	SPME direct	SPME glass	Syringe direct	Syringe glass
1,4-Cyclohexadiene	80		2.07	$\checkmark$	Х	$\checkmark$	$\checkmark$
Benzene	78		2.27	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Toluene	92		3.70		$\checkmark$	$\checkmark$	$\checkmark$
Styrene	104		5.91		$\checkmark$	$\checkmark$	$\checkmark$
Ethyl benzene	106		7.63	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Indene	116		8.64	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Benzene, propyl	120		8.68	$\checkmark$	$\checkmark$	Х	$\checkmark$
Indane	118		9.34	Х	$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene, 1,2dihydro	130		10.36	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene	128		10.95		$\checkmark$	$\checkmark$	$\checkmark$
Naphthalene, 1-methyl	142		12.59		$\checkmark$	$\checkmark$	$\checkmark$
Biphenylene	152		14.72	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Biphenyl	154		13.74	х	$\checkmark$	х	$\checkmark$
Anthracene	178		18.64	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$
Pyrene	202		22.45	$\checkmark$	$\checkmark$	$\checkmark$	$\checkmark$

tubes were not reproducible, while tedlar bags were plugged in all cases after only 10 seconds from the beginning of experiment. Table 5 shows the aromatic hydrocarbons identified in all experiments carried out at 800°C, using the different sampling methods mentioned in Figure 3. As reported above, the best results for collection of aromatic hydrocarbons resulting from LDPE burning were obtained using SPME and syringe, which allowed detection of 13 to 15 hydrocarbons. In contrast, the intensity of all peaks extracted using midget impinger with both acetonitrile and toluene was very low, in comparison with the results of collection by SPME and syringe. In case of using deionized water for sampling, only styrene was detected. To summarize, the most convenient method was sampling by syringe with a glass vessel which enabled detection of fifteen aromatic hydrocarbons, as reported in Table 5. Among the identified combustion products produced by burning LDPE at 800°C, three are considered as priority aromatic hydrocarbons: naphthalene, anthracene and pyrene. Their carcinogenic toxic equivalency factor (TEF) values are 0.001, 0.01 and 0.001, respectively [11].

#### 3.2. Aromatic Hydrocarbons Produced at 600°C

As mentioned earlier, the results obtained after LDPE burning at 800°C were not satisfactory when using sampling with sorption tube, tedlar bag and gas-solution absorbers. Moreover, these sampling techniques were time consuming and their setting needs more efforts, thus they were not used for experiments carried out at 600°C. Therefore, the only sampling methods conducted at 600°C which correspond to well-ventilated conditions were SPME and syringe, either direct or with glass vessel. **Figure 4** shows the results obtained by SPME and syringe, using both methods.

As it was observed in Figure 3, direct sampling using both techniques showed that intensity of volatile compounds was higher than that of the heavy products, whereas using glass sampling vessel gave more intense peaks for the heavier hydrocarbons. On the other hand, the intensity of all hydrocarbon peaks observed after burning at 600°C was higher than that obtained at 800°C; this is clearly due to the higher combustion rate of LDPE at 600°C, corresponding to well-ventilated conditions, because the burning time was longer and availability of oxygen was higher. Table 6 shows the aromatic hydrocarbons identified in all experiments carried out at 600°C, using the different sampling methods mentioned in Figure 4. As reported above, the best results for collection of aromatic hydrocarbons resulting from LDPE burning were obtained using SPME and syringe, which allowed detection of 13 to 15 aromatic hydrocarbons. To summarize, the most convenient method was sampling by syringe with a glass vessel which showed the presence of fifteen aromatic hydrocarbons. These results are in agreement with those obtained at burning LDPE at 800°C. Combustion of LDPE at 600°C vielded several aromatic hydrocarbons, among which three are listed as priority aromatic hydrocarbons: naphthalene, anthracene and pyrene whose carcinogenic toxic equivalency factor (TEF) values are 0.001, 0.01 and 0.001, respectively [11].

## 4. Conclusions

The aromatic hydrocarbons emitted from burning 1 g of LDPE have been investigated by using five different sampling methods under various fire conditions. The main gas phase components were sampled and analysed in small-scale tests using a toxicity tube furnace. The used sampling methods were: solid phase micro-extraction (SPME), syringe, sorption tubes, gas-solution absorbers (midget impingers) and tedlar bags, while the combustion products were analysed by GC-MS and Py-GC-MS. Collection of the fire effluents by syringe and SPME was carried out either by direct sampling or using a glass vessel. In this case, the identification of aromatic hydrocarbons produced showed that fifteen hydrocarbons were detected in significant concentrations, some of them having a potentially negative impact on the environment, or human health. The sampling method using syringe with a glass vessel showed the best results, as it gave the highest detection of aromatic hydrocarbons at 800 and 600°C; then followed by solid-phase microextraction. However, the intensity of the volatile detected hydrocarbons was higher at 600°C than for 800°C which corresponds to under-ventilated conditions. Gas-solution absorber (midget impinger) was also used with toluene, acetonitrile and deionised water as solvents with increasing polarity; it showed poorer results compared with syringe and SPME methods.

In addition, some experiments were performed using tedlar bags and sorption tubes, but they did not give satisfactory results, either because of plugged orifice or non-reproducible results. After burning LDPE and analysing its combustion products, benzene, naphthalene, anthracene and pyrene were among the identified hydrocarbons. They are considered as carcinogenic or possibly carcinogenic compounds.

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## **Conflicts of Interest**

The authors declare no conflicts of interest regarding the publication of this paper.

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