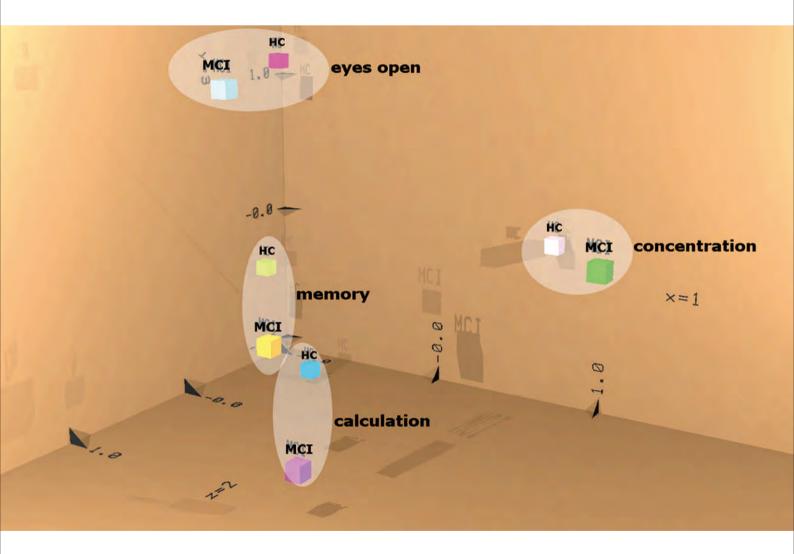


Advances in Alzheimer's Disease





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The Drug Development Based on Pathogenetic Research in Alzheimer's Disease

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Abstract

Neuropathologically, Alzheimer's disease is characterized by the presence of extracellular deposits of amyloid- β peptides, intracellular neurofibrillary tangles and atrophy of the basal forebrain cholinergic neurons. The research of pathogenesis of Alzheimer's disease inspirits potential clinical drugs for treatment. To block the progression of the disease, drugs under development have to interfere with the pathogenic steps responsible for the clinical symptoms, including cholinergic deficit, calcium dysregulation, inflammation and oxidative damage, and the deposition of amyloid- β plaques and of neurofibrillary tangles. In this review, the pertinent literature about drugs targeted on relieving symptoms above is reviewed. We aim to discuss possible research priorities in the future.

Keywords

Alzheimer's Disease, A β , Protein Tau, ULMWH, MT-Stabilizing Agents

1. Introduction

Alzheimer's disease (AD) is described as one of the most common neurodegenerative disorders, with a prevalence of 5 percent after sixty-five years of age, increasing to almost 30 percent in people over age eighty-five [1]. The clinical symptoms of AD include loss of memory, progressive cognitive impairment, various behavioral disturbances and neurological disorders. Typically, AD start with mild memory deficits, then gradually progress to severe dementia and stupor. Generally speaking, about nine years after clinical diagnosis, the AD patients may face to death caused by respiratory complications [2]. Neuropathologically, AD is characterized by senile plaques (SP) composed by amyloid- β peptides (A β), neurofibrillary tangles (NFTs) generated by hyper-phosphorylated forms of protein tau [3] and degeneration or atrophy of the basal forebrain cholinergic neurons.

APP is overexpressed in AD [4]. After the cleavage of APP by two proteases, denoted as β -secretase (BACE1) and γ -secretase, the s equential action will a ggregates i nto to xic A β . The two major subtypes ar e A $\beta_{1.40}$ and A $\beta_{1.42}$. The shorter one comes from the cutting of typical APP in endoplasmic reticulum, while the longer one is formed across the Golgi network. Compared with each other, A $\beta_{1.40}$ is more common, but A $\beta_{1.42}$ is even more relevant to the disease. Tau is a component of microtubules; it stabilizes growing axons and is necessary for neuritis [4]. In AD, tau is abnormally hyperphosphorylated and forms insoluble fibrils, which contribute to the earliest cytoskeletal changes in NFTs formation. This abnormal spiral structure interfere the normal function of neurons, ultimately leading to neuronal cell death. Cholinesterase is the key enzyme in biological nerve conduction. In the cholinergic synaptic cleft, it degrades acetylcholine, induces the termination of excitability role of the neurotransmitter on postsynaptic membrane, which ensures normal convery of nerve signals *in vivo*. Acetylcholinesterase catalyzes the cleavage reaction of acetylcholine, resulting in the lack of acetylcholine, thus interfere nerve signal transmission.

Alzheimer's disease pathology changes major involve cholinergic nerve pathways from frontal base to the cerebral cortex and hippocampus. As we all known, these pathways are associated with attention, learning ability, memory and other cognitive processes. The loss of basal forebrain cholinergic cells in AD patients leads to reduction of synaptic availability of acetylcholine. So, AD patients always suffer the cognitive impairment.

Scientists have be en working on the pathophysiological processes of Alzheimer's disease for more than a century. A large number of theories have been discovered to explain what is happening in the brain of AD patients. For example, $A\beta$ causes calcium dysregulation and oxidative stress in central nervous system cells, inflammatory changes can be observed in the brain, even diabetes and insulin-resistance may have connection with AD through GSK3 β . Nevertheless, there is a long way to go before we discover the exactly and comprehensive functions of $A\beta$ and tau in the process of AD. In other hand, the significantly effective drugs for AD clinical treatment are less than enough. In this article, we focus on the molecular and cellular alterations involved in neuronal dysfunctions caused by $A\beta$ and tau in AD. More importantly, the drug development based on pathogenetic research in AD, and possible research priorities in the future will be mentioned.

2. Cholinergic Drugs

Neurochemical research shows that the brain in AD patients have obvious shortage of central cholinergic neurotransmitter, causing loss of memory, directional force, behavior and personality change as a r esult. Enhancing cholinergic effects is an important way for the treatment of AD. Current research focuses on acetylcholinesterase inhibitors (AChEI), which can increase ACh concentration in the synaptic cleft by reducing its degradation, thus improve the central activity of choline. It is by far the most commonly used and is considered to be one of the most promising drug treatments of AD. The second generation of AChEI is widely applied in clinical treatment, that is Donepezil hydrochloride (aricept) [5], Rivastigmine Tartrate (exelon) [6], Galantamine and Huperzine A.

Donepezil hydrochloride is a highly selective AChEI. It is approved for use in mild-to-moderate AD patients [7] [8]. It has shown some benefit in slowing hippocampal atrophy and protecting nerve cells. Rivastigmine Tartrate has easy BBB permeability and is approved for mild-to-moderate AD [9]. Rivastigmine Tartrate can selectively enhance the effect of acetylcholine in cerebral cortex and hippocampus. Furthermore, cholinesterase inhibitors can slow down the the formation of the amyloid precursor protein (APP) fragment. After combining with its target enzyme into covalent compounds, Rivastigmine Tartrate causes a temporary loss of activity of the enzyme. Galantamine has allosteric nicotinic receptor modulation properties. Galantamine may increase ACh release by regulating brain external nicotinic acid receptor. Research showed that galantamine may be safe for the treatment of elderly patients with severe AD, which improves cognitive function in patients. However, everyday life parameters change is not obvious. Another ChEI, Huperzine A, is considered to be one of the treatment for memory disorders [10]. However, for its lacking of proprietary patent for the treatment of AD, it is considered as a nutraceutical supplement in the US. Clinical and preclinical toxicities are to be established in the future.

AD is a multifactorial disease, so the innovative model is to achieve the goal of "one molecule, multiple targets". Ideally, hybrids can provide parent compounds more potency such as BBB permeability, additional receptors or epitopes [11]. Human studies are planned.

3. Calcium Regulation as a Therapeutic Approach for AD

There is considerable evidence that $A\beta$ induces calcium dysregulation in neuron. Sequential cleavages of APP by secretases generate sAPP α , A β and AICD. Intracellular APP domain (AICD) can modify nuclear gene transcription and further more perturb Ca²⁺ homeostasis. On the other hand, sAPP α , generated from APP by α -secretase, is normally produced to active K^+ channels, thereby hyperpolarizing the membrane and reducing Ca^{2+} influx [12]. The production of sAPP α plays a protecting role in neurons; however, amyloidgenic processing may prevent this program. A β oligomers enhance calcium ion influx by Ca²⁺-permeable channels, which can be facilitated by binding to phosphatidylserine (PtdS). Cell-surface exposure of PtdS is usually indicative of apoptotic cells. Mitochondrial, the energy supply station of cells, once effected by $A\beta$, will induce superoxide anion radical (O2 \bullet) production, Ca²⁺ overload, and decreased ATP production. In this condition, PtdS will flip from the inner portion to the cell surface of the plasma membrane. In turn, neurons with lower ATP level are particularly susceptible to A β toxicity [13]. A β can also generate hydroxyl radical (OH) in the presence of Fe²⁺ and Cu⁺ [14] [15]. As a result, the function of ATPases dependent ion channels (Na⁺ and Ca²⁺ pumps) can be impaired by toxic aldehydes generated by membrane lipid peroxidation (LP). Therefore, the membrane becomes depolarized and toxic amounts of Ca²⁺ flux into the cytoplasm through the open channels, glutamate receptor channels (N-methyl-D-aspartate receptor, NMDAR) and voltage-dependent Ca^{2+} channels (VDCC) open. Inside the neurons, Presenilins (PS) functions as an endoplasmic reticulum (ER) Ca^{2+} leak channel. In familial Alzheimer's disease, PS mutations cause excessive accumulation of Ca^{2+} and then enhance Ca^{2+} release by inositol 1, 4, 5-trisphosphate receptors (IP₃R) channels and ryanodine receptor (RyR). A β can also block the response of nicotinic acetylcholine receptors (nAChRs) and induces sustained Ca^{2+} levels increases in presynaptic through IP₃ [16] (Figure 1).

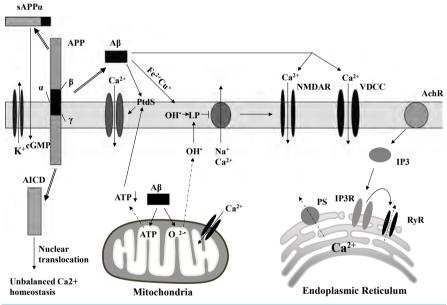


Figure 1. Molecular and cellular alterations involved in neuronal dysfunction in AD. The amyloid- β precursor protein (APP) can be cleaved by β -secretase and γ -secretase, resulting in the l iberation of s APP α , the a myloid- β peptitde (A β) and i ntracellular A PP d omain (AICD). AICD can translocate to the nucleus and perturb Ca²⁺ homeostasis. sAPP α may active K⁺ channels. A β oligomers enhance calcium ion influx into the cell by the formation of Ca²⁺-permeable channels. A β will also induce superoxide anion radical (O₂•⁻) production, Ca²⁺ overload, and decreased ATP production in mitochondrial. A β can also interact with Fe²⁺ and Cu⁺ to generate hydrogen peroxide and hydroxyl radical (OH). As a result, the function of Na⁺ and Ca²⁺ pumps can be impaired by toxic aldehydes generated by membrane lipid peroxidation (LP), resulting the Ca²⁺ flux through N-methyl-D-aspartate receptor (NMDAR) and voltage-dependent Ca²⁺ channels (VDCC). In familial Alzheimer's disease, Presenilins (PS) mutations cause excessive accumulation of Ca²⁺ in the endoplasmic reticulum (ER) and then enhance Ca²⁺ release through inositol 1,4,5-trisphosphate receptors (IP₃R) channels and ryanodine receptor (RyR). A β can also induce sustained nAChR-mediated increases in presynaptic Ca²⁺ levels through IP₃.

Inspired by the above theory, methods to stabilize neural intracellular calcium homeostasis may be one of the treatments for AD. So far, a variety of drugs have therapeutic potential *in vivo* or *in vitro* experiments.

Since ex cessive act ivation of g lutamate s ystem ca uses ex citotoxic n euronal d eath, NMDA o pen ch annel blocker, can antagonize glutamate excitotoxicity. Memantine, which belongs to NMDA receptor antagonist, is different from the cholinesterase inhibitor. It is the first and only drug that is approved for treatment of moderate to severe AD. Clinical studies have demonstrated that it b enefits in c ognitive and behavioral outcomes in patients, either as monotherapy or in combination with donepezil [17]. It also shows good effect to main types of dementia: Alzheimer's dementia, vascular dementia and AIDS dementia, which indicate that it has an advangtage o ver o ther kinds of AD drugs on the market. A nimal experiments s howed that N imodipine, an L -type VGCC in hibitor, plays a r ole as cal cium an tagonist. It d ecreases the intracellular cal cium i on concentration, promotes the regeneration of injured neurons, enhance the plasticity of the aging central nervous system. In the clinical treatment, Nimodipine has obvious curative effect on memory impairment due to AD [18]. Recently, nimodipine was reported to selectively stimulate secretion of $A\beta_{1.42}$ slightly [19]. But the exact mechanism has not yet been elucidated.

In vitro experiments, ultra-low-molecular-weight heparin (ULMWH) partly reduced the $[Ca^{2+}]i$ increase induced by glutamate, this suggests that ULMWH may inhibit external Ca^{2+} influx mediated by NMDA receptor. In addition, the IP₃R induced $[Ca^{2+}]i$ was significantly suppressed ULMWH, suggests that ULMWH can regulate calcium balance by inhibiting calcium ion release. So ULMWH may be speculated as a specific IP₃R antagonist just like heparin. In this way, less IP₃ binds to IP₃Rs and the increase of $[Ca^{2+}]i$ was blocked. Therefore, factors such as ULMWH are expected to have good effect on AD.

4. Anti-Amyloid Therapies

As $A\beta$ is the heart of the amyloid hypothesis of AD, the formation of $A\beta$ oligomers is a directly AD killer. The mainly function of β -secretase and γ -secretase is to produce toxic $A\beta_{1-42}$. Currently the focus of scientists is the inhibition of activity of β -secretase and γ -secretase. β -secretase (BACE1) initiates the amyloidogenic pathway. Activation of nuclear peroxisome proliferator-activated receptor γ (PPAR γ) can suppress expression of β -secretase inhibitors by stimulating PPAR γ [20]. The therapeutic effects of PPAR γ agonists in AD may be caused by their effect of increasing insulin sensitivity and reducing concentrations of insulin.

Present study is generally believed that insulin can not only pass through the blood brain barrier, but also be synthesized within the brain tissue. The insulin receptor (IR) and receptor signal transduction molecules exist in brain tissue [21]. Insulin can not only regulate sugar metabolism and energy metabolism, but also have various biological functions. The function of brain insulin is c omplex, including s upporting the surviving of mature neurons. Cascade control apoptosis is one of the important roles of insulin. The common pathological mechanism of diabetes encephalopathy and AD is an obstacle of insulin signal transduction pathways, namely the insulin resistance, which can cause metabolic disorders and cognitive dysfunction. In a word, insulin signaling induces the phosphorylation and inhibition of glycogen synthase kinase 3 (GSK3). It would therefore promote tau phosphorylation, leading to aggregation and tangle formation, as well as contributing to $A\beta$ peptide production and plaque formation [22] (Figure 2). The related content remains further research.

Using $A\beta$ antibody in treatment of AD is a current research hot spot. Active and passive immunization clinical research is ongoing in several pharmaceutical companies. According to animal tests, vaccination of synthetic $A\beta_{1.42}$ immunization can produce $A\beta_{1.42}$ antibody, thus causes activation of monocyte or microglia. As a result, immune therapy has become a new method for treatment of AD, which has entered clinical trials in the United States. Preclinical and early clinical trial results show $A\beta$ immunotherapy has great potential to overcome the AD. Active immunizations include injection of synthetic $A\beta$ peptide, $A\beta$ fragment joined with carrier protein or adjuvant. It will stimulate the host to produce antibodies against $A\beta$. Passive immunization is to directly inject $A\beta$ specific antibody into the host, thus activate the host immune system. The common basic principle of active and passive immunotherapy is removing $A\beta$ from the brain. In clinical trials of AN-1792 vaccine, the number of $A\beta$ has decreased in the brains of patients. However, some patients appeared subacute aseptic meningoencephalitis within II period clinical trials [23]. Better than AN-1792 vaccine, CAD-106 vaccine, did not show such side effect in the early human trials [24]. A new strategy to increase the security of active immunization is to optimize the drug delivery way. It has been proved that, intranasal administration of $A\beta$ peptide, without adjuvant,

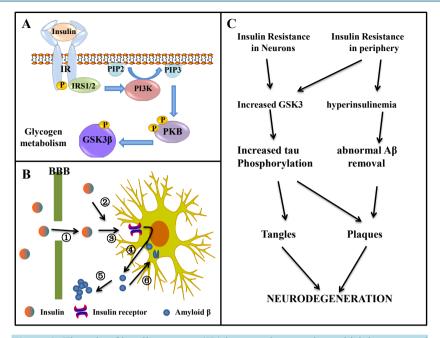


Figure 2. The role of insulin receptor (IR) in neurodegeneration, which is expressed in both neurons and glia. (A) Schematic representation of insulin signaling; (B) The potential r ole of insulin in the pathogenesis of A D. Insulin is transported a ctively across the blood-brain barrier (1), it can also be produced locally in the brain (2). Insulin act through cerebral insulin receptors (3). Thus stimulates the secretion of A β into the extracellular space (4) where it can aggregate into senile plaques (5). Alternatively, excessive A β can be cleared through endocytosis (6); (C) The connection between insulin resistance and neurodegeneration.

leads to appropriate immunoreaction. Gene immunotherapy, a kind of active immunization in essence, has much potential. Initially, $A\beta$ -DNA vaccine was coded by adenovirus or adeno-associated virus vector. Researchers are focusing on the development of non-viral plasmid vector, because of the large-scale low-cost production and no risk of infection or transfection [25].

As for passive immunization, many monoclonal antibody is in development and testing, such as Bapineuzumab (AAB-001) [26] and Solanezumab (LY2062430) [27] [28]. But the production of monoclonal antibodies cost great and need repeated injection. What's worse, it is hard to choose the appropriate target and pass through the BBB. It also has a risk of bleeding.

5. Drugs to Target Tau Protein

Tau protein is a kind of microtubule associated protein whose primary function is to stabilize the conformation of microtubules (MTs). The phosphorylation level of tau protein in AD patients' brain is three times higher than normal. Excessive phosphorylated tau protein may dissociate from microtubules and aggregates into nerve toxic NFTs [29]. As a consequence, microtubule is disintegrated and cytoskeleton is damaged, too.

There are two main therapeutic approaches aiming at tau protein: inhibitors of phosphorylase kinase of tau protein and c ompounds that inhibit tau a ggregation or promote its disassembly. Inhibition of the excessive phosphorylation of tau protein is the major research direction for the treatment of AD. GSK3, and specifically its beta isoform (GSK3 β), have been shown to closely connected with phosphorylate tau [30]. Animal studies have confirmed that in the early pathological changes of tao protein, lithium salt, one kind of non-specific GSK3 inhibitor [31], can prevent excessive phosphorylation of tau protein and block the further progress of the disease. But lithium salt has little effect during the late stage, so researchers have turned to develop specific inhibitors of phosphorylase kinase of tau protein. According to the research results, some kind of small molecular compounds can prevent the interaction of tau protein, thus inhibit its aggregation. For example, methylene blue, a widely used histology dye, can not only prevent polymerization of tau protein into oligomers, but also reduce $A\beta$ level

in the brain [32]. Methylene blue also has antioxidant properties and was effective in improving learning deficits either used alone or in combination with rivastigmine in animals [33]. So it is considered to be a promising new AD treatment. It has a high bioavailability when tested in a phase 2 study in patients with moderate AD [34] and its safety and clinical efficacy need to be further confirmed.

In addition to the two kinds of medicine mentioned above, new drugs designed on counteracting the functional loss of tau protein are promising. As we all know, the primary function of tau protein is to stabilize the conformation of MTs. Over the past decades, several classes of MT-stabilizing products have been used as antineoplastic drug. However, people are worried about dose-limiting toxicities if this class of therapeutics is used in long-term treatment of tauopathy patients. Paclitaxel, which belongs to taxanes natural products, binds to the lumen of the MT at β -tubulin subunit [35]. Importantly, paclitaxel is found to having a function of promoting MT stabilization instead of tau protein. [36]. The actin mode of epothilones is similar to paclitaxel. In vitro, both paclitaxel [37]-[41] and epothilones [42]-[44] have been found to protect neurons against neurotoxicity mediated by tau protein or A β . However, paclitaxel could not cross the BBB while epothilone D shows to be a brain-penetrant MT-stabilizing agent [45]. Considering that tau pathology is primarily in the brain, only epothilone D can be suitable as a therapeutic candidate for human tauopathies. As to the consideration of dose-limiting toxicities, one important observation made in vivo studies in epothilone D [46] indicated that low doses of epothilone D may produce optimal therapeutic effects. Overstabilization of MTs by high dose of agents on the other hand become counterproductive and may be accompanied by side effects. As a growing number of MT-stabilizing products are being discovered, a particular attention should be paid to these agents to find more useful treatment for AD and other tauopathies.

6. Summary and Prospective

The study of the pathophysiological processes of AD has been attracting people's attention for more than a century. $A\beta$ and NFTs have been observed to have extensively damage to normal function of central nervous system, such as cholinergic deficit in the CNS, intracellular Ca²⁺ disequilibrium, inordinate oxidative stress and inflammatory processes. These pathophysiological processes of AD provide multifarious targets for therapeutic or preventative agents. Besides those five drugs which are currently ratified for use in the treatment of AD (done-pezil (Aricept[®]), galantamine (Reminyl[®]), rivastigmine (Exelon[®]), tacrine (Cognex[®]) and memantine (Namenda[®])), many other kinds of drugs aiming at processes mentioned above have potentials in AD treatment, for example, GAG mimetic, NSAIDs, and hypoglycemic agents (Table 1).

According to the past failures we have met in AD drug clinical trials that were conducted over the last decades, we can come to the conclusion that it is questionable to conducting large clinical development programs in AD blindly. AD is the result of the interaction of pathogenic factors. It is necessary to be better appreciating the complicacy of the disease. In our way to find significant treatment for AD, it is important to better understand the relationship between tau, $A\beta$ and other factors. In a word, the competing risk factors, physiological factors such as age and genetics, and environmental factors all play non-ignorable roles in the progress of AD. As a result, drugs with wide range of targets always appear little effect in the treatment.

The failures in AD drug clinical trials also forced people to turn to another approach to the treatment of AD. There are three kinds of behaviors that are considered to reduce the risk of AD [56]. Those are cognitive stimu-

Table 1. Different classes of products and then stages of development as potential candidates for AD.							
Main mechanisms of action	Candidate drugs	Stage of development in trails					
Acetylcholinesterase inhibitors	Huperzine A	RCTs completed [10]					
Specific IP ₃ R antagonist	ULMWH	Vitro experiments [47]					
β -secretase inhibitors	Rosiglitazone	RCTs completed [48]					
,	Pioglitazone	Phase 2 RCT in MCI [48]					
Active immunotherapy that increase $A\beta$ clearance	AN1792 and CAD-106	RCTs completed [49] [50]					
Passive immunotherapy that increase $A\beta$ clearance	AAB-001 and LY2062430	RCTs completed [51] [52]					
Non-specific GSK3 inhibitor	Lithium salt	RCTs completed [53]					
Anti-tau that decrease tau fibrillization	Methylene blue	RCTs completed [54]					
MT-stabilizing products	Paclitaxel	Vitro animal model [55]					
	Epothilone D	Phase 1b clinical trail [46]					

Table 1. Different classes of products and their stages of development as potential condidates for AD

RCT: Randomized Controlled Trial. MCI: Mild Cognitive Impairment.

lation, mental and physical exercise, and dietary energy restriction. They are thought to show beneficial effect by activating ne urotransmitters, growth factors, and hormone receptors. In this way, the synaptic a ctivity in nerve cell networks is increased and neurons will be protected against oxidative and metabolic stress.

What's more, most compounds showed some benefits in mild AD, either than moderate AD. These discoveries inspire us that the early we carry out therapeutic trials, the more possible it will be to block the course of the disease. As a consequence, the identification of more accurate tools for early diagnosis is needed. If we have new markers that can detected in the blood or other body fluids at the mild phase of AD, it will be a useful aid for the diagnosis and management of patients with AD.

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References

- Walsh, D.M. and Selkoe, D.J. (2004) Deciphering the Molecular Basis of Memory Failure in Alzheimer's Disease. *Neuron*, 44, 181-193. <u>http://dx.doi.org/10.1016/j.neuron.2004.09.010</u>
- Heneka, M.T. and O'Banion. M.K. (2007) Inflammatory Processes in Alzheimer's Disease. Journal of Neuroimmunology, 184, 69-91. <u>http://dx.doi.org/10.1016/j.jneuroim.2006.11.017</u>
- [3] Lee, V.M., Goedert, M. and Trojanowski, J.Q. (2001) Neurodegenerative Tauopathies. *Annual Review of Neuroscience*, 24, 1121-1159. <u>http://dx.doi.org/10.1146/annurev.neuro.24.1.1121</u>
- [4] Sue, W. and Griffin, T. (2006) Inflammation and Neurodegenerative Diseases. *The American Journal of Clinical Nutrition*, 83, 4708-4748.
- [5] Jackson, S., Ham, R.J. and Wilkinson, D. (2004) The Safety and Tolerability of Donepezil in Patients with Alzheimer's Disease. *British Journal of Clinical Pharmacology*, 58, 1-8. <u>http://dx.doi.org/10.1111/j.1365-2125.2004.01848.x</u>
- [6] Kumar, V., Anand, R., Messina, J., Hartman, R. and Veach, J. (2000) An Efficacy and Safety Analysis of Exelon® in Alzheimer's Disease Patients with Concurrent Vascular Risk Factors. *European Journal of Neurology*, 7, 159-169. http://dx.doi.org/10.1046/j.1468-1331.2000.00046.x
- [7] Petersen, R.C., et al. (2005) Vitamin E and Donepezil for the Treatment of Mild Cognitive Impairment. New England Journal of Medicine, 352, 2379-2388. <u>http://dx.doi.org/10.1056/NEJMoa050151</u>
- [8] Salloway, S., et al. (2004) E fficacy of Donepezil in Mild Cognitive Impairment A Randomized Placebo-Controlled Trial. Neurology, 63, 651-657. <u>http://dx.doi.org/10.1212/01.WNL.0000134664.80320.92</u>
- [9] Corey-Bloom, J., Anand, R. and Veach, J.F. (1998) A Randomized Trial Evaluating the Efficacy and Safety of ENA 713(Rivastigmine Tartrate), a New Acetylcholinesterase Inhibitor, in Patients with Mild to Moderately Severe Alzheimer's Disease. *International Journal of Geriatric Psychopharmacology*, 1, 55-65.
- Bai, D. (2007) Development of Huperzine A and B for Treatment of Alzheimer's Disease. *Pure and Applied Chemistry*, 79, 469-479. <u>http://dx.doi.org/10.1351/pac200779040469</u>
- [11] Camps, P., et al. (2008) Novel Donepezil-Based Inhibitors of Acetyl-And Butyrylcholinesterase and Acetylcholinesterase-Induced β-Amyloid Aggregation. Journal of Medicinal Chemistry, 51, 3588-3598. http://dx.doi.org/10.1021/jm8001313
- [12] Sahu, S.K., Gummadi, S.N., Manoj, N. and Aradhyam, G.K. (2007) Phospholipid Scramblases: An Overview. Archives of Biochemistry and Biophysics, 462, 103-114. <u>http://dx.doi.org/10.1016/j.abb.2007.04.002</u>
- [13] Simakova, O. and Arispe, N.J. (2007) The Cell-Selective Neurotoxicity of the Alzheimer's Aβ Peptide Is Determined by Surface Phosphatidylserine and Cytosolic ATP Levels. Membrane Binding Is Required for Aβ Toxicity. *The Journal of Neuroscience*, 27, 13719-13729. http://dx.doi.org/10.1523/JNEUROSCI.3006-07.2007
- [14] Hensley, K., et al. (1994) A Model for Beta-Amyloid Aggregation and Neurotoxicity Based on Free Radical Generation by the Peptide: Relevance to Alzheimer Disease. Proceedings of the National Academy of Sciences, 91, 3270-3274. <u>http://dx.doi.org/10.1073/pnas.91.8.3270</u>
- [15] Huang, X., et al. (1999) The Aβ Peptide of Alzheimer's Disease Directly Produces Hydrogen Peroxide through Metal ion Reduction. Biochemistry, 38, 7609-7616. <u>http://dx.doi.org/10.1021/bi990438f</u>
- [16] Dougherty, J.J., Wu, J. and Nichols, R.A. (2003) β-Amyloid Regulation of Presynaptic Nicotinic Receptors in Rat Hippocampus and Neocortex. *The Journal of Neuroscience*, 23, 6740-6747.
- [17] Emre, M., Mecocci, P. and Stender, K. (2008) Pooled Analyses on Cognitive Effects of Memantine in Patients with Moderate to Severe Alzheimer's Disease. *Journal of Alzheimer's Disease*, 14, 193-199.

- [18] Fritze, J. and Walden, J. (1994) Clinical Findings with Nimodipine in Dementia: Test of the Calcium Hypothesis. Journal of Neural Transmission. Supplementum, 46, 439-453.
- [19] Facchinetti, F., Fasolato, C., Giudice, E.D., Burgo, A., Furegato, S., *et al.* (2006) Nimodipine Selectively Stimulates β-Amyloid 1-42 Secretion by a Mechanism Independent of Calcium Influx Blockage. *Neurobiology of Aging*, 27, 218-227. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2005.02.006</u>
- [20] Landreth, G., Jiang, Q., Mandrekar, S. and Heneka, M. (2008) PPARy Agonists as Therapeutics for the Treatment of Alzheimer's Disease. *Neurotherapeutics*, 5, 481-489. <u>http://dx.doi.org/10.1016/j.nurt.2008.05.003</u>
- [21] Abbott, M.A., Wells, D.G. and Fallon, J.R. (1999) The Insulin Receptor Tyrosine Kinase Substrate p58/53 and the Insulin Receptor Are Components of CNS Synapses. *The Journal of Neuroscience*, 19, 7300-7308.
- [22] Cole, A.R., Astell, A., Green, C. and Sutherland, C. (2007) Molecular Connexions between Dementia and Diabetes. *Neuroscience & Biobehavioral Reviews*, **31**, 1046-1063. <u>http://dx.doi.org/10.1016/j.neubiorev.2007.04.004</u>
- [23] Orgogozo, J.M., Gilman, S., Dartigues, J.F., Laurent, B., Puel, M., et al. (2003) Subacute Meningoencephalitis in a Subset of Patients with AD after Abeta42 Immunization. *Neurology*, 61, 46-54. <u>http://dx.doi.org/10.1212/01.WNL.0000073623.84147.A8</u>
- [24] Winblad, B.G., Minthon, L., Floesser, A., Imbert, G., Dumortier, T., et al. (2009) Results of the First-in-Man Study with the Active Aβ Immunotherapy CAD106 in Alzheimer Patients. Alzheimer's & Dementia, 5, P113-P114. http://dx.doi.org/10.1016/j.jalz.2009.05.356
- [25] Rinne, J.O., Brooks, D.J., Rossor, M.N., Fox, N.C., Bullock, R., et al. (2010) ¹¹C-PiB PET Assessment of Change in Fibrillar Amyloid-*β* Load in Patients with Alzheimer's Disease Treated with Bapineuzumab: A Phase 2, Double-Blind, Placebo-Controlled, Ascending-Dose Study. *The Lancet Neurology*, **9**, 363-372. http://dx.doi.org/10.1016/S1474-4422(10)70043-0
- [26] Seubert, P., Barbour, R., Khan, K., Motter, R., Tang, P., et al. (2008) Antibody Capture of Soluble Aβ Does Not Reduce Cortical Aβ Amyloidosis in the PDAPP Mouse. Neurodegenerative Diseases, 5, 65-71. http://dx.doi.org/10.1159/000112834
- [27] Siemers, E.R., Friedrich, S., Dean, R.A., Gonzales, C.R., Farlow, M.R., Paul, S.M. and DeMattos, R.B. (2010) Safety and Changes in Plasma and Cerebrospinal Fluid Amyloid [beta] after a Single Administration of an Amyloid [beta] Monoclonal Antibody in Subjects with Alzheimer Disease. *Clinical Neuropharmacology*, 33, 67-73. http://dx.doi.org/10.1097/WNF.0b013e3181cb577a
- [28] DaSilva, K.A., Brown, M.E., and McLaurin, J. (2009) Reduced Oligomeric and Vascular Amyloid-β Following Immunization of TgCRND8 Mice with an Alzheimer's DNA Vaccine. *Vaccine*, 27, 1365-1376. http://dx.doi.org/10.1016/j.vaccine.2008.12.044
- [29] Himmler, A., Drechsel, D., Kirschner, M.W. and Martin, D.W. (1989) Tau Consists of a Set of Proteins with Repeated C-Terminal M icrotubule-Binding Domains and V ariable N-Terminal Domains. *Molecular and Cellular Biology*, 9, 1381-1388.
- [30] Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P. and Kivipelto, M. (2010) Alzheimer's Disease: Clinical Trials and Drug Development. *The Lancet Neurology*, 9, 702-716. <u>http://dx.doi.org/10.1016/S1474-4422(10)70119-8</u>
- [31] Noble, W., Planel, E., Zehr, C., Olm, V., Meyerson, J., et al. (2005) Inhibition of Glycogen Synthase Kinase-3 by Lithium Correlates with Reduced Tauopathy and Degeneration in Vivo. Proceedings of the National Academy of Sciences of the United States of America, 102, 6990-6995. http://dx.doi.org/10.1073/pnas.0500466102
- [32] Wischik, C.M., Edwards, P.C., Lai, R.Y., Roth, M. and Harrington, C.R. (1996) Selective Inhibition of Alzheimer Disease-Like Tau Aggregation by Phenothiazines. *Proceedings of the National Academy of Sciences of the United States* of America, 93, 11213-11218. <u>http://dx.doi.org/10.1073/pnas.93.20.11213</u>
- [33] Deiana, S., Harrington, C.R., Wischik, C.M. and Riedel, G. (2009) Methylthioninium Chloride Reverses Cognitive Deficits Induced by Scopolamine: Comparison with Rivastigmine. *Psychopharmacology*, 202, 53-65. <u>http://dx.doi.org/10.1007/s00213-008-1394-2</u>
- [34] Wischik, C. (2009) Rember: Issues in Design of a Phase 3 Disease Modifying Clinical Trial of Tau Aggregation Inhibitor Therapy in Alzheimer's Disease. *Alzheimer's & Dementia*, 5, P74. <u>http://dx.doi.org/10.1016/j.jalz.2009.05.175</u>
- [35] Nogales, E., Wolf, S.G., Khan, I.A., Ludueña, R.F. and Downing, K.H. (1995) Structure of Tubulin at 6.5 Å and Location of the Taxol-Binding Site. *Nature*, **375**, 424-427. http://www.nature.com/nature/journal/v375/n6530/pdf/375424a0.pdf
- [36] Amos, L.A. and Löwe, J. (1999) How Taxol® Stabilises Microtubule Structure. Chemistry & Biology, 6, R65-R69. http://dx.doi.org/10.1016/S1074-5521(99)89002-4
- [37] Shemesh, O.A. and Spira, M.E. (2011) Rescue of Neurons from Undergoing Hallmark Tau-Induced Alzheimer's Disease Cell Pathologies by the Antimitotic Drug Paclitaxel. *Neurobiology of Disease*, 43, 163-175. http://dx.doi.org/10.1016/j.nbd.2011.03.008
- [38] Das, V. and Miller, J.H. (2012) Microtubule Stabilization by Peloruside A and Paclitaxel Rescues Degenerating Neu-

rons from Okadaic Acid-Induced Tau Phosphorylation. *European Journal of Neuroscience*, **35**, 1705-1717. http://dx.doi.org/10.1111/j.1460-9568.2012.08084.x

- [39] Michaelis, M.L., Ranciat, N., Chen, Y., Bechtel, M., Ragan, R., et al. (1998) Protection against Beta-Amyloid Toxicity in Primary Neurons by Paclitaxel (Taxol). Journal of Neurochemistry, 70, 1623-1627. http://dx.doi.org/10.1046/j.1471-4159.1998.70041623.x
- [40] Michaelis, M.L., Chen, Y., Hill, S., Reiff, E., Georg, G., Rice, A. and Audus, K. (2002) Amyloid Peptide Toxicity and Microtubule-Stabilizing Drugs. *Journal of Molecular Neuroscience*, **19**, 101-105. http://dx.doi.org/10.1007/s12031-002-0018-2
- [41] Michaelis, M.L., Ansar, S., Chen, Y., Reiff, E.R., Seyb, K.I., et al. (2005) β-Amyloid-Induced Neurodegeneration and Protection by Structurally Diverse Microtubule-Stabilizing Agents. Journal of Pharmacology and Experimental Therapeutics, 312, 659-668. <u>http://dx.doi.org/10.1124/jpet.104.074450</u>
- [42] Brunden, K.R., Yao, Y., Potuzak, J.S., Ferrer, N.I., Ballatore, C., et al. (2011) The Characterization of Microtubule-Stabilizing Drugs as Possible Therapeutic Agents for Alzheimer's Disease and Related Tauopathies. *Pharmacological Research*, 63, 341-351. <u>http://dx.doi.org/10.1016/j.phrs.2010.12.002</u>
- [43] Hoffmann, J., Fichtner, I., Lemm, M., Lienau, P., Hess-Stumpp, H., et al. (2009) Sagopilone Crosses the Blood-Brain Barrier in Vivo to Inhibit Brain Tumor Growth and Metastases. Neuro-Oncology, 11, 158-166. http://dx.doi.org/10.1215/15228517-2008-072
- [44] O'Reilly, T., Wartmann, M., Brueggen, J., Allegrini, P.R., Floersheimer, A., Maira, M. and McSheehy, P.M. (2008) Pharmacokinetic Profile of the Microtubule Stabilizer Patupilone in Tumor-Bearing Rodents and Comparison of Anti-Cancer Activity with Other MTS in Vitro and in Vivo. Cancer Chemotherapy and Pharmacology, 62, 1045-1054. http://dx.doi.org/10.1007/s00280-008-0695-9
- [45] Ballatore, C., Brunden, K.R., Huryn, D.M., Trojanowski, J.Q., Lee, V.M.Y. and Smith III, A.B. (2012) Microtubule Stabilizing Agents as Potential Treatment for Alzheimer's Disease and Related Neurodegenerative Tauopathies. *Jour*nal of Medicinal Chemistry, 55, 8979-8996. <u>http://dx.doi.org/10.1021/jm301079z</u>
- [46] Barten, D.M., Fanara, P., Andorfer, C., Hoque, N., Wong, P.Y.A., *et al.* (2012) Hyperdynamic Microtubules, Cognitive Deficits, and Pathology Are Improved in Tau Transgenic Mice with Low Doses of the Microtubule-Stabilizing Agent BMS-241027. *The Journal of Neuroscience*, **32**, 7137-7145. <u>http://dx.doi.org/10.1523/JNEUROSCI.0188-12.2012</u>
- [47] Hao, L., Zhang, Q., Yu, T., Yu, L. and Cheng, Y. (2011) Modulation of Ultra-Low-Molecular-Weight Heparin on [Ca²⁺]i in Nervous Cells. *Brain Research Bulletin*, 86, 355-359. <u>http://dx.doi.org/10.1016/j.brainresbull.2011.08.018</u>
- [48] Vellas, B., Sol, O., Snyder, P.J., Ousset, P.J., Haddad, R., et al. (2011) EHT0202 in Alzheimers Disease: A 3-Month, Randomized, Placebo-Controlled, Double-Blind Study. Current Alzheimer Research, 8, 203-212. <u>http://dx.doi.org/10.2174/156720511795256053</u>
- [49] Mangialasche, F., Solomon, A., Winblad, B., Mecocci, P. and Kivipelto, M. (2010) Alzheimer's Disease: Clinical Trials and Drug Development. *The Lancet Neurology*, 9, 702-716. <u>http://dx.doi.org/10.1016/S1474-4422(10)70119-8</u>
- [50] Winblad, B., Andreasen, N., Minthon, L., Floesser, A., Imbert, G., *et al.* (2012) Safety, Tolerability, and Antibody Response of Active Aβ Immunotherapy with CAD106 in Patients with Alzheimer's Disease: Randomised, Double-Blind, Placebo-Controlled, First-in-Human Study. *The Lancet Neurology*, **11**, 597-604. http://dx.doi.org/10.1016/S1474-4422(12)70140-0
- [51] Scheltens, P., Sperling, R., Salloway, S. and Fox, N. (2012) Bapineuzumab IV Phase 3 Results. 5th Conference Clinical Trials on Alzheimer's Disease. *The Journal of Nutrition, Health & Aging*, 16, 795-872.
- [52] Doody, R. (2012) Safety and Efficacy of Solanezumab in Patient with Mild to Moderate Alzheimer's Disease: Results from Phase 3. 5th Conference Clinical Trials on Alzheimer's Disease. *The Journal of Nutrition, Health & Aging*, 16, 801-802.
- [53] Hampel, H., Ewers, M., Bürger, K., Annas, P., Mörtberg, A., et al. (2009) Lithium Trial in Alzheimer's Disease: A Randomized, Single-Blind, Placebo-Controlled, Multicenter 10-Week Study. *The Journal of Clinical Psychiatry*, 70, 922-931. <u>http://dx.doi.org/10.4088/JCP.08m04606</u>
- [54] Wischik, C.M., Bentham, P., Wischik, D.J. and Seng, K.M. (2008) O3-04-07: Tau Aggregation Inhibitor (TAI) Therapy with Rember[™] Arrests Disease Progression in Mild and Moderate Alzheimer's Disease over 50 Weeks. *Alzheimer's & Dementia*, 4, T167. <u>http://dx.doi.org/10.1016/j.jalz.2008.05.438</u>
- [55] Zhang, B., Maiti, A., Shively, S., Lakhani, F., McDonald-Jones, G., et al. (2005) Microtubule-Binding Drugs Offset Tau Sequestration by Stabilizing Microtubules and Reversing Fast Axonal Transport Deficits in a Tauopathy Model. Proceedings of the National Academy of Sciences of the United States of America, 102, 227-231. http://dx.doi.org/10.1073/pnas.0406361102
- [56] Texel, S.J. and Mattson, M.P. (2011) Impaired Adaptive Cellular Responses to Oxidative Stress and the Pathogenesis of Alzheimer's Disease. *Antioxidants & Redox Signaling*, 14, 1519-1534. <u>http://dx.doi.org/10.1089/ars.2010.3569</u>



Neurophysiological Biomarker of Mild Cognitive Impairment*

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Abstract

Mild cognitive impairment is sometimes regarded as related to aging. However, statistically every second case turns into full dementia, which still is resistant to any treatment. It is therefore desirable to recognize deviations from normality as early as possible. This might be feasible by using quantitative EEG analysis in the presence of mental work. The present retrospective data analysis revealed a new quantitative biomarker indicating the degree of impairment. Current source density was calculated from 16 channel EEG using CATEEM® software. Four different conditions were analyzed: relaxed state, performing a d2-concentration test, a calculation performance test and a memory test for 5 min each. Subjects older than 40 years were divided into two groups according to their DemTect score: 13 - 18 (HC; n = 44) or 8 - 12 (MCI; n = 45). Spectral power was chopped into six frequency ranges (delta, theta, alpha 1, alpha 2, beta 1 and beta 2). Average spectral power was enhanced in the MCI group in comparison to healthy subjects with respect to delta (p = 0.05) during relaxed state when all electrode positions were regarded. With respect to EEG recording during performance of three different psychometric tests it was recognized that mainly spectral changes during performance of the d2-concentration test were related to mild cognitive impairment. With regard to all electrode positions statistically significantly lower spectral power values were reached during the d2-test for delta (p = 0.001), theta (p = 0.0001) and alpha 1 waves (p = 0.001) 0.08) in impaired subjects in comparison to healthy subjects. Regarding regions of interest increases of delta and theta power were seen in the fronto-temporal brain during performance of the d2-concentration test. These increases disappeared when looking at MCI data. In the centro-parietal region decreases of alpha and beta 1 power emerged, which were even larger in MCI subjects. No MCI-dependent changes were observed in the other two tests. A correlation was found between psychometric performance of the d2-test and the DemTect score (r = 0.51). MCI subjects had statistically significant worse performance in all three mental challenges in comparison to healthy volunteers. It is concluded that MCI can be characterized at an early stage by EEG recording in the relaxed state. High spectral delta and theta power in general and specifically at fronto-

^{*}Data was presented as poster at the 6th World Congress on Controversies in Neurology, Vienna, Austria, March 8-11, 2012 and awarded the best one in the category dementia.

temporal electrode positions (especially at T₃) was recognized as a biomarker for MCI. A DemTect score of 8-12 was validated as indicative for MCI.

Keywords

DemTect, Cognition, Psychometry, EEG, Source Density, Mild Cognitive Impairment (MCI), Alzheimer's Disease, CATEEM

1. Introduction

Mild cognitive impairment is regarded as a possible transitional stage during the development of Alzheimer's disease [1]. According to literature about every second subject suffering from this impairment develops manifest dementia [2]-[4]. Since progression from mild cognitive impairment to dementia occurs fluently, the problem arises to find a quantitatively defined diagnosis of this stage in order to recognize possible parameters for the risk of development of dementia later on. This early brain dysfunction can be described by considering three different ap proaches. The first ap proach consists i n u sing a n in teractive questionnaire like mini-mental state (MMS [5]) or DemTect [6] [7]. The second approach is represented by use of psychometric tasks (cognitive testing) like the d2-test for concentration, a memory test or performance of arithmetic calculations [8]. The third approach very often is covered by a neurophysiological methodology, usually recording of quantitative electroencephalography [9] [10] or biochemical parameters from cerebrospinal fluid [11]. Thus, a combination of measurements on these three levels should be able to provide a quantitative definition of mild cognitive impairment and fulfil the criteria for an early diagnosis and recognition of the risk potential. Results should also provide the quantitative base for testing of drugs or food supplements aiming at the improvement of these early deficits.

The present investigation deals with the comparison of d ata from 45 he althy control subjects (HC group; DemTect score 13 - 18) with 44 subjects rated as cognitively impaired on the base of a DemTect score between 8 and 12 (MCI group). Concomitant use of psychometric performance and current source density imaging [12] [13] is used to validate this s core range for the d iagnosis of m ild c ognitive i mpairment as p roposed by the DemTect guideline in comparison to being mentally healthy. With respect to EEG, basic conditions (recording during the relaxed eyes open condition) as well as event related EEG during performance of mental tasks [14] are used in order to learn more on possible deficits with respect to special demands. The analysis is based on data from 89 subjects using recordings of the first visit in the laboratory during several clinical studies (EudraCT-Nr.: 2007-004753-29 and EudraCT-Nr. 2009-015827-97).

2. Methods

2.1. Subjects

Eighty-nine subjects were taken from five consecutive clinical studies. They were asked to perform an interactive questionnaire developed for recognition of mild cognitive impairment (DemTect) before they entered the study consisting of a combined technology of EEG recording in the presence of mental performance of three different psychometric tasks. Volunteers were grouped according to the proposal of the developers of the test and according to the validation results published. Subjects having a score from 8 - 12 were assembled into the cognitive impaired group (MCI), those scoring higher from 13 - 18 were taken as healthy control group (HC). Both groups were compared with respect to electric power under the different recording conditions and with respect to psychometric performance.

2.2. Experimental Procedure

Subjects (HC group 17 male/27 female, average age 50.5 and 47 years, respectively; MCI group 25 male/20 female, average age 58.1 and 56.6 years, respectively) were sitting alone in a quiet separate room in a comfortable easy chair. The light was dimmed. Baseline recording of 6 min under the condition of eyes open was followed by the performance of the d2-test, a mathematical calculation task and a memory test. All experiments took place at the same time of the day (starting at 8 am).

2.3. EEG Recording

The EEG was recorded bipolarly from 17 surface electrodes according to the international 10/20 system [15] against a common average reference calculated from Cz against all other electrodes as proposed by Lehmann [16] (Computer ai ded topographical el ectroencephalometry: CATEEM[®] from MEWICON CATEEM-Tec GmbH, A-4164 Schwarzenberg, Austria) using an electrocap. EEG recording was performed as reported earlier [17]. Setting was kept constant for each individual throughout the experiment.

2.4. Current Source Density Analysis

In this study the EEG was processed not in the potential mode based on voltage [18], but in a surface charge mode obtained by Laplacian estimates also known as current source density analysis (CSD), [12] [13]. Charge is the 2^{nd} derivation of the potential and gives the curvature of the potential curve according to space. Under the condition of using a homogenous, steadily conducting medium surface charge mode provides the source density of the electric flow on the cortex surface. Whereas the EEG in the potential mode tends to produce a more extensive and diffuse picture of changes, Laplacian estimate acts as a spatial filter emphasizing local sources over distant sources (for review see [19]). There is a sharply contrasted display of cortex areas with highly activated generators in the depth of the brain and brain areas with less intensely working generators. Harmony *et al.* [12] were able to demonstrate, that spectral parameters obtained from the CSD showed higher correlations with computer tomography measures than those calculated from the potential mode of the EEG. We therefore used this methodology in order to describe the focal changes of brain activity.

Brain Imaging was achieved by conversion of numerical values of spectral EEG power into spectral colours and additive colour mixture according to RGB as used in TV settings [20]. Data acquisition and analysis were carried out simultaneously and provide topographical maps displayed on-line on the computer screen. The maps show the relative, time averaged changes of electrical brain activity of each recording condition during mental work in % of the reference period during relaxation with open eyes.

2.5. Psychometric Testing

The d2 attention test is a well-known standardized validated test. Number of correct answers and number of lines were evaluated as performance index including quality and quantity of answers. Arithmetic test (Concentration Performance Test (CPT) was carried out as described by Düker and Lienert, [8]. Number of solved tasks and correctness gave a performance index. The memory test was applied according to the following schema: a combination of 8 numbers and/or letters (for example: (Dv8L3oPX) was presented on the screen for 4 s. After this no information was given for 10 s. Finally, a fourfold multiple choice including the correct answer was presented for decision. Number of tasks and correctness were evaluated to give a performance index. Each test was presented for 5 minutes. The row of order was kept constant for the sake of direct comparisons of the results under identical conditions.

2.6. Statistics

Since EEG data are not normally distributed, the non-parametrical Wilcoxon test was chosen for comparison between the two groups. Data were averaged for each of the recording periods of 5 minutes during eyes open and the different challenges (*i.e.* in separate for each psychometric test). Statistics gave p values, which are presented at the appropriate site. The absolute power values under the recording condition "eyes open" were taken as r eference values and s et to 1 00% when p sychometric tests were p erformed. Thus, p ossible p hysiological changes during test performance are given in % of these reference values.

In order to differentiate results from healthy subjects and those suffering from mild cognitive impairment data were fed into linear discriminant analysis according to Fischer. Results from the first three discriminant functions were depicted in space (x, y and z coordinates). Results from the 4^{th} to 6^{th} functions were transformed into colour according to the RGB mode (like in TV).

3. Results

3.1. Electric Power during the Recording Condition "Eyes Open"

Comparison of the absolute electric power values with respect to all 17 electrode-positions of the source density EEG of subjects with mild cognitive impairment (MCI; n = 45) with the healthy control group (HC; n = 44) revealed massive differences. Higher values of electric power at single electrodes were generally seen in the group of subjects with MCI in comparison to healthy subjects. Global median values regarding all electrode positions were higher in MCI subjects than in HC subjects but did not reach such high statistical significance as with respect to delta (Table 1).

Table 1. Absolute power values for each electrode position under the recording condition of "eyes open" in relaxed position for every frequency range from delta, through theta, alpha 1, alpha 2, beta 1 and beta 2. Electrode positions: C = central, P = parietal; F = frontal; T = temporal; O = o ccipital. Even numbers represent the right hemisphere, uneven numbers the left hemisphere. Statistically significant differences with respect to median values regarding all electrodes from healthy subjects (HC; n = 44) in comparison to individuals with mild cognitive impairment (MCI; n = 45) are marked by stars before the number. * = p < 0.05; ** = p < 0.01; *** = p < 0.001.

Eyes Open [µV ²]													
Electrode	Delta		Theta		Alp	Alpha 1		Alpha 2		Beta 1		Beta 2	
	НС	MCI	НС	MCI	НС	MCI	НС	MCI	НС	MCI	НС	MCI	
Cz	2.71	3.23	0.58	0.58	0.55	0.66	0.36	0.56	0.62	0.67	0.89	1.15	
Fz	3.03	3.29	0.72	0.74	0.74	0.86	0.49	0.67	0.67	0.78	0.96	1.33	
F3	2.93	*3.89	0.64	0.81	0.76	0.97	0.51	0.66	1.04	1.17	2.08	2.49	
C3	2.10	*3.65	0.52	0.56	0.58	0.77	0.63	1.04	1.24	1.40	1.76	2.04	
Р3	1.71	*2.21	0.42	0.40	0.52	0.63	0.48	0.65	0.88	0.72	0.77	0.63	
Pz	2.49	2.48	0.54	0.53	0.58	0.75	0.52	0.69	0.67	0.70	0.66	0.62	
P4	1.73	2.17	0.35	0.42	0.44	0.55	0.44	0.73	0.76	0.71	0.73	0.74	
C4	2.36	2.89	0.50	0.55	0.57	0.78	0.70	1.04	1.39	1.45	1.96	2.31	
F4	3.52	4.12	0.72	0.78	0.86	0.91	0.62	0.82	1.11	1.33	2.60	3.75	
F7	7.07	10.59	1.39	1.41	1.38	1.79	1.15	1.42	1.98	1.87	4.75	3.16	
Т3	4.16	4.62	0.88	0.89	1.22	1.31	1.23	1.32	2.44	1.78	3.36	3.10	
Т5	3.19	3.79	1.10	0.96	1.74	1.51	1.37	1.25	2.07	1.70	1.84	1.34	
01	3.10	***5.88	0.73	*0.88	0.68	1.17	0.70	*0.99	1.30	1.60	1.85	2.65	
02	4.00	**5.46	0.79	0.86	0.90	0.91	0.87	1.10	1.49	1.83	2.20	2.40	
T6	3.07	2.88	0.77	0.69	1.14	1.51	1.04	1.36	1.86	1.50	1.72	1.70	
T4	3.79	4.12	0.93	0.89	1.20	1.38	1.18	1.11	2.37	2.13	3.31	3.35	
F8	7.84	*11.88	1.57	1.60	1.61	1.92	1.42	1.40	2.67	2.01	6.18	3.66	
global median	3.14	*3.62	0.71	0.74	0.83	0.91	0.75	0.86	1.33	1.43	1.81	1.92	

3.2. Spectral Power during Performance of the d2-Concentration Test

Regarding the median of all electrode positions during performance of the d2-concentration test statistically different values for the MCI subjects in comparison to healthy control were obtained when recordings in the relaxed s tate were t aken as r efference (100%). H ighest s tatistical significance for the d ifference b etween the healthy and the cognitively impaired group is reached in the delta (p < 0.001) and theta frequency range (p < 0.001). But also lower values for alpha 2 power (p < 0.08) were seen during this challenge. Lower spectral power in the alpha 1 frequency range did not reach statistical significance. No statistically significant differences between MCI subjects and healthy controls were observed with the calculation performance test (CPT) or the memory test (Table 2).

During performance of the d2-test **healthy** volunteers (HC-group) were able to **increase** fronto-temporal delta and theta power taking the recording condition "eyes open" as reference (100%). Highest statistical significance was reached by increases of theta power in frontal and temporal areas of the brain as documented in **Figure 1** by statistical analysis for each location in separate. At the same time **attenuation** of electric power were observed with respect to alpha waves mainly in central areas of the brain.

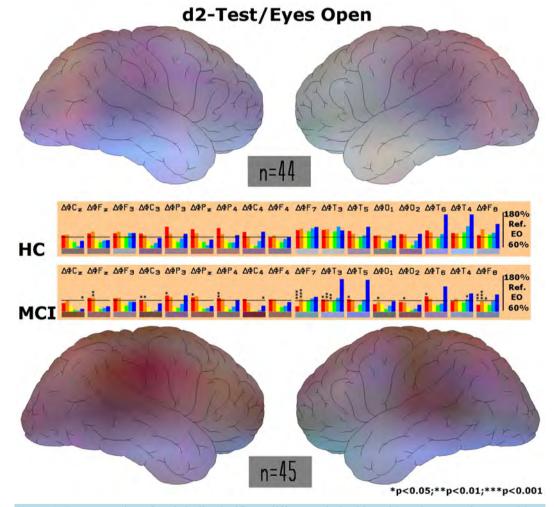


Figure 1. Documentation of statistically significant differences in healthy subjects in comparison to subjects suffering from MCI with respect to every single electrode position under the recording condition of the d2-test. Statistically significant differences between H C and M CI are documented by s tars. B rain maps were constructed using the recording condition eyes open as reference. Differences are shown between healthy subjects (HC) and s ubjects with m ild c ognitive impairment (MCI) during performance of the d2-test. P lease note fronto-temporal brightening in the left hemisphere (right side of upper map) in healthy people compared to lack of such feature in mild cognitive impairment (lower map).

Opposite to this, subjects diagnosed to have **MCI** according to "DemTect" score produced significant less electric power regarding delta and theta frequencies during performance of this test. Differences in changes of power with respect to single electrode locations are depicted in Figure 2 for the HC control group (upper panel) and the MCI group (lower panel), respectively. Statistical significance is given in the lower part for each location of recording.

During the course of brain research it has become obvious that electric activity depends on the region where one looks at. Different mental challenges have been recognized to induce quite different patterns of electric activity. This is also seen during performance of a concentration test (**Figure 1**). Under this recording condition eminent higher delta spectral power values are observed at parietal areas (electrode positions $P_{3,4}$) as well as delta and theta spectral power in fronto-temporal areas represented by the electrode positions $F_{7,8,72}$ and $T_{3,4}$. In MCI subjects these increases disappear nearly completely in a highly statistically significant manner. In addition, alpha waves are depressed in the parietal region during the d2test. This decrease became even more pronounced in mentally impaired subjects. This difference also becomes visible when looking at the electric maps

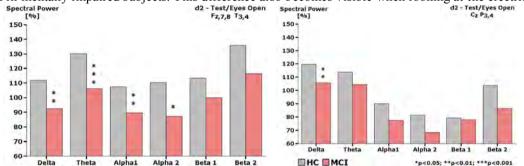


Figure 2. Differences in spectral EEG power between healthy subjects (HC) and subjects suffering from mild c ognitive i mpairment (MCI) with r espect t o t wo r egions of interest (ROI) when performing the d2-concentration t est. D ata f rom t he r elaxed s tate are t aken as r efference. F ronto-temporal r egion is represented by the electrode positions $F_{z_27,8}$ and centro-parietal region is represented by the positions C_z and P_{3x4} .

Table 2. Differences of frequency changes (test condition in % relative to eyes open condition) between healthy subjects (HC) and the MCI group are given for each psychometric test as median values taking all electrode positions in consideration. Statistical significance is given as p-values for the comparison between healthy people (HC) and subjects suffering from mild cognitive impairment (MCI).

d2-Test/Eyes Open [%]												
	Delta		Theta		Alpha 1		Alpha 2		Beta 1		Beta 2	
	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI
global median	116.26	98.41	110.67	99.66	93.77	88.74	86.82	76.67	88.07	81.75	118.80	100.54
p <	0.001		0.001		0.076							
	CPT/Eyes Open [%]											
	Delta		Theta		Alpha 1		Alpha 2		Beta 1		Beta 2	
	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI
global median	117.49	113.91	102.32	106.54	90.32	81.79	80.66	72.70	85.53	86.86	98.19	103.84
p <												
				N	lemory/E	yes Open	[%]					
	Delta		Theta		Alpha 1		Alpha 2		Beta 1		Beta 2	
	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI	HC	MCI
global median	116.60	98.50	97.72	88.91	88.08	81.03	90.39	75.68	92.32	90.30	101.95	94.97
p <												

calculated for both groups (Figure 1). Due to the absence of theta power a large difference is seen in fronto-

temporal areas. Thus, under the recording condition "d2-concentration test" massive differences with respect to spectral power in fronto-temporal and parietal brain regions are visible. The numerical values for these two regions of interest are given in separate as median of the respective fronto-temporal and parietal electrode positions (Figure 2).

3.3. Electric Power during the Recording Condition "CPT"

During performance of the calculation-performance-test (CPT) **healthy** volunteers were able to increase frontal delta and theta power in comparison to the recording condition "eyes open" (set to 100%). As depicted in the middle bar chart of **Figure 3**, electrode locations F_7 , F_8 , T_5 and T_6 show increases of delta and theta power. With respect to this hardly a difference was observed in cognitively impaired volunteers. However, when looking at the brain map depicted in **Figure 3**, left frontal increases of slow power were attenuated in mildly impaired subjects. Despite the impression of lower production of delta and theta waves in the impaired group (as also documented in **Figure 4**) this difference was not statistically significant. The same is true for beta power. Global

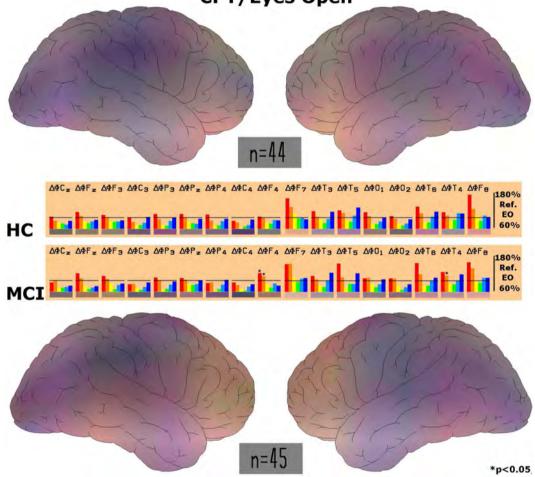


Figure 3. Documentation of statistically significant differences in healthy subjects in comparison to subjects suffering from MCI with respect to every single electrode position under the recording condition of the calculation performance test (CPT). Statistically, significant differences between HC and MCI are documented by a star. Brain maps constructed using the recording condition eyes open as reference. Differences are shown between healthy subjects (HC) and subjects with mild cognitive impairment (MCI) during performance of the d2-test. Please note that only marginal differences between healthy people subjects suffering from mild cognitive impairment are seen.

CPT/Eyes Open

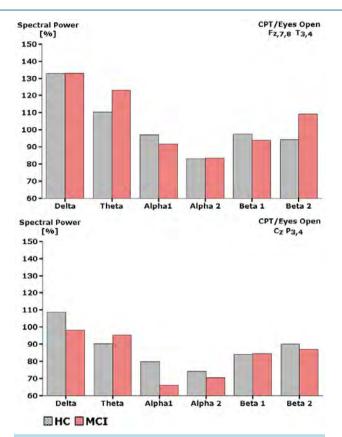


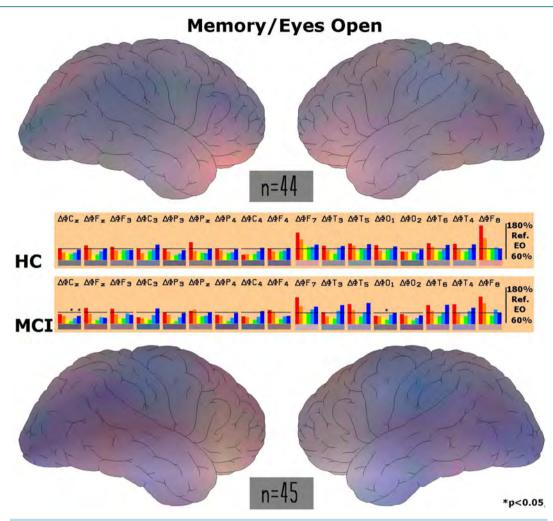
Figure 4. Differences in s pectral E EG power b etween h ealthy subjects (HC) and s ubjects s uffering from mild c ognitive i mpairment (M CI) with respect to two regions of interest (RO I) when p erforming the cal culation p erformance t est. D ata from the relaxed state are taken as reference. Fronto-temporal region is r epresented by the el ectrode p ositions $F_{z_57.8}$ and ce n-tro-parietal region is represented by the positions C_z and $P_{3,4}$.

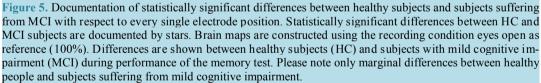
median values of electric power with regard to all electrode positions during the performance of the calculation test are given in Table 2 (middle part). Despite some further reduction of alpha 1 and alpha 2 power in mildly impaired subjects this difference in comparison to healthy subjects did not become statistically significant.

This result is underlined by the spectral changes as observed in the two regions of interest. Despite some lower value with respect to alpha 1 power in mildly impaired subjects, the difference did not become statistically significant as depicted in Figure 4.

3.4. Electric Power during the Recording Condition "Memory Test"

During performance of the memory test, **healthy** volunteers were able to increase frontal delta power (F_{7,8}) and to some extent also theta power in comparison to the recording condition "eyes open" (set to 100% in **Figure 5**). A similar feature was observed in cognitively impaired volunteers. But with respect to delta and theta spectral power cognitively impaired subjects produced somewhat less spectral power (bar chart in the middle of **Figure 5**). Electric maps in mildly impaired subjects reveal less red colour in the frontal brain due to some but statistically not significant delta decreases. Differences between healthy controls and impaired subjects are also obvious from the regions of interest in **Figure 6**. Obviously, the lower spectral delta power is statistically not significantly different between the two groups. However, alpha 2 spectral power was significantly less in impaired subjects. When regarding global median values (calculated from all electrode positions) also a t endency of attenuation of alpha 2 spectral power emerged, but which did not become statistically significant at this global measurement (**Table 2**).





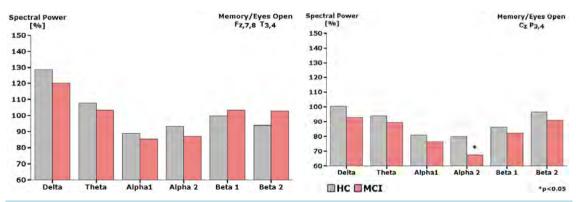


Figure 6. Differences in spectral EEG power between healthy subjects (HC) and subjects suffering from mild cognitive impairment (MCI) with respect to two regions of interest (ROI) when performing the memory test. Data from the relaxed state are taken as reference (100%). The frontotemporal region is represented by the electrode positions $F_{z_37,8}$ and the centroparietal region is represented by the positions C_z and $P_{3,4}$.

3.5. Comparison by Means of Discriminant Analysis

Discriminant analysis is a mathematical tool, which allows statistical evaluation of a large set of parameters. Since this quantitative EEG analysis consists of 102 parameters (17 electrode positions \times 6 frequency bands) absolute spectral power data from all participants of the studies were fed into this type of analysis. As is documented in **Figure 7** healthy controls and impaired subjects can be discriminated from each other with respect to all 4 recording conditions. It is also obvious that the different recording conditions lead to different types of electric brain states. For example the state of "eyes open" can easily discriminated from the state during performance of these psychometric tests.

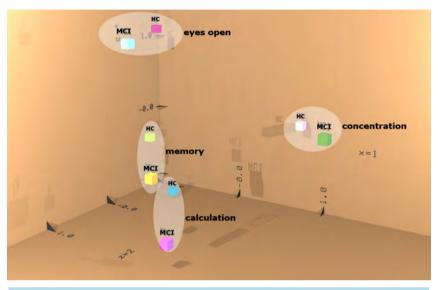


Figure 7. Documentation of results of discriminant analysis for both groups of subjects (HC = healthy controls; M CI = mild cognitive impairment) with r espect t o p erformance of p sychometric t ests and t he r ecording condition r elaxed s tate with "ey es open". Results from the first three discriminant functions are depicted in space (x, y and z coordinates). Results from the 4th to 6th discriminant functions are depicted using the RGB mode (additive colour mixture of r ed, green and b lue for the result of each function). Single test conditions are marked by arbitrarily brightening.

3.6. Evaluation of Psychometric Results

Psychometric performance was documented for each of the three mental tests according to the definition given under "Material and Methods". As described in **Table 3** performance of cognitively impaired subjects was always significantly lower in all three tests with respect to an index calculated on the base of quality and quantity of the answers.

In addition, a statistically significant correlation was observed between the psychometric performance during these tests and the score of the "DemTect". These data confirm the cognitive impairment as indicated by the "DemTect" score, since a correlation was observed between the DemTect score and these psychometric results (r = 0.53; p = 0.000001) as depicted in Figure 8.

and deviation are given besides the statistical significance (p-values) on the left side.										
	Mean HC	SEM HC	Mean MCI	SEM MCI	Statistical Significance					
d2-test	11.20	0.48	8.21	0.43	p < 0.0001					
СРТ	4.46	0.59	2.93	0.49	p < 0.03					
Memory	10.38	0.49	8.36	0.64	p < 0.02					

Table 2 Deput for neurophemotric testing (details under motorial and methods). Mean values

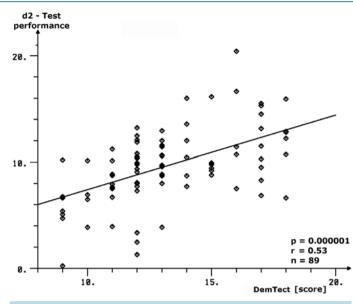


Figure 8. Statistically significant correlation between DemTect score values and psychometric performance in the d2-test. The rank correlation value according to Spearman is given with statistical significance.

4. Discussion

Two groups of subjects as defined according to the result of the interactive DemTect questionnaire already differed with respect to absolute voltage in the delta frequency range providing higher values for the MCI group. Largest differences were seen in frontal delta and theta power (electrode positions F_7 and F_8). This feature of higher delta and theta power has been reported using the same methodology for demented patients in comparison to normal aged matched controls [21]. Other authors also have recently documented higher delta values for subjects suffering from mild cognitive impairment in comparison to healthy volunteers and reported on further alterations typical for those seen in Alzheimer's disease [22]. Higher power in the delta frequency range indicates lower activity of the cholinergic transmitter system [23]. In animal lesion studies, increased delta activity has been reported after destruction of the Nucleus Basalis [24], the main production site of acetylcholine [25]. A comparison of cortical source EEG with MR-based measurements of lobar brain volume (white and grey matter) revealed a n egative correlation between the frontal white matter and the amplitude of the delta sources across MCI and Alzheimer subjects (AD), which support a transition hypothesis of brain structural and functional continuity between MCI and AD [26].

Changes of delta and theta waves were also related to pathological changes in cholinergic brain regions. A significant increase of these frequencies was found in patients with the highest total cholinergic burden as well as in patients with highest capsular pathway damage [27]. Abnormal high frontal delta waves **under basic re-laxed recording conditions** can therefore be regarded as indicative for a biochemical and pathological brain dysfunction involving the cholinergic transmitter system.

This also applies to theta waves, which also have been reported to be significantly higher in demented patients than in controls and related to decreased performance in all cognitive domains [28]. In this analysis increase of theta p ower was observed in m ost of the b rain r egions and the d ifference b etween the M CI g roup and the healthy control group (HC) was however not statistically significant with respect to global median values. Very strong evidence for theta power increase as indicator for cognitive decline comes also from longitudinal studies in normal elderly with subjective complaints [1]. With respect to EEG frontal theta power a negative linear correlation was reported to hippocampal volumes for patients suffering from MCI or AD compared to control also indicating fluent transitions for these d iseases [29]. These d ata cl early contradict i mpressions from n uclear magnetic resonance pictures suggesting only one frontal brain area to be involved in cognitive function. In addition, electric features of brain activity recorded under different performance conditions provide evidence for the involvement of different transmitter activities within different brain regions. There is evidence that theta waves are under the control of the norepinephrine alpha 2 receptor [30]. Also changes of beta frequency ranges have

been used to successfully predict the clinical status of subjects with MCI over a two years period [31]. However, changes of beta activity did not become statistically significant in my analysis.

Thus, E EG source density measurements in the presence of a r elaxed state confirm deviations of electric power within several frequency ranges reported in the literature so far. But like an engine can only be tested under load, brain function should be tested also under "mental" load in order to characterize possible dysfunctions in a more relevant manner. This is possible by recording the EEG in the presence of cognitive performance. Using a battery of different psychometric tests it was recognized long ago that frontal delta and theta power increase under those conditions in healthy volunteers and were related to the difficulty of the task [14]. Interestingly, it was then observed, that demented patients were only able to produce power increases, which were significantly lower than those observed in age matched healthy controls [32]. At the same time a significant correlation was recognized to the severity of the disease as indicated by the interactive questionnaires MMS and ADAS. The present analysis reveals a similar picture. All three psychometric tests induced fronto-temporal increases of delta and theta power in healthy controls. Increases of fronto-central electric theta power have also been observed during other memory demands [33]. Reflection of cognitive and memory performance in the EEG has also been described in detail by others. For example, retrieval of lexical semantic information was linked to theta increases [34]. Furthermore, it was suggested that the encoding of new information is reflected by theta oscillations in hippocampal-cortical feedback loops, whereas search and retrieval processes in (semantic) long-term memory are reflected in upper alpha (alpha 2) oscillations in thalamo-cortical feedback loops [35]. According to animal data alpha 2 frequencies are under the control of dopamine.

In the group of mild cognitive impairment, however, performance of the d2-test led to considerable smaller increases of delta and theta power. Similar results were described for individuals with mild cognitive impairment in the literature [36]. As already recognized in demented patients and confirmed by a correlation analysis between basic theta and event related induced theta [32] this lacking production of fronto-temporal theta waves presumably derives from too high baseline values during the relaxed eyes-open condition. There is obviously a ceiling effect, which prevents further increase of theta power after reaching a physiologically limited maximum. Interestingly, low performance in attention testing was reported to be associated with reduced grey matter density of the left inferior frontal gyrus [37]. In demented patients under the condition of mental load, theta changes were related to the MMS questionnaire. In subjects suffering from mild cognitive impairment a close correlation between theta changes and the score of the DemTect is now observed. This parallel feature speaks in favor of fluent transient states from being healthy via mild cognitive impairment to dementia. But, according to the literature only about every second individual suffering from a decline in cognition develops dementia. In summary, there is compelling evidence, that this change in theta power reactivity can be taken as an indicator for decline of cognition. A longitudinal analysis of future recordings from our subjects will tackle this question.

It can be concluded from our data, that deficits in concentration seem to be the first and most important sign of mild cognitive impairment represented by aberrations in theta activity, followed by already some deficits in memory recognized by deviations in alpha 2 reactivity, whereas arithmetic deficits are not so obvious at this early stage of cognitive impairment. In summary, cognition is a rather complex process, which involves several parts of the brain with increases of electric power in frontal delta and theta waves but also decreases of power in central alpha 2 waves governed by different neurotransmitters. Which of the differences between healthy and mildly impaired subjects are indicative for final development into dementia will hopefully be discovered in future longitudinal studies. But we have now clear neurophysiological parameters to follow in future measurements.

5. Conclusion

The present analysis of current source density of the EEG resulted in the detection of quantitative parameters, which are suitable to diagnose mild cognitive impairment at a very early stage. Lower production of theta waves during performance of the d2-test as paper pencil version seems to be the most sensitive neurophysiologic indicator of a cognitive decline. This parameter can now also be used as a non-invasive biomarker for early diagnosis and for testing new drugs aiming at the prevention of development of MCI into dementia.

Competing Interest

There is no conflict of interest.

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References

- Prichep, L.S., John, E.R., Ferris, S.H., Rausch, L., Fang, Z., Cancro, R., Torossian, C. and Reisberg, B. (2006) Prediction of Longitudinal Cognitive Decline in Normal Elderly with Subjective Complaints Using Electrophysiological Imaging. *Neurobiology of Aging*, 27, 471-481. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2005.07.021</u>
- [2] Schofield, P.W., Marder, K., Dooneief, G., Jacobs, D.M., Sano, M. and Stern, Y. (1997) Association of Subjective Memory Complaints with Subsequent Decline in Community-Dwelling Elderly Individuals with Baseline Cognitive Impairment. *American Journal of Psychiatry*, **154**, 609-615.
- [3] Wang, L., van Belle, G., Crane, P.K., Kukull, W.A., Bowen, J.D., McCormick, W.C. and Larson, E.B. (2004) Subjective Memory Detoriation and Future Dementia in People Aged 65 and Older. *Journal of the American Geriatrics Society*, 52, 2045-2051. <u>http://dx.doi.org/10.1111/j.1532-5415.2004.52568.x</u>
- [4] Treves, T.A., Verchovsky, R., Klimovitzky, S. and Korczyn, A.D. (2005) Incidence of Dementia in Patients with Subjective Memory Complaints. *International Psychogeriatrics*, 17, 265-273. http://dx.doi.org/10.1017/S1041610205001596
- [5] Folstein, M.F., Folstein, S.E. and McHugh, P.R. (1975) Mini-Mental State. A Practical Method for Grading the Cognitive State of Patients for the Clinician. J Psychiatr Res, 12, 189-198.
- [6] Kessler, J., Calabrese, P., Kalbe, E. and Berger, F. (2000) DemTect. Ein neues Screening-Verfahren zur Unterstützung der Demenzdiagnostik. *Psycho*, 26, 343-347.
- [7] Kalbe, E., Kessler, J., Calabrese, P., Smith, R., Passmore, A.P., Brand, M. and Bullock, R. (2004) DemTect: A New, Sensitive Cognitive Screening Test to Support the Diagnosis of Mild Cognitive Impairment and Early Dementia. *International Journal of Geriatric Psychiatry*, **19**, 136-143. <u>http://dx.doi.org/10.1002/gps.1042</u>
- [8] Düker, H. and Lienert, G.A. (1965) Der Konzentrationsleistungstest (KLT). Göttingen, Hofgrefe.
- [9] Jelic, V., Johansson, S.E., Almkvist, O., Shigeta, M., Julin, P., Nordberg, A., Winblad, B. and Wahlund, L.O. (2000) Quantitative Electroencephalography in Mild Cognitive Impairment: Longitudinal Changes and Possible Prediction of Alzheimer's Disease. *Neurobiology of Aging*, 21, 533-540. <u>http://dx.doi.org/10.1016/S0197-4580(00)00153-6</u>
- [10] Alexander, D.M., Arns, M.W., Paul, R.H., Rowe, D.L., Cooper, N., Esser, A.H. and Fallahpour, K. (2006) EEG Markers for Cognitive Decline in Elderly Subjects with Subjective Memory Complaints. *Journal of Integrative Neuroscience*, 5, 49-74. <u>http://dx.doi.org/10.1016/S0197-4580(00)00153-6</u>
- [11] Stomrud, E., Hansson, O., Minthon, L., Blennow, K., Rosen, I. and London, E. (2010) Slowing of EEG Correlates with CSF Biomarkers and Reduced Cognitive Speed in Elderly with Normal Cognition over 4 Years. *Neurobiology of Aging*, 31, 215-223. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2008.03.025</u>
- [12] Harmony, T., Fernandez-Bouzas, A., Marosi, E., Fernandez, T., Bernal, J., Rodriguez, M., Reyes, A., Silva, J., Alonso, M. and Casian, G. (1993) Correlation between Computed Tomography and Voltage and Current Source Density Spectral Parameters in Patients with Brain Lesions. *Electroencephalography and Clinical Neurophysiology*, **87**, 196-205. <u>http://dx.doi.org/10.1016/0013-4694(93)90019-R</u>
- [13] Dimpfel, W., Hofmann, H.C., Prohaska, A., Schober, F. and Schellenberg, R. (1996) Source Density Analysis of Functional Topographical EEG: Monitoring of Cognitive Drug Action. *European Journal of Medical Research*, 1, 283-290.
- [14] Schober, F., Schellenberg, R. and Dimpfel, W. (1995) Reflection of Mental Exercise in the dynamic Quantitative Topographical EEG. *Neuropsychobiology*, 31, 98-112. <u>http://dx.doi.org/10.1159/000119179</u>
- [15] Jasper, H.H. (1958) The Ten-Twenty Electrode System of the International Federation. *Electroencephalography and Clinical Neurophysiology*, 10, 371-375.
- [16] Lehmann, D. (1987) Principles of Spatial Analysis. In: Gevins, A.S. and Remond, A., Eds., *Handbook of Electroence-phalography and Clinical Neurophysiology*, Rev. Series, Vol. 1: Methods of Analysis of Brain and Magnetic Signals, Elsevier, Amsterdam, 309-354.
- [17] Dimpfel, W., Kler, A., Kriesl, E., Lehnfeld, R. and Keplinger-Dimpfel, I.K. (2006) Neurophysiological Characterization of a Functionally Active Drink Containing Extracts of Ginkgo and Ginseng by Source Density Analysis of the Human EEG. *Nutritional Neuroscience*, 9, 213-224. <u>http://dx.doi.org/10.1080/10284150601043713</u>
- [18] Berger, H. (1929) Über das Elektroenkephalogramm des Menschen. Archiv für Psychiatrie und Nervenkrankheiten, 87, 527-570. <u>http://dx.doi.org/10.1007/BF01797193</u>

- [19] Brunet, D., Murray, M.M. and Michel, C.M. (2011) Spatiotemporal Analysis of Multichannel EEG: CARTOOL. Computational Intelligence and Neuroscience, 2011, Article ID: 813870. <u>http://dx.doi.org/10.1155/2011/813870</u>
- [20] Dimpfel, W., Kler, A., Kriesl, E., Lehnfeld, R. and Keplinger-Dimpfel, I.K. (2007) Source Density Analysis of the Human EEG after Ingestion of a Drink Containing Decaffeinated Extract of Green Tea Enriched with L-Theanine and Theogallin. *Nutritional Neuroscience*, 10, 169-180. <u>http://dx.doi.org/10.1080/03093640701580610</u>
- [21] Schellenberg, R., Todorova, A., Hofmann, H.C., Dimpfel, W. and Schober, F. (1995) Differentiation of Demented Patients and Healthy Subjects by Means of Quantitative-Topological EEG—A Classification Approach. *Alzheimer's Research*, **1**, 23-28.
- [22] Babiloni, C., Frisoni, G., Steriade, M., Bresciani, L., Binetti, G., Percio, C.D., Geroldi, C., Miniussi, C., Nobili, F., Rodriguez, G., Zappasodi, F., Carfagna, T. and Rossini, P.M. (2006) Frontal White Matter Volume and Delta EEG Sources Negatively Correlate in Awake Subjects with Mild Cognitive Impairment and Alzheimer's Disease. *Clinical Neurophysiology*, **117**, 1113-1129. <u>http://dx.doi.org/10.1016/j.clinph.2006.01.020</u>
- [23] Dimpfel, W. (2005) Pharmacological Modulation of Cholinergic Brain Activity and Its Reflection in Special EEG Frequency Ranges from Various Brain Areas in the Freely Moving Rat (Tele-Stereo-EEG). *European Neurospycho-pharmacology*, 15, 673-682. <u>http://dx.doi.org/10.1016/j.euroneuro.2005.03.006</u>
- [24] Buzsaki, G., Bickkford, R.G., Ponomareff, G., Thal, L.J., Mandel, R. and Gage, F.H. (1988) Nucleus Basalis and Thalamic Control of Neocortical Activity in the Freely Moving Rat. *Journal of Neuroscience*, **11**, 4007-4026.
- [25] Riekkinen Jr., P., Sirviö, J. and Riekkinen, P. (1990) Relationship between the Cortical Choline Acetyltransferase Content and EEG Delta-Power. *Neuroscience Research*, 8, 12-20. <u>http://dx.doi.org/10.1016/0168-0102(90)90052-g</u>
- [26] Babiloni, C., Visser, P.J., Frisoni, G., DeDeyn, P.P., Bresciani, L., Jelic, V., Nagels, G., Rodriguez, G., Rossini, P.M., Vecchio, F., Colombo, D., Verhey, F., Wahlund, L.O. and Nobili, F. (2010) Cortical Sources of Resting EEG Rhythms in Mild Cognitive Impairment and Subjective Memory Complaint. *Neurobiology of Aging*, **31**, 1787-1798. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2008.09.020</u>
- [27] Moretti, D.V., Pievani, M., Fracassi, C., Geroldi, C., Calabria, M., De Carli, C.S., Rossini, P.M. and Frisoni, G.B. (2008) Brain Vascular Damage of Cholinergic Pathways and EEG Markers in Mild Cognitive Impairment. *Journal of Alzheimer's Disease*, 15, 357-372.
- [28] Van der Hiele, K., Vein, A.A., Reijntjes, R.H., Westendorp, R.G., Bollen, E.L., van Buchem, M.A., van Dijk, J.G. and Middelkoop, H.A. (2007) EEG Correlates in the Spectrum of Cognitive Decline. *Clinical Neurophysiology*, **118**, 1931-1939. <u>http://www.ncbi.nlm.nih.gov/pubmed/17604688</u> http://dx.doi.org/10.1016/j.clinph.2007.05.070
- [29] Grunwald, M., Busse, F., Hensel, A., Kruggel, F., Riedel-Heller, S., Wolf, H., Arendt, T. and Gertz, H.J. (2001) Correlation between Cortical Theta A ctivity and Hippocampal V olumes in Health, Mild Cognitive Impairment, and Mild Dementia. *Journal of Clinical Neurophysiology*, 18, 178-184. <u>http://www.ncbi.nlm.nih.gov/pubmed/11435810</u> <u>http://dx.doi.org/10.1097/00004691-200103000-00010</u>
- [30] Dimpfel, W. and Schober, F. (2001) Norepinephrine, EEG Theta Waves and Sedation. Brain Pharmacol, 1, 89-97.
- [31] Baker, M., Kwaku, A., Schiffer, R. and O'Boyle, M.W. (2008) EEG Patterns in Mild Cognitive Impairment (MCI) Patients. *The Open Neuroimaging Journal*, 2, 52-55. <u>http://www.ncbi.nlm.nih.gov/pubmed/19018315</u> http://dx.doi.org/10.2174/1874440000802010052
- [32] Schellenberg, R., Todorova, A., Dimpfel, W. and Schober, F. (1997) Pathophysiology and Psycho-Pharmacology of Dementia—A New Study Design. I. Diagnosis Comprising Subjective and Objective Criteria. *Neuropsychobiology*, 32, 81-97. <u>http://dx.doi.org/10.1159/000119219</u>
- [33] Grunwald, M., Weiss, T., Krause, W., Beyer, L., Rost, R., Gutberlet, I. and Gertz, H.J. (1999) Power of Theta Waves in the EEG of Human Subjects Increases during Recall of Haptic Information. *Neuroscience Letters*, 260, 189-192. http://dx.doi.org/10.1016/S0304-3940(98)00990-2
- [34] Bastiaanson, M.C., Oostenveld, R., Jensen, O. and Hagoort, P. (2008) I See What You Mean: Theta Power Increases Are Involved in the Retrieval of Lexical Semantic Information. *Brain and Language*, 106, 15-28. <u>http://dx.doi.org/10.1016/j.bandl.2007.10.006</u>
- [35] Klimesch, W. (1999) EEG Alpha and Theta Oscillations Reflect Cognitive and Memory Performance: A Review and Analysis. Brain Research Reviews, 29, 169-195. <u>http://dx.doi.org/10.1016/S0165-0173(98)00056-3</u>
- [36] Deiber, M.P., Ibañez, V., Missonnier, P., Herrmann, F., Fazio-Costa, L., Gold, G. and Giannakopoulos, P. (2009) Abnormal-Induced Theta Activity Supports Early Directed-Attention Network Deficits in Progressive MCI. *Neurobiology* of Aging, 30, 1444-1452. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2007.11.021</u>
- [37] Leyhe, T., Ethofer, T., Bretscher, J., Künle, A., Säuberlich, A.L., Klein, R., Gallwitz, B., Häring, H.U., Fallgatter, A., Klingberg, A., Saur, R. and Müssig, K. (2013) Low Performance in Attention Testing Is Associated with Reduced Grey Matter Density of the Left Inferior Frontal Gyrus in Euthyroid Patients with Hashimoto's Thyroiditis. *Brain, Be-havior, and Immunity*, 27, 33-37. <u>http://dx.doi.org/10.1016/j.bbi.2012.09.007</u>



Lipolysaccharide-Induced Neuroinflammation Is Associated with Alzheimer-Like Amyloidogenic Axonal Pathology and Dendritic Degeneration in Rats

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Abstract

Chronic neuroinflammation is thought to play an etiological role in Alzheimer's disease (AD) which is characterized pathologically by amyloid and tau formation, as well as neuritic dystrophy and synaptic degeneration. The causal relationship between these pathological events is a topic of ongoing research and discussion. Recent data from transgenic AD models point to a tight spatiotemporal link between neuritic and amyloid pathology, with the obligatory enzyme for β -amyloid (A β) production, namely β -secretase-1 (BACE1), being overexpressed in axon terminals undergoing dystrophic change. However, the axonal pathology inherent with BACE1 elevation seen in transgenic AD mice may be secondary to increased soluble $A\beta$ in these genetically modified animals. Further, it is unclear whether the inflammation seen in AD is the result of , or the cause of

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neuritic dystrophy. Here we explored the occurrence of AD-like axonal and dendritic pathology in adult rat brains affected by LPS-induced chronic neuroinflammation. Unilateral intracerebral LPS injection induced prominent inflammatory response in glial cells in the ipsilateral cortex and hippocampal formation. BACE1 protein levels were elevated in the ipsilateral hippocampal lysates in the LPS-treated animals relative to controls. BACE1 immunoreactive dystrophic axons appeared in the LPS-treated ipsilateral cortex and hippocampal formation, colocalizing with increased β -amyloid precursor protein and $A\beta$ antibody (4G8) immunolabeling. Quantitative Golgi studies revealed reduction of dendritic branching points and spine density on cortical layer III and hippocampal CA3 pyramidal neurons in the LPS-treated ipsilateral cerebrum. These findings suggest that Alzheimer-like amyloidogenic axonal pathology and dendritic degeneration occur in wildtype mammalian brain in partnership with neuroinflammation following LPS injection.

Keywords

Amyloid Pathogenesis, Neuritic Dystrophy, Neurodegeneration, Neuroplasticity, Synaptic Pathology

1. Introduction

Neuroinflammation has been linked to many neuropsychiatric disorders, including neurodegenerative diseases such as Alzheimer's diseases (AD), Parkinson's disease (PD), multiple sclerosis and traumatic brain injury [1]-[8]. Aging is a major risk factor for many age-related diseases, and is associated with a certain degree of chronic inflammation [9] [10]. In general, chronic inflammation is considered to mount lasting stress on neurons and synapses, and may lead to brain dysfunction, including cognitive deficits [5] [11]-[14]. The causal relationship between chronic inflammation and some of the hallmark pathological lesions in neurological diseases is under intensive investigation. For example, oxidative or inflammatory stress is suggested to promote cerebral amyloid pathology via i nereased pr oduction and/or i mpaired clearance of $A\beta$, involving b oth n eurons and gl ial c ells [15]-[19]. $A\beta$ products including soluble and aggregated variants may also act as proinflammatory factors [20] [21].

Loss of synapses and their connectivity best correlate with cognitive deficits in AD [22]-[29]. The density of dendritic spines appears to be reduced readily at prodromal stages of the disease [30]. Besides synaptic degeneration, axonal elements including presynaptic terminals undergo aberrant sprouting and dystrophic expansion [30]-[32]. Recent data from transgenic AD models, nonhuman primates and human subjects show that upregulation of the amyloidogenic proteins, especially the rate-limiting enzyme β -secretase-1 (BACE1), appears to be an molecular cascade tightly associated with axonal sprouting and dystrophy, suggestive of a driving role for amyloidogenic axonal pathology in plaque formation [33]-[35].

The bacterial endotoxin lipopolysaccharide (LPS) can induce chronic neuroinflammation in rodents [36]-[38]. LPS administration also causes learning and memory deficits in the animals, providing an excellent model system for studying cognitive dysfunction associated with chronic neuroinflammation [39]-[41]. Neuroinflammation is considered to play an early or inductive role in the development of AD pathologies, although the anatomic evidence remains to be better formulated. Therefore, the present study was set to address whether intracerebral LPS injection in adult rats may induce axonal and dendritic pathologies similar to that seen in AD [32]. Specifically, we aimed to identify whether dystrophic axonal pathology inherent with amyloidogenic modulation and degenerative dendritic/spine changes occur on cerebral principal neurons in LPS-treated adult rats.

2. Materials and Methods

2.1. Animals and Intracerebral Injection

In-house bred male adult Sprague-Dawley rats (n = 24) weighing ~200 g (Animal Center of Central South University) were used. Rats were maintained in temperature (20° C - 26° C), humidity (30% - 60%) and lighting (12:12 hours light/dark cycle) controlled rooms, with food and water freely available. For intracerebral injections, rats were placed in a stereotaxic apparatus under sodium pentobarbital an esthesia (50 mg/kg, i.p.). LPS

from Escherichia coli serotype 055:B5 (L2637, Sigma-Aldrich, St. Louis, MO, USA) was dissolved ($2.5 \mu g/\mu l$) in sterile phosphate-buffered saline (PBS, 0.01 M, pH 7.2) (vehicle) [41]. LPS (10 μg in 4 μl) was injected through the neocortex into the right hippocampus with a microsyringe in each animal (n = 12), using the following coordinates: 3.5 mm lateral to the sagittal cranial suture; 5 mm caudal to the bregma and 3 mm below the dura mater. Controls (n = 12) were injected with the same amount of PBS. Brain examination was carried out 30 days post intracortical injection. The experimental protocol was approved by the Animal Care and Use Committee of the Central South University and were in compliance with NIH guidelines.

2.2. Immunoblot

Hippocampi ipsilateral to the injection side were dissected out following decapitation (n = 4/group). Tissue was weighed and homogenized in a commercial protein extraction buffer at 1 to 10 w/v ratio (CW0883, Kangwei Century Company, Beijing), followed by centrifuge at 15,000 g. Protein concentrations in the supernatants were determined by DC protein assay (Bio-Rad Laboratories, Hercules, CA, USA). A total of 25 µg protein from each sample was separated electrophoretically in 10% SDS-polyacrylamide gel and transferred to polyvinylidene fluoride membrane. Membranes were blocked with 1% non-fat milk and 5% bovine serum albumin (BSA) in 0.1 M Tris-buffered saline containing 0.1% Tween-20 (TBS-T) for 2 hours at room temperature. Membranes were then incubated overnight at 4 °C in the same buffer containing rabbit anti-BACE1 (1:2000) [33] [34] [42] [43], rabbit anti-glial fibrillary acidic protein (GFAP) (G9269, Sigma-Aldrich, St Louis, MO, USA, 1:4000) or rabbit anti- β -tubulin (Sigma-Aldrich, T2200, 1:10000). Membranes were washed thoroughly with TBS-T, then were incubated for 2 h ours at room temperature with horseradish peroxidase-conjugated goat anti-rabbit IgGs (1:20,000; Bio-Rad Laboratories). B ound antibodies were detected by enhanced chemiluminescence (ECL kit, GE Healthcare Life Sciences, Piscataway, NJ, USA). The membranes were exposed to X-ray films developed subsequently in a darkroom. The films were scanned, with optical density (OD) of immunoblot bands measured using Image-J, followed by standardization to the internal references.

2.3. Immunohistochemistry

Rats were perfused transcardially with PBS followed by 4% phosphate-buffered (0.1M) paraformaldehyde (pH 7.4). B rains were r emoved and postfixed for 12 h ours and transferred into 30% sucrose for c ryoprotection. Thirty micrometer-thick coronal sections were cut in a cryostat, with 12 sets of sections collected for cresyl violet stain and for immunohistochemical studies. For immunohistochemistry with the avidin-biotin complex (ABC) method, free-floating sections were soaked in 1% H_2O_2 in PBS for 30 minutes to diminish endogenous peroxidase activity, and preincubated in PBS buffer containing 0.2% Triton X-100 and normal horse serum for 1 hour. The sections were then reacted with one of the following primary antibodies: 1) Mouse monoclonal antibody against major histocompatibility complex class II molecules (MHC-II) at 1:1000 (ab55152, Abcam, Cambridge, MA, USA); 2) Rabbit anti-human BACE1 at 1:2000; 3) mouse anti- β -amyloid precursor protein (APP) monoclonal antibody 22C11 (MAB348, EMD Millipore, Billerica, MA, USA, 1:4000); 4) mouse anti- $A\beta$ 17-24 monoclonal antibody 4G8 (#39240, Signet, Dedham, USA, 1:4000); 5) rabbit anti-GFAP (G9269, Sigma-Aldrich, 1:2000). The sections were further reacted with a biotinylated pan-specific secondary antibody (BA-1300, Vector Laboratories, Burlingame, CA, USA, 1:400) for 2 hours, and subsequently with the ABC reagents (PK-6100, Vector L aboratories, 1: 400) for on e hour. I mmunoreactivity was visualized in 0.05% 3,3'-diaminobenzidine (DAB) and 0.003% H₂O₂.

Selected sections were processed for double immunofluorescence beginning with blocking nonspecific reactivity by incubation in PBS buffer containing 5% normal donkey serum. Sections were further reacted overnight at 4°C with a pair of primary antibodies raised in different species in PBS containing 0.2% Triton X-100 and the blocking s erum. T he a ntibody pa irs included: 1) mouse a nti-MCH-II (1:1000) a nd r abbit a nti-CD11b (MABT149, EMD Millipore, 1:1000); 2) mouse anti-MCH-II and rabbit anti-GFAP (G9269, 1:2000); 3) rabbit anti-BACE1 and mouse anti-APP 22C11; 4) rabbit anti-BACE1 and mouse anti-A β 4G8; 5) rabbit anti-BACE1 and mouse a nti-synaptophysin (MAB329, E MD M illipore, 1: 4000); 6) r abbit a nti-BACE1 a nd mouse a nti-microtubule associated protein-2 (MAP2) (M9942, Sigma-Aldrich, 1:2000). On the second day, the sections were rinsed with PBS and incubated at room temperature for 2 hours with Alexa Fluor® 488 and Alexa Fluor[®] 594 conjugated donkey anti-mouse and anti-rabbit IgGs (1:200, Invitrogen, Carlsbad, CA, USA). Sections were then c ounter-stained with bi sbenzimide (Hoechst 33342, 1:50000, C atalog #B2261, S igma-Aldrich), w ashed thoroughly, and mounted with anti-fading medium before microscopic examination.

2.4. Rapid Golgi-Cox Stain

Brains were removed following a vascular rinse with PBS. Blocks containing the middle 1/3 cerebrum of both hemispheres were rinsed briefly in double distilled water, and processed with the FD Rapid Golgi stain TM Kit (FD Neuro Technologies, Ellicott City, MD) following the manufacturer's instruction. The brain blocks were immersed in freshly made mixture of Solutions A and B (1:1) in the dark at room temperature for 2 weeks, and then in Solution C at 4°C in the dark for 3 days. After silver impregnation, the blocks were cut slowly into frontal sections at 100 μ m thickness in a vibratome. Sections were collected alternatively in 10 sets in Solution C, mounted on gelatin-coated microslides, dehydrated through ascending concentrations of ethanol, cleared in xylene, and sealed with PermountTM mounting medium.

2.5. Imaging and Data Analysis

An Olympus (BX53) microscope equipped with i maging system (CellSens Standard, Olympus) was used for examining sections stained with the ABC and fluorescent methods. Double immunofluorescence was also imaged on a confocal microscope (Nikon, DIGITAL ECLIPSE C1 plus, 5 µm thickness scan). Immunolabeling in sections around the level of injection in the rostrocaudal dimension was comparatively examined between the ipsilateral and contralateral cerebral hemispheres, using the needle track as a reference. Golgi-impregnated sections were examined on a Zeiss Axioplan microscope equipped with the Neurolucida and a high-resolution motorized stage for 3D neuronal reconstruction (MicroBrightField China). Two sections nearest to the injection coordinates (in the rostrocaudal dimension) were selected from each brain for neuronal morphometric analysis. Golgi-stained pyramidal neurons that met the following criteria were selected for reconstruction: 1) they were located in layer III of the parietal cortex overlying the mid-hippocampus and in the middle portion of CA3 (*i.e.*, around the dorsal to ventral turning area); 2) they were among the labeled cells with the widest dendritic field by overall visual judgment; 3) they were well separated from other impregnated cells such that their dendritic tree was not or minimally overlapped with the processes from other cells; 4) the somata and dendritic processes were well-impregnated throughout the section thickness, with no apparent truncation of the dendritic arbor. Ten pyramidal neurons per region/brain were reconstructed with the aid of the Neurolucida software. Subsequently, somal area, total length of the dendritic tree, branching nodes and spine density (per 10 µm length) of the apical and basal dendrites, were obtained from a given selected neuron.

2.6. Statistical Analysis

Imaging and numerical data for comparing groups were processed, with the mean \pm SD calculated. Means were statistically a nalyzed by S tudent-*t* test or one-way A NOVA with posthoc Duncan's multi-group comparisons when applicable. P < 0.05 was considered statistically significant. Figures were assembled with Photoshop 7.1, with brightness and contract adjusted as needed.

3. Results

3.1. LPS Injection Induced Immunoinflammatory Cellular/Molecular Changes

To confirm the occurrence of chronic neuroinflammation, cerebral sections from the LPS and PBS groups were processed under identical conditions for the detection of immunoinflammatory proteins. Compared to PBS controls, increased immunoreactivity for MCH-II (Figures 1(A)-(E)), CD11b (not shown) and GFAP (not shown) occurred in the ipsilateral cortex and hippocampal formation in the LPS-injected brain sections. Specifically, MCH-II immunoreactive cells appeared to be largely glial cells (Figure 1(D) and Figure 1(E)). Double immunofluorescence showed that the majority of MCH-II labeled cells co-expressed the microglial marker CD11b (Figures 1(F)-(H)), although a few also colocalized with GFAP immunoreactivity, suggestive of a colocalization in astrocytes (Figures 1(I)-(K)).

3.2. LPS Injection Elevated BACE1 Protein Levels

BACE1 protein levels were immunoblotted with a well-characterized r abbit an tibody, which d etects mature BACE1 protein migrating at ~70 kd [33] [34] [42] [43]. In the lysates of the ipsilateral hippocampi, immunob-

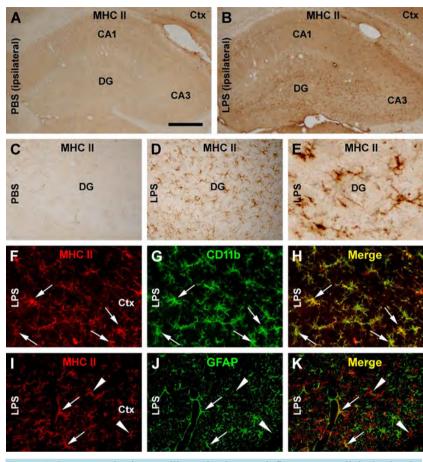


Figure 1. Representative images illustrating immunoinflammatory cellular/molecular changes in r at c ortex and hippocampal formation following i ntracerebral lipolysaccharide (LPS) injection. Panel (A) shows minimal immunoreactivity of major histocompatibility complex class II molecules (MHC II) in the hippocampal CA1 to CA3 areas, the dentate gyrus (DG) and the overlaying cortex (Ctx) in a P BS-injected control rat, with the area of the dentate gyrus enlarged as (C). MHC II immunolabeling is increased in both the cortex and hippocampal formation in the LPS-injected an imal (B), with the labeled c ellular profiles appeared as glial cells at high magnifications ((D), (E)). Confocal double immunofluorescence shows a great extent of colocalization of MHC II reactivity a mong CD11b labeled microglial cells in the LPS-treated cerebrum ((F)-(H)). A small amount of MHC II labeled c ells exhibit glial fibrillary acidic p rotein (GFAP) immunoreactivity, suggestive of a colocalization in activated astrocytes ((I)-(K)). Scale bar = 500 μ m in A applying to B; equivalent to 250 μ m for ((C), (D)); and to 75 μ m for ((E)-(K)).

lotted BACE1 signal was significantly increased relative to PBS-treated counterparts (Figure 2(A)). The mean optic density of BACE1 in the LPS group increased one fold (199.0% \pm 52.8%) relative to control (100.0% \pm 12.2%) (P = 0.043, t = 3.40, df = 3; two-tailed paired t-test) (Figure 2(A) and Figure 2(B)). Serving as an experimental as well as assay control [36]-[38], GFAP levels in the same cerebral samples were checked. In the ipsilateral hippocampal lysates, levels of GFAP were significantly (P = 0.027, t = 4.03, df = 3; two-tailed paired t-test) elevated in the LPS-treated (141.7% \pm 11.4%) relative to PBS-treated (100% \pm 13.4%) samples (Figure 2(A) and Figure 2(C)).

3.3. LPS Injection Induced Axonal Pathology with Enhanced Amyloidogenic Protein Expression

In normal mammalian brains BACE1 immunoreactivity is largely expressed in the neuropil in a diffuse pattern

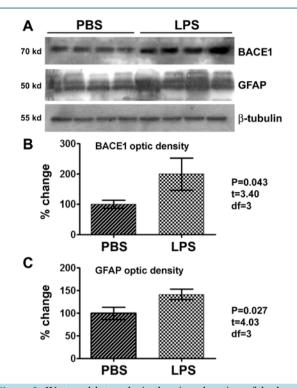
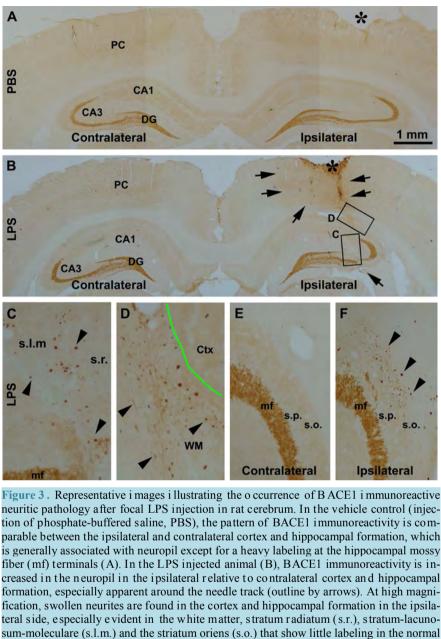


Figure 2. Western blot analysis showing elevation of the levels of β -secretase-1 (BACE1) and GFAP in the LPS-injected relative to PBS-treated (control) brains (n = 4/group), as as sayed using h ippocampal 1 ysates ip silateral to the injection. B ACE1 levels in the LPS-treated group are about 2 times of that in the control ((A), (B)), while G FAP levels in the L PS g roup a pproach to 1.5 times of the control ((A), (C)), with the changes being significant for both proteins (paired two-tail t-tests).

except for a distinct heavy labeling at selected neuronal terminal fields including the mossy fiber terminals and olfactory glomeruli. In human and transgenic animal brains with amyloid plaque pathology, BACE1 immunoreactivity is increased preferentially in swollen and sprouting a xonal terminals or a xonal dystrophic neurites [33]-[35] [42]. In the present study, BACE1 labeling in both cerebral hemispheres in the PBS injected brains and the contralateral cerebrum in the LPS-injected animals exhibited the aforementioned normal distribution pattern (Figure 3(A) and Figure 3(B)). On the contrary, BACE1 labeled neuritic structures emerged in the ipsilateral hemisphere of the LPS-treated brains in a site-specific manner. Thus, increased BACE labeling appeared in both the cortex and hippocampal formation especially evident around the needle track. At high magnification, these labeled profiles appeared as swollen sphericles and neuritic processes (Figures 3(B)-(F)). In the white matter, these neuritic processes appeared to spread from the needle track for a considerable distance (Figure 3(B) and Figure 3(D)). In the ipsilateral hippocampus, the BACE1 labeled swollen neurites occurred in the stratum pyramidale (s.p.) to stratum-lacunosum-moleculare (s.l.m.) of CA3 and the adjacent CA1 area (Figure 3(C)). A large amount of swollen sphericles extended along the stratum oriens (s.o.) of CA3 in the LPS-injected animals (Figure 3(E) and Figure 3(F)). At high magnifications, these BACE1 labeled neurites were mostly round or oval in shape, and varied in size. Some smaller sphericles were sometimes arranged in chains resembling enlarged axonal varicosities (Figure 3(C), Figure 3(D) and Figure 3(F)).

In double immunofluorescence, the BACE1 immunoreactive swollen neurites exhibited a great extent of colocalization with APP (**Figures 4(A)-(H)**). In addition, the $A\beta$ antibody 4G8 (which detects mouse APP, $A\beta$ and likely APP β -C-terminal fragments, ref. 34) showed increased immunoreactivity in the same area with BACE1 labeled neurites (**Figures 4(I)-(K)**). A colocalization of BACE1 and 4G8 labeling was detected among some individual neuritic profiles. Notably, there existed a considerable amount of diffuse extracellular $A\beta$ labeling (not colocalized with BACE1 r eactivity), some of which ap peared p unctate (**Figure 4(L)**). The BACE1 labeled



formation, especially apparent around the needle track (outline by arrows). At high magnification, swollen neurites are found in the cortex and hippocampal formation in the ipsilateral side, especially evident in the white matter, stratum radiatum (s.r.), stratum-lacunosum-moleculare (s.l.m.) and the striatum oriens (s.o.) that show little labeling in the normal conditions ((C), (D), (F)). Panels ((E), (F)) are taken from the section that is ~480 µm (12 × 30 µm) apart from (B), showing the distinct difference regarding the neuritic pathology between the two sides. The needle entry at the cortical surface is marked with an asterisk (*). Ctx: cortex (grey matter); DG: dentate gyrus; PC: parietal cortex; s.l.m.: stratum-lacunosum-moleculare; s.p.: stratum pyramidale. WM: white matter; Scale bar = 1 mm in (A) applying to (B); equivalent for 400 µm for ((C), (D)) and 200 µm for (E).

sphericles and swollen processes were partially colocalized with the presynaptic marker synaptophysin (Figures 4(M)-(P)). In contrast, the BACE1 labeled swollen neurites did not colocalize with MAP2, which is clearly expressed in dendrites and somata of nearby pyramidal neurons (Figures 4(R)-(U)).

3.4. LPS Injection Induced Somotodendritic Changes in Cerebral Principal Neurons

In our Golgi preparations, pyramidal neurons around layer III were consistently impregnated in all brain samples (Figure 5(A)). In the hippocampal proper, the curving area of CA3, whereby the s.p. continues dorsally from the

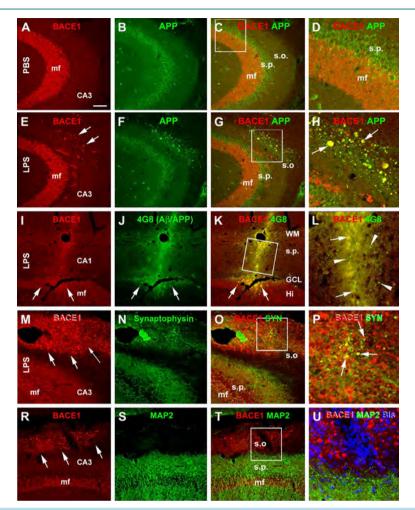


Figure 4. Confocal double immunofluorescent images showing LPS induced axonal pathology as sociated with increased labeling of amyloidogenic proteins. All images are taken from the CA3 area of the hippocampus ipsilateral to the intracerebral injection of PBS ((A)-(C)) or LPS ((E)-(U)). Antibody markers and color channels are as indicated. Panels ((A)-(C)) show double labeling of BACE1 and β -amyloid precursor protein (APP) in ipsilateral CA3 of the control animal, with the former expressed predominantly in the mossy fiber (mf) terminals, and the latter largely in somata of CA3 pyramidal neurons. Note that no abnormal neurites are present in the strum or iens (s.o.). Panels ((E)-(H)) illustrate the occurrence of BACE1/APP double labeled swollen neurites (examples are indicated by arrows) in the s.o. of the ipsilateral hippocampus of the LPS injected rat. Panels (I-L) show that these BACE1 positive neurites are locally associated with increased 4G8 labeling within (pointed by arrows) as well as outside (arrowheads) the swollen terminals in the cortex and CA1. Panels ((M)-(P)) show that a partial coexpression of synaptophysin (SYN) among the BACE1 labeled swollen neurites in the s.o. of the LPS injected ipsilateral hippocampus, which appear in yellow in the merged image (arrows, P). Panels (R-U) show that there is no colocalization of the microtubule associated protein-2 (MAP2) in the BACE1 labeled swollen neurites. MAP2 labeling is distinctly associated with the somata and dendrites of pyramidal neurons in the stratum pyramidale (s.p.). DAPI counterstain is included in panels (L) and (U), showing that the BACE1 labeled elements are not somatic. Scale bar = 200 µm in (A) applying to ((B), (C), (E)-(G) and (R)-(T)), equivalent to 100 µm for (I)-(K), (M)-(O)), 50 µm for ((D), (H), (U)) and 25 µm for ((L), (P)).

CA1 direction and t urns ventrally t oward t he d entate g yrus, could be d efined s ystematically across b rains. Therefore, we decided to use Golgi-stained pyramidal neurons in cortical layer III and around the dorsoventral turning area of CA3 for automated Neurolucida reconstruction and morphometry. Measurements from reconstructed layer III and CA1 pyramidal neurons in the ipsilateral hemisphere of the LPS and PBS treated brains

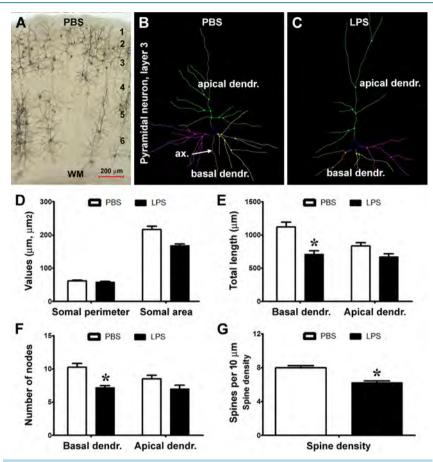


Figure 5. Quantitative Golgi study of somal and dendritic changes in cortical layer III pyramidal ne urons in L PS-injected r elative t o PBS-injected r ats. Panel (A) s hows an example of Golgi stain in the parietal neocortex (ipsilateral to PBS injection), note that layer III pyramidal neurons are relatively well-labeled. Panels ((B), (C)) show representative N eurolucida r econstructions of i mpregnated layer I II pyramidal ne urons from a LPS (B) and a PBS (C) injected brains, with their apical and basal dendrites and dendritic branching points illustrated in color. Bar graphs ((D)-(G)) summarize the parameters obtained from 10 c onstructed neurons in each brain (n = 4/group). The means of somal perimeter and somal area show a trend of reduction in the LPS relative to PBS (control) groups, although the difference is n ot s tatistically significant (E). The m eans of to tal dendritic length tends to reduce in the LPS group relative to control, with significant difference for that of the basal dendritic tree (G). Overall spine density is significantly reduced on t he dendritic tree of G olgi-impregnated c ortical pyramidal ne urons i n the LPS group relative to control (H). Scale bar = 200 μ m in (A).

(10 cells per region per brain) were compared quantitatively (Figures 5(A)-(C) and Figures 6(A)-(D)).

The somal perimeters of the layer III pyramidal neurons were comparable between the LPS ($58.6 \pm 7.4 \mu m$, mean \pm S.D., same format below) and PBS ($61.7 \pm 9.9 \mu m$) groups (P > 0.05, two-tail paired t-test; same statistical test below) (Figure 5(D)). The mean somal area of layer III pyramidal neurons was reduced in the LPS ($168.4 \pm 21.6 \mu m^2$) relative to the PBS ($216.5 \pm 45.3 \mu m^2$) groups (P < 0.001). The total length of the dendritic processes of the layer III pyramidal neurons tended to be reduced in the LPS relative to PBS groups (Figure 5(E)). Thus, the total length of the apical dendrites was $647.4 \pm 182.9 \mu m$ in the LPS group as compared to $834.9 \pm 228.8 \mu m$ in the PBS group, with the means not statistically different (P > 0.05). The means of the total length of the basal dendrites were $711.4 \pm 226.9 \mu m$ and $1122.0 \pm 320.7 \mu m$ in the LPS and PBS groups, respectively, showing a significant difference (P < 0.001). The means of the branching points (nodes) on the apical dendrites were 7.0 ± 2.5 and 8.5 ± 2.4 in the LPS and the PBS groups, respectively, while they were 7.2 ± 1.3

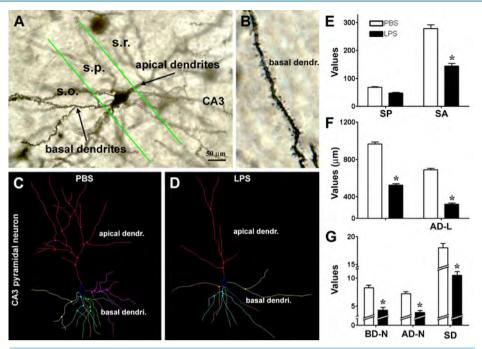


Figure 6. Quantitative Golgi study of somal and dendritic changes in in CA3 pyramidal neurons in LPS-injected relative to PBS-injected rat brains. Panel (A) shows an example of Golgi-stained CA3 pyramidal neurons located around dorsal to ventral turning area of the stratum pyramidale (s.r.). B asal dendrites in the stratum oriens (s.o.) and apical dendrites in the stratum radiatum (s.r.) are visible, with dendritic spines clearly labeled at high resolution (B). Panels ((C), (D)) show examples of Neurolucida reconstructions of CA3 pyramidal neurons from LPS (C) and a PBS (D) injected brains. The reduction in mean somal area (SA) in the LPS relative to PBS groups is statistically significant, whereas the reduction in the somal perimeter (SP) is not (E). The reduction in the total dendritic length in the LPS relative to PBS groups is significant for both the basal and apical trees (F). Scale bar = 50μ m in (A) and 5μ m for (B).

and 10.6 ± 2.7 for the two groups on the basal dendrites. Statistical tests reported a significant difference between the two groups for the means of the basal dendritic but not the apical measurements (Figure 5(F)). The number of spines per unit length of dendrite was reduced significantly on the layer III pyramidal neurons in the LPS relative to PBS groups (Figure 5(G)). Data from the apical and basal dendrites combined, showed spine density to be 6.3 ± 0.9 vs. 8.0 ± 1.1 for the LPS relative to PBS groups (P < 0.0001).

For the reconstructed CA3 pyramidal neurons (**Figures 6(A)-(D)**), both the somal perimeter and somal area of the pyramidal neurons tended to reduce in the LPS relative to PBS groups (**Figure 6(E)**). Thus, the mean somal perimeters were $47.8 \pm 11.0 \ \mu\text{m}$ (mean \pm S.D., same format below) and $67.8 \pm 12.3 \ \mu\text{m}$ in the LPS and PBS treated animals, respectively (P > 0.05, two-tail paired t-test, same statistical test below). The reduction in the somal area of CA3 pyramidal neurons was significant (P < 0.001) in the LPS group ($144.8 \pm 40.4 \ \mu\text{m}^2$) compared to the PBS group ($278.3 \pm 60.2 \ \mu\text{m}^2$). There were significant decreases in the total lengths of the apical dendrites ($320.0 \pm 52.6 \ \text{vs}$. $690.0 \pm 63.6 \ \mu\text{m}$; P < 0.005) and the basal dendrites ($534.6 \pm 70.6 \ \text{vs}$. $967.0 \pm 90.6 \ \mu\text{m}$; P < 0.001) in the LPS compared to PBS groups (**Figure 6(F)**). The number of branching points on the apical adendrites ($4.3 \pm 2.1 \ \text{vs}$. 8.3 ± 2.1 ; P < 0.001) and basal dendrites ($3.9 \pm 1.5 \ \text{vs}$ 7.2 ± 1.7 ; P < 0.001) of CA1 pyramidal neurons was significantly reduced in the LPS relative to PBS groups (**Figure 6(G)**). Finally, the density of spines on the apical and basal dendrites of the CA3 pyramidal neurons was significantly reduced (P < 0.001) in the LPS animals (10.5 ± 3.2) relative to PBS controls (18.0 ± 3.3) (**Figure 6(H)**).

4. Discussion

4.1. The LPS Model for Studying Chronic Neurodegeneration and Alzheimer's Disease

The LPS model is increasingly used in mechanistic and translational studies into chronic neurodegenerative dis

eases such as AD and PD [15] [16] [38] [44]-[50]. This relatively simple experimental approach offers yet a number of advantages in elucidating the neurobiology of aging and age-relative diseases. This model recapitulates many cellular/molecular events of neuroinflammation, which is considered an early, perhaps fundamental, causal factor in chronic neuronal and synaptic degeneration [8]-[14] [36]-[41]. The model also reproduces some defining clinical manifestations of human neurodegenerative diseases in rodents [44]-[46]. For instance, peripheral and central LPS administration induces learning/memory deficits in experimental animals analogous to AD-like co gnitive d ecline [39]-[41]. Importantly, LPS t reatment el icits n europathological and b ehavioral changes in *wild-type* animals [47]-[50]. As neurodegenerative diseases occur mostly in a sporadic nature, the LPS model serves an ideal system to address pathogenic mechanisms underlying common human neurological disorders. Specifically, this s tudy e xamined whether inflammation is a consequence or cau se of d ystrophic amyloidogenic neuritis.

We have shown in a recent study that unilateral intracerebral injection of ~10 μ g LPS is sufficient to induce spatial le arning a nd memory i mpairments i n a dult r ats. U pregulation of i mmunoinflammatory molecules i ncluding PirB and GFAP is evident in the LPS-treated brains, occurring largely in the ipsilateral cortex and hippocampal formation [41]. In the present study we observe that this same dose of LPS injection dramatically induces MHC II expression in the ipsilateral neocortex and hippocampal formation. This modulation appears to be largely associated with microglial activation, a key finding in the initial studies that establish this model of neurological diseases [36] [39]. Consistent with other reports [51], LPS-induced MHC II upregulation also appears to occur to a lesser extent in activated astrocytes that express high levels of GFAP. Thus, our data are consistent with the n otion th at in tracranial LPS injection i nduces profound g lial i mmunoinflammatory r esponses i n the brain.

4.2. Axonal and Dendritic Pathology in LPS-Induced Neuroinflammation

Besides glial responses, neuronal and synaptic alterations have been reported in rodent LPS models anatomically and electrophysiologically. Local LPS infusion causes increased cell/neuronal death in the entorhinal cortex [52] and substantia nigra [53] in rats as well as the hippocampus in ICR mice [54]. In BALB/c mice, the length and branching points of apical dendrites of CA1 pyramidal neurons are reduced in old but not young adults 72 hours following intraperitoneal LPS injection, while no change in spine density is found on these neurons [55]. In C57BL/6 mice, an increase in the density of thin spines on the dendrites of adult-born granule cells is found in the inner but not outer molecular layer 28 days following intrahippocampal LPS injection [56]. In adult rats, intrahippocampal LPS injection does not alter the intrinsic membrane properties, dendritic arborization or spine formation of the newly generated granule cells, but affect the overall network activity in the hippocampal neural circuitries [57]. LPS treatment can alter hippocampal synaptoplasticity in rats as measured by long-term potential and long-term depression [52] [58]-[60].

Here we demonstrate pre- and post-synaptic alterations among cortical and hippocampal neurons in LPS relative to PBS injected rats. Microscopically evident axonal pathology is featured by aberrant sprouting and swelling, and increased expression of APP, BACE1 and synaptophysin, but not MAP2. These abnormal axonal profiles seen following LPS treatment resemble the dystrophic neurites seen in models of injury that exhibit an increase in APP or A-beta [61] [62], although they do not arrange in typical rosette-like clusters as seen around established compact plaques [32]. Our Golgi data indicate that focal LPS injection causes a significant decrease in dendritic length and nodes on the basal tree as well as a reduction of spine density on the entire dendritic tree of layer III cortical principal neurons. The impact of focally injected LPS appears to be greater on CA3 pyramidale neurons, including significant reductions of somal size, lengths of basal and apical dendrites, branching points of the basal and apical dendrites, and overall spine density. The vulnerability of CA3 pyramidale neurons to LPS toxicity might relate to an intrinsic property of these cells in regard to synaptoplasticity. The dendrites and spines of CA3 pyramidal neurons belong to one of most plastic network systems in the brain-they receive major presynaptic inputs from the mossy fiber terminals that are constantly renewed in the process of adult neurogenesis of the dentate granule cells [63]. Alternatively, the observed variable somotodendritic changes between the cortical and hippocampal pyramidal neurons may reflect a potential spatial or dose-related effect by the injected toxin (*i.e.*, the proximity to the injection site), which can be expected since the inflammation and BACE1 upregulation are not evident in the contralateral hemisphere.

4.3. Amyloid Pathogenesis in LPS-Induced Neuroinflammation

LPS a dministration h as b een s hown t o i nfluence a myloid p athogenesis vi a mixed c ellular modulations i n wild-type and transgenic rodents. Chronic intraventricular LPS infusion in rats induces intraneuronal A β immunoreactivity in the hippocampus [37]. Repeated intraperitoneal LPS injections in ICR mice result in increased intraneuronal and extracellular A β 42 immunoreactivity in the cortex and hippocampus, with biochemical data indicating reduced α -secretase activity, increased γ -secretase activity, and elevated BACE1 protein and activity in tissue lysate [64]. In transgenic models of AD, LPS administration is shown to increase APP and A β immunoreactivity in ne uronal s omata [65], el evate γ -secretase activity [66] and accelerate p laque d eposition [67]. However, other reports show reduced amyloid deposition via enhanced A β clearance by activated microglia after LPS administration [68] [69].

Being the obligatory enzyme for $A\beta$ genesis, BACE1 is at the crossing point potentially linking cellular and signaling substrates to amyloid pathogenesis. BACE1 elevation is associated with axonal pathology during plaque development in the brains of transgenic AD models as well as aged and AD human subjects [32] [70]. As elaborated in the preceding section, we have identified axonal sprouting and swelling associated with enhanced APP, BACE1 and $A\beta$ antibody reactivity in the ipsilateral cortex and hippocampal formation in LPS treated rat brains, especially evident around the needle track. This finding clearly indicates that chronic neuroinflammation can act as an inductive factor for amyloidogenic axonal pathology in wild-type adult mammalian brain.

More basic questions can be asked as to why axonal sprouting/dystrophy, dendritic/spine degeneration and glial activation develop in partnership under neuroinflammatory conditions. One possibility is that the toxin interrupts axonal transport causing accumulation of the amyloidogenic proteins and other neuronal molecules [31]. This is consistent with the observation that the presynaptic marker synaptophysin is present only among a subset of the BACE1 immunoreactive swollen neurites. One may also speculate that this pathogenic correlation potentially involves the triadic organization of the synapse—presynaptic and postsynaptic terminals engulfed by glial processes. Neuroinflammation could initially induce some disengagement between the synaptic terminals and glial scaffold. For instance, inflammation-associated structural and functional changes in the glial cells may disrupt the fitness, integrity and plasticity of the pre- and post-synaptic terminals. This may eventually lead to major degenerative changes in the postsynaptic components, resulting in dendritic shrinkage and spine loss. The presynaptic terminals might otherwise undergo an aggressive regenerative attempt because of the loss of appropriate postsynaptic targets. In this sense, the aberrant sprouting and dystrophy of the axonal terminals may be considered as a form of maladaptive neuronal regeneration, with the activated amyloidogenic machinery being a part of the its complex molecular dyshomeostasis.

In summary, the present study demonstrates that chronic neuroinflammation induced by intracerebral LPS injection promotes the amyloidogenic cascade in dystrophic neurites by upregulation of BACE1 and APP in the brain. This modulation is evident in axon terminals that exhibit dystrophic-like morphology. LPS injection also induces d egenerative c hanges i n p ostsynaptic c omponents manifested a s d endritic s hrinkage a nd s pine l oss among cortical and hippocampal pyramidal neurons. Together with the immunoinflammatory responses of the glial cells, the LPS model recapitulates multiple cellular and molecular deficits seen in aging and AD brains. The synaptic triad could be the initial site as well as center of neural degeneration and aberrant regeneration, with its progression ultimately leading to overt neuritic amyloid pathology and cognitive deficits.

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References

- [1] McGeer, E.G. and McGeer, P.L. (2010) Neuroinflammation in Alzheimer's Disease and Mild Cognitive Impairment: A Field in Its Infancy. *Journal of Alzheimer's Disease*, **19**, 355-361.
- [2] Qian, L., Flood, P.M. and Hong, J.S. (2010) Neuroinflammation is a Key Player in Parkinson's Disease and a Prime

Target for Therapy. Journal of Neural Transmission, 117, 971-979. http://dx.doi.org/10.1007/s00702-010-0428-1

- [3] Broussard, G.J., Mytar, J., Li, R.C. and Klapstein, G.J. (2012) The Role of Inflammatory Processes in Alzheimer's Disease. *Inflammopharmacology*, **20**, 109-126.
- [4] Luessi, F., Siffrin, V. and Zipp, F. (2012) Neurodegeneration in Multiple Sclerosis: Novel Treatment Strategies. *Expert Review of Neurotherapeutics*, **12**, 1061-1076. <u>http://dx.doi.org/10.1586/ern.12.59</u>
- [5] Rao, J.S., Kellom, M., Kim, H.W., Rapoport, S.I. and Reese, E.A. (2012) Neuroinflammation and Synaptic Loss. *Neurochemical Research*, 37, 903-910. <u>http://dx.doi.org/10.1007/s11064-012-0708-2</u>
- [6] Blandini, F. (2013) Neural and Immune Mechanisms in the Pathogenesis of Parkinson's Disease. Journal of NeuroImmune Pharmacology, 8, 189-201. <u>http://dx.doi.org/10.1007/s11481-013-9435-y</u>
- [7] Breunig, J.J., Guillot-Sestier, M.V. and Town, T. (2013) Brain Injury, Neuroinflammation and Alzheimer's Disease. Frontiers in Aging Neuroscience, 11, 26.
- [8] Enciu, A.M. and Popescu, B.O. (2013) Is There a Causal Link between Inflammation and Dementia? *BioMed Research International*, 2013, 316495. <u>http://dx.doi.org/10.1155/2013/316495</u>
- [9] Ownby, R.L. (2010) Neuroinflammation and Cognitive Aging. Current Psychiatry Reports, 12, 39-45. http://dx.doi.org/10.1007/s11920-009-0082-1
- [10] Pizza, V., Agresta, A., D'Acunto, C.W., Festa, M. and Capasso, A. (2011) Neuroinflammation and Ageing: Current Theories and an Overview of the Data. *Reviews on Recent Clinical Trials*, 6, 189-203. http://dx.doi.org/10.2174/157488711796575577
- [11] Cagnin, A., Brooks, D.J., Kennedy, A.M., Gunn, R.N., Myers, R. and Turkheimer, F.E. (2001) *In-Vivo* Measurement of Activated Microglia in Dementia. *Lancet*, 358, 461-467. <u>http://dx.doi.org/10.1016/S0140-6736(01)05625-2</u>
- [12] Butcher, S.K. and Lord, J.M. (2004) Stress Responses and Innate Immunity: Aging as a Contributory Factor. Aging Cell, 3, 151-160. <u>http://dx.doi.org/10.1111/j.1474-9728.2004.00103.x</u>
- [13] Eikelenboom, P., van Exel, E., Hoozemans, J.J., Veerhuis, R., Rozemuller, A.J. and van Gool, W.A. (2010) Neuroinflammation—An Early Event in Both the History and Pathogenesis of Alzheimer's Disease. *Neurodegenerative Diseases*, 7, 38-41. <u>http://dx.doi.org/10.1159/000283480</u>
- [14] Wang, X., Wang, W., Li, L., Perry, G., Lee, H.G. and Zhu, X. (2013) Oxidative Stress and Mitochondrial Dysfunction in Alzheimer's Disease. *Biochimica et Biophysica Acta*, S0925-4439(13)00323-2.
- [15] Katafuchi, T., Ifuku, M., Mawatari, S., Noda, M., Miake, K., Sugiyama, M. and Fujino, T. (2012) Effects of Plasmalogens on Systemic Lipopolysaccharide-Induced Glial Activation and β-Amyloid Accumulation in Adult Mice. Annals of the New York Academy of Sciences, 1262, 85-92. http://dx.doi.org/10.1111/j.1749-6632.2012.06641.x
- [16] Krstic, D., Madhusudan, A., Doehner, J., Vogel, P., Notter, T., Imhof, C., Manalastas, A., Hilfiker, M., Pfister, S., Schwerdel, C., Riether, C., Meyer, U. and Knuesel, I. (2012) Systemic Immune Challenges Trigger and Drive Alzheimer-Like Neuropathology in Mice. *Journal of Neuroinflammation*, 9, 151. <u>http://dx.doi.org/10.1186/1742-2094-9-151</u>
- [17] Kraft, A.W., Hu, X., Yoon, H., Yan, P., Xiao, Q., Wan, Y., Gil, S.C., Brown, J., Wilhelmsson, U., Restivo, J.L., Cirrito, J.R., Holtzman, D.M., Kim, J., Pekny, M. and Lee, J.M. (2013) Attenuating Astrocyte Activation Accelerates Plaque Pathogenesis in APP/PS1 Mice. *FASEB Journal*, 27, 187-198. <u>http://dx.doi.org/10.1096/fj.12-208660</u>
- [18] Medeiros, R. and LaFerla, F.M. (2013) Astrocytes: Conductors of the Alzheimer Disease Neuroinflammatory Symphony. *Experimental Neurology*, 239, 133-138. <u>http://dx.doi.org/10.1016/j.expneurol.2012.10.007</u>
- [19] Zhao, J., O'Connor, T. and Vassar, R. (2011) The Contribution of Activated Astrocytes to Aβ Production: Implications for Alzheimer's Disease Pathogenesis. *Journal of Neuroinflammation*, 8, 150. http://dx.doi.org/10.1186/1742-2094-8-150
- [20] Sondag, C.M., Dhawan, G. and Combs, C.K. (2009) Beta Amyloid Oligomers and Fibrils Stimulate Differential Activation of Primary Microglia. *Journal of Neuroinflammation*, 6, 1. <u>http://dx.doi.org/10.1186/1742-2094-6-1</u>
- [21] Maezawa, I., Zimin, P.I., Wulff, H. and Jin, L.W. (2011) Amyloid-Beta Protein Oligomer at Low Nanomolar Concentrations Activates Microglia and Induces Microglial Neurotoxicity. *Journal of Biological Chemistry*, 286, 3693-3706. <u>http://dx.doi.org/10.1074/jbc.M110.135244</u>
- [22] DeKosky, S.T. and Scheff, S.W. (1990) Synapse Loss in Frontal Cortex Biopsies in Alzheimer's Disease: Correlation with Cognitive Severity. *Annals of Neurology*, 27, 457-464. <u>http://dx.doi.org/10.1002/ana.410270502</u>
- [23] Terry, R.D., Masliah, E., Salmon, D.P., Butters, N., Deteresa, R., Hill, R., Hansen, L.A. and Katzman, R. (1991) Physical Basis of Cognitive Alterations in Alzheimer's Disease: Synapse Loss Is the Major Correlate of Cognitive Impairment. *Annals of Neurology*, **30**, 572-580. <u>http://dx.doi.org/10.1002/ana.410300410</u>
- [24] Masliah, E., Mallory, M., Alford, M., DeTeresa, R., Hansen, L.A., McKeel Jr., D.W. and Morris, J.C. (2001) Altered Expression of Synaptic Proteins Occurs Early during Progression of Alzheimer's Disease. *Neurology*, 56, 127-129. <u>http://dx.doi.org/10.1212/WNL.56.1.127</u>

- [25] Heinonen, O., Soininen, H., Sorvari, H., Kosunen, O., Paljarvi, L., Koivisto, E. and Riekkinen, P.J. (1995) Loss of Synaptophysin-Like Immunoreactivity in the Hippocampal Formation Is an Early Phenomenon in Alzheimer's Disease. *Neuroscience*, 64, 375-384. <u>http://dx.doi.org/10.1016/0306-4522(94)00422-2</u>
- [26] Reddy, P.H., Mani, G., Park, B.S., Jacques, J., Murdoch, G., Whetsell Jr., W., Kaye, J. and Manczak, M. (2005) Differential Loss of Synaptic Proteins in Alzheimer's Disease: Implications for Synaptic Dysfunction. *Journal of Alzheimer's Disease*, 7, 103-117.
- [27] Scheff, S.W., Price, D.A., Schmitt, F.A., Scheff, M.A. and Mufson, E.J. (2011) Synaptic Loss in the Inferior Temporal Gyrus in Mild Cognitive Impairment and Alzheimer's Disease. *Journal of Alzheimer's Disease*, **24**, 547-557.
- [28] D'Amelio, M. and Rossini, P.M. (2012) Brain Excitability and Connectivity of Neuronal Assemblies in Alzheimer's Disease: From Animal Models to Human Findings. *Progress in Neurobiology*, 99, 42-60. http://dx.doi.org/10.1016/j.pneurobio.2012.07.001
- [29] Berchtold, N.C., Coleman, P.D., Cribbs, D.H., Rogers, J., Gillen, D.L. and Cotman, C.W. (2013) Synaptic Genes Are Extensively Downregulated across Multiple Brain Regions in Normal Human Aging and Alzheimer's Disease. *Neurobiology of Aging*, 34, 1653-1661. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2012.11.024</u>
- [30] Knobloch, M. and Mansuy, I.M. (2008) Dendritic Spine Loss and Synaptic Alterations in Alzheimer's Disease. *Molecular Neurobiology*, 37, 73-82. <u>http://dx.doi.org/10.1007/s12035-008-8018-z</u>
- [31] Kanaan, N.M., Pigino, G.F., Brady, S.T., Lazarov, O., Binder, L.I. and Morfini, G.A. (2013) Axonal Degeneration in Alzheimer's Disease: When Signaling Abnormalities Meet the Axonal Transport System. *Experimental Neurology*, 246, 44-53. <u>http://dx.doi.org/10.1016/j.expneurol.2012.06.003</u>
- [32] Yan, X.X., Ma, C., Gai, W.P., Cai, H. and Luo, X.G. (2014) Can BACE1 Inhibition Mitigate Early Axonal Pathology in Neurological Diseases? *Journal of Alzheimer's Disease*, **38**, 705-718.
- [33] Zhang, X.M., Cai, Y., Xiong, K., Cai, H., Luo, X.G., Feng, J.C., Clough, R.W., Struble, R.G., Patrylo, P.R. and Yan, X.X. (2009) β-Secretase-1 Elevation in Transgenic Mouse Models of Alzheimer's Disease Is Associated with Synaptic/ Axonal Pathology and Amyloidogenesis: Implications for Neuritic Plaque Development. *European Journal of Neuroscience*, **30**, 2271-2283. <u>http://dx.doi.org/10.1111/j.1460-9568.2009.07017.x</u>
- [34] Cai, Y., Zhang, X.M., Macklin, L.N., Cai, H., Luo, X.G., Oddo, S., Laferla, F.M., Struble, R.G., Rose, G.M., Patrylo, P.R. and Yan, X.X. (2012) BACE1 Elevation Is Involved in Amyloid Plaque Development in the Triple Transgenic Model of Alzheimer's Disease: Differential Aβ Antibody Labeling of Early-Onset Axon Terminal Pathology. *Neurotoxicity Research*, 21, 160-174. <u>http://dx.doi.org/10.1007/s12640-011-9256-9</u>
- [35] Li, J.M., Xue, Z.Q., Deng, S.H., Luo, X.G., Patrylo, P.R., Rose, G.W., Cai, H., Cai, Y. and Yan, X.X. (2013) Amyloid Plaque Pathogenesis in 5XFAD Mouse Spinal Cord: Retrograde Transneuronal Modulation after Peripheral Nerve Injury. *Neurotoxicity Research*, 24, 1-14. <u>http://dx.doi.org/10.1007/s12640-012-9355-2</u>
- [36] Xu, J. and Ling, E.A. (1994) Upregulation and Induction of Surface Antigens with Special Reference to MHC Class II Expression in Microglia in Postnatal Rat Brain Following Intravenous or Intraperitoneal Injections of Lipopolysaccharide. *Journal of Anatomy*, 184, 285-296.
- [37] Hauss-Wegrzyniak, B., Dobrzanski, P., Stoerh, J.D. and Wenk, G.L. (1998) Chronic Neuroinflammation in Rats Reproduces Components of the Neurobiology of Alzheimer's Disease. *Brain Research*, 780, 294-303. http://dx.doi.org/10.1016/S0006-8993(97)01215-8
- [38] Sugaya, K., Chou, S., Xu, S.J. and McKinney, M. (1998) Indicators of Glial Activation and Brain Oxidative Stress after Intraventricular Infusion of Endotoxin. *Molecular Brain Research*, 58, 1-9. <u>http://dx.doi.org/10.1016/S0169-328X(97)00365-3</u>
- [39] Tanaka, S., Ide, M., Shibutani, T., Ohtaki, H., Numazawa, S., Shioda, S. and Yoshida, T. (2006) Lipopolysaccharide-Induced Microglial Activation Induces Learning and Memory Deficits without Neuronal Cell Death in Rats. *Journal of Neuroscience Research*, 83, 557-566. <u>http://dx.doi.org/10.1002/jnr.20752</u>
- [40] Lee, J.W., Lee, Y.K., Yuk, D.Y., Choi, D.Y., Ban, S.B., Oh, K.W. and Hong, J.T. (2008) Neuro-Inflammation Induced by Lipopolysaccharide Causes Cognitive Impairment through Enhancement of Beta-Amyloid Generation. *Journal of Neuroinflammation*, 5, 37. <u>http://dx.doi.org/10.1186/1742-2094-5-37</u>
- [41] Deng, X.H., Ai, W.M., Lei, D.L., Luo, X.G., Yan, X.X. and Li, Z. (2012) Lipopolysaccharide Induces Paired Immunoglobulin-Like Receptor B (PirB) Expression, Synaptic Alteration, and Learning-Memory Deficit in Rats. *Neuroscience*, 209, 161-170. <u>http://dx.doi.org/10.1016/j.neuroscience.2012.02.022</u>
- [42] Laird, F.M., Cai, H., Savonenko, A.V., Farah, M.H., He, K., Melnikova, T., Wen, H., Chiang, H.C., Xu, G., Koliatsos, V.E., Borchelt, D.R., Price, D.L., Lee, H.K. and Wong, P.C. (2005) BACE1, a Major Determinant of Selective Vulnerability of the Brain to Amyloid-β Amyloidogenesis, Is Essential for Cognitive, Emotional, and Synaptic Functions. *Journal of Neuroscience*, 25, 11693-11709. <u>http://dx.doi.org/10.1523/JNEUROSCI.2766-05.2005</u>

- [43] Xiong, K., Cai, H., Luo, X.G., Struble, R.G., Clough, R.W. and Yan, X.X. (2007) Mitochondrial Respiratory Inhibition and O xidative Stress E levate β-Secretase (BACE1) Proteins and A ctivity in Vivo in the R at R etina. *Experimental Brain Research*, **181**, 435-446. <u>http://dx.doi.org/10.1007/s00221-007-0943-y</u>
- [44] Dutta, G., Zhang, P. and Liu, B. (2008) The Lipopolysaccharide Parkinson's Disease Animal Model: Mechanistic Studies and Drug Discovery. *Fundamental & Clinical Pharmacology*, 22, 453-464. http://dx.doi.org/10.1111/j.1472-8206.2008.00616.x
- [45] Lee, D.C., Rizer, J., Selenica, M.L., Reid, P., Kraft, C., Johnson, A., Blair, L., Gordon, M.N., Dickey, C.A. and Morgan, D. (2010) LPS-Induced Inflammation Exacerbates Phospho-Tau Pathology in rTg4510 Mice. *Journal of Neuroinflammation*, 7, 56. <u>http://dx.doi.org/10.1186/1742-2094-7-56</u>
- [46] Hoban, D.B., Connaughton, E., Connaughton, C., Hogan, G., Thornton, C., Mulcahy, P., Moloney, T.C. and Dowd, E. (2013) Further Characterisation of the LPS Model of Parkinson's Disease: A Comparison of Intra-Nigral and Intra-Striatal Lipopolysaccharide Administration on Motor Function, Microgliosis and Nigrostriatal Neurodegeneration in the Rat. Brain, Behavior, and Immunity, 27, 91-100. http://dx.doi.org/10.1016/j.bbi.2012.10.001
- [47] Ifuku, M., Katafuchi, T., Mawatari, S., Noda, M., Miake, K., Sugiyama, M. and Fujino, T. (2012) Anti-Inflammatory/ Anti-Amyloidogenic Effects of Plasmalogens in Lipopolysaccharide-Induced Neuroinflammation in Adult Mice. *Journal of Neuroinflammation*, 9, 197. <u>http://dx.doi.org/10.1186/1742-2094-9-197</u>
- [48] Martin, S.A., Pence, B.D., Greene, R.M., Johnson, S.J., Dantzer, R., Kelley, K.W. and Woods, J.A. (2013) Effects of Voluntary Wheel Running on LPS-Induced Sickness Behavior in Aged Mice. *Brain, Behavior, and Immunity*, 29, 113-123. <u>http://dx.doi.org/10.1016/j.bbi.2012.12.014</u>
- [49] Samanani, S., Mishra, M., Silva, C., Verhaeghe, B., Wang, J., Tong, J. and Yong, V.W. (2013) Screening for Inhibitors of Microglia to Reduce Neuroinflammation. CNS & Neurological Disorders-Drug Targets, 12, 741-749. <u>http://dx.doi.org/10.2174/18715273113126660177</u>
- [50] Asti, A. and Gioglio, L. (2014) Can a Bacterial Endotoxin Be a Key Factor in the Kinetics of Amyloid Fibril Formation? *Journal of Alzheimer's Disease*, 39, 169-179.
- [51] Morga, E., Faber, C. and Heuschling, P. (1999) Regional Heterogeneity of the Astroglial Immunoreactive Phenotype: Effect of Lipopolysaccharide. *Journal of Neuroscience Research*, 57, 941-952. <u>http://dx.doi.org/10.1002/(SICI)1097-4547(19990915)57:6<941::AID-JNR20>3.0.CO;2-Z</u>
- [52] Hauss-Wegrzyniak, B., Lynch, M.A., Vraniak, P.D. and Wenk, G.L. (2002) Chronic Brain Inflammation Results in Cell Loss in the Entorhinal Cortex and Impaired LTP in Perforant Path-Granule Cell Synapses. *Experimental Neurology*, **176**, 336-341. <u>http://dx.doi.org/10.1006/exnr.2002.7966</u>
- [53] Burguillos, M.A., Hajji, N., Englund, E., Persson, A., Cenci, A.M., Machado, A., Cano, J., Joseph, B. and Venero, J.L. (2011) Apoptosis-Inducing Factor Mediates Dopaminergic Cell Death in Response to LPS-Induced Inflammatory Stimulus: Evidence in Parkinson's Disease Patients. *Neurobiology of Disease*, **41**, 177-188. http://dx.doi.org/10.1016/j.nbd.2010.09.005
- [54] Lee, J.W., Lee, Y.K., Yuk, D.Y., Choi, D.Y., Ban, S.B., Oh, K.W. and Hong, J.T. (2008) Neuro-Inflammation Induced by Lipopolysaccharide Causes Cognitive Impairment through Enhancement of Beta-Amyloid Generation. *Journal of Neuroinflammation*, 5, 37. <u>http://dx.doi.org/10.1186/1742-2094-5-37</u>
- [55] Richwine, A.F., Parkin, A.O., Buchanan, J.B., Chen, J., Markham, J.A., Juraska, J.M. and Johnson, R.W. (2008) Architectural Changes to CA1 Pyramidal Neurons in A dult and A ged Mice after Peripheral Immune Stimulation. *Psychoneuroendocrinology*, 33, 1369-1377. <u>http://dx.doi.org/10.1016/j.psyneuen.2008.08.003</u>
- [56] Chugh, D., Nilsson, P., Afjei, S.A., Bakochi, A. and Ekdahl, C.T. (2013) Brain Inflammation Induces Post-Synaptic Changes during Early Synapse Formation in Adult-Born Hippocampal Neurons. *Experimental Neurology*, 250, 176-188. <u>http://dx.doi.org/10.1016/j.expneurol.2013.09.005</u>
- [57] Jakubs, K., Bonde, S., Iosif, R.E., Ekdahl, C.T., Kokaia, Z., Kokaia, M. and Lindvall, O. (2008) Inflammation Regulates Functional Integration of Neurons Born in Adult Brain. *Journal of Neuroscience*, 28, 12477-12488. <u>http://dx.doi.org/10.1523/JNEUROSCI.3240-08.2008</u>
- [58] Commins, S., O'Neill, L.A. and O'Mara, S.M. (2001) The Effects of the Bacterial Endotoxin Lipopolysaccharide on Synaptic Transmission and Plasticity in the CA1-Subiculum Pathway in Vivo. Neuroscience, 102, 273-280. <u>http://dx.doi.org/10.1016/S0306-4522(00)00498-X</u>
- [59] Jo, J.H., Park, E.J., Lee, J.K., Jung, M.W. and Lee, C.J. (2001) Lipopolysaccharide Inhibits Induction of Long-Term Potentiation and Depression in the Rat Hippocampal CA1 Area. *European Journal of Pharmacology*, **422**, 69-76. <u>http://dx.doi.org/10.1016/S0014-2999(01)01075-5</u>
- [60] Min, S.S., Quan, H.Y., Ma, J., Lee, K.H., Back, S.K., Na, H.S., Han, S.H., Yee, J.Y., Kim, C., Han, J.S. and Seol, G.H. (2009) Impairment of Long-Term Depression Induced by Chronic Brain Inflammation in Rats. *Biochemical and Bio-physical Research Communications*, 383, 93-97. <u>http://dx.doi.org/10.1016/j.bbrc.2009.03.133</u>

- [61] Gentleman, S.M., Nash, M.J., Sweeting, C.J., Graham, D.I. and Roberts, G.W. (1993) β-Amyloid Precursor Protein (βAPP) as a Marker for Axonal Injury after Head Injury. *Neuroscience Letters*, 160, 139-144. <u>http://dx.doi.org/10.1016/0304-3940(93)90398-5</u>
- [62] Chen, X.H., Johnson, V.E., Uryu, K., Trojanowski, J.Q. and Smith, D.H. (2009) A Lack of Amyloid Beta Plaques Despite Persistent Accumulation of Amyloid β in Axons of Long-Term Survivors of Traumatic Brain Injury. *Brain Pathology*, 19, 214-223. <u>http://dx.doi.org/10.1111/j.1750-3639.2008.00176.x</u>
- [63] Mongiat, L.A. and Schinder, A.F. (2011) Adult Neurogenesis and the Plasticity of the Dentate Gyrus Network. European Journal of Neuroscience, 33, 1055-1061. <u>http://dx.doi.org/10.1111/j.1460-9568.2011.07603.x</u>
- [64] Lee, J.W., Lee, Y.K., Yuk, D.Y., Choi, D.Y., Ban, S.B., Oh, K.W. and Hong, J.T. (2008) Neuro-Inflammation Induced by Lipopolysaccharide Causes Cognitive Impairment through Enhancement of Beta-Amyloid Generation. *Journal of Neuroinflammation*, 5, 37. <u>http://dx.doi.org/10.1186/1742-2094-5-37</u>
- [65] Sheng, J.G., Bora, S.H., Xu, G., Borchelt, D.R., Price, D.L. and Koliatsos, V.E. (2003) Lipopolysaccharide-Induced-Neuroinflammation I ncreases I ntracellular A ccumulation of A myloid Precursor Protein and A myloid β Peptide in APPswe Transgenic Mice. *Neurobiology of Disease*, 14, 133-145. http://dx.doi.org/10.1016/S0969-9961(03)00069-X
- [66] Joshi, Y.B., Giannopoulos, P.F., Chu, J. and Praticò, D. (2014) Modulation of Lipopolysaccharide-Induced Memory Insult, γ-Secretase, and Neuroinflammation in Triple Transgenic Mice by 5-Lipoxygenase. *Neurobiology of Aging*, 35, 1024-1031. <u>http://dx.doi.org/10.1016/j.neurobiolaging.2013.11.016</u>
- [67] Qiao, X., Cummins, D.J. and Paul, S.M. (2001) Neuroinflammation-Induced Acceleration of Amyloid Deposition in the APPV717F Transgenic Mouse. *European Journal of Neuroscience*, 14, 474-482. http://dx.doi.org/10.1046/j.0953-816x.2001.01666.x
- [68] DiCarlo, G., Wilcock, D., Henderson, D., Gordon, M. and Morgan, D. (2001) Intrahippocampal LPS Injections Reduce Aβ Load in APP+PS1 Transgenic Mice. *Neurobiology of Aging*, 22, 1007-1012. http://dx.doi.org/10.1016/S0197-4580(01)00292-5
- [69] Herber, D.L., Mercer, M., Roth, L.M., Symmonds, K., Maloney, J., Wilson, N., Freeman, M.J., Morgan, D. and Gordon, M.N. (2007) Microglial Activation Is Required for Aβ Clearance after Intracranial Injection of Lipopolysaccharide in APP Transgenic Mice. *Journal of Neuroimmune Pharmacology*, 2, 222-231. <u>http://dx.doi.org/10.1007/s11481-007-9069-z</u>
- [70] Kandalepas, P.C., S adleir, K.R., E imer, W.A., Z hao, J., N icholson, D.A. and Vassar, R. (2013) The A lzheimer's β-Secretase BACE1 Localizes to Normal Presynaptic Terminals and to Dystrophic Presynaptic Terminals Surrounding Amyloid Plaques. Acta Neuropathologica, 126, 329-352. <u>http://dx.doi.org/10.1007/s00401-013-1152-3</u>



Prevalence of Dementia and Its Associated Factors in Cotonou Teaching Hospital, Benin

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Abstract

Introduction: Dementia constitutes a public health hazard in developing countries. There is little data in the sub-Saharan region of African especially in Benin. Objective: Determining dementia hospitalization prevalence and identifying its associated factors in CNHU-HKM, Cotonou. Method: It was a cross-sectional, prospective, descriptive and analytical research conducted from October 2012 to July 2013 in the neurology department; it involved 251 patients aged 50 and above. Dementia screening was conducted using a modified and adapted Mini Mental Scale Examination (MMSE). Dementia clinical and etiological diagnoses were respectively conducted based on DMS-IV and HACHINSKI criteria. Results: Patients were averagely aged 60.9 ± 8.1 . Sex ratio (Male/Female) was 1.07. Dementia prevalence was 8.8%. This rate increased proportionally with age, from 5.3% with patients aged below 60 to 12.7% with patients aged above 60. Degenerative dementia was the most predominant type (50%). Following multi-varied analysis, smoking (RC = 6.05 [IC 95% = 1.26 - 29.38] p = 0.0001) and stroke past records (RC = 6.05 [IC 95% = 1.26 - 29.38] p = 0.001) revealed to be the factors associated with dementia. Conclusion: This research showed that dementia affects a significant part of the aging population in CNHU-HKM. It is imperative to combat its associated factors so as to defuse its prevalence.

Keywords

Prevalence, Dementia, Elders, MMSE, DSMIV, Cotonou, Benin

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1. Introduction

In sub-Saharan Africa, the gradual improvement of living conditions is correlated with life span increase. The end result is the increasing emergence of certain pathologies associated with age, among others, dementia [1]. Dementia syndrome is defined as a global cognitive decline in a person with a normal state of consciousness. Its occurrence and development are progressive. The Pathophysiology is dependent on the cause. Dementia occurs through accelerating apoptosis. AD is the commonest among neurodegenerative dementias; it is predominantly cortical. There are two types of brain damage resulting in AD: senile plaques (amyloidogenesis) and neurofibrillary tangles. Preferentially, the lesions affect the limbic system. In parallel, during fibrillary degeneration, pathologic filaments comprised of TAU proteins build up around the cell bodies of neurons. Several factors such as ageing, environment and genetic interrelate then causing dementia [2]. Dementia diagnosis is referred to a progressive and acquired memory deterioration associated with a disorder of at least one of the other cognitive functions. It results in social and occupational impairment, as well as difficulties in carrying out daily activities, not forgetting loss of autonomy [3]. It has become a public health hazard. In 2005 the number of dementia patients was estimated at 24.3 million worldwide, and 4.6 million new cases recorded yearly. This figure will reach 81.1 million by 2040 [2]. Worldwide, dementia is one of the major causes of disability and dependency in elderly population. It has physical, psychological, social and economic consequences on care givers, families and society. There is hardly any epidemio-clinical data on dementia in Africa [3] [4] particularly in Benin [5] [6]. So, this research was initiated to determine dementia hospitalization prevalence and identify its associated factors.

2. Method

It consists in a cross-sectional, prospective, descriptive and analytical research conducted from October 1, 2012 to July 30, 2013 in Hubert Koutoukou Maga teaching hospital of Cotonou Benin. Benin is a French-speaking country located in West African (Figure 1), sharing border with Nigeria on the east. It boasts of 9 million inhabitants with an area of 112,622 km². The population subjected to the research consisted of patients who went through consultation in the neurology department during research period. The sample size was calculated using Daniel Schwartz formulae $n = Z\alpha^2 pq/i^2 = 182$ with p = 13.79% (dementia prevalence in neurology department of CNHU-HKM in 1991 [5], $\alpha = 5\%$ and I = 5%. The total number of subjects enrolled in the research was 251. But during the study period, we performed a systematic enrollment of all patients who met the inclusion criteria and consented to participate in the research, up to the number expected.

Inclusion criteria

- Be aged 50 and above.
- Having been consulted in the neurology department during research period. *Exclusion criteria*
- Having experienced language disorder.
- Having been diagnosed as experiencing continuous confusional episode, chronic psychosis and depressive syndrome.



Figure 1. Position of Benin in Africa.

Diagnosis Criteria

Patients were subjected to their cognitive functions evaluation through Mini Mental State Examination (MMSE) in its modified version which is adapted to the research cultural era. Patients who recorded MMSE $\leq 24/30$ are likely to have dementia and subjected to DSM IV criteria [7] for dementia diagnosis. Its severity is appreciated in conformity with MMSE score. In fact,

- a score 20/30 ≤ MMSE ≤ 24/30 with a patient matching DMS IV criteria, is considered as "moderate dementia".
- a score MMSE \leq à 19/30 with a patient matching DMS IV criteria, is considered as "severe dementia". HACHINSKI score was used to determine the nature of the effect:
- >7: vascular dementia.
- Between 4 and 7: mixed dementia.
- <7 degenerative dementia.

The Variables Studied Were

Dependent variable: dementia.

Independent variables: they were:

- Socio-demographic: age, gender, ethnicity, profession, residence, marital status, number of children, education level.
- Clinical: Hypertension, diabetes, obesity, stroke, cardiomyopathy, sinusitis, epilepsy hyperlipidemia.
- Types of dementia: curable dementia, vascular dementia, degenerative dementia, mixed dementia.

Data Processing and Analysis

Data processing was conducted through EPI-DATA. Data analysis was done using statistics software STATA/ IC 11.0. A descriptive analysis was completed with regard to the variables which were studied. So, for qualitative variables, frequencies and proportions were determined. Either chi 2 or FISHER test was used if only expected values are lower than 5. For quantitative values, averages together with their typical gaps, medians, minima and maxima have all been described. STUDENT test was utilized for comparisons purpose. The study of associated factors was conducted using logistical regression model in unvaried and multi-varied analyses. The multi-varied a nalysis was conducted by i nserting into the model all variables, of which p value in unvaried analysis is less or equal to 20%, due to the exploratory nature of the study. The break-even point in terms of significance was 5% and confidence gap rated at 95%.

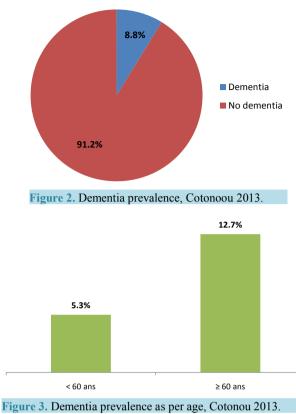
Ethical Considerations

Each patient or his/her next of kin submitted a written notice of consent upon explanation of the research objectives and modalities. Moreover, patients' right as well as confidentiality was highly respected.

3. Results

All in all, 251 patients were enrolled. They were aged between 50 to 86 years with an average of 61 years ± 8.1 . There were 130 (51.8%) males against 121 females (48.2%) meaning a sex-ratio of 1.07. The other socio-de-mographic characteristics are shown in Table 1.

Clinically, 59 (23.5%) complain of mental disorders. 173 (68.9%) and 73 (29.1%) respectively had hypertension and stroke past records. **Table 2** shows the population clinical characteristics. 28 (11.2%) out of the 251 patients enrolled had MMSE score below or equal to 24. Indeed, 22 out of the 28 patients matched DSM IV dementia criteria, representing 8.8% (**Figure 2**). The prevalence was 5.3% with patients aged below 60 and 12.7% with those aged above 60 (**Figure 3**). All the 22 patients suffering from dementia were aged between 55 and 84, meaning an average age of 66 ± 9.1 . There were 11 males (50%) and 11 females (50%). 18 (81.2%) were married; 21 (95.5%) lived in a family and 9 (41.0%) already retired from work. **Table 3** shows the classification of other s ocio-demographic characteristics of d ementia patients. 1 8.2% r evealed to have b ehavioral and mental disorders. **Table 4** shows clinical characteristics of d dementia patients. Total MMSE score varied from 7 to 23 with a median of 22. **Table 5** shows the classification of MMSE various items of dementia patients. 19 (86.3%) showed memory disorders against 13% showing temporal and spatial disorders. **Table 6** shows the prevalence rate of c ognitive di sorders of dementia. **Figure 4** shows classification of dementia etiologies. Within curable etiologies, there were three chronic hydrocephaly cases, two chronic subdural hematomas and a single cerebral tumor (**Figure 5**).



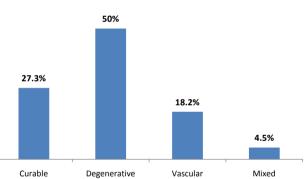


Figure 4. Classification of dementia etiologies, Cotonou 2013.

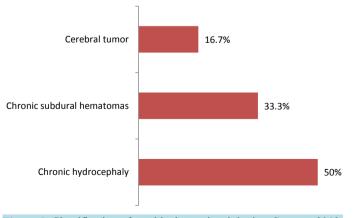


Figure 5. Classification of curable dementia etiologies, Cotonou 2013.

Table 1. Socio-demographic characteristics of the population, Cotonou 2013.		
	Percentage	
Marital status		
Married	75.7%	
Widower	15.9%	
Single	4.3%	
Divorced	1.1%	
Life pattern		
Lives in family	88.8%	
Lives alone/single life	11.2%	
Residence		
Urban locality	85.3%	
Rural locality	14.7%	
Level of education		
Uneducated	10.4%	
Primary	25.9%	
Secondary	38.6%	
University	25.1%	
Profession		
Civil servant	24.7%	
Private worker	11.2%	
Craftsman	9.6%	
Housewife	17.5%	
Retired	30.7%	
Trader	63%	

Table 2. Clinical characteristics of the population, Cotonou 2013.

Clinical characteristics	Total	Percentage
Motive of consultation		
Mental disorders	59	23.5%
Behavioral disorders	21	8.4%
Past records		
Hypertension	173	68.9%
Diabetes	42	16.7%
Obesity	58	23.1%
Hyperlipidemia	59	23.5%
Cardiomyopathy	21	8.4%
Epilepsy	10	3.9%
Stroke	73	29.1%
Smoking	17	6.8%
Alcoholism	71	28.3%

socio-demographic characteristics of dementia patients, Cotoniou 2013.		
	Total	Percentage (%)
Matrimonial status		
Single	2	9.1%
Married/concubine	18	81.8%
Widower	2	9.1%
Life pattern		
Lives alone/Single	1	4.5%
Lives in family	21	95.5%
Profession		
Civil servant	1	4.5%
Private worker	3	13.6%
Craftsman	2	9.1%
Housewife	6	27.3%
Retired	9	41.0%
Trader	1 4.5%	
Residence		
Urban locality	18	81.8%
Rural locality	4	18.2%
Other	7	31.8%
Level of education		
Uneducated	5	22.7%
Primary	6	27.3%
Secondary	6	27.3%
Higher	5	22.7%

Table 3. Socio-demographic characteristics of dementia patients, Cotonou 2013.

Table 4. Clinical characteristics of dementia patients, Cotonou 2013.

Clinic characteristics	Total	Percentage
Motive of consultation		
Mental disorders	4	18.2%
Behavioral disorders	4	18.2%
Past records		
Hypertension	16	72.7%
Diabetes	3	13.6%
Obesity	6	27.3%
Hyperlipidemia	5	22.7%
Cardiomyopathy	3	14.3%
Epilepsy	1	4.5%
Stroke	14	63.6%
Smoking	4	18.2%
Alcoholism	5	22.7%

	ina patientis, cotonou 2015.
MMSE items	Median score [Mini - Max]
Time orientation	4 [0 - 5]
Space orientation	4 [0 - 5]
Learning	3 [2 - 3]
Attention and calculation	0 [0 - 2]
Memory retention	0 [0 - 1]
Language	7 [2 - 8]
Constructive practices	1 [0 - 1]

 Table 5. MMSE characteristics of dementia patients, Cotonou 2013.

 Table 6. Classification of cognitive disorders in dementia population, Cotonou 2013.

Types of disorder	Total	Percentage
Time orientation	13	59.1%
Space orientation	13	59.1%
Learning	1	4.6%
Attention and calculation	21	95.4%
Memory retention	19	86.3%
Language	14	63.6%
Constructive practices	6	27.3%

3.1. Factors Associated with Dementia

The only one socio-demographic factor associated with dementia was age exceeding 60 years (IC 95% = 2.61 [1.03 - 6.67], p = 0.036). Table 7 shows such data.

Clinically, stroke past records (IC 95% = 5.04 [2.01 - 12.62], p = 0.0001) and smoking (IC 95% = 3.69 [1.09 - 12.50], p = 0.049) were closely associated with dementia as shown in **Table 8**.

3.2. Predictive Factors of Dementia

After multi-varied analysis of associated variables in unvaried analysis, those which are individually associated with mortality were stroke past records (IC 95% = 7.66 [2.46 - 23.85], p = 0.0001), and smoking (IC 95% = 6.05 [1.26 - 29.38], p = 0.001). Table 9 shows these data.

4. Discussion

It was a cr oss-sectional, descriptive and analytical research. The method we implemented is really adapted to this kind of research in the sense that, there was no plan to follow-up the patients; instead the strategy was to conduct a questionnaire after consultation. During the research period, we exhaustively screened all patients fulfilling our inclusion criteria. This enabled us incorporate 251 patients. However, only 22 patients showed dementia based on DMS IV diagnosis criteria. Such a small number of patients did not allow us highlight some factors associated with dementia, which were however observed in the paper.

4.1. Prevalence

In our research dementia prevalence was 8.8%. This result is close to 8.1% obtained by Mbelesso *et al.* [8] in a cross-sectional research conducted in the 3rd district of Bangui (Central African Republic) in 2012, on dementia epidemiology in elderly population. Similarly, Molero *et al.* [9] recorded in 2007 a prevalence of 10.3% in a re-

	Dementia n (%)	No dementia n (%)	RC [IC 95%]	p valu
Age				0.036
<60 years	7 (5.3%)	126 (94.7%)	1	
≥60 years	15 (12.7%)	103 (87.3%)	2.61 [1.03 - 6.67]	
Sex				0.860
Male	11 (8.5%)	119 (91.5%)	1	
Female	11 (9.1%)	110 (90.1%)	1.08 [0.45 - 2.59]	
Marital status				0.390
Single	2 (18.2%)	9 (81.8%)	1	
Married	18 (9.5%)	172 (90.5%)	2.12 [0.42 - 1059]	
Widower	2 (5%)	38 (95%)	0.50 [0.11 - 2.25]	
Divorced	0 (0%)	10 (100%)	-	
Life pattern				0.250
Lives alone	1 (3.6%)	27 (96.4%)	1	
Lives in family	21 (9.4%)	202 (90.6%)	2.80 [0.36 - 21.71]	
Profession				0.160
Civil servant	1 (1.6%)	61 (98.4%)	1	
Private worker	3 (10.7%)	25 (89.4%)	7.32 [0.73 - 73.78]	
Craftsman	2 (8.3%)	22 (91.7%)	5.54 [0.47 - 64.22]	
Housewife	6 (13.6%)	38 (86.7%)	9.63 [1.11 - 83.13]	
Retired	9 (11.7%)	68 (88.3%)	4.35 [0.25 - 73.98]	
Trader	1 (6.2%)	15 (93.7%)	8.07 [0.99 - 65.58]	
Education level				0.295
Uneducated	5 (19.2%)	21 (80.8%)	1	
Primary	6 (9.2%)	59 (90.1%)	4.31 [0.35 - 63.98]	
Secondary	6 (6.2%)	91 (93.8%)	2.80 [0.36 - 21.71]	
Higher	5 (7.9%)	58 (92.1%)	0.50 [0.11 - 2.25]	

Table	7. Socio-demograp	hic factors	associated with	dementia	Cotonou 2013
1 ant	1. Docio-acinograp	me nacions	associated with	uomontia.	Colonou 2015.

*Significant result.

search conducted on elderly population living on the Caribbean coast of Venezuela. In 2012, Coume M. *et al.* [7] recorded 10.8% as prevalence rate in a cross-sectional research on the estimation of cognitive prevalence within an elderly Senegalese population. Our record of 8.8% dementia prevalence is lower than 24.3% registered by Uwakwe R. *et al.* [10] in a cross-sectional research conducted in 2009 on dependency epidemiology in elderly Nigerian population. It is also lower than 43.2% recorded by Ndiaye *et al.* [11] in a research conducted from January 2004 to June 2005 on the operation of a Senegalese memory clinic in Fann teaching hospital psychiatry department (Dakar, Senegal). In fact, many reasons may justify this difference in terms of the prevalence rate. First of all, in Uwakwe R. *et al.* [10] research, patients were aged 65 and above and the sample was made up of 1238 patients who went through 10/66 protocol; knowing that the specificity and sensitivity of such a protocol are higher than those of MMSE we implemented. The same goes with Ndiaye *et al.* [11] who utilized in their research Senegal Test, which sensitivity and specificity are higher than those of MMSE.

	Dementia n (%)	No dementia n (%)	RC [IC 95%]	p value
	Dementia ii (70)	No dementia il (70)	Re [le 95/0]	
Hypertension				0.68
Yes	16 (9.3%)	157 (90.7%)	1.22 [0.46 - 3.23]	
No	6 (7.7%)	72 (92.3%)	1	
Diabetes				0.67
Yes	3 (7.1%)	39 (92.9%)	0.76 [0.22 - 2.73]	
No	19 (9.1%)	190 (90.9%)	1	
Obesity				0.63
Yes	6 (10.3%)	52 (89.7%)	1.28 [0.45 - 3.42]	
No	16 (8.3%)	177 (91.7%)	1	
Epilepsy				0.89
Yes	1 (10%)	9 (90%)	1.2 [0.14 - 9.64]	
No	21 (8.7%)	220 (91.3%)	1	
Stroke past records				0.0001*
Yes	14 (19.2%)	59 (80.8%)	5.04 [2.01 - 12.62]	
No	8 (4.5%)	170 (95.5%)	1	
Smoking				0.049*
Yes	4 (23.5%)	13 (76.5%)	3.69 [1.09 - 12.50]	
No	18 (7.7%)	216 (92.3%)	1	
Alcoholism				0.53
Yes	5 (7.0%)	66 (93.0%)	0.72 [0.26 - 2.05]	
No	17 (9.4%)	163 (90.6%)	1	

*Significant result.

Table 9. Dementia predictive factors. Results of multi-varied analysis, Cotonou 2013.

Variables	Total	Dementia (n%)	RC _{Gross} [IC _{95%}]	RC _{adjusted} [IC _{95%}]	p value
Stroke					0.0001*
Yes	73	14 (19.2)	5.04 [2.01 - 12.62]	7.66 [2.46 - 23.85]	
No	178	8 (4.5)	1	1	
Smoking					0.001*
Yes	17	4 (23.5)	3.69 [1.09 - 12.50]	6.05 [1.26 - 29.38]	
No	234	18 (7.7)	1	1	

search, enrolled patients (132 in total) were selected among those who showed memory disorders. All this could explain the higher rate recorded in these two researches. Notwithstanding our 8.8% prevalence is higher than the figure recorded by other authors. Thus, Paraïso et al. [6] recorded 2.6% in rural area (Djidja) and 3.7% in urban area (Cotonou) in a door-to-door research conducted in Benin in 2010 on dementia epidemiology with patients aged 65 and above, whilst Guerchet et al. [12] recorded 6.7% in a cross-sectional research conducted in the year 2012 on dementia epidemiology within elderly population living in Brazzaville (Congo). Longdon et al. [13] registered 6.4% prevalence in a cross-sectional community research on dementia prevalence in rural Tanzania. El Tallawy *et al.* [14] also registered 2.3% in a research conducted in 2011 on dementia prevalence in Kharga (Egypte) in 2012, whilst Y usuf *et al.* [15] estimated dementia prevalence at 2.8% in a community research on dementia prevalence and subtypes dementia within elderly population living in northern Nigeria. These prevalence rates which are very lower than what we recorded in our research could be explained by the fact that most of these researches were conducted within a vast population with samples above 1000 patients (1198, 8173, and 1139 patients were respectively enrolled by Longdon, El Tallawy and Paraïso). In addition, the test utilized in these researches is CSID, which has a weaker specificity than MMSE.

4.2. Socio-Demographic Characteristics

In our research dementia patients average age was 66 years \pm 9.1. This result is close to 67 years \pm 17 observed by Ndiaye et al. [11] as well as 67 years \pm 7.5% recorded by Coume et al. [7]. The same result is lower to 76 years \pm 7.1 recorded by Guerchet *et al.* [12]. This result is explained by the fact that the patients enrolled in the research were 65 years and above against 50 years and above in our research. It comes out that age is the sole risk factor which was corroborated. In our research, 62% of dementia patients were aged beyond 60 years. Moreover, d ementia p revalence i ncreased with a ge. The figure varied from 5.3% b elow 60 years to 12.7% beyond 60 years. El Tallawy [14] recorded that dementia prevalence of 2.3% increased drastically to 18.5% with subjects aged beyond 80 years. Libre et al. [16] confirmed that beyond 65 years, dementia prevalence doubles anytime age increases by 5 years. Other authors such as Longdon, Coume, Yusuf et Paraïso [6] [7] [13] [15] recorded similar results. Out of 22 dementia patients 11 were males; meaning a sex ratio of 1. This result is dissimilar to the ratio recorded by most researches which revealed a female predominance. Thus, Mbeleso et al. [8] recorded 82.35% female predominance whilst Stewart et al. [17] recorded 60.8%. Letenneur et al. [18] as well as Nitrini et al. [19] corroborated this female predominance which according to Stewart et al. [17] might be explained by the biological and hormonal differences between the two sexes, particularly estrogens. Our result which is quite different from others' could be explained by the fact that the population was male biased and not large enough. Indeed 18 (81.8%) of dementia patients were married. This result is similar to that of Ndiaye et al. (76.5) and Coume et al. (79%). However, it is different from most authors' findings. Thus, Fratiglioni [20] maintains t hat b eing a widower o r a d ivorcee i mplies an u nder-performance r egarding MM SE; s uch u nder-performance is not recorded with elderly married persons. We could then bring up the hypothesis of marriage playing a role of social balance. Van Gelder et al. [21] in a research conducted in 2006 in Finland noted that men who lost their partner, those who weren't married and those who live single life or lived single life for at least 5 years were much more exposed to cognitive decline, unlike married men or men who lived with a partner. This predominance of married persons has its explanation in our socio-cultural realities. In fact, it is rare to see persons aged 50 and above who are not married. Out of the 22 dementia patients, 21 (95.5%) lived in a family. Fratiglioni et al. [20] proved that single patients were twice exposed to dementia comparing to those living in a family. They also found out that a closed social network increases by 60% the risk. Dementia risk is therefore correlated with the significance of social network. Baiyewu et al. [3] exposed that in Africa el derly persons live in extended family made up of several generations. 6.7% live alone; which contrast the prevalence rate in Europe (35% - 39%). Our results reflect the importance of social network in Africa particularly in Benin. Such a rich social network may explain the low dementia prevalence in our societies. Unfortunately, this social network tends to ebb with a progressive urbanized lifestyle leading to individualism. However, we should not forget that these maladies are under-diagnosed. 5 (22.7%) of the dementia patients were uneducated. Among the 17 (77.3%) who were educated, 6 (27.3%) had primary education only. In Egypt, El Tallawy et al. [14] conducted a research which revealed that dementia prevalence rate was significantly higher among uneducated persons (6.35%) than literate persons (0.6%). They asserted that the low education level played a predominant role in dementia occurrence in elderly population especially when they did not reach elementary school. This assertion is supported by Guerchet et al. who demonstrated that, not acquiring elementary education was significantly associated with dementia. On the contrary, a research conducted in Kenya revealed that there was no proof of little education being associated with dementia occurrence [22].

4.3. Dementia Etiologies

Results from our research revealed that degenerative dementia cases (50%) were mostly prevalent, followed by

vascular dementia (31.8%) then curable dementia (27.3%) and finally mixed dementia (18.2%). This result is similar to that of Shelley et al. [23] who recorded 57.1% and 26.9% respectively for degenerative and vascular dementia. Alzheimer disease (AD) is the commonest neurodegenerative dementia; it is cortically predominant and represents dementia major cause in elderly population. AD is fundamentally caused by two types of cerebral lesions: senile plaques and fibrillary neurodegeneration. Senile plaques are extracellular deposits of amyloidal substance, mostly made up of peptide polymer A β (or β -amyloidal). This substance settles down progressively in the brain predominantly in the gray substance. These insoluble deposits are not metabolized by the body and destroy progressively adjacent nervous fibers. Parallel to this, in the fibrillary degeneration process, pathologic filaments made of protein build up around the cell body of the neurons. These neurons depolymerize then aggregate in the cytoplasm. Generally, this degeneration begins from the hippocampus and spreads across temporal regions. Other researches also revealed a high prevalence of AD. Thus, Yusuf et al. [15] recorded 66.7% AD while Alladi et al. [24] and Shelley et al. [23] respectively recorded 38.39% and 52.6%. Vascular dementia (VD) was the second etiology of dementia syndromes after degenerative dementia of Alzheimer type. V ascular dementia encompasses the whole dementia states secondary to cerebrovascular lesions. El Tallawy et al. [14] recorded 28.7% of v ascular d ementia cas es. They are cau sed by the consequences of cerebrovascular affection (blockage of vessels or cerebral hemorrhage). In general they worsen discretely (during a new stroke occurrence). Neuro-imaging plays a predominant role in diagnosing by showing lesions of vascular origin. Curable dementia cases are those of which the etiology was identified. The most frequent causes are neurosurgical: cerebral tumors, adult hydrocephaly, and subdural hematoma. These causes are easily revealed by the neuro-imaging. The systematic results of research about infectious dementia (mainly syphilis, HIV-Aids or hardly ever Whipple disease), endocrine or deficient dementia are rarely productive because these types of dementia can totally be reversible. The discovery of curable cause for dementia does not always guarantee a complete recovery from the disease. However, systematic results from the research allow finding and treating reversible concomitant affections and disorders which can contribute to worsening dementia.

4.4. Factors Associated with Dementia

In our research two factors were revealed to be associated with dementia. There were stroke past records and smoking. 63.6% had stroke. This result is similar to the record of the University of Oxford in the United Kingdom [25], following a research conducted by the department of research on diseases prevention with stroke inpatients. The prevalence of dementia with these patients was comparable to the rate of prevalence recorded with stroke free patients who were 10 years older. Only one stroke occurrence provokes no diffuse decline of cognitive functions leading to dementia. It is the repetitive occurrence of such incidents with the same patient which can provoke cognitive decline. We then refer to multiple infarction dementia or multi-infarction dementia. In a research conducted by Altieri et al. [26], about 30% of stroke survivors suffered from dementia thereafter. The risk is high after stroke occurrence, but it is higher even after the embolic thrombosis occurrence. Post stroke dementia risk factors are not well known. However, three theoretical reasons could explain post stroke dementia occurrence. First of all, dementia could be the direct consequence of cerebral lesions of vascular origin. Then it could be due to the combination of degenerative lesions of Alzheimer type in a pre-clinical stage; some post stroke dementia states are progressive. Finally, white substance anomalies could also contribute to the decline in the sense that, they are associated with a high risk of vascular relapses and could stimulate neuropsychological disorders. V ascular lesions, white substance and lesions of A lzheimer type a nomalies all put together, could stimulate post stroke dementia occurrence [27]. 18% were addicted to smoking. This result is similar to Juan et al. [28] findings, as they maintained that smoking revealed to be a risk factor. Likewise, Rusanen et al. [29] in 2011 established that smoking increases Alzheimer disease risk occurrence by 2. These Finnish scientists conducted a broad research between 1978 and 2008. In this research, 21,123 patients aged 50 to 60 years were thoroughly followed during 23 years. They diagnosed 5367 senile dementia cases of which 1136 Alzheimer diseases. Having reviewed different risk factors associated with dementia, researchers discovered that smoking played an important role. In fact, dementia risk increased by hundred percent (100%) concerning heavy smokers consuming over two packets of cigarette a day. Globally, smokers presented a risk of vascular dementia multiplied by 2.72. The same researchers showed that smoking affected the brain directly and increased both oxidative stress and inflammation which were two risk factors related to Alzheimer disease [29]. Nonetheless, the effect of smoking in dementia occurrence is not well explained. We only know that smoking has cerebrovascular effects and amplifies the cholinergic metabolism by dysfunctioning brain nicotine receptacles. A Chinese research [30] proved that second-hand smoking constitutes a risk factor which could increase by 29% the occurrence of severe dementia.

5. Conclusion

Dementia with elderly population aged 50 and above, remains a pathology to which very little attention is paid in Benin health system. This research through its theme originality highlights dementia in the intention of developing adapted and effective national strategies to combat it. There is a need to conduct other researches about this theme in developing countries, where, just like in the case of Benin no consideration is given to this pathology.

References

- Napon, C., Traore, S., Idris, S., Niakara, A., Ouango, G.G., Kabré, A., *et al.* (2009) Dementia in Sub-Saharan Africa: Clinic and Etiological Aspects in Ouagadougou Hospital (Burkina Faso). *African Journal of Neurological Sciences*, 28, 1-5.
- [2] Ferri, C.P. (2005) Global Prevalence of Dementia. *Delphy Consensus Study*, **366**, 2112-2117.
- [3] Baiyewu, O., Bella, A.F., Adeyemi, J.D., Bamgboye, E.A., Ikuesan, B.A. and Jegede, R.O. (1997) Health Problems and Socio-Demographic Findings in Elderly Nigerians. *African Journal of Medicine and Medical Sciences*, 2, 13-17.
- [4] Touré, K., Coumé, M., Ndiaye-Ndongo, N.D., Thiam, M.H., Zunzunegui, M.V., Bacher, Y., et al. (2008) Dementia Prevalence in a Senegalese Elderly Population. *African Journal of Neurological Sciences*, 27, 15-22.
- [5] Avode, D.G., Gandaho, P., Da-Cruz, P.C., Ahyi, R.G. and Zohoun, Th. (1998) Senile Dementia in Cotonou Hospital. Le Bénin Médical, 7, 23-27.
- [6] Paraïso, M.N. (2010) Dementia Epidemiology in Elderly Benin Population Aged 65 and Above (Afrique de l'Ouest). Ph.D. Heath Science Biology. University of Limoges, Limoges, 486p.
- [7] Coume, M., Touré, K., Thiam, M.H., Zunzunegui, M.V., Bacher, Y., Diop, T.M., et al. (2012) Evaluation of Cognitive Deficiency P revalence in E Iderly P opulation in a Senegalese H ealth C enter "I nstitution R etraite". Geriatr P sychol Neuropsychiatr Vieil, 10, 39-46.
- [8] Mbelesso, P., Tabo, A., Guerchet, M., Mouanga, A.M., Bandzouzi, B., Houinato, D., et al. (2012) Dementia Epidemiology in Elderly Population Living in Bangui 3rd Municipality (Central A frica Republic)]. Bulletin de la Société de pathologie exotique, 105, 388-395. <u>http://dx.doi.org/10.1007/s13149-012-0247-8</u>
- [9] Molero, A.E., Pino-Ramirez, G. and Maestre, G.E. (2007) High Prevalence of Dementia in a Caribbean Population. *Neuroepidemiology*, **29**, 107-112. <u>http://dx.doi.org/10.1159/000109824</u>
- [10] Uwakwe, R., Ibeh, C.C., Modebe, A.I., Bo, E., Ezeama, N., Njelita, I., *et al.* (2009) Dependence Epidemiology in Elderly Population in Nigeria: Prevalence, Determinants, Informal Care and the Use of Health Services; Research Group 10/66 Dementia. *Journal of the American Geriatrics Society*, **57**, 1620-1627. http://dx.doi.org/10.1111/j.1532-5415.2009.02397.x
- [11] Ndiaye, N.N.D., Sylla, A., Touré, K., Thiam, M.H. and Gueye, M. (2011) Operations Assessment in a S enegalese Memory Clinic, F ann Teaching Hospital Psychiatric Department (Dakar, Senegal). *African Journal of Neurological Sciences*, 30, 2-10.
- [12] Guerchet, M., Houinato, D., Paraiso, M.N., von-Ahsen, N., Nubukpo, P., Otto, M., et al. (2009) Dementia and Cognitive Di sorders in Elderly Population Living in Benin Rural Areas, West Africa. Dementia and Geriatric Cognitive Disorders, 27, 34-41. <u>http://dx.doi.org/10.1159/000188661</u>
- [13] Longdon, A.R., Paddick, S.M., Kisoli, A., Dotchin, C., Gris, W.K., Dewhurst, F., et al. (2013) Dementia Prevalence in Rural Tanzania: A Community Cross-Sectional Research. International Journal of Geriatric Psychiatry, 28, 728-737. http://dx.doi.org/10.1002/gps.3880
- [14] El-Tallawy, H.N., Farghly, W.M., Shehata, G.A., Rageh, T.A., Hakeem, N.A., Abo-Elfetoh, N., et al. (2012) Dementia Prevalence in Kharga District Al, New Valley Governorate, Egypt. Neuroepidemiology, 38, 130-137. <u>http://dx.doi.org/10.1159/000335655</u>
- [15] Yusuf, A.J., Baiyewu, O., Sheikh, T.L. and Shehu, U.A. (2011) Prevalence of Dementia and Sub-Types of Dementia in a Northern Nigerian Elderly Population. *International Psychogeriatrics*, 3, 379-386. <u>http://dx.doi.org/10.1017/S1041610210001158</u>
- [16] Llibre-Rodriguez, J.J., Ferri, C.P., Acosta, D., Guerra, M., Huang, Y., Jacob, K.S., et al. (2008) 10/66 Dementia Research Group. Prevalence of Dementia in Latin America, India, and China: A Population-Based Cross-Sectional Sur-

vey. Lancet, 372, 464-474. http://dx.doi.org/10.1016/S0140-6736(08)61002-8

- [17] Stewart, R., Kim, J., Shin, S. and Yoon, J. (2003) Education and the Association between Vascular Risk Factors and Cognitive Function. A Cross-Sectional Study in Older Koreans with Cognitive Impairment. *International Psychogeriatrics*, 15, 27-36. <u>http://dx.doi.org/10.1017/S1041610203008731</u>
- [18] Letenneur, L., Gilleron, V., Commenges, D., Helmer, C., Orgogozo, J.M. and Dartigues, J.F. (1999) Are Sex and Educational L evel I ndependent Predictors of Dementia and A lzheimer's Disease? I ncidence Data from the P AQUID Project. *Journal of Neurology, Neurosurgery & Psychiatry*, 66, 177-183. <u>http://dx.doi.org/10.1136/jnnp.66.2.177</u>
- [19] Nitrini, R., Caramelli, P., Herrera, E., Bahia, V.S., Caixeta, L.F., Radanovic, M., *et al.* (2004) Incidence of Dementia in A Community-Dwelling Brazilian Population. *Alzheimer Disease and Associated Disorders*, **18**, 241-246.
- [20] Fratiglioni, L., Paillard-Borg, S. and Winblad, B. (2004) An Active and Socially Integrated Lifestyle in Late Life Might Protect against Dementia. *The Lancet Neurology*, 3, 343-353. <u>http://dx.doi.org/10.1016/S1474-4422(04)00767-7</u>
- [21] Van-Gelder, B,M., Tijhuis, M., Kalmijn, S., Giampaoli, S., Nissinen, A. and Kromhou, D. (2006) Marital Status and Living Situation During a 5-Year Period Are Associated With a Subsequent 10-Year Cognitive Decline in Older Men: The FINE Study. *The Journals of Gerontology*, 61, 213-219. <u>http://dx.doi.org/10.1093/geronb/61.4.P213</u>
- [22] Chen, C.H., Mizuno, T., Elston, R., Kariuki, M.M., Hall, K., Unverzagt, F., et al. (2010) Comparative Study to Screen Dementia and APOE Genotypes in an Ageing East African population. *Neurobiology Aging*, 31, 732-740. http://dx.doi.org/10.1093/geronb/61.4.P213
- [23] Shelley, B.P. and Al-Khabouri, J. (2007) Dementia Spectrum: Prevalence, Causes and Clinic Profile. A National Reference; Research in Oman Hospital. *Dementia and Geriatric Cognitive Disorders*, 24, 280-287. http://dx.doi.org/10.1159/000107494
- [24] Alladi, S., Mekala, S., Chadalawada, S.K., Jala, S., Mridula, R. and Kaul, S. (2011) Dementia Subtypes Divisions: A Study from an Indian Memory Clinic. *Dementia and Geriatric Cognitive Disorders*, **32**, 32-38. http://dx.doi.org/10.1159/000329862
- [25] Pendlebury, S.T. (2012) Dementia in Patients Hospitalized with Stroke: Rates, Time Course, and Clinico-Pathologic Factors. *International Journal of Stroke*, **7**, 570-581.
- [26] Altieri, M., Di-Piero, V., Pasquini, M., Gasparini, M., Vanacore, N., Vicenzini, E., et al. (2004) Delayed Post Stroke Dementia. A 4-Year Follow-Up Study. *Neurology*, 62, 2193-2197. http://dx.doi.org/10.1212/01.WNL.0000130501.79012.1A
- [27] Pasquier, F. and Leys, D. (1997) Post Stroke Dementia Mechanism. Blood Thrombosis Vessels, 9, 220-226.
- [28] Juan, D., Zhou, D.H., Li, J., Wang, J.Y., Gao, C. and Chen M.A. (2004) 2-Year Follow-Up Study of Cigarette Smoking and Risk of Dementia. *European Journal of Neurology*, **11**, 277-282. http://dx.doi.org/10.1212/01.WNL.0000130501.79012.1A
- [29] Rusanen, M., Kivipelto, M., Quesenberry, C.P. and Whitmer, R.A. (2011) Heavy Smoking in Midlife and Long-Term Risk of Alzheimer Disease and Vascular Dementia. *JAMA Internal Medicine*, 171, 333-339.
- [30] Dong, M.J., Peng, B., Lin, X.T., Zhao, J., Zhou, Y.R. and Wang, R.H. (2007) The Prevalence of Dementia in the People's Republic of China: A Systematic Analysis of 1980-2004 Studies. *Age Ageing*, 36, 619-624. http://dx.doi.org/10.1093/ageing/afm128

Abbreviation List

Abbreviations	Full meaning
CNHU-HKM	Centre National Hospitalier et Universitaire Hubert Koutoukou Maga
MMSE	Mini Mental State Examination (MMSE)
DSM-IV	Diagnostic and Statistical Manuel of mental disorders, 4th edition
AD	Alzheimer Disease
VD	Vascular Dementia
HIV	Human Immunodéficiency Virus



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