# Comparative Study of the Effects of Metformin and Garlic Extract on Hippocampal Na+/K+ ATPase, Ca<sup>2+</sup> ATPase and Glutamine Synthetase Activities in Type II Diabetic Wistar Rat

Abass Alao Safiriyu<sup>1</sup>, Ibrahim Semuyaba<sup>1</sup>, Sodiq Kolawole Lawal<sup>2</sup>, Muhammad Olanrewaju Buhari<sup>3</sup>, Emmanuel Ayikobua Tiyo<sup>1</sup>, Muhamudu Kalange<sup>1</sup>, Alfred Omachonu Okpanachi<sup>1</sup>, Miriam Nansunga<sup>1,4</sup>

Correspondence to: Abass Alao Safiriyu, alaoabass@gmail.com

 $\textbf{Keywords:} \ ATP ase, Diabetic \ Mellitus, \ Garlic, \ Glutamine \ Synthetase, \ Hippocampus, \ Metformin, \ Na^+/K^+ \ ATP ase$ 

Received: April 5, 2018 Accepted: September 26, 2018 Published: September 29, 2018

Copyright © 2018 by authors and Scientific Research Publishing Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

http://creativecommons.org/licenses/by/4.0/



Open Access

#### **ABSTRACT**

Diabetes mellitus has not ceased to be on rise in spite of the continuous research on its management. Brain dysfunction associated with Diabetes mellitus especially Type II has been the great concern. The aim of this study was to investigate the effect of insulin sensitizing drug metformin and ethanolic extract of garlic on membrane bound enzymes Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and glutamate-glutamine cycle enzyme, Glutamine Synthetase activities in the hippocampus of streptozotocin-Nicotinamide induced Type II Diabetic rats. Twenty four male wistar rats weighted 120 - 150 g were used and divided into four groups with six rats in each group. Group A was non-diabetic (Control) and Groups B, C and D were diabetic. Group B received no treatment (DNT) while Groups C and D were treated with 1000 mg/kg of ethanolic garlic extract (EGE) and 50 mg/kg of metformin (MET) respectively orally for three weeks. All the groups were fed on standard rat chow with water ad libitum. Blood glucose was monitored weekly. Animals were sacrificed and the brains were removed and hippocampi were carefully excised and homogenate were obtained. Homogenate was analyzed for Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and Glutamine Synthetase (GS) activities. MET and EGE significantly reduced the blood glucose levels. There was a significant increase in the activities of hippocampal Na+/K+ ATPase, Ca<sup>2+</sup> ATPase and GS in MET and

<sup>&</sup>lt;sup>1</sup>Department of Physiology, Faculty of Biomedical Sciences, Kampala International University, Ishaka, Uganda;

<sup>&</sup>lt;sup>2</sup>Department of Anatomy, St. Francis University College of Health and Applied Sciences, Ifakara, Tanzania;

<sup>&</sup>lt;sup>3</sup>Department of Anatomy, Faculty of Biomedical Sciences, Kampala International University, Ishaka, Uganda;

<sup>&</sup>lt;sup>4</sup>Department of Physiology, Faculty of Biomedical Sciences, St. Augustine International University, Kampala, Uganda

EGE when compared to DNT. The results suggest that both MET and EGE increase the activities of hippocampal Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and GS which were reduced by diabetes mellitus, thus garlic and metformin administration exhibiting neuroprotective effect during hippocampal-related disorders associated with diabetes mellitus.

#### 1. INTRODUCTION

Diabetes has become a global epidemic in the general population. The estimated number of African population that would be diabetic by 2030 according to World Health Organisation is approx. 18 million. Diabetes mellitus as a metabolic disease posed neuronal impairment, cognitive deficits and promote progressive neurodegeneration [1]. Insulin is one of the hormones that regulate the activity of Na<sup>+</sup>/K<sup>+</sup> ATPase. Complications associated with diabetes has been shown to alter the activity Na<sup>+</sup>/K<sup>+</sup> ATPase [2]. Study has shown decreased Na<sup>+</sup>/K<sup>+</sup> ATPase activity in animal models of T2DM [3]. Diabetes has been shown to affect neurotransmitter synthesis or release in several brain regions [4]. Ca<sup>2+</sup> ATPase regulates Ca<sup>2+</sup> pump activity which acts as a second messenger in the control of cellular processes that plays a central role in mediating neurosecretion. Inhibition of Ca<sup>2+</sup> ATPase activity has shown to increase intracellular concentration of Ca<sup>2+</sup> which in turn altered the signal transduction pathways and cellular fluidity and eventually resulted in cell death [5]. A decrease in Ca<sup>2+</sup> ATPase activity of diabetic brain has been reported [6]. Glutamatergic neurotransmission is important for hippocampal modulation of learning and memory processing [7]. The conversion of glutamate to glutamine by glutamine synthetase that takes place within the astrocytes, represents a key mechanism in the regulation of excitatory neurotransmission [8]. Study has shown that healthy neuronal cells require both intracellular and extracellular glutamine [9]. Development of diabetes-associated cognition decline in db/db mice has been suggested to be due to disturbance of glutamate-glutamine shuttling between neurons and astrocytes in hippocampus [10]. Metformin is an antidiabetic agent that is being widely used in the management of diabetes mellitus especially Type II. Several studies have shown that metformin improved learning and memory behaviours in diabetic model [11, 12], by protecting hippocampal neurons [13, 14], by activating AMPK and Na<sup>+</sup>/K<sup>+</sup> ATPase system [15]. Metformin treatment is neuroprotective against the detrimental effects of  $\beta$ -Amyloid and high fat diets on hippocampal synaptic plasticity [16]. Several studies have investigated the anti-diabetic potential of both fresh and aged garlic extract and reported it to have antihyperglycemia, hypolipidemic, renoprotective effects and its ability to scavenging many reactive oxygen species (ROS) in both animal models and humans [17-21]. Fresh and cooked but not aged garlic extracts has been shown to increase both short and long term memory in both diabetic male and female rats [22]. Ethanolic extract of garlic has previously been reported to enhance spatial working memory by increasing hippocampal Na<sup>+</sup>/K<sup>+</sup> and Ca<sup>2+</sup> ATPases activities in wistar rats [23] and improve memory in Type II diabetic rats by increasing Na<sup>+</sup>/K<sup>+</sup>, Ca<sup>2+</sup>ATPases and glutamine synthetase activities in the hippocampus [24]. This study aimed to compare the effect of metformin, a standard anti-diabetic drug and ethanolic extract of garlic on hippocampal Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and Glutamine synthetase activities in Type II Diabetic rat model. Our findings showed that both metformin and garlic increased the activities of these enzymes, though increased effect posed by metformin was more than that of garlic thus metformin may be a better anti-diabetic drug, among others in managing hippocampal-dependent dysfunction associated with diabetes mellitus.

## 2. MATERIALS AND METHODS

#### 2.1. Chemical

Streptozotocin was purchased from Sigma Aldrich (St. Louis, MO, USA). Nicotinamide and metformin were purchased from Nono Pharmaceutics Ltd. (Kampala, Uganda).

#### 2.2. Ethanol Extraction of Garlic

Extraction was done using cold maceration at the laboratory in the department of Pharmacology, Kampala International University Western Campus Uganda. Peeled garlic weighing 500 g was cut into small pieces and homogenized in 70 ml of cold sterile 0.9% NaCl. The paste material was suspended in 80% ethanol for 48 hours in air tight glass jar using a rubber stopper, and the suspension was shaken periodically for three times a day at 5 minute interval. After 2 days, the suspension was filtered using Whitman filter paper to remove residue. Filtration was repeated 3 times and clear filtrate was obtained. The filtrate was concentrated using rotary evaporator at a bath temperature of 40°C. The extract concentrate obtained was then transferred to a cornical flask and further evaporated in oven drier at 50°C to obtain ultimately a gel like mass for the study [23, 25].

#### 2.3. Animals

Male Wistar rats weighing (120 - 150 g) were used in this experiment. The animals were obtained from the Animal House of College of Medicine, Mbarara University of Science and Technology, Uganda. The animals were housed in a well-ventilated room maintained under standard conditions of light, feeding and temperature of research laboratory of Kampala International University Western Campus Uganda. The study was conducted in accordance with the standards established by the Guide for the Care and Use of Laboratory Animals.

#### 2.4. Induction of Diabetes

After acclimatization of animals for five days, diabetes mellitus type II was induced in rats fasted overnight on day six with a single injection of 60 mg/kg streptozotocin dissolved in 0.05 ml of citrate buffer (pH 4.5) intraperitoneally which was followed by administration of 120 mg/kg of nicotinamide dissolved in 0.5 ml of normal saline intraperitoneally 15 min later. Elevated levels of blood glucose (Hyperglycemia) was measured from a drop of tail blood of rats 3 days and a week after injection using One-Touch Ultra-Easy Glucometer (LifeScan, UK). Rats confirmed hyperglycemic with blood glucose concentration (>250 mg/dl) was used as diabetic rats for this study [24, 26].

#### 2.5. Grouping

The rats were divided into 4 groups (n = 6) as follows:

Group A: Non-diabetic Control-received normal saline 1 ml/kg body weight

Group B: Diabetic non-treated (DNT)-received normal saline 1 ml/kg body weight

Group C: Diabetic + garlic (EGE) received 1000 mg/kg body weight of ethanolic extract of garlic

Group D: Diabetic + metformin (MET)-received 50 mg/kg body weight of metformin

Administration or ally was carried out daily between 8 - 10 am for period of three weeks.

### 2.6. Measurement of Na+/K+ ATPase Activity

The hippocampal homogenates was analyzed for Na<sup>+</sup>/K<sup>+</sup> ATPase according to the method of Tirri *et al.* [27]. Assay medium used consist of (in mM) 30 Tris-HCl buffer (pH 7.4), 50 NaCl, 6 MgCl<sub>2</sub>, 5 KCl and 50 μg of protein in the presence and absence of ouabain, 0.1 EGTA, in a final volume of 350 μL. The reaction was started by the addition of ATP to a final concentration of 3 mM. After 30 min at 37 °C, the reaction was stopped by the addition of 50% (w/v) trichloroacetic acid (70 μL). The saturating substrate concentrations was used, and reaction was in linear with protein and time. Some controls was included in the assays for non-enzymatic hydrolysis of ATP. The Pi (amount of inorganic phosphate) released was quantified calorimetrically, as described Fiske and Subba [28], using 300 KH<sub>2</sub>PO<sub>4</sub> as reference standard. Specific Na<sup>+</sup>/K<sup>+</sup> ATPase activity was calculated from the overall activity (in the absence of ouabain) and was recorded as Pi/min/mg of protein in nmol [23].

## 2.7. Measurement of Ca2+ ATPase Activity

The method of Desaiah and Ho [29] was used to assay  $Ca^{2+}$  ATPase in hippocampal homogenates. Pi (Inorganic phosphates) was estimated by the method of [28]. The assay medium had a final volume of 200  $\mu$ L. It consisted of (in mM), 30 Tris-HCl, 100  $\mu$ g of protein in the presence or absence of 0.4 CaCl<sub>2</sub>, buffer (pH 7.4), 3 MgCl<sub>2</sub> and 0.1 EGTA. The reaction was started by the addition of ATP to a final volume of 3 mM. 60 min after at 37°C, the reaction was stopped by the addition of 50% (w/v), 70  $\mu$ L of trichloroacetic acid. Substrate concentrations was used, and reaction was in linear with time and concentration of protein. Some controls was included in the assays to assess non-enzymatic ATP hydrolysis. The Pi (concentration of inorganic phosphate) released was quantified colorimetrically, as described [28], using KH<sub>2</sub>PO<sub>4</sub> as a reference standard. The Ca<sup>2+</sup> ATPase activity was determined by subtracting the activity measured from absence of Ca<sup>2+</sup> (no added 0.1 mM EGTA and Ca<sup>2+</sup>) and expressed as Pi/min/mg protein in nmol [23].

#### 2.8. Measurement of Glutamine Synthetase Activity

The method of Rowe *et al.* [30] was used in the enzymatic assay of glutaminesynthetase. In this method, 0.1 mL homogenates solubilized in 140 mM KCl was added to 0.1 mL of the reaction mixture in mM and incubated for 15 min (37°C). The reaction was stopped by 0.4 mL addition of a solution containing (in mM): 370 ferric chloride, 200 TCA and 670 HCl. The absorbance of the supernatant was measured at 530 nm after centrifugation and standard quantities of ferric chloride reagent treated with c-glutamylhy-droxamate was compared to the absorbance generated. Results were expressed as percentages of the control condition [24].

#### 2.9. Statistical Analysis

All statistical analyses were performed using SPSS version 20 and GraphPad Prism software version 6 was for graphs. All values were presented as means  $\pm$  SEM (standard error of mean). ANOVA which was followed by Post Hoc Bonferroni test was done and statistically significant differences were accepted at p < 0.05.

#### 3. RESULTS

## 3.1. Effect of Metformin and Ethanolic Extract of Garlic on Hippocampal Na+/K+ ATPase Activity in Diabetic Rats

There was a decrease activity of hippocampal Na $^+$ /K $^+$  ATPase (µmol of pi liberated/min/mg protein) in diabetic control DNT when compared with normal rats. Na $^+$ /K $^+$  ATPase activity in the hippocampus of diabetic rats in groups C and D *i.e.* EGE and MET were found to be significantly (p < 0.001) higher when compared with the diabetic control rats. However, there was a significant (p < 0.05) increase in the activity of hippocampal Na $^+$ /K $^+$  ATPase in MET group when compared with EGE group (**Figure 1**).

# 3.2. Effect of Metformin and Ethanolic Extract of Garlic on Hippocampal Ca<sup>2+</sup> ATPase Activity in Diabetic Rats

There was a decrease activity of hippocampal  $Ca^{2+}$  ATPase (µmol of pi liberated/min/mg protein) in diabetic control DNT when compared with normal rats.  $Ca^{2+}$  ATPase activity in the hippocampus of diabetic rats in groups C and D *i.e.* EGE and MET were found to be significantly (p < 0.001) higher when compared with the diabetic control rats. There was also a significant (p < 0.05) increase in the activity of hippocampal  $Ca^{2+}$  ATPase in MET group when compared with EGE group (Figure 2).

# 3.3. Effect of Metformin and Ethanolic Extract of Garlic on Hippocampal Glutamine Synthetase Activity in Diabetic Rats

Figure 3 shows the significant (p < 0.05) decrease in the glutamine synthetase activity (mMol of

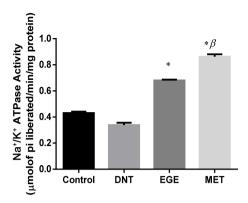


Figure 1. Effect of Metformin (MET) and ethanolic garlic extract (EGE) on hippocampal Na<sup>+</sup>/K<sup>+</sup> ATPase activity in diabetic rats. Data are expressed as mean  $\pm$  SEM (n = 6). \*P < 0.001 when compared with diabetic non-treated (DNT) group.  $\beta$  denotes significant difference when compared with EGE group.

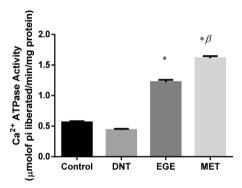


Figure 2. Effect of Metformin (MET) and ethanolic garlic extract (EGE) on hippocampal Ca<sup>2+</sup> ATPase activity in diabetic rats. Data are expressed as mean  $\pm$  SEM (n = 6). \*P < 0.001 when compared with diabetic non-treated (DNT) group.  $\beta$  denotes significant difference when compared with EGE group.

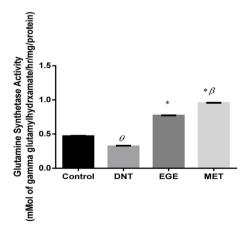


Figure 3. Effect of Metformin (MET) and ethanolic garlic extract (EGE) on hippocampal Glutamine Synthetase activity in diabetic rats. Data are expressed as mean  $\pm$  SEM (n = 6). \*P < 0.001 when compared with diabetic non-treated (DNT) group.  $\beta$  denotes significant difference when compared with EGE group.  $\theta$  denotes significant difference when compared with non-diabetic rats (Control).

gamma glutamylhydroxamate/hr/mg/protein) of hippocampus of diabetic non-treated group when compared with normal control group. Hippocampal glutamine synthetase activity was significantly (p < 0.001) higher in EGE and MET groups when compared with diabetic control group. Activity of glutamine synthetase in hippocampus was found to be significantly (p < 0.05) higher in MET group when compared with EGE group.

#### 4. DISCUSSION

In the present study, we investigated the effect of metformin, an insulin sensitizing drug and ethanolic extract of garlic on hippocampal membrane bound enzymes Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase activities and glutamate-glutamine cycle enzyme, glutamine synthetase activity in Type II diabetic rat model. Both metformin and garlic increased the activities of Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and glutamine synthetase in the hippocampus of diabetic rat. Diabetes mellitus showed reduction in the activities of these enzymes which is in line with the study of Vague et al. [3] for Na<sup>+</sup>/K<sup>+</sup> ATPase, Kamboj et al. [6] for Ca<sup>2+</sup> ATPase and Zheng et al. [10] for glutamine synthetase [24]. Administration of metformin and garlic to diabetic rats raised the level of activities of these enzymes in the hippocampus more than non-diabetic rats. However, metformin posed higher effects compared with garlic extract. Our previous studies reported increased hippocampal Na<sup>+</sup>/K<sup>+</sup> ATPase and Ca<sup>2+</sup> ATPase activities in normal rats [23] and in Type II diabetic rats [24] following garlic extract consumption. Diabetes in pregnancy has been shown to induce neuronal cell apoptosis in offspring hippocampus which could be normalized by controlling the maternal glycemic state with insulin treatment [31]. Study has shown that garlic administration during pregnancy and lactation protected the rat offspring against lead-induced neuronal damage in the hippocampus [32]. Metformin has been reported to reduce the decrease loss of neurons in the hippocampus of diabetic animals [14]. The increased activities of Na<sup>+</sup>/K<sup>+</sup> ATPase, Ca<sup>2+</sup> ATPase and Glutamine synthetase in the hippocampus by metformin revealed by our present study could be mechanism through which loss of neurons in hippocampus is prevented in diabetic rats. Metformin has been shown to recruit neural stem cells and enhance neural function by activating an a PKC-CBP pathway [33]. Neuroprotective properties of glucagon-like peptide-1, alone and in combination with metformin, in Type II diabetes has been shown by Lennox et al. [34] where metformin combination enhanced learning and memory, improved long-term potentiation in the hippocampal CA1 region with reduced hippocampal levels of 8-oxoguanine and glia fibriallary acidic protein. The results of our present study could be added to the established mechanisms through which metformin mediates its neuroprotective functions especially hippocampal—dependent cognitive functions.

#### 5. CONCLUSION

In conclusion, the present study suggested that the neuroprotective effect of metformin could be found by increasing the activities of  $Na^+/K^+$ ,  $Ca^{2+}$  ATPases and glutamine synthetase in the hippocampus of Type II diabetic rats, thus makes the metformin a better drug in the management of Type II diabetes that is associated with complication of cognitive dysfunction.

#### **CONFLICTS OF INTEREST**

The authors declare that there is no conflict of interest regarding the publication of this paper.

#### REFERENCES

- Sacai, H., Sasaki-Hamada, S., Sugiyama, A., Saitoh, A., Mori, K., Yamada, M. and Oka, J. (2014) The Impairment in Spatial Learning and Hippocampal LTD Induced through the PKA Pathway in Juvenile-Onset Diabetes Rats Are Rescued by Modulating NMDA Receptor Function. *Neuroscience Research*, 81-82, 55-63. <a href="https://doi.org/10.1016/j.neures.2014.02.002">https://doi.org/10.1016/j.neures.2014.02.002</a>
- 2. Stevens, M.J., Dananberg, J., Feldman, E.L., Lattimer, S.A., Kamijo, M., Thomas, T.P., Shindo, H., Sima, A.A. and Greene, D.A. (1994) The Linked Roles of Nitric Oxide, Aldose Reductase and (Na+- K+)- ATPase in the

- Slowing of Nerve Conduction in the Streptozotocin Diabetic Rat. *Journal of Clinical Investigation*, **94**, 853-859. https://doi.org/10.1172/JCI117406
- 3. Vague, P., Coste, T.C., Jannot, M.F., Raccah, D. and Tsimaratos, M. (2004) C-Peptide, Na<sup>+</sup>, K<sup>+</sup>-ATPase, and Diabetes. *Experimental Diabesity Research*, **5**, 37-50. <a href="https://doi.org/10.1080/15438600490424514">https://doi.org/10.1080/15438600490424514</a>
- 4. Welsh, B. and Wecker, L. (1991) Effects of Streptozotocin-Induced Diabetes on Acetylcholine Metabolism in Rat Brain. *Neurochemical Research*, **16**, 453-460. https://doi.org/10.1007/BF00965566
- 5. Aubier, M. and Viires, N. (1998) Calcium ATPase and Respiratory Muscle Function. *The European Respiratory Journal*, **11**, 758-766.
- Kamboj, S.S., Chopra, K. and Sandhir, R. (2009) Hyperglycemia-Induced Alterations in Synaptosomal Membrane Fluidity and Activity of Membrane Bound Enzymes: Beneficial Effect of N-Acetylcysteine Supplementation. Neuroscience, 162, 349-358. https://doi.org/10.1016/j.neuroscience.2009.05.002
- 7. Khakpai, F., Nasehi, M., Haeri-Rohani, A., Eidi, A. and Zarrindast, M.R. (2012) Scopolamine Induced Memory Impairment Possible Involvement of NMDA Receptor Mechanisms of Dorsal Hippocampus and/or Septum. *Behavioural Brain Research*, **231**, 1-10. https://doi.org/10.1016/j.bbr.2012.02.049
- 8. Szatkowski, M. and Attwell, D. (1994) Triggering and Execution of Neuronal Death in Brain Ischaemia: Two Phases of Glutamate Release by Different Mechanisms. *Trends in Neurosciences*, **17**, 359-365. https://doi.org/10.1016/0166-2236(94)90040-X
- 9. Chen, J. and Herrup, K. (2012) Glutamine Acts as a Neuroprotectant against DNA Damage, Beta-Amyloid and H2O2-Induced Stress. *PLoS ONE*, **7**, e33177. https://doi.org/10.1371/journal.pone.0033177
- 10. Zheng, Y., Yang, Y., Dong, B., Zheng, H., Lin, X., Du, Y., Li, X., Zhao, L. and Gao, H. (2016) Metabonomic Profile Delineate Potential Role of Glutamate-Glutamine Cycle in db/db Mice with Diabetes-Associated Cognitive Decline. *Molecular Brain*, **18**, 9-40. https://doi.org/10.1186/s13041-016-0223-5
- 11. Pintana, H., Apaijai, N., Pratchayasakul, W., Chattipakorn, N. and Chattipakorn, S.C. (2012) Effects of Metformin on Learning and Memory Behaviors and Brain Mitochondrial Functions in High Fat Diet Induced Insulin Resistant Rats. *Life Sciences*, **91**, 409-414. https://doi.org/10.1016/j.lfs.2012.08.017
- 12. Yuan, X., Chen, Y.B., Gan, D.N., Cheng, Y.F. and Xu, J.P. (2014) Metformin Improves Learning and Memory of Rats Induced by High-Fat Diet. *Military Medical Sciences*, **38**, 17-21.
- 13. Zhang, W.H., Liu, Z., Shi, F.G. and Chen, Z.M. (2012). Huanglianwendan Tang in Mice with Type 2 Diabetes and the Impact on the Learning and Memory and the Form of Hippocampal Nerve Cells. *Lishizhen Medicine and Materia Medica Research*, **23**, 948-949.
- 14. Oliveira, W.H., Nunes, A.K., Franca, M.E., Santos, L.A., Los, D.B., Rocha, S.W., Barbosa, K.P., Rodrigues, G.B. and Peixoto, C.A. (2016) Effect of Metformin on Inflammation and Short-Term Memory in Streptozoto-cin-Induced Diabetic Mice. *Brain Research*, **1644**, 149-160. https://doi.org/10.1016/j.brainres.2016.05.013
- 15. Mo, W. and Qian, G.F. (2014) Metformin for High Fat Diet Induced Insulin Resistance the Effect of Space Cognition, Learning and Memory and Cerebral Energy Metabolism. *Chinese Journal of Gerontology*, **34**, 2813-2815.
- 16. Asadbegi, M., Yaghmaei, P., Salehi, I., Ebrahim-Habibi, A. and Komaki, A. (2016) Neuroprotective Effects of Metformin against a β-Mediated Inhibition of Long-Term Potentiation in Rats Fed a High-Fat Diaet. *Brain Research Bulletin*, **121**, 178-185. <a href="https://doi.org/10.1016/j.brainresbull.2016.02.005">https://doi.org/10.1016/j.brainresbull.2016.02.005</a>
- 17. Anwar, M. and Meki, A. (2003) Oxidative Stress in Streptozotocin-Induced Diabetic Rats: Effects of Garlic Oil and Melatonin. *Comparative Biochemistry and Physiology Part A*, **135**, 539-547. https://doi.org/10.1016/S1095-6433(03)00114-4
- 18. Thomson, M., Al-Amin, Z.M., Al-Qattan, K.K., Shaban, L.H. and Ali, M. (2006) Anti-Diabetic and Hypolipi-daemic Properties of Garlic (*Allium sativum*) in Streptozotocin Induced Diabetic Rats. *International Journal of*

- Diabetes and Metabolism, 15, 108-115.
- 19. Drobiova, H., Thomson, M., Al-Qattan, K., Peltonen-Shalaby, R., Al-Amin, Z. and Ali, M. (2011) Garlic Increases Antioxidant Levels in Diabetic and Hypertensive Rats Determined by a Modified Peroxidase Method. *Evidence-Based Complementary and Alternative Medicine*, **2011**, Article ID: 703049.
- Liu, C.T., Hse, H., Lii, C.K., Chen, P.S. and Sheen, L.Y. (2005) Effects of Garlic Oil and Diallyl Trisulfide on Glycemic Control in Diabetic Rats. *European Journal of Pharmacology*, 516, 165-173. https://doi.org/10.1016/j.ejphar.2005.04.031
- 21. Shiju, T.M., Rajesh, N.G. and Viswanathan, P. (2013) Renoprotective Effect of Aged Garlic Extract in Strepto-zotocin-Induced Diabetic Rats. *Indian Journal of Pharmacology*, **45**, 18-23.
- 22. Sarkaki, A., Valipour, S.C., Farbood, Y., Mohammad, S., Mansouri, T., Naghizadeh, B. and Basirian, E. (2013) Effects of Fresh, Aged and Cooked Garlic Extracts on Short- and Long-Term Memory in Diabetic Rats. *Avicenna Journal of Phytomedicine*, **3**, 45-55.
- 23. Safiriyu, A.A., Semuyaba, I., RemónFigueredo, N., Etibor, T.A. and Ajibola, M.I. (2017) Garlic Extract (*Allium sativum*) Enhances Spatial Working Memory in Wistar Rats: Involvement of Hippocampal Na<sup>+</sup>/K<sup>+</sup> ATPase and Ca<sup>2+</sup> ATPase Activities. *Journal of Behavioral and Brain Science*, 7, 31-40. https://doi.org/10.4236/jbbs.2017.72004
- 24. Semuyaba, I., Safiriyu, A.A., Tiyo, E.A. and RemónFigueredo, N. (2017) Memory Improvement Effect of Ethanol Garlic (*A. sativum*) Extract in Streptozotocin-Nicotinamide Induced Diabetic Wistar Rats Is Mediated through Increasing of Hippocampal Sodium-Potassium ATPase, Glutamine Synthetase, and Calcium ATPase Activities. *Evidence-Based Complementary and Alternative Medicine*, **3**, 1-7. https://doi.org/10.1155/2017/3720380
- 25. Homayounfar, H., BaluchnejadMojarad, T., Roghani, M., Hosseini, M. and Kamalinejad, M. (2003) Effect of Aqueous Garlic (*Allium sativum* L.) Extract on Acetylcholine and Isosorbide-Induced Relaxation of Isolated Aorta in Rat. *Iranian Biomedical Journal*, 7, 23-27.
- 26. Marudamuthu, A.S. and Leelavinothan, P. (2008) Emerging Targets Effect of Pterostilbene on Lipids and Lipid Profiles in Streptozotocin-Nicotinamide Induced Type 2. *Journal of Applied Biomedicine*, **6**, 31-37.
- 27. Tirri, R., Lagrspetz, K.Y.H. and Kohomen, J. (1973) Temperature Dependence of the ATPase Activity in Brain Homogenates during the Postnatal Development of Rat. *Comparative Biochemistry and Physiology*, **44**, 473-480.
- 28. Fiske, C.H. and Subba Row, Y. (1925) The Colorimetric Determination of Phosphates. *The Journal of Biological Chemistry*, **66**, 375-400.
- Desaiah, D. and Ho, I.K. (1979) Effect of Acute and Continuous Morphine Administration on Catecholamine-Sensitive Adenosine Triphosphatase in Mouse Brain. *Journal of Pharmacology and Experimental Thera*peutics, 208, 80-85.
- 30. Rowe, W.B., Ronzio, R.A., Wellner, V.P. and Meister, A. (1970) Glutamine Synthetase (Sheep Brain) Methods. *Enzymology*, **17**, 900-910. <a href="https://doi.org/10.1016/0076-6879(71)17304-1">https://doi.org/10.1016/0076-6879(71)17304-1</a>
- 31. Lotfi, N., Hami, J., Hosseini, M., Haghir, D. and Haghir, H. (2016) Diabetes during Pregnancy Enhanced Neuronal Death in the Hippocampus of Rat Offspring. *International Journal of Developmental Neuroscience*, **51**, 28-35. https://doi.org/10.1016/j.ijdevneu.2016.04.009
- 32. Sadeghi, A., Ebrahimzadeh Bideskan, A.R., Alipour, F., Fazel, A.R. and Haghir, A. (2013) The Effect of Ascorbic Acid and Garlic Administration on Lead-Induced Neural Damage in Rat Offspring's Hippocampus. *Iranian Journal of Basic Medical Sciences*, **16**, 157-164.
- 33. Wang, J., Gallagher, D., DeVito, L.M., Cancino, G.I., Tsui, D., He, L., Keller, G.M., Frankland, P.W., Kaplan, D.R. and Miller, F.D. (2012) Metformin Activates an Atypical PKC-CBP Pathway to Promote Neurogenesis and

- Enhance Spatial Memory Formation. Cell Stem Cell, 11, 23-35. https://doi.org/10.1016/j.stem.2012.03.016
- 34. Lennox, R., Porter, D.W., Flatt, P.R., Holscher, C., Irwin, N. and Gault, V.A. (2014) Comparison of the Independent and Combined Effects of Sub-Chronic Therapy with Metformin and a Stable GLP-1 Receptor Agonist on Cognitive Function, Hippocampal Synaptic Plasticity and Metabolic Control in High-Fat Fed Mice. *Neuro-pharmacology*, **86**, 22-30. https://doi.org/10.1016/j.neuropharm.2014.06.026