

Biodiesel Production from Oils Extracted from Date Pits

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Abstract

Biodiesel production had received a considerable attention as a green, nontoxic and renewable alternative to petroleum diesel. To avoid using vegetable oils, which are expensive and compete with food, as feedstock, waste oils have been proposed. However, these waste materials contain a large amount of free fatty acids that complicates the production process. In this work, biodiesel production using an alternative feedstock; namely oils from date-pits, has been investigated. These oils have the same favorable features of straight oils, and at the same time are considered waste, since they are extracted from a waste material. The yield of oils extracted using *n*-hexane in a Soxhlet apparatus was compared to that of oils extracted using methanol-chloroform solvent mixture. The extracted oils were then converted to biodiesel via transesterification with methanol in presence of Novozym^{*}435 or Eversa^{*}Transform. The highest oils extraction yield of 11.7%, per dry weight sample, was obtained using Soxhlet extraction apparatus compared to 8.9% using methanol-chloroform mixture. The highest biodiesel production yield was 30% of the oil used, achieved after 6 hours using Novozym^{*}435 at 40°C, 5:1 methanol to oil molar ratio and 10wt% enzyme loading. By using a chemical alkaline catalyst, NaOH, at the same conditions, the yield was 27%.

Keywords

Date Pits, Soxhlet, Lipase, Biodiesel, Enzymatic Reaction

1. Introduction

Biodiesel, a mixture of fatty acid methyl esters (FAME), is commonly produced from oil-rich feedstocks, such as vegetable oils, animal fats, waste cooking oils

and microalgae lipids. Commercially, the production takes place via transesterification using alkali catalysts. Although high yields have been reported, the main drawback of such process is in the soap formation when a feedstock with high free fatty acids (FFAs) content is used. Acid pre-treatment therefore become necessary prior to alkali transesterification. Lipases, on the other hand, can catalyze both FFAs esterification and triglycerides transesterification to produce high quality biodiesel [1] [2]. This process has been extensively studied using Novozym[®]435, which is the immobilized form of *Candida antarctica* lipase B.

The selection of the proper feedstock depends on the contents of water and FFAs present in the extracted oil. Date palm trees are commonly cultivated in United Arab Emirates, and used for date production [3]. Date seeds (pits) are the main waste products from the date industry, which are currently used either as animal feed or directly burned for heating. Utilization of date pits in the production of high value products has received considerable attention. For example, they have been used to produce activated carbons for wastewater treatment [4] [5] [6] [7].

It was reported that about 10% of pits chemical composition is oil, with less than 1% FFAs [8] [9] [10], which suggests their suitability to be used for biodiesel production. Although several studies had considered the date-pits characterization, Jamil *et al.* [11] was one of a few researchers who tested the production of biodiesel from oils extracted from date pits using potassium hydroxide (KOH). In the present study oil was extracted from date pits using a single solvent, *n*-hexane, in a dynamic Soxhlet system, and the extraction yield was compared to that achieved using methanol-chloroform solvent mixture in a static extraction process. The extracted oil was used then as a feedstock for biodiesel production catalyzed by lipase, and the production yield was compared to that achieved using conventional alkaline catalyst, sodium hydroxide (NaOH). A comparison between immobilized and free lipase has also been considered.

2. Materials and Methods

2.1. Chemicals and Enzymes

All chemicals used in this work were of analytical grade and purchased from *Sigma* Aldrich, US. The reference standards of FAMEs containing; 4% myristic acid, 10% palmitic acid, 6.0% stearic acid, 35% oleic acid, 36.0% linoleic acid, 5.0% archidonic acid and 4% behenic acid were also obtained from Sigma Aldrich, US. High purity gases of helium (He), hydrogen (H₂) and zero-air were supplied by Abu-Dhabi Oxygen Oxygen Company, UAE. Novozym^{*}435 of 10,800 PLU·g⁻¹ (propyl laurate units per gram) enzyme activity and 2% water content was purchased from Novozyms A/S, Denmark. Samples of Eversa^{*}Transform lipase formulation, produced by submerged fermentation of a genetically modified *Aspergillus oryzae* of 100,000 PLU·g⁻¹, which is close to that of Novozym^{*}435, was a kind gift from the same supplier. Both enzymes were stored below 8°C and above 0°C according to the supplier's instructions.

2.2. Date Pits Approximate Analysis

The proximate analysis of the date pits was performed using Thermo Gravimetric Analyzer (TGA-Q50). The sample approximate analysis provides information about the percentage of moisture, volatiles, fixed carbon and Ash present in the sample. After warming up and tarring procedures, a 12 mg date pits sample was tested under constant nitrogen purging at 100 mL·min⁻¹. A custom five steps heating cycle was used: Step 1: temperature was held constant at 25°C for 5 minutes, Step 2: temperature was ramped to 125°C in 5 minutes, Step 3: temperature was held at 125°C for 5 minutes, Step 4: temperature was ramped to 900°C in 38.75 minutes and Step 5: temperature was held at 900°C for 15 minutes.

2.3. Oil Extraction

Fruits of mature palm date, Khalas type, were obtained from local palm trees. Date pits were manually isolated from the fruits, socked and washed many times with tap water to remove adhering date flesh, followed by sun drying for two days. The samples were then grinded into small particles using a grinder (AR1043 Moulinex, China) for a short period of 15 s, and then screened based on their particle size using sieve analysis (USA Standard Testing Sieves, W.S. Tyler Incorporated, USA) of 2.36, 0.5 and 0.35 mm mesh sizes.

Two solvent extraction methods were investigated to extract the oils, namely Soxhlet apparatus using *n*-hexane and methanol-chloroform solvent mixture. The weight of extracted oils was determined gravimetrically and used to calculate the extraction yield, as shown in Equation (1). All tests were performed in triplicate, and the results reported were the average values.

$$\%Y = \frac{m_{oil}}{m_{sample}} \times 100 \tag{1}$$

where, m_{ail} is the extract weight and m_{sample} is the sample dry weight.

1) Soxhlet Apparatus

The dynamic extraction of oils was performed using 75 ml n-hexane in a Soxhlet apparatus (Lab-line instruments, Inc., Melrose Park, Illinois, USA). The ground and dried date pits (3 g) were placed in a porous cellulose thimble, which was initially dipped into *n*-hexane so that the solvent diffused completely into the sample. After 10 min, the thimble was subjected to the system that contained *n*-hexane at 80°C for 12 h. At the end of the extraction process, the oils dissolved in *n*-hexane were recovered using a rotary evaporator at 40°C.

2) Methanol-Chloroform

Oils were also extracted from 1 g of biomass using a mixture of chloroform and methanol at 1:1 v/v, according to Bligh and Dyer [12] method. The samples were mixed with 40 ml of mixture, and the extraction was allowed to equilibrate for 30 min, under continuous ultra-sonication using Branson Sonifier 450 (Danbury, USA). This was followed by centrifuging the mixture at 3000 r·min⁻¹ for 5 min using IEC CL31 multispeed centrifuge (Thermo Scientific, USA). The upper aqueous layer was discarded and the lower chloroform layer containing the oils



was separated from the solid biomass. The oils were then recovered from the chloroform layer using the rotary evaporator at 40°C and the oils is the solids biomass left at the bottom of extraction tube were extracted using the same procedure two more times.

2.4. Fatty Acids Profile of Date-Pits Oils

Oil profile was determined after derivatization to esters by methylation with 14% BF_3 -methanol mixture, according to Rule [13] protocol. 0.25 ml of 14% BF_3 -methanol mixture were added to 10 mg of extracted oils, and the mixture was heated in a water bath for 45 min at 100°C and then cooled-down, followed by addition of 1 ml of distilled water and 2 ml of *n*-hexane, eddying and centrifuging at a low speed. The upper phase containing esters was collected and *n*-hexane was evaporated. Produced esters were then analyzed using Gas Chromatograph.

2.5. Transesterification

The production of biodiesel production was carried out using oils extracted via Soxhlet apparatus. Extracted oils (1 ml) were transesterified with methanol at 40°C and 5:1 methanol to oil molar ratio. The reaction vials were placed in a temperature controlled water bath (Daihan Labtech, Korea) set at 40°C at 90 r·min⁻¹. In the alkali based transesterification, the extracted oils were mixed with methoxide solution, prepared by dissolving 0.1 g (correspond to 10% catalysts) of NaOH in methanol. In enzymatic reactions, the 0.1 g NaOH was replaced with 0.1 g of enzyme, either Novozym^e435 or Eversa[®]Transformlipase. Due to methanol negative effect on lipase activity, the specified amount of oil and enzyme were first added to the reaction vial, followed by the methanol addition. After six hours, *n*-hexane was added to dissolve the produced biodiesel, which was then separated by centrifugation in at 3000 r·min⁻¹ for 5 min.

2.6. Analysis

Fatty acid compositions of extracted oils via Soxhlet was determined using Varian Gas Chromatography (GC) (CP-3800, USA) equipped with flame ionization detector (FID) and CP-Sil 88 FAME capillary column (100 m \times 0.25 mm \times 0.2 μ m, Varian, USA) with auto-injector (CP 8410. Varian, USA). The column temperature was programmed from 150°C to 220°C at 4°C/min and the injector and detector temperature were set at 260°C.

The amount of produced FAME during transesterification reactions were also determined using the GC. The identification and quantification of the products were achieved by comparing them with FAME standards. Then the biodiesel yield % was found using Equation (2).

$$%Y_{FAME} = \frac{m_{FAME}}{m_{oil}} \times 100$$
 (2)

where, m_{FAME} is the total produced esters found in produced sample.

3. Results and Discussion

3.1. TGA Results

The TGA results can be used to determine the proximate analysis of the date pits and can also be used to quantify the oils content. Figure 1 shows the percentage of weight loss as a function of temperature and its corresponding thermogravimetric derivative analysis (DTG).

The pyrolysis underwent three steps: 1) dehydration, 2) devolatilization and 3) solid decomposition. The first dehydration stage began from the room temperature to around 220°C, where moisture was released and weight loss of around 3% was recorded. This low value is expected, as dried date pits were used. The major biomass reduction was observed in the second devolatilization stage at temperatures between 220°C and 500°C, with weight loss of about 75%. It is found that the devolatilization went through the two steps demonstrated by the two peaks. The first peak, at around 300°C, is attributed to protein and carbohydrate devolatilization, and shows that their combined content was (55% - 3%)= 52%). The second peak, at around 390°C, is attributed to lipids, and shows that its content was (65% - 55% = 10%). This result agrees with the gravitational determination of the oils in the date pits. In the third solids decomposition stage, the carbonaceous solids were degraded.

3.2. Date-Pits Oil Extraction Yield and Profile

For comparison, the dynamic extraction in the Soxhlet apparatus using *n*hexane, which is commonly used as a baseline to evaluate extraction processes efficiency was compared to that of static extraction using solvents mixture, namely methanol-chloroform (1:1). Sample of different sizes were tested. It is important to note that the in the static mode, the extraction was assisted by ultra-sonication, which was not the case in Soxhlet tests. Table 1 shows the percentage of extracted oils from the different particle sizes. The performances of

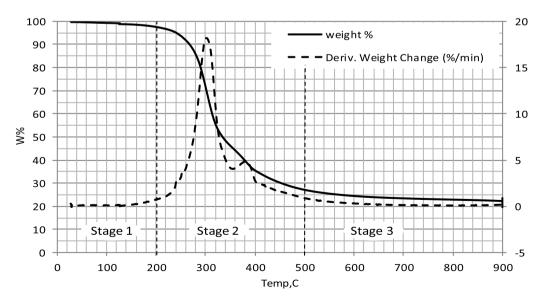


Figure 1. TGA and DTG curves of date pits.



both methods were comparable, with Soxhlet extraction showing better performance for smaller sizes samples. When large particles were used, Soxhlet extraction was less effective than the static method. This was expected as ultrasoniccation enhanced the extraction efficiency by a caustic-induced cavitation that can disrupt the sample and facilitate solvent penetration, and therefore enhanced the contact between the solvent and the solute. The comparison revealed that dynamic extraction was 18% and 31% higher than the static mode when medium and small particle size samples were used, respectively. By decreasing samples particle sizes, the extraction yield increased dramatically for more than 3 folds, reaching 11.67% when n-hexane was used with small size particles, compared to the large particles. The effect was less when the static mode was tested, where only 8.9% yield of oil was obtained with small size samples (equivalent to 75% of total extractable oils using *n*-hexane). Generally, by reducing particle size, the increased specific surface area enhances oils exposure to the solvent, therefore dissolving and extracting more oils. The results found in this work are in agreement with previously reported oil contents in date pits [8] [9] [10] [11].

The composition of extracted oils was determined, and the results are shown in **Table 2**. As shown, extracted oils contained mainly of trans-9-elaidic and linoleic acids. These together accounts for more than 45% of the total fatty acids present in the oil. Palmitic acid is the second highest acid in accounting for around 20% followed by myristic and stearic acids. The results found in this work were in agreement with previous results on date pits [10] [14] and other potentials biodiesel feed stocks, such as microalgae lipids [15].

 Table 1. Effect of samples particle size on extracted oil yield.

Size of sample (PS)	Extraction yield	
	Soxhlet	Methanol:Chloroform
Large (PS > 2.36 mm)	2.38%	3.19%
Medium (2.36 mm > PS > 0.5 mm)	7.95%	6.72%
Small (0.5 mm > PS > 0.35 mm)	11.67%	8.9%

Table 2. Date-pits oil composition.

Fatty Acid	Composition, w/w%	
Unidentified	11.3	
Myristic	9.4	
Palmitic	19.8	
Stearic	8.6	
Trans-9-Elaidic	32.6	
Linoleic	14.0	
Linolenic	2.9	
Archidic	0.2	
Behenic	1.1	

3.3. Biodiesel Production

The production of biodiesel from extracted date pits oils was tested using different catalysts; namely NaOH, Eversa^{*}Transform and Novozym^{*}435. Reactions were carried out with methanol to oil molar ratio of 5:1 for six hours. Produced FAMEs were dissolved in 10 ml *n*-hexane before being analyzed. Comparable yields were obtained when the reaction was carried using Novozym^{*}435 and NaOH. As shown in **Figure 2**, the highest yield, reaching 30%, was achieved in the presence of Novozym^{*}435. A slightly lower yield of 27% was obtained in NaOH. By using Eversa^{*}Transform a yield of only 11% was achieved.

Despite the comparable yields obtained using Novozym^{*}435 and NaOH, the profiles of the produced esters were different, as shown in **Figure 2**. It was found that the selectivity of NaOH was high toward transesterifyingtrans-9-elaidic acids compared to other acids. Whereas Novozym^{*}435 equally transestrified most acids present in the oil sample. Similar observation was also found using the liquid Eversa^{*}Transform lipase. It is important to mention that this is the first work to test the use of this lipase in biodiesel production

4. Conclusion

The potential of using waste date-pits as a new source for oil to be used in biodiesel production was examined. From grinded date pits, in size ranging of 0.5 -0.35 mm, maximum extraction yields of 11% and 7.9% were achieved using Soxhlet and methanol-chloroform solvent mixture extraction methods, respectively. The effectiveness of using lipase from different sources in producing the biodiesel was investigated, and compared to that of conventional alkaline catalyst. The highest biodiesel yield of 30% was obtained at 40°C, 5:1 methanol to molar ratio after 6 hrs using Novozym[®]435. The possibility of using liquid

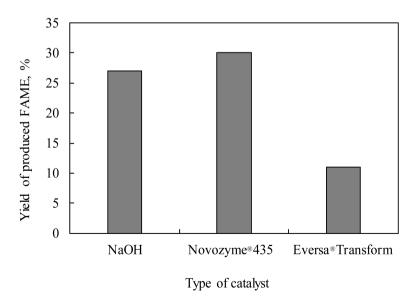


Figure 2. Biodiesel production yield after six hours from oils extracted from date pits using different enzymes.



Eversa^{*}Transform lipase was also examined; however, a lower yield was obtained.

References

- Taher, H., Al-Zuhair, S., Al-Marzouqi, A.H., Haik, Y. and Farid, M.M. (2011) A Review of Enzymatic Transesterification of Microalgal Oil-Based Biodiesel Using Supercritical Technology. *Enzyme Research*, 2011, 25 p.
- [2] Yadav, G.D. and Devi, K.M. (2004) Immobilized Lipase-Catalysed Esterification and Transesterification Reactions in Non-Aqueous Media for the Synthesis of Tetrahydrofurfuryl Butyrate: Comparison and Kinetic Modeling. *Chemical Engineering Science*, **59**, 373-383. <u>https://doi.org/10.1016/j.ces.2003.09.034</u>
- Chisti, Y. (2007) Biodiesel from Microalgae. *Biotechnology Advances*, 25, 294-306. https://doi.org/10.1016/j.biotechadv.2007.02.001
- [4] Ahmed, M.J. (2016) Preparation of Activated Carbons from Date (*Phoenix Dactyli-fera* L.) Palm Stones and Application for Wastewater Treatments: Review. *Process Safety and Environmental Protection*, **102**, 168-182. https://doi.org/10.1016/j.psep.2016.03.010
- [5] El-Naas, M.H., Al-Zuhair, S. and Alhaija, M.A. (2010) Reduction of Cod in Refinery Wastewater through Adsorption on Date-Pit Activated Carbon. *Journal of Hazard*ous Materials, 173, 750-757. <u>https://doi.org/10.1016/j.jhazmat.2009.09.002</u>
- [6] Reddy, K.S.K., Al Shoaibi, A. and Srinivasakannan, C. (2012) Activated Carbon from Date Palm Seed: Process Optimization Using Response Surface Methodology. *Waste and Biomass Valorization*, 3, 149-156. <u>https://doi.org/10.1007/s12649-011-9104-4</u>
- [7] Girgis, B.S. and El-Hendawy, A.-N.A. (2002) Porosity Development in Activated Carbons Obtained from Date Pits under Chemical Activation with Phosphoric Acid. *Microporous and Mesoporous Materials*, **52**, 105-117. https://doi.org/10.1016/S1387-1811(01)00481-4
- [8] Hamada, J.S., Hashim, I.B. and Sharif, F.A. (2002) Preliminary Analysis and Potential Uses of Date Pits in Foods. *Food Chemistry*, 76, 135-137. <u>https://doi.org/10.1016/S0308-8146(01)00253-9</u>
- [9] Nehdi, I., Omri, S., Khalil, M.I. and Al-Resayes, S.I. (2010) Characteristics and Chemical Composition of Date Palm (*Phoenix Canariensis*) Seeds and Seed Oil. *Industrial Crops and Products*, **32**, 360-365. <u>https://doi.org/10.1016/j.indcrop.2010.05.016</u>
- [10] Besbes, S., Blecker, C., Deroanne, C., Drira, N.-E. and Attia, H. (2004) Date Seeds: Chemical Composition and Characteristic Profiles of the Lipid Fraction. *Food Chemistry*, 84, 577-584. <u>https://doi.org/10.1016/S0308-8146(03)00281-4</u>
- [11] Jamil, F., Al-Muhtaseb, A.a.H., Al-Haj, L., Al-Hinai, M.A., Hellier, P. and Rashid, U.
 (2016) Optimization of Oil Extraction from Waste "Date Pits" for Biodiesel Production. *Energy Conversion and Management*, **117**, 264-272. https://doi.org/10.1016/j.enconman.2016.03.025
- Bligh, E.G. and Dyer, W.J. (1959) A Rapid Method of Total Lipid Extraction and Purification. *Canadian Journal of Biochemistry and Physiology*, **37**, 911-917. <u>https://doi.org/10.1139/o59-099</u>
- [13] Rule, D.C. (1997) Direct Transesterification of Total Fatty Acids of Adipose Tissue, and of Freeze-Dried Muscle and Liver with Boron-Trifluoride in Methanol. *Meat Science*, 46, 23-32. <u>https://doi.org/10.1016/S0309-1740(97)00008-9</u>
- [14] Al-Hooti, S., Sidhu, J.S. and Qabazard, H. (1997) Physicochemical Characteristics of

Five Date Fruit Cultivars Grown in the United Arab Emirates. Plant Foods for Human Nutrition, 50, 101-113. https://doi.org/10.1007/BF02436030

[15] Taher, H., Al-Zuhair, S., Al-Marzouqi, A.H., Haik, Y. and Farid, M. (2014) Effective Extraction of Microalgae Lipids from Wet Biomass for Biodiesel Production. Biomass and Bioenergy, 66, 159-167. https://doi.org/10.1016/j.biombioe.2014.02.034

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