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Chemical and Microbiological Characterization of Egyptian Cultivars for Some Spices and Herbs Commonly Exported Abroad

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Abstract

The present study highlighted some chemical, nutritional, microbiological and essential oil characteristics of the Egyptian traditional cultivars for seeds of cumin (Cuminum cyminum) and coriander (Coriandum sativum) spices as well as for basil whole herb (Ocimum basilicum) collected from different Egyptian export centers as being ready for export. The found values for humidity in dry seeds of cumin (7.4%) and coriander (6.4%) as well as total ash and ash insoluble in acid (in cumin 7.7% and 0.74%, but in coriander 5.3% and 0.55%, respectively) were lower than the maximum limits indicated by the Egyptian Specification Standards (ES) and by International Standards Organization (ISO) for cumin and coriander seeds. Analysis of essential minerals in seed spices and herbs indicated that they were are rich in K, Ca, Na, Fe and Zn. Total bacterial count was low content in seeds of cumin and coriander as well as fresh whole basil herb. The microbiological load in all tested seed spices and herbs was found lower than those indicated by the ES and ISO for cumin and coriander seeds. Yields in hydro-distilled essential oils (EOs) were the highest in cumin seeds (3.762%), while both coriander and basil herb had lower amounts (0.285% and 0.686%, respectively). EOs contents were found higher than the maximum limits for cumin (1.5% - 2.5%) on dry weight basis), but the within the limits for coriander (0.1% - 0.5%) on dry weight basis) as indicated by the ES and ISO for cumin and coriander seed oils. Gas chromatography of extracted EOs from seeds of cumin and coriander as well as basil herbs indicated the presence of 41, 35 and 47 compounds, respectively, where cuminaldehyde was the major component in cumin volatiles, but was linalool in volatiles of both coriander seeds and basil herbs. EOs of basil herbs grown in Egypt, were of the high linalool-chemotype which were characterized by high contents of linalool and relatively lower amounts of eugenol. However, the major compounds in the

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three tested EOs from seeds or herbs grown in Egypt are in accordance with literature reports from different parts of the world. Volatile oil components in EOs of the three tested Egyptian spices and herbs were classified into groups, based on the relative area (%). The proportion of the major and the other main components in EOs from seeds of cumin and coriander cultivars were within the ranges indicated by both the ES and ISO for cumin seed oils (cuminaldehyde between 15% - 46%) and for coriander seed oils (linalool between 65% - 78%). No Egyptian Specification Standards are established yet for fresh basil herbs and for basil oil of Linalool chemotype, but only present for basil oil of methylchavicol-chemotype.

Keywords

Cumin, Coriander, Basil, GC-Fractionation, Chemical, Physical, Microbiological Analysis, Spices, Herbs

1. Introduction

Spices and herbs, commonly known as aromatic plants, are an important group of agricultural commodities being used by many civilizations all over the world to add flavor, taste, nutritional values and increase shelf life to food as well as to heal various physical, mental, emotional problems and to restore human health [1]. However, each spice or herb is characterized by a peculiar quail-quantitative composition for its essential oil and all of these oils contain compounds with established biological activity [2]. Cumin (*Cuminum cyminum*, family Umbelliferae) is among the large number of spices used to flavor foods and beverages in the world, especially India and Mediterranean regions, and occupies a place of prominence [3]. Cumin seeds possess an aromatic odor and have a spicy and bitter taste and largely used in the Egyptian kitchen and is locally known as "Kammoun" [4]. Coriander (*Coriandum sativum* L., family Apiaceae) is among many of the aromatic plants that actually gathered when they have finished flowering, with the leaves being referred to as an herb, and the dried seeds as a spice [5]. Although plant can be grown throughout the year, coriander is processed to increase its palatability, profitability and facilitate international trade where processing of fruits and leaves of coriander is the best way to preserve this herb [1].

Basil (*Ocimum basilicum* L., Lamiaceae family) or sweet basil is one of the most important well-known herbs to many cultures [6], and is a very versatile and popular annual herb with abundantly aromatic leaves which are used fresh or dried [7]. Basil is called as "reyhan" in Turkish [8]. The word "Reyhan" has been mentioned in two suras, Ar-Rehman and Al-Waqia (once in each) of the holy Qura'n [9]. In fact, basil is multipurpose plant species used as a decorative, seasoning, medicinal plant and used to relish many kinds of foods [10] [11].

Besides being used as spices and herbs in Egyptian trade, cumin, coriander and basil are widely cultivated for export abroad due to the continuous and increased demands for their products and their distinguished economic importance throughout the world [12]. They are greatly valued as medicinal plants and have attracted enormous attention of researchers worldwide to experimentally validate their pharmacological activities and the therapeutic use as they have documented in several indigenous healing systems [13].

Therefore, the present study was lunched to highlight some chemical, nutritional and microbiological characteristics for seeds and essential oils for the Egyptian traditional cultivars of cumin and coriander spices as well as for basil herb collected from different Egyptian export centers as being ready for export.

2. Material and Methods

2.1. Materials

Cumin seeds (*Cuminum cyminum*) and coriander seeds (*Coriandum sativum*) were obtained from an Egyptian local store market for bulk and retail spices commerce and their export (Harraz market for Seeds and Pesticides, Bab El-Khalksquare, Cairo, Egypt).

Basil herbs (*Ocimum basilicum*) were obtained from Egyptian Baladi basil cultivar which is a hybrid between native and American basil types at ratio of 2:1. Such cultivar was grown a private farm (run with organic plantation system) at Abshoway Village in El-Fayoum Governorate, Egypt.

2.2. Preparation of Spices and Herbs

Spices of cumin and coriander seeds were already prepared into dry seed form in the same aforementioned purchasing place, as usually practiced in Egypt for the preparation of dry spices for export abroad. Basil plants were cultivated, collected and the aerial parts of herbs were harvested at the start of flowering period and prepared in the aforementioned private farm under strict managing and precaution as usually practiced for the preparation of green basil herbs. Seeds were ground to powder form and sieved while basil herbs were minced into very small pieces.

2.3. Analytical Methods

2.3.1. Determination of Chemical Composition of Spices and Herbs

Moisture, crude protein, ether extract, total ash, insoluble ash in acid, crude fibers were determined as described by A.O.A.C. methods [14]. Total carbohydrates were determined by difference [15].

2.3.2. Determination of Essential Minerals

- a) Method of digestion for mineral analysis: Digestion of dry seed spices and fresh herbs for mineral analysis by photometric and colorimetric was performed according to A.O.A.C. methods [14].
- b) Flame photometric determination: The concentration of potassium, calcium and sodium in digested ashes of the fresh and dry spices and herbs according to methods described previously by Brown and Lillel and [16].
- c) Spectrophotometric determination of phosphorus: Determination was performed according to the method of Murphy and Riley [17].
- d) Atomic absorption spectrophotometeric determination: Iron, zinc, manganese, cupper and magnesium were determined in ashed samples of dry seed spices and fresh herbs using atomic absorption Spectrophotometer (Model AA 4000) according to A.O.A.C. method [14].

2.4. Determination of Microbiological Characteristics

The microbiological examinations of dried seed spices and fresh herbs samples included the determination of total aerobic counts, total anaerobic bacteria, yeast and mold counts, and *Coliform* group which were determined according to APHA [18], while detection for *Salmonella sp.* was performed according to IAEA [19].

2.5. Chemical Characterization of Essential Oils

- a) Methods for extraction of essential oils: The essential oils of dried cumin and coriander seeds and fresh basil herbs were extracted through hydro distillation by using a Clevenger type apparatus according to El-Ghorab *et al.* [20].
- b) Determination of yield in essential oils: Percentage of volatile oil extracted was calculated on fresh and dry weight basis in replicate distillations from the tested spices and herbs according to the following Equation:

Volatile oil (%) =
$$\frac{\text{Weight of volatile oil recovered in the receiver}}{\text{Weight of sample}} \times 100\%$$

The volatile oil was removed from the receiver with ether and dried overnight using anhydrous sodium sulphate before removing the ether. The obtained volatile oil was stored in the dark at a temperature of -18° C until requiring for analysis.

- c) Gas chromatography of essential oils:
- 1) Apparatus and conditions for separation

Volatile compounds in essential oils of dried cumin and coriander seeds and fresh basil herbs were identified by comparison with kovats gas chromatographic retention index [21] and by the mass spectral fragmentation pattern of each GC component compared with authentic compounds. Agas chromatograph (Hewlett Packard model 6890) equipped with a DB5 capillary column (30 m \times 0.25 mm i.d. \times 0.25 μ m df.), FID detector was used. The analysis was carried out under the following conditions: injector temperature 200°C and detector temperature 250°C. The column was programmed from 35°C to 220°C at 30°C/min and held for 40 min. The heliumcarrier gas flow rate was 29 cm/sec. Injections were in the split less mode.

2) Identification and quantitation

Kovat's indices were determined by co-injection of the sample with a solution containing homologous series of n-hydrocarbons (C_6 - C_{26}) under the same conditions as described above. The separated components were identified by matching with N1ST mass-spectral library data, and by comparison of Kovat's indices with those of authentic components and with published data of Adams *et al.* [22]. The quantitative determination was carried out based on peak area integration.

3. Results and Discussion

3.1. Comparative Chemical Composition of Tested Spices and Herbs

The fresh basil leaves showed the highest moisture content, which reached to 80.35 ± 0.12 , compared to samples of cumin and coriander seeds (7.44 ± 0.13 and 6.47 ± 0.16 , respectively) (**Table 1**). Moisture values for dry cumin seeds and coriander as collected from the Egyptian export centers were found to be lower than the maximum humidity limits for cumin seeds (9% - 13% according to quality grades) and for coriander (9%) which were indicated by the Egyptian Specification Standards for dry seeds of cumin and coriander (ES: 1930/2008 and ES: 2095/2005, respectively) and by International Standards Organization [(ISO: 9301/2003) and (ISO: 3516/1997), respectively]. No Egyptian Specification Standards or International Standards Organization is established yet for fresh basil herbs.

With regard to cumin dry seeds, although higher moisture values were reported of 7% - 22% [23] and of 12.8% [24], the maximum humidity in cumin seeds must be no more than 9%, which confirm with the International Standards Specifications [25].

The found moisture values for coriander seeds were comparable to values of 6.65% [26] and 6.2% [5]. Various studies abroad indicated that moisture content of fresh whole basil herbs was found to be in the range of 80% - 88.25%, which is consistent or slightly higher than that found in the present results [27] [28].

The second major component of all samples was carbohydrates content. On as is basis, coriander seeds showed the highest total carbohydrates (62.32%) followed by cumin (55.58%). In contrast, when calculation was made in dry weight basis, fresh whole basil herbs showed the highest total carbohydrates (68.07%). The found carbohydrates values were higher than those reported in literature for cumin seeds of 48.01% [29] for coriander seeds of 24.0% [30] or 52.10% [31] and whole fresh basil of 7.0% [32], of 7.02% [33], of 6.6% [34] and of 9.3% [27].

Protein content, on both fresh weight and dry weight basis, was the highest in cumin seeds (18.40% and 19.88%, respectively), followed by coriander seeds (15.39% and 16.46%, respectively) while fresh basil herb showed the lowest content (1.78% and 9.04%, respectively). The found protein content values were comparable to those reported in literature for cumin seeds of 19% [35], 18.7% [24] and of 17.7% [29], but higher (15.7%) than that reported others [36].

Table 1. Comparative chemical composition of some spices and herbs collected from different Egyptian export centers (as ready for export).

Component	Cumin seeds		Coriander seeds		Whole fresh basil herb	
	On wet weight	On dry weight	On wet weight	On dry weight	On wet weight	On dry weight
Moisture	$7.44 \pm 0.13^*$	-	6.47 ± 0.16	-	80.35 ± 0.12	-
Crude protein	18.40 ± 0.16	19.88 ± 0.20	15.39 ± 0.18	16.46 ± 0.20	1.78 ± 0.03	9.04 ± 0.12
Ether extract	11.44 ± 0.20	12.36 ± 0.23	10.84 ± 0.12	11.59 ± 0.15	1.08 ± 0.03	5.50 ± 0.15
Crude fibers	21.82 ± 0.13	23.57 ± 0.13	27.23 ± 0.14	29.11 ± 0.20	2.89 ± 0.08	14.71 ± 0.29
Total ash	7.14 ± 0.10	7.71 ± 0.10	4.98 ± 0.18	5.33 ± 0.18	3.42 ± 0.11	17.39 ± 0.46
Ash (insoluble in acid)	0.69 ± 0.18	0.74 ± 0.19	0.51 ± 0.12	0.55 ± 0.13	0.10 ± 0.03	0.52 ± 0.15
Total carbohydrate**	55.58	60.05	62.32	66.62	13.37	68.07

 $^{^*}$ Mean of triplicate determination \pm standard deviation. ** Total carbohydrate calculated by difference.

However, protein values were found higher than those reported by many researchers for coriander seeds of 11.49% [37], 11% [38], 11.75% [26], 12.58% [5] and 12.58% [1] as well as for basil herb either on dry weight basis of 22.2% [39] or on fresh weight basis of 3.3% [32], 3.16% [33], 3.8%, [34] and 4.2% [28].

With the same trend of protein, cumin seeds had the highest ether extract content, on both fresh weight and dry weight basis (11.44% and 12.36%, respectively), followed by dried coriander seeds (10.84% and 11.59%, respectively) while fresh basil herb showed the lowest content (1.08% and 5.50%, respectively). Comparable literature values to those found in the present study for ether extract were reported for cumin seeds of 10% [35] [40], for coriander seeds of 9.8% [26] or 9.12% [1] and for whole basil herbs on fresh basis of 1.2% [32] or 1.05% [33].

Crude fiber content data, on both fresh weight and dry weight basis, indicated that dry coriander seeds exhibited the highest values (27.23% and 29.11%, respectively) followed by cumin seeds (21.82% and 23.57%, respectively) while fresh basil herbs had the lowest content (2.89% and 14.71%, respectively).

High amounts of fibers than those found inseeds of the Egyptian cumin cultivar were reported in Pakistani cultivars of 37.2% [36] depending upon the varieties. The found crude fibers values for seeds of the tested coriander cultivar compared well with those reported of 28.43% for Italian cultivars [37] or nearly 30% [38]. In contrast, higher value (37.14%) for Indian cultivars was found [1].

Crude fibers contents in the tested whole basil herbs, on fresh basis, were found (2.89%) higher than those reported of 2.0% [32] or of 1.0 % [34], but lower on dry basis (14.71%) than of 19.07% [27] or 33.3% [39].

Total ash content data, on fresh weight basis, indicated that dry cumin seeds had the highest values (7.14%) followed by coriander seeds (4.98%) while fresh basil herbs exhibited the lowest content (3.42%), but when calculation was made on dry weight basis the fresh basil herbs showed the highest total ash value (17.39 %) followed by cumin seeds (7.71%), while coriander seeds showed the lowest content (5.33%).

In contrast, ash insoluble in acid content showed different results, on both fresh weight and dry weight basis, where cumin had the highest values (0.69% and 0.74%, respectively) followed by coriander (0.51% and 0.55%, respectively) while fresh basil herb showed the lowest content (0.10% and 0.52%, respectively).

The found total ash and as hinsoluble in acid values were found lower than the maximum limits indicated for dry cumin seeds (8.5% - 12% and 1.5% - 4% according to quality grades, respectively) by the Egyptian Specification Standards (ES: 1930/2008) and by the International Standards Organization (ISO: 9301/2003) and those indicatedfordry coriander seeds (7% and 1.5% for all quality grades, respectively) by the Egyptian Specification Standards (ES: 2095/2005) and by the International Standards Organization (ISO: 2255/1996).

Moreover, values found for total ash and ash insoluble in acid were lower than those reported for dry cumin seeds by others of 9.5% total ash and a maximum 2% acid insoluble ashes [25] as well as that value reported for coriander seeds grown in India [31] of 14.02%. In contrast, the found values for total ash in basil herbs consisted well with those indicated in literature of 3.32% [27], but lower than those reported on fresh weight basis of 2.0% [32] and of 1.8% [34] and higher than those reported by others of 10.18% [39] or 8.7% [41] on dry weight basis.

It should be mentioned that the chemical composition of the various spices and herbs were reported to vary significantly according to the variety and species in plant herb, cultivation practices, plantation season, number and time of cuts, plant development stage and the climatic conditions [30].

3.2. Comparative Content in Essential Minerals of Tested Spices and Herbs

Table 2. Essential minerals composition of some spices and herbs previously collected from different Egyptian export centers as ready for export.

Component -	Cumin seeds		Coriano	Coriander seeds		Whole fresh basil herb		
	On wet weight	On dry weight	On wet weight	On dry weight	On wet weight	On dry weight		
	Macro elements (g/100g)							
K	3.25 ± 0.35	3.51 ± 0.52	3.71 ± 0.30	3.97 ± 0.26	0.96 ± 0.30	4.88 ± 0.82		
Na	0.10 ± 0.10	0.11 ± 0.01	0.24 ± 0.06	0.26 ± 0.13	0.06 ± 0.00	0.31 ± 0.08		
Ca	0.20 ± 0.03	0.22 ± 0.02	0.18 ± 0.02	0.19 ± 0.04	1.23 ± 0.68	6.25 ± 1.41		
Mg	0.21 ± 0.05	0.23 ± 0.05	0.15 ± 0.04	0.17 ± 0.03	0.28 ± 0.02	1.42 ± 0.02		
P	0.52 ± 0.01	0.56 ± 0.01	0.73 ± 0.02	0.78 ± 0.02	0.16 ± 0.01	0.81 ± 0.05		
	Micro elements (mg/100g)							
Fe	330 ± 8.16	357 ± 7.24	369 ± 9.20	394 ± 8.05	141 ± 2.20	718 ± 13.70		
Mn	65 ± 2.10	70 ± 2.22	53 ± 1.14	57 ± 1.15	49 ± 1.15	249 ± 3.71		
Zn	64 ± 1.30	69 ± 1.13	80 ± 1.25	85 ± 2.28	24 ± 1.26	122 ± 1.13		
Cu	30 ± 1.20	33 ± 1.34	32 ± 1.01	34 ± 1.02	10 ± 0.89	51 ± 1.45		

3.3. Comparative Microbiological Characteristics of Spices and Herbs

Data in **Table 3** showed that total bacterial count was low content in seeds of cumin and coriander as well asfresh whole basil herb, where its log number was 3.72, 3.73 and 3.78, respectively. However, anaerobic bacteria, moulds and yeasts and Coliform group were of higher content. The log number of anaerobic bacterial count was: 3.65, 3.55 and 3.47 incumin, coriander and basil, respectively, for moulds and yeasts were: 2.97, 2.39 and 2.39, respectively. In addition, the log number of Coliform group count was: 3.00, 3.14 and 3.32, respectively. On the other hand, the three tested seed spicesand herbs (cumin, coriander and basil) were nil from Salmonella spp. bacteria. The microbiologicalload in all tested seed spices and herbs were lower than those indicated by the Egyptian Specification Standards (ES: 1930/2008 and ES: 2095/2005) as well as by the International Standards Organization (ISO: 9301/2003 and ISO: 2255/1996) for cumin and coriander seeds, respectively.

3.4. Comparative Yield in Essential Oils Extracted from Spices and Herbs

Yields of the different essential oils (EOs), extracted by hydro distillation (**Table 4**) showed that, on dry weight basis, cumin seeds had the highest amount of volatiles ($3.762\% \pm 0.18\%$), while both coriander and basil samples had lower amounts ($0.285\% \pm 0.01\%$ and $0.686\% \pm 0.03\%$, respectively).

Furthermore, EOs contents of the tested cumin seeds are in the range reported in literature in Bulgaria of 5.3% [45], in China of 2.608% to 4.062% [46] and in Pakistan of $2.52\% \pm 0.11\%$ [36]. However, it is higher than the maximum limits (1.5% - 2.5% on dry weight basis according to quality grades) as indicated by the Egyptian Specification Standards (ES: 1930/2008) and by the International Standards Organization (ISO: 9301/2003) for cumin seed oil. Moreover, it is higher than other cultivars grown in different locations in Iran of 1.4% - 2.2% [47], in Turkey of 1.4% to 2.8% [48], in Tunisia of 1.6% [49], in India of 1.21% or 1% [50] [51].

Results indicated that EOs content of coriander seeds cultivar grown in Egypt is higher and/or comparable to many varieties or cultivars grown in different locations (in India of 0.5% - 1% [52], in Pakistan of 0.15% [53] and 0.03% to 2.7% [5], in Iran of 0.1% to 0.36% [54], in Bulgaria of 0.1% - 0.5% [30], in Argentina of 0.40% [55], in Turkey between 0.03% and 2.6% [56]). However, it is within the limits (0.1% - 0.5% on dry weight basis according to quality grades) indicated by both the Egyptian Specification Standards (ES: 2095/2005) and by International Standards Organization (ISO: 2255/1996) for coriander seeds.

The found EOs content of whole basil herbs from the cultivar grown in Egypt, was sometimes either higher or comparable to many varieties or cultivars grown in different locations (in Turkey of 0.15% - 1.59% [57], in India of 0.2% in stems to 1.2% in leaves on a fresh weight basis [58], in Algeria of 0.4% [59], in Iran of 0.5% [60], in Pakistan of 0.5% to 0.8% [61], in Sudan of 0.33% to 0.47% in fresh leaves [62], in Romania 0.2% and 1%

Table 3. Comparative microbiological characteristics of some spices and herbs previously collected from different Egyptian export centers as ready for export (as is basis).

Microbiological examina	tion	Cumin seeds	Coriander seeds	Whole fresh basil herb
Total bacterial count	No.	5.3×10^{3}	5.4×10^{3}	6.1×10^{3}
Total bacterial count	Log No.	3.72	3.73	3.78
A	No.	4.6×10^3	3.6×10^3	5.5×10^3
Anaerobic bacterial count	Log No.	3.65	3.55	3.47
W 11 1	No.	9.5×10^2	2.5×10^2	2.5×10^2
Moulds and yeasts	Log No.	2.97	2.39	2.39
C-1:f	No.	1.0×10^3	$1.4\times10^{\ 3}$	2.1×10^3
Coliform group count	Log No.	3.00	3.14	3.32
G 1 11	No.	-	-	-
Salmonella spp.	Log No.	-	-	-

^{(-):} Not detected.

Table 4. Comparison of yield of essential oils extracted from some spices and herbs previously collected from different Egyptian export centers as ready for export (on as is and dry basis).

6-1	Essential oils extracted (%)			
Spices and herbs —	Before dried	After dried		
Cumin seeds	3.52 ± 0.17	3.762 ± 0.18		
Coriander seeds	0.27 ± 0.01	0.285 ± 0.01		
Whole fresh basil herb	0.14 ± 0.01	0.686 ± 0.03		

[63], of 0.171% in Omani basil [64]).

It should be mentioned that yields of EOs extracted from spices and herbs were found to vary, not only with varieties, season, cuttings and agricultural practices, but also according to parts of herbs(whole, flowers, leaves and stems for basil herbs [65] [66] and for seed spices [67].

3.5. Comparative Composition of EOs Extracted from Spices and Herbs

Chemical identification of the oil constituents dry seeds of cumin and coriander as well as fresh whole basil herbs collected from different Egyptian export centers as being ready for export abroad was conducted using gas chromatography (GC) based on their retention indices (RI). The volatile oil components in EOs of the three tested Egyptian spices and herbs were classified into groups, based on the relative area %.

3.5.1. GC Identification of Essential Oil Composition of Cumin Seeds

GC chromatograms indicated the presence of 41 compounds, which accounted for 98.78% of EOs hydrodistilled from seeds of the cumin cultivar grown in Egypt (**Table 5**). The major compounds in cuminessential oil were cumin aldehyde (35.25%), tetradecene (12.25%), γ -terpenene (12%), β -ocimene (9.72%), p-mentha-2-en-ol (9%), α -terpinyl acetate (5.32%), α -terpinolene (3%), Lmonine (0.5%), myrcene (0.2%), β -pinene (0.9%) and α -pinene (0.19%).

Results indicated that the major component and also its proportion in the tested cumin Eos resembled those values found by various investigators in different locations in the world e.g., of 36.31% in Chinese cultivars [68] [40], 36% in Bulgarian cultivar [45], 39.48% in Tunisian variety [4]. However, there were differences in proportion of the major component (of 22.76% in Chinese cultivar [69], 27.7% in Pakistani cultivar [36], 30.2% [70]) and in number of separated compounds in GC chromatograms (21 components in Tunisian cultivar [4], 19 components [70] or 38 compounds [71]) and also the identity of major component (as found in china [69], in Tunisia

Table 5. Effect of storage conditions on gas chromatographic profile of essential oils extracted from dried seeds of Egyptian cultivar of cumin (*Cuminum cyminum*) stored for six months before exportation abroad (as is basis).

Peak No.	Compounds	KI	% Area	Method of identification	Type of component	
1	α -pinene	939	0.19	KI & MS	M	
2	Sabinene	976	0.12	KI & MS	M	
3	β -pinene	980	0.9	KI & MS	M	
4	Octanone <2->	987	0.07	KI & MS	LOC	
5	Myrcene	991	0.2	KI & MS	M	
6	Octanol <2->	995	0.35	KI & MS	LOC	
7	α -phellandrene	1005	0.82	KI & MS	M	
8	Limonene	1031	0.5	KI & MS	M	
9	1, 8-cineole	1033	0.49	KI & MS	LOC	
10	(E)- β -ocimene	1042	9.72	KI & MS & ST	M	
11	γ-terpenine	1064	12	KI & MS & ST	M	
12	Para cymene	1089	0.1	KI & MS	M	
13	α -terpinolene	1096	3	KI & MS & ST	LOC	
14	3 linalool	1098	0.10	KI & MS & ST	LOC	
15	cis-sabinene hydrate	1104	0	KI & MS	LOC	
16	p-menth-2-en-1-ol	1130	9	KI & MS & ST	LOC	
17	Terpin-4-oL	1177	0.1	KI & MS & ST	LOC	
18	Cumin aldehyde	1239	35.25	KI & MS & ST	LOC	
19	Geraniol	1254	0.63	KI & MS	HOC	
20	Phellandral	1273	0	KI & MS	HOC	
21	2-caren-10-al	1281	1.32	KI & MS	HOC	
22	Methyl geranate	1326	0.24	KI & MS	HOC	
23	α -Terpinyl acetate	1344	5.32	KI & MS	HOC	
24	Tetradecene <1->	1392	12.25	KI & MS & ST	S	
25	β -caryophyllene	1467	0	KI & MS	S	
26	Carotol	1549	0.1	KI & MS	HOC	
27	Germacrene-D-4-ol	1573	0.25	KI & MS	HOC	
28	Humulene epoxide II	1604	0	KI & MS	HOC	
29	Dill apiole	1624	0	KI & MS	HOC	
30	Cubenol	1639	0	KI & MS	HOC	
31	Acetocyclohexanedione	1713	0.22	KI & MS	HOC	
32	Sesquilavandulyl acetate <e-></e->	1740	0	KI & MS	HOC	
33	α -sinensal	1751	0	KI & MS	HOC	
34	Ethyl tetradecanoate	1795	0	KI & MS	HOC	
35	Bisabolol acetate <epi-alpha-></epi-alpha->	1805	0	KI & MS	HOC	
36	α -vetivone	1830	0	KI & MS	HOC	
37	Farnesyl acetate <e-e></e-e>	1838	0.05	KI & MS	HOC	
38	Laurenene <epi-></epi->	1892	0.15	KI & MS	S	
39	Occidol acetate	1967	0	KI & MS	HOC	
40	Manoyl oxide	1987	0.63	KI & MS	HOC	
41	Phynyl ethyl anthranilate-2-	2110	4.70	KI & MS	HOC	
	Chemical classes					
	Monoterpene (M)	M	24.55			
Light	oxygenated compound (LOC)	LOC	48.37			
Heavey	oxygenated compound (HOC)	HOC	13.46			
	Sesqterpene (S)	S	12.40			

[59] and in Iran [51]).

However, the name and proportion of the major and other main components in cumin EOs were within the ranges indicated by both the Egyptian Specification Standards (ES: 2034/2007) and by International Standards Organization (ISO: 9301/2003) for essential oils of cumin seeds (cumin aldehyde between 15% - 46% of oil and other components, with the exception of the proportion of β -pinene).

The volatile oil components in EOs from the tested Egyptian cultivar were classified into four groups (based on the relative area %), viz. mono terpenes [M] (24.55%), lightly oxygenated compounds [LOC] (48.37%) sesquiterpenes [S] (12.4%), and heavily oxygenated compounds [HOC] (13.46%). The LOC and M were found in higher amounts in comparison with HOC and S, respectively. The most significant compounds in LOC and M were cumin aldehyde and g-terpinene, respectively, while S and HOC were tetradecene and α -terpinyl acetate, respectively. However, different groups for volatile components than those found in EOs of the Egyptian cultivar were reported in literature [72].

3.5.2. Identification of Essential Oil Composition of Coriander Seeds

GC Chromatograms revealed the presence of 35 peaks in Eos hydro-extracted from the tested coriander cultivar grown in Egypt of which nineteen compounds constituted 99.97% OF EOs. The major constituents were: translinalool (72.6%), sabinene hydrate (4.53%), α -Pinene (3%), ethylhexanoic acid <2-> (5.19%), p-cymen-8-ol (4.51%), nerol (1.1%), caryophyllene <9-epi-E> (2.71%) a-thujene (3.28%), camphor (0.27%) and Limonine (0.13%). In addition, the coriander seeds essential oil also contained considerable amounts of various minor constituents whose contribution was <0.7% (Table 6).

Results indicated that major compounds identified in Eos from seeds or tested coriander cultivar grown in Egypt are in accordance with the findings on various investigators for cultivars from different parts of the world, e.g., 68.14% in Argentina [55], 68.00% in Russian [73], 70.5% in Korea [74], 75.30% in India [75], 65% to 70% in Pakistan [53]. In contrast, there were reported differences inlinalool proportion [54.57% in Cuba [76], 87.54% in India [77], of 63.9% - 66.2% in Canada [78], of 37.65% in Bangladesh [79], of 58.22% [80] in Brazil, of 53.79% [81] in South Korea, of 55.59% in Pakistan [5], and in the number of separated compounds in GC chromatograms (35 compounds in Cuba [76], 24 components in India [82] and 53 compounds in Bangladesh [79]).

However, the major component and the other main components in EOs of seeds of the tested coriander cultivar were within the ranges for Linalool (between 65% - 78% of oil) and other components as indicated by both the Egyptian Specification Standards (ES: 2037/2007) and by International Standards Organization (ISO: 3516/1979) for coriander oil seeds. Regarding the groups of chemical constituents, EOs of the Egyptian coriander seeds mainly consisted of light oxygenated compounds (88.93%), followed by monoterpene hydrocarbons (8.27%), sesquiterpene hydrocarbons (2.7%) and heavy oxygenated compounds (0.06). Linalool was the main light oxygenated compounds, while thujene (3.28%) was the major monoterpene.

In agreement to the found results about groups of chemical constituents in EOs of the Egyptian coriander seeds, it was stated that EOs of seeds from Indian cultivar of coriander rich in oxygenated monoterpenes, while the oxygenated monoterpenes, monoterpene hydrocarbon amounted to 80.47%, 6.45%, respectively [77]. Furthermore, EOs of seeds from Pakastani cultivar of coriander mainly comprised of oxygenated monoterpene hydrocarbons (80.83%), followed by monoterpene hydrocarbons (8.00%), sesquiterpene hydrocarbons (0.47%) and oxygenated sesquiterpene hydrocarbons (0.35%) [53]. In contrast, the presence of sesquiterpene hydrocarbons at a concentration over 13% in EOs of coriander seeds has been detected [83].

3.5.3. Identification of Essential Oil Composition of Whole Basil Herbs

GC Chromatograms of essential oils from Egyptian cultivar of basil whole herb revealed that forty seven compounds, representing 99.08% of the GC profile, were identified, where the major identified constituents (**Table7**) were: linalool (54.01%), Kessane (10.02%), Germacrene D (4.4%), Terpin-4-ol (2.19%), Eugenol (1.75%), β -selinene (4.4%), Cadina-1, 4-diene (1.54%), α -cadinene (0.84%), elmicine (1.46%), caryophyllene oxide (1.66%), viridiflorol (1.19%), humuleneepoxide II (2.44%), 10-epi-g-eudesmol (4.01%) and α -sinensal (3.09%).

The EOs of basil herbs grown in Egypt, were of the high Linalool-chemotype which are characterized by high contents of linalool and relatively lower amounts of eugenol (in the present they were 54.01% and 1.75%, respectively). It has been always reported that basil oils have very variable chemical composition [65]. On this basis of the oil composition, basil accessions were divided into seven groups: 1) high-linalool chemotype, 2) linalool-eugenolchemotype, 3) methyl chavicolchemotype and no linalool 4) methyl chavicol-linalool chemotype,

Table 6. Gas chromatographic characteristics of essential oils extracted from dried seeds of coriander (*Coriandum sativum*) previously collected from different Egyptian export centers as ready for export (as is basis)*.

D1- N-	Composed to	1/1	0/ 1	M-4-4-6:4-4:6:-4:	T
Peak No.	Compounds .	KI	% Area	Method of identification	Type of component
1	α-pinene	939	3	MS & KI	M
2	α-thujene	995	3.28	MS & KI	M
3	β -pinene	980	0.1	MS & KI	M
4	(delta3) δ 3-carene	1012	0.65	MS & KI	M
5	P-cymene	1026	0.2	MS & KI & ST	M
6	Limonene	1031	0.13	MS & KI & ST	M
7	(Z)- β -ocimene	1041	0.7	MS & KI & ST	M
8	γ -terpinene	1061	0.21	MS & KI & ST	M
9	Sabinene hydrate trans	1097	4.53	MS & KI & ST	LOC
10	Linalool	1098	72.6	MS & KI & ST	LOC
11	Ethyl hexanoic acid <2->	1129	5.19	MS & KI & ST	LOC
12	Camphor	1151	0.27	MS & KI & ST	LOC
13	Borneol	1168	0.32	MS & KI	LOC
14	P-cymen-8-ol	1184	4.51	MS & KI & ST	LOC
15	Geranial	1275	0.18	MS & KI	LOC
16	Nerol	1228	1.1	MS & KI & ST	LOC
17	Carvacrol	1301	0.23	MS & KI	LOC
18	Geranyl acetate	1379	0	MS & KI	HOC
19	Humulene <alpha-></alpha->	1464	0	MS & KI	S
20	Caryophyllene <9-epi-(E)->	1471	2.71	MS & KI	S
21	Geranylisobutyrate	1514	0	MS & KI	HOC
22	Caryophyllene-oxide	1589	0.06	MS & KI	HOC
23	Ethyl tetradecanoate	1793	0	MS & KI	HOC
24	Bisabolol acetate <epi-alpha-></epi-alpha->	1803	0	MS & KI	HOC
25	Santalol acetate <(z)-epi-beta->	1807	0	MS & KI	HOC
26	Vetivone <alpha></alpha>	1836	0	MS & KI	S
27	Farnesyl acetate <e-e></e-e>	1841	0	MS & KI	HOC
28	Laurenene <epi-></epi->	1891	0	MS & KI	S
29	Isophytol	1941	0	MS & KI	HOC
30	Phytol	1956	0	MS & KI	HOC
31	Occidol acetate	1970	0	MS & KI	HOC
32	Manoyl oxide	1987	0	MS & KI	HOC
33	Laurenan-3-one	2104	0	MS & KI	HOC
34	Phynyl ethyl anthranilate-2-	2117	0	MS & KI	HOC
35	Incensole	2155	0	MS & KI	HOC
	Chemical classes				
Monoterpene (M) M 8.27					
	ygenated compound (LOC)	LOC	88.93		
	xygenated compound (HOC)	HOC	0.06		
	Sesqterpene (S)	S	2.71		

Table 7. Gas chromatographic characteristics of essential oils extracted from green whole basil herb (*Ocimum basilicum*) previously collected from different Egyptian export plantation centers as ready for export (as is basis)*.

Peak No.	Compounds	KI	% Area	Method of identification	Type of component
1	Myrcene	994	0.08	KI & MS	M
2	1,8 cineol	1033	0.62	KI & MS	LOC
3	(Z)- β -ocimene	1038	0.14	KI & MS & ST	M
4	γ-terpinene	1057	0.06	KI & MS & ST	M
5	Linalool	1098	54.01	KI & MS & ST	LOC
6	Limonene oxide	1130	0.09	KI & MS & ST	LOC
7	Camphor	1150	0.04	KI & MS & ST	LOC
8	Borneol	1167	0.07	KI & MS & ST	LOC
9	Terpinen-4-ol	1177	2.19	KI & MS & ST	LOC
10	Nerol	1231	0.82	KI & MS & ST	LOC
11	Exo-fenchyle acetate	1237	0.04	KI & MS	LOC
12	Geraniol	1255	0.05	KI & MS	LOC
13	Geranial	1268	0.26	KI & MS	LOC
14	Thymol	1292	0.9	KI & MS	LOC
15	Carvacrol	1298	0.64	KI & MS	LOC
16	Eugenol	1358	1.75	KI & MS & ST	LOC
17	β -caryophyllene	1421	0.09	KI & MS	S
18		1421	0.09	KI & MS	S S
	β -gurjunene				
19	α-humulene	1451	0.03	KI & MS	S
20	cis-muurola-4(14)5-diene	1465	0.43	KI & MS	S
21	Germacrene D	1476	0.8	KI & MS	S
22	β -selinene	1483	4.4	KI & MS	S
23	α -muurolene	1502	0.21	KI & MS	S
24	γ-cadinene	1514	0.41	KI & MS	S
25	7 epi-α-salinene	1517	0.61	KI & MS	S
26	δ -cadinene	1521	0.25	KI & MS	S
27	Kessane	1528	10.02	KI & MS	S
28	Cadina-1,4-diene	1532	1.54	KI & MS	S
29	α -cadinene	1538	0.84	KI & MS	S
30	Elemicine	1553	1.46	KI & MS	S
31	Germacrene-B	1557	0.4	KI & MS	S
32	Germacrene-D-4-ol	1574	0.65	KI & MS	HOC
33	Spathulenol	1579	0.01	KI & MS	HOC
34	Caryophyllene oxide	1582	1.66	KI & MS	HOC
35	Viridiflorol	1597	1.19	KI & MS	HOC
36	Humulene epoxide II	1606	2.44	KI & MS	HOC
37	10-epi-γ-eudesmol	1618	4.01	KI & MS	HOC
38	Dill apiole	1622	1.15	KI & MS	HOC
39	γ-eudesmol	1630	0.42	KI & MS	HOC
40	Cadinol, epi- α	1642	0.08	KI & MS	HOC
41	α-cadinol	1662	0.12	KI & MS	HOC
42	α -bisabolol	1686	0.2	KI & MS	HOC
43	β -sinensal	1695	0.07	KI & MS	HOC
44	-	1747			HOC
	Sesquilavandulyl acetate <e-></e->		0.29	KI & MS	
45	α-sinensal	1766	3.09	KI & MS	HOC
46	Ethyl tetradecanoate	1770	0.02	KI & MS	HOC
47	Phytol	1950	0.15	KI & MS	HOC
	Chemical classes				
	Monoterpene (M)	M	0.28		
	Sesqterpene (S)	S	21.77		
Light	oxygenated compound (LOC)	LOC	61.48		
_	oxygenated compound (HOC)	HOC	15.55		

5) methyl eugenol-linalool chemotype, 6) methyl cinnamate-linalool chemotype, and 7) bergamotenechemotype [84].

The obtained findings for the Egyptian basil cultivar that linalool was the main component and also having the highest proportion are in good agreement with various literature reports about Linalool-chemotype cultivars grown in different parts of the world where Linalool was the main compound and of high content (54% - 60%) in EOs of different basil types or cultivars (from Italy [10], from Bangladesh [85], from Bulgaria [86], from Spain [87] and from Pakistan [61]).

Lower or higher linalool percentage than that found in EOs from the Egyptian basil cultivar were reported in literature for Linalool-chemotype basil oil [of 49.7% in Brazilian basil cultivars [88], of 71.4% in Bulgarian cultivar [89], of 41.2% in Turkish basil [90], of 43.8% in Algerian cultivar [59], of 44.18% in an Egyptian cultivar [91], 69.9% for Omani basil [64]. Moreover, the number of components isolated in GC chromatograms from EOs of the Egyptian basil cultivar (47 components) was either lower number (29 compounds in Pakistan [61], 33 components [92]) or higher number (59components in Algeria [59] and 75 in Omani basil [64]).

The major component, its proportion and proportion of the other main components in EOs of the tested fresh basil herb cultivar were found different than those indicated by the Egyptian Specification Standards (ES: 1359/2007) and by International Standards Organization (ISO: 11043/1998) for basil oil of methyl-chavicol type. No specifications are present yet for basil oil of Linalool-chemo type.

Regarding the groups of chemical constituents, the majority of compounds in EOs from whole basil herbs grown in Egypt were light oxygenated monoterpene compounds (LOC) (61.48%), sesquiterpene (S) (21.34%) and heavy oxygenated compounds (HOC) (15.98%). Monoterpenes (0.28%) were also present. The found results for classified chemical groups in Egyptian basil cultivar volatile oils compared well with literature reports that the oxygenated compounds were also the major constituent in Turkish *basilicum* volatile oils [93]. Pakistani basil EOs mainly consisted of oxy-genated monoterpenes followed by sesquiterpenes hydrocarbons and oxygenated sesquiterpenes [61]. In addition, basil oil was found richer in oxygenated monoterpenes (49.15%), where linalool representing the most important compound in the genus [63].

In addition, it was stated that sweet basiloil is mainly composed of monoterpenes, sesquiterpenes and phenyl-propanoids [94]. The presence of monoterpene hydrocarbons, oxygenated monoterpene, sesquiterpene hydrocarbons, oxygenated sesquiterpene, etc. in basil oils was ensured [9]. In fact, basil volatiles were characterized by the prevalence of oxygenated monoterpenoid compounds, being the main constituent's linalool, eugenol and eucalyptol [95].

The observed differences in the identity of major components, their proportion and in group classification of these components between results of the three tested Egyptian cultivars of spices and herbs and those in literature reports could be attributed to various reasons and factors. For example, geographic origin and region of the tested spices and herbs including plant part, harvest time, extraction method, type of cultivar, storage conditions, climatic effects on the plants eventually affects the chemical composition [61] [70].

4. Conclusion

The tested seed spices and basil herbs and their extracted EOs, commonly cultivated and exported abroad in Egypt as collected from the export centers were chemically, microbiologically and chromatographically characterized. Their characteristics were compared with those cultivated in different parts of the world and also to the Egyptian Standards and International Specifications (ISO) for seeds and EOs of cumin and coriander seeds as well as for basil herb and were found to be within the limits for these specifications.

References

- [1] Bhat, S., Kaushal, P., Kaur, M. and Sharma, H.K. (2014) Coriander (*Coriandrumsativum* L.): Processing, Nutritional and Functional Aspects. *African Journal of Plant Science*, **8**, 25-33. http://dx.doi.org/10.5897/AJPS2013.1118
- [2] Stefanini, I., Piccaglia, R., Marotti, M. and Biavati, B. (2006) Characterization and Biological Activity of Essential Oils from Fourteen Labiatae Species. *Acta Horticulturae*, **723**, 221-226.
- [3] Milan, K.S.M., Dholakia, H., Tiku, P.K. and Vishveshwaraiah, P. (2008) Enhancement of Digestive Enzymatic Activity by Cumin (*Cuminum cyminum* L.) and Role of Spent Cumin as a Bionutrient. *Food Chemistry*, 110, 678-683. http://dx.doi.org/10.1016/j.foodchem.2008.02.062
- [4] Hajlaoui, H., Mighri, H., Noumi, E., Snoussi, M., Trabelsi, N., Ksouri, R. and Bakhrouf, A. (2010) Chemical Compo-

- sition and Biological Activities of Tunisian *Cuminum cyminum* L. Essential Oil: A High Effectiveness against *Vibrio* spp. Strains. *Food and Chemical Toxicology*, **48**, 2186-2192. http://dx.doi.org/10.1016/j.fct.2010.05.044
- [5] Shahwar, M.K., El-Ghorab, A.H., Anjum, F.M., Butt, M.S., Hussain, S. and Nadeem, M. (2012) Characterization of Coriander (*Coriandrumsativum* L.) Seeds and Leaves: Volatile and Nonvolatile Extracts. *International Journal of Food Properties*, 15, 736-747. http://dx.doi.org/10.1080/10942912.2010.500068
- [6] Sullivan, C. (2009) Herbs, 09 in College Seminar 235-Food for Thought. The Science, Culture, and Politics of Food in Spring.
- [7] Janicked, J.E., Mario, R., Morales, A., Winthrop, B., Phippen, R., Fontes, V. and Zhigang, H. (1999) Basil: A Source of Aroma Compounds and a Popular Culinary and Ornamental Herb. Reprinted from: Perspectives on New Crops and New Uses, ASHS Press, Alexandria.
- [8] Akgul, A. (1993) Spice Science and Technology. Turkish Association of Food Technology, Ankara, 15.
- [9] Marwat, S.K., Khan, M.A., Fazal-ur-Rehman, A.H., Shoaib, M. and Shah, M.A. (2011) Interpretation and Medicinal Potential of Ar-Rehan (Ocimum basilicum L.)—A Review. American-Eurasian Journal of Agricultural and Environmental Science, 10, 478-484.
- [10] Marotti, M., Piccaglia, R. and Giovanelli, E. (1996) Differences in Essential Oil Composition of Basil (*Ocimum basilicum* L.) Italian Cultivars Related to Morphological Characteristics. *Journal of Agricultural and Food Chemistry*, 14, 3926-3929. http://dx.doi.org/10.1021/jf9601067
- [11] Ravid, U., Putievsky, E., Katzir, I. and Lewinsohn, E. (1997) Enantiomeric Composition of Linalool in the Essential Oils of *Ocimum* Species and in Commercial Basil Oils. *Flavour Fragrance Journal*, **12**, 293-296. http://dx.doi.org/10.1002/(SICI)1099-1026(199707)12:4<293::AID-FFJ648>3.0.CO;2-3
- [12] Omer, E.A., Said-Al-Ahl, H.A.H. and Hendawy, S.F. (2008) Production, Chemical Composition and Volatile Oil of Different Basil Species/Varieties Cultivated under Egyptian Soil Salinity Conditions. *Research Journal of Agriculture and Biological Sciences*, **4**, 293-300.
- [13] Deepak (2013) Importance of Cuminum cyminum L. and Carum carvi L. in Traditional Medicaments—A Review. Indian Journal Traditional Knowledge, 12, 300-307.
- [14] AOAC (2005) Official Methods of Analysis of AOAC international. In: Horwitz, W., Ed., Association of Official Analytical Chemists, 17th Edition, AOAC International Suite 500, Gaithersburg.
- [15] Nielsen, S.S. (1994) Chemical Composition and Characteristics of Foods. In: *International to the Chemical Analysis of Foods*, Jones and Bartlett, Inc., Boston, 142.
- [16] Brown, J.D. and Lilleland, O. (1964) Rapid Determination of Potassium Calcium and Sodium in Plant Material and Soil Extracts Flaw Phosphorus. *Proceedings of the American Society for Horticultural Sciences*, **48**, 341-346.
- [17] Murphy, J. and Riely, J.P. (1962) A Modified Single Solution Methods for the Determination of Phosphate in Natural Wastes. *Analytica Chimica Acta*, **27**, 31-36.
- [18] APHA (1992) Standard Methods for the Examination of Dairy Products. American Public Health Association, Washington DC.
- [19] IAEA (1970) Microbiological Specifications and Testing Methods for Irradiated Food. Technical Reports Series, IAEA, Vienna.
- [20] El-Ghorab, A., Fadel, H., Marx, F. and El-Sawy, A. (1999) Effect of Extraction Techniques on the Chemical Composition and Antioxidant Activity of Eucalptus camaldulensis var. Brevirostris Leaves Oil. Zeitschrift fur Lebensmittal-Untersuchung und—Forschung A, 208, 212-216. http://dx.doi.org/10.1007/s002170050405
- [21] Kovats, E. (1965) Gas Chromatographic Characterization of Organic Substances in the Retention Index System. *Advances in Chromatography*, **1**, 229-247.
- [22] Adams, R.P. (1995) Identification of Volatile Oil Components by Gas Chromatography/Mass Spectroscopy. Allured Publishing Corporation, Carol Stream.
- [23] Singh, K.K. and Goswam, T.K. (1998) Mechanical Properties of Cumin Seed (*Cuminum cyminum* Linn) under Compressive Loading. *Journal of Food Engineering*, **36**, 311-321. http://dx.doi.org/10.1016/S0260-8774(98)00056-9
- [24] Emam, O.A. (2001) The Quality of Cumin Treated with Gamma and Microwave Irradiation. *Annals of Agricultural Science, Moshtohor*, **39**, 1601-1614.
- [25] Kafi, M. (2002) Cumin (*Cuminum cyminum*) Production and Processing. Ferdowsi University of Mashhad Press, Mashhad
- [26] Khan, A.L., Javid, H., Rehman, N., Khan, F., Hussain, S.T. and Shinwari, Z.K. (2009) Proximate and Nutrient Investigations of Selected Medicinal Plants Species of Pakistan. *Pakistan Journal of Nutrition*, 8, 620-624. http://dx.doi.org/10.3923/pjn.2009.620.624

- [27] Morsy, N.F.S. (2004) Quality Evaluation of the Essential Oil of Basil Plant Dried by Different Methods. Ph.D. Thesis, Cairo University, Giza.
- [28] Sarfraz, S., Anjum, F., Khan, M., Arshad, M. and Nadeem, M. (2011) Characterization of Basil (*Ocimum basilicum* L.) Parts for Antioxidant Potential. *African Journal of Food Science and Technology*, **2**, 204-213.
- [29] Farhath, K., Sudarshanakrishna, K.R., Semwal, A.D. and Vishwanathan, K.R. (2001) Proximate Composition and Mineral Contents of Spices. *Indian Journal of Nutrition and Dietetics*, 38, 93-97.
- [30] Sharma, M.M. and Sharma, R.K. (1999) Coriander. Handbook of Herbs and Spices. Wood Head Publishing Ltd., Sawston, 1-6.
- [31] Verghese, J. (2001) Coriander. *Indian Spices*, 38, 8.
- [32] Duke, J.A. (1985) CRC Handbook of Medicinal Herbs. CRC Press, Boca Raton.
- [33] Rinzler, C.A. (1990) The Complete book of Herbs, Spices and Condiments. Fact on File, Inc., New York.
- [34] Anon (1997) Food and Nutrition. Middle East Regional Office of World Health Organization. Academia International, Lebanan.
- [35] Gopalan, C., Rama Sastri, B.V. and Balasubramanian, S.C. (1989) Nutritive Value of Indian Foods. National Institute of Nutrition, ICMR, Hyderabad, 59-93.
- [36] El-Ghorab, A.H., Nauman, M., Anjum, M.F., Hussain, S. and Nadeem, M. (2010) A Comparative Study on Chemical Composition and Antioxidant Activity of Ginger (*Zingiber officinale*) and Cumin (*Cuminum cyminum*). *Journal of Agricultural and Food Chemistry*, **58**, 8231-8237. http://dx.doi.org/10.1021/jf101202x
- [37] Diederichsen, A. (1996) Promoting the Conservation and Use of Underutilized and Neglected Crops: Coriander (*Coriandrum sativum L.*). International Plant Genetic Resources Institute (IPGRI), Rome.
- [38] Peter, K.V. (2004) Handbook of Herbs and Spices. Vol. 2. Wood Head Publishing Ltd., Abnigton Hall, 158-174.
- [39] Pennington, J.A.T. and Church, H.N. (1985) Food Values of Portions Commonly Used. 14th Edition, J. B. Lippinsott Company, Philadelphia.
- [40] Li, R. and Jiang, Z.T. (2004) Chemical Composition of the Essential oil of Cuminum cyminum L. from China. Flavour and Fragrance Journal, 19, 311-313. http://dx.doi.org/10.1002/ffj.1302
- [41] Bihari, G.C., Manaswini, B., Prabhat, J. and Kumar, T.S. (2011) Pharmacognostical and Phytochemical Investigation of Various Tulsi Plants Available in South Eastern Odisha. *International Journal of Research in Pharmaceutical and Biomedical Sciences*, **2**, 605-610.
- [42] Hamad, O., Al-Obaidi, S., Ezzat, M.O. and Mordi, M.N. (2012) Isolation and Biological Activity Study of Some Active Substances and Elements Quantification of the Water, Alcoholic and Oil Extracts of *Cuminum cyminum*. Science International (Lahore), 24, 27-29.
- [43] Khan, S.A., Ahmad, I. and Mohajir, M.S. (2006) Evaluation of Mineral Contents of Some Edible Medicinal Plants. *Pakistan Journal Pharmaceutical Sciences*, **19**, 148-152.
- [44] Hossein, M.S. (2010) Determination of Mineral Nutrients in Some Leafy Vegetables. Acta Horticulturae, 877, 1229-1232.
- [45] Jirovetz, L., Buchbauer, G., Stoynanava, A., Georgiev, E. and Damianova, S. (2005) Composition, Quality Control and Antimicrobial Activity of the Essential Oil of Cumin (*Cuminum cyminum* L.) Seeds from Bulgaria That Had Been Stored for up 36 Years. *International Journal of Food Science Technology*, 40, 305-310. http://dx.doi.org/10.1111/j.1365-2621.2004.00915.x
- [46] Wu, S., Zhang, W.M., Sun, X.M., Yang, Y. and Zhang, F.L. (2011) Analysis of Flavors and Flavonoid in Cumin Seeds from Different Producing Area. *China Condiment*, **3**, 96-98.
- [47] Hashemian, N., Pirbalouti, A.G., Hashemi, M., Golparvar, A. and Hamedi, B. (2013) Diversity in Chemical Composition and Antibacterial Activity of Essential Oils of Cumin (*Cuminum cyminum* L.) Diverse from Northeast of Iran. *Australian Journal of Crop Sciences*, 7, 1752-1760.
- [48] Beis, S.H., Azcan, N., Ozek, T., Kara, M. and Baser, K.H. (2000) Production of Essential Oil from Cumin Seeds. *Journal Chemistry of Natural Compounds*, **36**, 265-268. http://dx.doi.org/10.1007/BF02238331
- [49] Rebey, I.B., Jabri-Karoui, I., Hamrouni-Sellami, I., Bourgou, S., Limam, F. and Marzouk, B. (2012) Effect of Drought on the Biochemical Composition and Antioxidant Activities of Cumin (*Cuminum cyminum L.*) Seeds. *Industrial Crops* and *Products*, 36, 238-245. http://dx.doi.org/10.1016/j.indcrop.2011.09.013
- [50] Bettaieb, I., Bourgou, S., Sriti, J., Msaada, K., Limam, F. and Marzouk, B. (2011) Essential Oils and Fatty Acids Composition of Tunisian and Indian Cumin (*Cuminum cyminum* L.) Seeds: A Comparative Study. *Journal Science Food and Agriculture*, 91, 2100-2107. http://dx.doi.org/10.1002/jsfa.4513
- [51] Mohammadpour, H., Moghimipour, E., Rasooli, I., Fakoor, M.H., Astaneh, S.A., Sara, S.S. and Zeynab, J. (2012)

- Chemical Composition and Antifungal Activity of *Cuminum cyminum* L. Essential Oil from Alborz Mountain against *Aspergillus* Species. *Jundishapur Journal of Natural Pharmaceutical Products*, **7**, 50-55.
- [52] Pokhrel, S., Singh, R., Gautam, P., Dixit, V.K. and Das, A.J. (2012) Comparison of Antimicrobial Activity of Crude Ethanolic Extracts and Essential Oils of Spices against Five Strains of Diarrhoea Causing *Escherichia coli*. *International Journal of Pharmacy and Life Sciences*, **3**, 1624-1627.
- [53] Anwar, F., Sulman, M., Hussain, A.I., Saari, N., Iqbal, S. and Rashid, U. (2011) Physicochemical Composition of Hydro-Distilled Essential Oil from Coriander (*Coriandrum sativum L.*) Seeds Cultivated in Pakistan. *Journal of Medicinal Plants Research*, 5, 3537-3544.
- [54] Ebrahimia, S.N., Hadianb, J. and Ranjbarc, H. (2010) Essential Oil Compositions of Different Accessions of Coriandrum sativum L. from Iran. Natural Product Research, 24, 1287-1294. http://dx.doi.org/10.1080/14786410903132316
- [55] Viturro, C.I., Molina, A.C., Villa, W.C., Saavedra, O.N., Zampini, M., Gozalvez, M. and Garcia, E. (1999) Preliminary Assays of Adaptation in Jujuy (Argentina) of *Satureja hortensis* L., *Ocimum basilicum* L., and *Coriandrum sativum* Linn. *Acta Horticulturae*, **500**, 47-50.
- [56] Coskuner, Y. and Karababa, E. (2007) Physical Properties of Coriander Seeds (Coriandrum sativum L.). Journal of Food Engineering, 80, 408-416. http://dx.doi.org/10.1016/j.jfoodeng.2006.02.042
- [57] Özcan, M. and Chalchat, J.C. (2002) Essential Oil Composition of *Ocimum basilicum* L. and *Ocimum minimum* L. in Turkey. *Czech Journal of Food Sciences*, **20**, 223-228.
- [58] Saikia, N. and Nath, S.C. (2004) Evaluation of Essential Oil Characters of Sweet Basil (Ocimum basilicum Linn) Growing at Assam Valley Condition of Northeast India. Bioprospecting of Commercially Important Plants. Proceedings of the National Symposium on Biochemical Approaches for Utilization and Exploitation of Commercially Important Plants, Jorhat, 12-14 November 2003, 73-78.
- [59] Dob, T., Benabdelkader, T. and Chelghoum, C. (2006) Essential Oil Content and Composition of *Ocimum basilicum* L. Cultivated in Algeria. *Rivista Italiana EPPOS*, **41**, 13-21.
- [60] Sajjadi, S.E. (2006) Analysis of the Essential Oils of Two Cultivated Basil (*Ocimum basilicum L.*) from Iran. *DARU*, 14, 128-130.
- [61] Hussain, A.I., Anwar, F., Sherazi, S.T.H. and Przybylski, R. (2008) Chemical Composition, Antioxidant and Antimicrobial Activities of Basil (*Ocimum basilicum*) Essential Oils Depends on Seasonal Variations. *Food Chemistry*, 108, 986-995. http://dx.doi.org/10.1016/j.foodchem.2007.12.010
- [62] Abduelrahman, A.H.N., Elhussein, S.A., Osman, N.A. and Nour, A.H. (2009) Morphological Variability and Chemical Composition of Essential Oils from Nineteen Varieties of Basil (*Ocimum basilicum* L.) Growing in Sudan. *International Journal of Chemical Technology*, 1, 1-10. http://dx.doi.org/10.3923/ijct.2009.1.10
- [63] Benedec, D., Oniga, I., Oprean, R. and Tamas, M. (2009) Chemical Composition of the Essential Oils of *Ocimum basilicum* L. Cultivated in Romania. *Farmacia*, **57**, 5.
- [64] Hanif, M.A., Al-Maskari, M.Y., Al-Maskari, A., Al-Shukaili, A., Al-Maskari, A.Y. and Al-Sabahi, J.N. (2011) Essential Oil Composition, Antimicrobial and Antioxidant Activities of Unexplored Omani Basil. *Journal of Medicinal Plants Research*, 5, 751-757.
- [65] Chalchat, J. and Ozcan, M.M. (2008) Comparative Essential Oil Composition of Flowers, Leaves and Stems of Basil (Ocimum basilicum L.) Used as Herb. Food Chemistry, 110, 501-503. http://dx.doi.org/10.1016/j.foodchem.2008.02.018
- [66] Koba, K., Poutouli, P.W., Raynaud, C., Chaumont, J. and Sanda, K. (2009) Chemical Composition and Antimicrobial Properties of Different Basil Essential Oils Chemotypes from Togo. *Bangladesh Journal of Pharmacology*, **4**, 1-8.
- [67] Amin, G. (1991) Popular Medicinal Plants of Iran. Health and Education of Iran, 116.
- [68] Li, X.M., Tian, S.L., Pang, Z.C., Shi, J.Y., Feng, Z.S. and Zhang, Y.M. (2009) Extraction of *Cuminum cyminum* Essential Oil by Combination Technology of Organic Solvent with Low Boiling Point and Steam Distillation. *Food Chemistry*, 115, 1114-1119. http://dx.doi.org/10.1016/j.foodchem.2008.12.091
- [69] Wang, L., Wang, Z.M., Zhang, H.H., Zhang, H.Q. and Li, X.Y. (2009) Ultrasonic Nebulization Extraction Coupled with Headspace Single Drop Microextraction and Gas Chromatography-Mass Spectrometry for Analysis of the Essential Oil in Cuminum cyminum L. Analytica Chimica Acta, 647, 72-77. https://dx.doi.org/10.1016/j.aca.2009.05.030
- [70] Oroojalian, F., Kasra-Kermanshahi, R., Azizi, M. and Bassami, M.R. (2010) Phytochemical Composition of the Essential Oils from Three Apiaceae Species and Their Antibacterial Effects on Food-Borne Pathogens. *Food Chemistry*, 120, 765-770. http://dx.doi.org/10.1016/j.foodchem.2009.11.008
- [71] Abulimiti, Y., Xie, X.G., Ma, Q.L. and Hajiakber, A. (2011) Study on Chemical Compositions and Biological Characteristics of Essential Oil from *Cuminum cyminum* L. Produced in Hotan of Xinjiang Province. *Medicinal Plant*, 2, 62-64.

- [72] Wanner, J., Bail, S., Jirovetz, L., Buchbauer, G., Schmidt, E., Gochev, V., Girova, T., Atanasova, T. and Stoyanova, A. (2010) Chemical Composition and Antimicrobial Activity of Cumin Oil (*Cuminum cyminum*, *Apiaceae*). *Natural Product Communications*, 5, 1355-1358.
- [73] Misharina, T.A. (2001) Influence of the Duration and Conditions of Storage on the Composition of the Essential Oil from Coriander from Coriander Seeds. *Applied Biochemistry and Microbiology*, 37, 622-628. http://dx.doi.org/10.1023/A:1012315403828
- [74] Jeong-Sook, B. and Gil Hah, K. (2005) Fumigant Toxicity of the Constituents of Coriander (*Coriandum sativum*) Oil to *Blattella germanica*. *Korean Journal of Applied Entomology*, **44**, 37-41.
- [75] Anju, V., Pandeya, S.N., Yadav, S.K., Singh, S. and Soni, P. (2011) A Review on *Coriandrum sativum* Linn: An Ayurvedic Medicinal Herb of Happiness. *Japhr*, 1, 3.
- [76] Pino, J.A., Rosado, A. and Fuentes V. (1996) Chemical Composition of the Seed Oil of *Coriandrum sativum* L. from Cuba. *Journal of Essential Oil Research*, **8**, 97-98. http://dx.doi.org/10.1080/10412905.1996.9700565
- [77] Pande, K.K., Pande, L., Pande, B., Pujari, A. and Sah, P. (2010) Gas Chromatographic Investigation of *Coriandrum sativum* L. from Indian Himalayas. *New York Sciences Journal*, **3**, 43-47.
- [78] Arganosa, G.C., Sosulski, F.W. and Slikard, A.E. (1998) Seed Yields and Essential Oil of Northern-Grown Coriander (*Coriandrum sativum* L.). *Journal of Herbs*, *Spices and Medicinal Plants*, **6**, 23-32. http://dx.doi.org/10.1300/J044v06n02_03
- [79] Bhuiyan, M.N.I., Begum, J. and Sultana, M. (2009) Chemical Composition of Leaf and Seed Essential Oil of *Corian-drum sativum* L. from Bangladesh. *Bangladesh Journal Pharmacology*, 4, 150-153. http://dx.doi.org/10.3329/bjp.v4i2.2800
- [80] Soares, B.V., Morais, S.M., Fontenelle, R.O., dos Santos Fontenelle, R.O., Queiroz, V.A., Vila-Nova, N.S., Pereira, C.M.C., Brito, E.S., Neto, M.A.S., Brito, E.H.S., Cavalcante, C.S.P., Castelo-Branco, D.S.C.M. and Rocha, M.F.G. (2012) Antifungal Activity, Toxicity and Chemical Composition of the Essential Oil of *Coriandrum sativum L. Fruits. Molecules*, 17, 8439-8448. http://dx.doi.org/10.3390/molecules17078439
- [81] Nagella, P., Kim, M.Y., Ahmad, A., Thiruvengadam, M. and Chung, I. M. (2012) Chemical Constituents, Larvicidal Effects and Antioxidant Activity of Petroleum Ether Extract. *Journal of Medicinal Plants Research*, 6, 2948-2954. http://dx.doi.org/10.5897/JMPR11.992
- [82] Bhattacharya, A.K., Kaul, P.N. and Rao, B.R.R. (1998) Chemical Profile of the Essential Oil of Coriander (*Coriandrum sativum* L.) Seeds Produced in Andhra Pradesh. *Journal of Essential Oil Bearing Plants*, 1, 45-50.
- [83] Mazza, G. (2002) Minor Volatile Constituents of Essential Oil and Extracts of Coriander (*Coriandrum sativum* L.) Fruits. *Sciences des Aliments*, 22, 617-627. http://dx.doi.org/10.3166/sda.22.617-627
- [84] Zheljazkov, V.D., Callahan, A. and Cantrell, C.L. (2008) Yield and Oil Composition of 38 Basil (*Ocimum basilicum* L.) Accessions Grown in Mississippi. *Journal of Agricultural and Food Chemistry*, 56, 241-245. http://dx.doi.org/10.1021/jf072447y
- [85] Mondello, L., Zappia, G., Cotroneo, A., Bonaccorsi, I., Chowdhury, J.U., Usuf, M., et al. (2002) Studies on the Chemical Oil-Bearing Plants of Bangladesh. Part VIII. Composition of some Ocimum Oils, O. basilicum L. Var. Purpurascens, O. sanctum L. Green; O. sanctum L. Purple; O. americanum L., Citral Type, O. americanum L., Camphor Type. Flavour and Fragrance Journal, 17, 335-340. http://dx.doi.org/10.1002/ffj.1108
- [86] Opalchenova, G. and Obreshkov, D. (2003) Comparative Studies on the Activity of Basil—An Essential Oil from Ocimum basilicum L.—Against Multidrug Resistant Clinical Isolates of the Genera Staphylococcus, Enterococcus and Pseudomonas by Using Different Test Methods Journal Microbiology Methods, 54, 105-110. http://dx.doi.org/10.1016/S0167-7012(03)00012-5
- [87] Díaz-Maroto, M.C., Sánchez Palomo, E., Castro, L., González Viñas, M.A. and Pérez-Coello, M.S. (2004) Changes Produced in the Aroma Compounds and Structural Integrity of Basil (*Ocimum basilicum* L.) during Drying. *Journal Sciences Food and Agriculture*, **84**, 2070-2076. http://dx.doi.org/10.1002/jsfa.1921
- [88] Vieira, R.F. and Simon, J.E. (2000) Chemical Characterization of Basil (*Ocimum* spp.) Found in the Markets and Used in Traditional Medicine in Brazil. *Economic Botany*, 54, 207-216. http://dx.doi.org/10.1007/BF02907824
- [89] Jirovetz, L. and Buchbauer, G. (2001) Analysis, Chemotype and Quality Control of the Essential Oil of Anew Cultivated Basil (*Ocimum basilicum* L.) Plant from Bulgaria. *Scientia Pharmaceutica*, **69**, 85-89.
- [90] Gurbuz, B., Ipek, A., Basalma, D., Sarihan, E.O., Sancak, C. and Ozcan, S. (2006) Effect of Diurnal Variability on Essential Oil Composition of Sweet Basil (*Ocimum basilicum L.*). *Asian Journal of Chemistry*, **18**, 285-288.
- [91] Ismail, M. (2006) Central Properties and Chemical Composition of *Ocimum basilicum* Essential Oil. Pharmaceutical Biology, **44**, 619-626. http://dx.doi.org/10.1080/13880200600897544
- [92] Politeo, O., Jukic, M. and Milos, M. (2007) Chemical Composition and Antioxidant Capacity of Free Volatile Agly-

- cones from Basil (*Ocimum basilicum* L.) Compared with Its Essential Oil. *Food Chemistry*, **101**, 379-385. http://dx.doi.org/10.1016/j.foodchem.2006.01.045
- [93] Telci, I., Bayram, E., Yilmaz, G. and Avci, B. (2006) Variability in Essential Oil Composition of Turkish Basils (Ocimum basilicum L.). Biochemical Systematics and Ecology, 34, 489-497. http://dx.doi.org/10.1016/j.bse.2006.01.009
- [94] Nagai, A., Duarte, L.M. and Santos, D.Y. (2011) Influence of Viral Infection on Essential Oil Composition of *Ocimum basilicum* (Lamiaceae). *Natural Product Communications*, **6**, 1189-1192.
- [95] Rojas, M.M., Sanchez, Y., Abreu, Y., Espinosa, I., Correa, T.M. and Pino, O. (2012) Chemical Characterization and Antibacterial Activity of Essential Oils of *Ocimum basilicum* L. and *Ocimum basilicum var. Genovese* L. *Revista de Protección Vegetal*, 27, 130-134.