

# Optimization of a Tiger Nut-Based Yoghurt Formulation by Response Surface Methodology

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## Abstract

Tiger nut is a comestible tuber which offers many under products such as yoghurt. The parameters influencing the quality of yoghurt being numerous, response surface methodology was used to optimize the formulation in order to reach a low intake of milk powder. The volume of tiger nut milk, mass milk powder and sugar mass are the factors monitored while the titratable acidity (AT), pH, dry soluble extract (ESS), ash, viscosity and color are the expected responses in these tests. The data are processed with a degree of confidence  $p < 0.05$  associated with statistical analysis by the software Statgraphic Centurion XVI version 16.2.04. The different tests show that the factors have overall significant effects ( $p < 0.05$ ) on the acidity, the ashes and the Whiteness index. The linear and quadratic factors of tiger nuts milk as well as those linears of the milk powder have significant effects ( $p < 0.05$ ) on the pH and the ashes. The linear factors of milk powder have significant effects on DSE, AT and pH. The optimal formulation yielded a volume of tiger nut milk equal to 3.7 L/kg, a mass of powder milk of 63.4 g and 75 g of sugar for one kilogram of yoghurt. With a desirability of 75%, this model is apt to explain the results and the experimental values fit with the predicted ones and are within the norms. The proximate analysis of optimal yoghurt formulation shows that fat and proteins contents are respectively 5.67% and 2.2%. Calcium, magnesium and potassium contents are respectively 160, 40 and 180 mg in 100 g of yoghurt.

## Keywords

Optimization, Tiger Nut, Yoghurt

## 1. Introduction

The inadequacy of the distribution of plant resources around the world and the lack of processing of available resources in some parts of the world mean that today a large part of the world's population still suffers from the consequences of malnutrition. The latter remains a real scourge and one of the major public health problems in the world despite the considerable progress made. It is thought to be responsible for almost 55% of all deaths among children aged from 0 to 5 years, and this proportion is higher in poor countries; particularly in Africa. Nowadays, one of the axes of research in the field of food sciences is the development of functional foods that provide health benefits [1] [2] [3]. Tiger nut is one of the underused tubers in West Africa, which presents an interesting amino acids profile [4] [5], displaying high levels of essential fatty acids [5], soluble fibers, vitamins, minerals [6] [7] and phytochemicals such as isoflavones, flavonoids, terpenoids, alkaloids and saponins [8] [9]. The latter are known for their biological properties. Recent studies have proved the biological effects exhibited by the tiger nut tubers, such as anti-diabetic, cholesterol-lowering, hepatoprotective, aphrodisiac [10] [11] [12] [13], antibacterial [9] and galactogenic [14]. The diversity of tiger nuts based-products allows populations to benefit from its properties. In fact, the development of new fermented products based on tiger nut would procure specific healthy advantages for populations, compared to the conventional yogurts sold in the market. For the sake of minimising the powdered milk input during the production of this tiger nut-based yogurt, this study has been performed by varying the required volume of the tiger nut milk from a constant mass (tiger nut milk concentration) and adding sugar using design of experiments to optimize the process of tiger nut-based yogurt.

## 2. Material and Methods

### 2.1. Plant Material

The tiger nut tubers (*Cyperus esculentus*) represent the basic raw material for this present study (Figure 1). Tiger nut samples were obtained from local market. The samples were separated from stalks and stem and washed thoroughly under running water to remove dirt and weighed.



Figure 1. Dry tiger nut.

## 2.2. Extraction of the Tiger Nut Milk

The tiger nut tubers and the powdered milk used in this work were bought from the local market. The tubers have been cleaned many times with tap water before being soaked in water for twelve hours. Tubers after soaking are represented in **Figure 2**. Various concentrations have been set following the formulations. The milk is obtained by using a mixer of the brand Moulinex (France) at the maximum vitesse for 40 s three times, mixing the tubers with water. The blended mixture is then filtering through a muslin cloth two times.

## 2.3. Preparation of the Yoghurt

The extracted tiger nut milk has been mixed with a mass of milk and sugar according to the different formulations described by the design of experiments in order to obtain a final mass of 1 kg of yogurt. The pasteurization is carried out at 75°C during 5 minutes and then the mix is transferred into jars for fermentation at 45°C during 4 hours in an incubator and they are removed to be cooled.

## 2.4. Experimental Design and Statistical Analysis

Box-Behnken design are used in this study for the conception of the design of experiments. The response surface methodology (RSM) has been applied to optimize the factors and study the influences of the volume of tiger nut milk ( $X_1$ ) (concentration), of the mass of powdered milk ( $X_2$ ) and the mass of sugar ( $X_3$ ) on the responses. The tiger nut milk volume extracted from 1 kg of tiger nut tuber ranges from 2 liter to 5 liter, the masse of milk powder varies between 20 g and 120 g and sugar mass ranges from 50 g to 75 g. These values are taken in order to produce a firm yoghurt. The pH, titratable acidity, total soluble dry extract (SDE), viscosity, ashes content and color of the yogurt from the different formulations were evaluated as responses for the factors studied. The real levels of the variables are indicated in **Table 1**. Statgraphic centurion XVI Version 16.2.04 (32-bits) software was used for analysis. A second order polynomial regression model was applied to express the dependent variables as a function of the independent variables where  $Y$  is the measured response associated to each



**Figure 2.** Tiger nut tuber after 12 hour soaking.

**Table 1.** Level and code of independent variables, volume (tiger nut milk), mass of sugar, mass of milk level used for Box-Behnken experimental design.

Independent variables	Symbol	Unit	Minimum	Level	Maximum
			-1	0	1
Volume (tiger nut milk)	$X_1$	l/kg	2	3.5	5
Mass of sugar	$X_2$	G	50	62.5	75
Mass of milk	$X_3$	G	20	70	120

Note:  $X_1$  (A),  $X_2$  (B),  $X_3$  (C),  $X_1X_2$  (AB),  $X_1X_3$  (AC),  $X_2X_3$  (CB),  $X_{11}$  (AA),  $X_{22}$  (BB) and  $X_{33}$  (CC).  $X_1$ ,  $X_2$  and  $X_3$  mean respectively the letter A, B and C in the response surface and PARÉTO graphic.

combination of factor levels;  $\beta_0$  is a constant;  $\beta_i$  is the regression coefficient calculated from the observed experimental values of  $Y_i$ . The terms  $X_i$ ,  $X_iX_j$  and  $X_i^2$  represent the interaction and the quadratic terms, respectively.

### Physicochemical Analysis

#### 1) pH and titratable acidity

The samples were analysed to get pH and titratable acidity as described by Olubamiwa *et al.* (2006). A pH-meter of the brand Hanna HI-98128 was utilized to evaluate the pH of the different formulations of yogurt. The acidity has been measured by titration of 10 ml of yogurt with NaOH 0.1 N; and was expressed in equivalent gram of lactic acid/100g.

#### 2) Viscosity and color

Yoghurt samples were gently stirred with a plastic spoon prior to viscosity measurements. The viscosity was measured at 7°C using a Brookfield digital viscometer and is expressed in Cp. The color was measured with a colorimeter (CM-3600d, MINOLTA Co.; Japan) of the model D25A-9. The coordinates CIE Lab was obtained with the illuminating observer D65/10°. The color parameters such as  $L$ ,  $a^*$ ,  $b^*$ ,  $c^*$  and  $h^*$  were given by the colorimeter directly and are used to calculate the whiteness index (WI) and total color (TC) according to [15] and [16],

$$WI = 100 - \sqrt{(100 - L)^2 + a^{*2} + b^{*2}} \quad (1)$$

$$TC = \sqrt{L^2 + a^{*2} + b^{*2}} \quad (2)$$

#### 3) Physicochemical properties analysis

The ash content was determined according the [17] method. Two grams of the sample flours were weighed into the crucible and the weight taken. The crucible containing the samples were placed into the muffle furnace and ignited at 550°C. This temperature was maintained for three hours. The dry soluble extract was determined by using refractometer (ATAGO B024889 JAPON). Fat was determined in the soxhlet apparatus (AOAC, 1990) using hexane as the solvent of extraction. The Kjeldahl procedure based on the AOAC (1990) method was used for the protein content determination.

#### 4) Resultats and discussion

The analysis results are set out in **Table 2** and show the different variations

**Table 2.** The experimental design and obtained values of the responses.

Sample	Volume (l)	Mass of Sugar (g)	Mass of Milk (g)	Titratable acidity (g lactic acid/100g) ( $Y_1$ )	SDE (brix) ( $Y_2$ )	Ph ( $Y_3$ )	Ash (%) ( $Y_4$ )	Viscosity (cp) ( $Y_5$ )	Color		
									TC ( $Y_6$ )	H ( $Y_7$ )	WI ( $Y_8$ )
1	5.0	50.0	70.0	0.74 ± 0.02	16.6 ± 0.50	4.69 ± 0.005	0.22 ± 0.00	38.76 ± 0.66	85.43 ± 0.05	120.31 ± 0.25	81.27 ± 0.00
2	5.0	75.0	70.0	0.75 ± 0.00	16.65 ± 0.65	4.67 ± 0.005	0.54 ± 0.00	39.69 ± 0.54	85.20 ± 0.10	120.13 ± 0.26	81.01 ± 0.02
3	5.0	62.5	120.0	1.13 ± 0.02	22.45 ± 0.05	4.88 ± 0.01	0.94 ± 0.00	200.17 ± 0.46	91.82 ± 0.02	101.83 ± 0.04	81.97 ± 0.01
4	5.0	62.5	20.0	0.45 ± 0.00	11.90 ± 0.10	4.47 ± 0.005	0.06 ± 0.00	288.68 ± 0.57	81.92 ± 0.01	109.89 ± 0.00	79.87 ± 0.01
5	3.5	62.5	70.0	0.63 ± 0.02	17.45 ± 0.35	4.57 ± 0.005	0.99 ± 0.08	30.21 ± 0.09	85.46 ± 0.00	116.83 ± 0.06	80.47 ± 0.00
6	3.5	75.0	20.0	0.49 ± 0.01	17.95 ± 0.15	4.39 ± 0.005	0.77 ± 0.14	30.53 ± 0.24	79.50 ± 0.33	122.67 ± 0.8	77.17 ± 0.18
7	3.5	75.0	120.0	1.17 ± 0.00	23.55 ± 0.25	4.69 ± 0.005	1.23 ± 0.14	109.67 ± 0.33	81.27 ± 0.00	103.43 ± 0.02	81.74 ± 0.00
8	3.5	62.5	70.0	0.81 ± 0.01	17.45 ± 0.50	4.61 ± 0.005	1.01 ± 0.05	176.23 ± 0.87	89.84 ± 0.00	109.86 ± 0.03	80.43 ± 0.01
9	3.5	50.0	20.0	0.50 ± 0.00	12.70 ± 0.00	4.45 ± 0.005	0.75 ± 0.00	219.87 ± 0.84	88.27 ± 0.01	109.44 ± 0.03	78.64 ± 0.00
10	3.5	62.5	70.0	0.80 ± 0.00	21.40 ± 0.15	4.66 ± 0.005	0.99 ± 0.02	267.89 ± 0.56	80.36 ± 0.01	103.10 ± 0.05	81.77 ± 0.00
11	3.5	50.0	120.0	1.21 ± 0.00	21.40 ± 0.35	4.77 ± 0.01	1.06 ± 0.04	298.90 ± 0.17	88.61 ± 0.01	113.39 ± 0.04	81.03 ± 0.00
12	2.0	62.5	120.0	0.72 ± 0.00	21.75 ± 0.15	4.68 ± 0.00	1.37 ± 0.01	74.36 ± 0.08	88.19 ± 0.20	98.82 ± 0.43	79.83 ± 0.06
13	2.0	75.0	70.0	0.86 ± 0.00	21.10 ± 0.10	4.97 ± 0.005	1.01 ± 0.02	74.52 ± 0.16	86.04 ± 0.01	109.37 ± 0.01	78.42 ± 0.00
14	2.0	50.0	70.0	0.68 ± 0.01	19.15 ± 0.45	4.80 ± 0.005	0.97 ± 0.01	266.16 ± 1.07	83.21 ± 0.00	104.87 ± 0.02	79.58 ± 0.01
15	2.0	62.5	20.0	0.51 ± 0.01	15.65 ± 0.05	4.88 ± 0.01	0.86 ± 0.02	296.04 ± 0.23	81.07 ± 0.01	113.78 ± 0.01	77.10 ± 0.00

Note:  $Y_1$  (Soluble dry extracts),  $Y_2$  (titratable acidity),  $Y_3$  (pH),  $Y_4$  (ashes),  $Y_5$  (viscosity),  $Y_6$  (total color),  $Y_7$  (angle h),  $Y_8$  (whiteness index). DSE (dry soluble extract), gram (g).

of the parameters studied. The results obtained from the ANOVA are shown in **Table 3**.  $R^2$  Values range from 42.63% to 98.18% and was established in **Table 3**. Responses on witch factor have global significant effects (linear, interaction, or quadratic) are marked (\*) (**Table 3**). The p-values below 0.05 indicate that the quadratic model of the yoghurt formulation is statistically significant at a 95% confidence interval. Titratable acidity, soluble dry extract, pH, ash, viscosity, total color, hue, and whiteness index are described respectively by  $Y_1$ ,  $Y_2$ ,  $Y_3$ ,  $Y_4$ ,  $Y_5$ ,  $Y_6$ ,  $Y_7$  and  $Y_8$  which are regression equation predicting the relationship between the factors (**Table 4**).

### 5) Titratable acidity

Titratable acidity is a very important factor in the characterization and acceptability of fermented dairy products. The variance analysis of the regression models gives a  $R^2$  value of 90.67%. The titratable acidity varies from 0.45% to 1.21%. Response surface plot of the relationship is shown as **Figure 3**. The effect of independent variables is represented in **Figure 4**. The high value is found in formulation (3.5 l; 50 g; 120 g) and the lowest value in formulation (5 l; 62.5 g; 20 g). The linear factor milk powder mass is the only one to have the most influence on the acidity of yogurts, with significant positive effects ( $p < 0.05$ ) (**Figure 4**). The larger the milk powder mass, the greater the effects on the titratable acidity. Milk is the main element used by lactic acid bacteria during

**Table 3.** ANOVA for the experimental variables.

Model	Titrateable Acidity	Soluble Dry Extracts	pH	ASH	Viscosity	TC	H	WI
Transformation	None	None	None	None	None	None	None	None
D.D.L. Model	9	9	9	9	9	9	9	9
Probability	0.0390*	0.0869	0.0511	0.0028*	0.7665	0.8823	0.3170	0.0008*
D.D.L. error	5	5	5	5	5	5	5	5
Standard error	0.125632	2.06723	0.0901018	0.101923	125.447	4.63919	6.22152	0.334627
Mean absolute error	0.062222	1.03444	0.0432222	0.0487778	60.9667	2.983	2.32433	0.156333
R-squared	90.67	86.57	89.47	96.95	51.63	42.63	74.11	98.18
Adjusted R-squared	73.88	62.38	70.51	91.47	0.00	0.00	27.50	94.90

\*Significant at 0.05.

**Table 4.** Equation of regression of response.

$Y_1$	$Y_1 = 1.58198 + 0.257926X_1 - 0.05016X_2 - 0.000177X_3 - 0.028704X_1^2 + 0.000483X_2^2 + 0.000008X_3^2 - 0.002267X_1X_2 + 0.00156667X_1X_3 - 0.000012X_2X_3$	(3)
$Y_2$	$Y_2 = 16.6632 + 2.67398X_1 - 0.318867X_2 + 0.085675X_3 - 0.500926X_1^2 - 0.0253333X_1X_2 + 0.0268333X_1X_3 + 0.00470667X_2^2 - 0.00124X_2X_3 - 0.000240833X_3^2$	(4)
$Y_3$	$Y_3 = 5.54418 - 0.536037X_1 + 0.00286X_2 - 0.00192833X_3 + 0.0714815X_1^2 + 0.000053X_2^2 - 0.000019X_3^2 - 0.002533X_1X_2 + 0.00203X_1X_3 - 0.000008X_2X_3$	(5)
$Y_4$	$Y_4 = -0.878881 + 0.186537X_1 + 0.0549X_2 - 0.00486X_3 - 0.101481X_1^2 + 0.00373333X_1X_2 + 0.001233X_1X_3 - 0.000533X_2^2 + 0.00006X_2X_3 + 0.000016X_3^2$	(6)
$Y_5$	$Y_5 = 66.0527 - 198.462X_1 + 26.6765X_2 - 5.203X_3 - 0.723889X_1^2 + 2.5676X_1X_2 + 0.4439X_1X_3 - 0.330872X_2^2 + 0.000044X_2X_3 + 0.0233325X_3^2$	(7)
$Y_6$	$Y_6 = 60.0084 + 1.218X_1 + 0.815993X_2 - 0.00922833X_3 + 0.115X_1^2 - 0.0354667X_1X_2 + 0.00926667X_1X_3 - 0.006904X_2^2 + 0.000572X_2X_3 - 0.0000795X_3^2$	(8)
$Y_7$	$Y_7 = 167.815 + 8.15278X_1 - 3.01338X_2 + 0.551525X_3 - 0.536111X_1^2 + 0.031656X_2^2 - 0.0010575X_3^2 - 0.0624X_1X_2 + 0.023X_1X_3 - 0.009276X_2X_3$	(9)
$Y_8$	$Y_8 = 74.6532 + 0.2445X_1 + 0.11616X_2 - 0.005905X_3 - 0.011667X_1^2 - 0.002072X_2^2 - 0.000281X_3^2 + 0.012X_1X_2 - 0.0021X_1X_3 + 0.001272X_2X_3$	(10)

\*Co-efficient are significant ( $p < 0.05$ ).

fermentation and therefore in the production of lactic acid and acidic compounds. As such, it plays a key role in the acidity of yogurts. The lactose contained milk is an ideal substrate for yogurt cultures. These microorganisms use milk lactose to release more lactic acid, which leads to the influence of milk on acidity. Similar results were found by [18], it was found that the addition of sodium caseinate increased the acidity of fermented tigernut. The acidic compounds of yoghurt are intermediates of several metabolic processes that

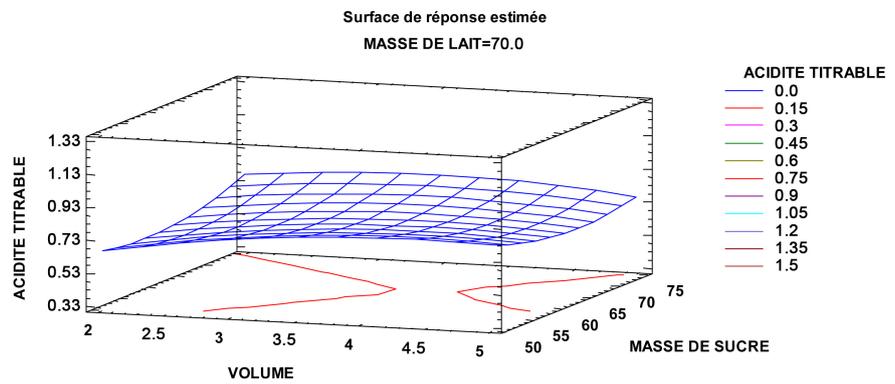


Figure 3. Titratable acidity response surface.

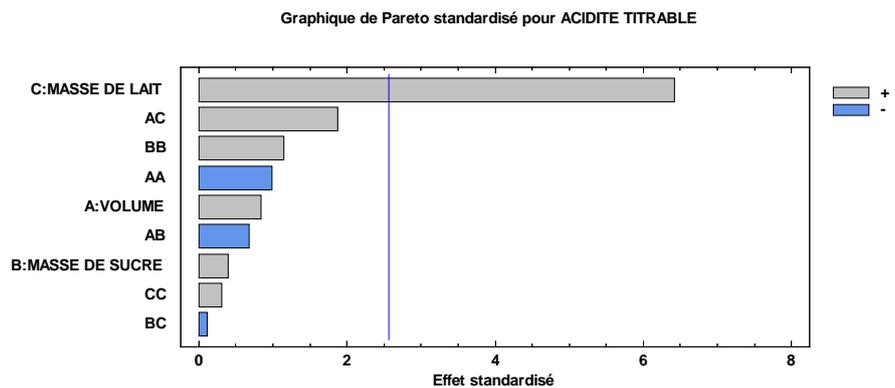


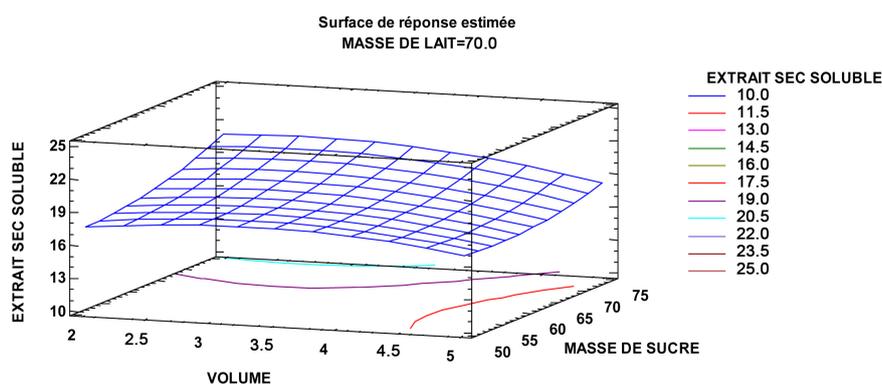
Figure 4. Pareto graph of the effects on titratable acidity.

influence the shelf life. However, given the fiber content of the tiger nut milk, it has been reported that the hydrolysis of these compound during the fermentation could increase glucose and maltose levels, which are converted into organic acids or alcohols by microbial activity and which directly influences the acidity of the environment [19]. On the other hand, pH and acidity can be affected by several factors such as the initial microbial load of milk, milk and handling hygiene conditions [20] but also the amount and type of materials solid, culture activity and fermentation period [21]. As for the volume of tiger nut milk, it has very little influence on the lactic acid content ( $p > 0.05$ ). The less the volume of tiger nut milk is concentrated, the lower the lactic acid content, because the tiger nut milk itself has a low lactic acidity of the order of 0.15% [22]. Interactions between milk nutmeg and milk powder, and sugar interactions, are also important but not significant ( $p > 0.05$ ). The co-efficient of determination predicting the mean score for aroma explained up to 92.41% (Table 3) variability in data. The regression Equation (3) predicting the relationship between titratable acidity and independent variables is established in Table 4.

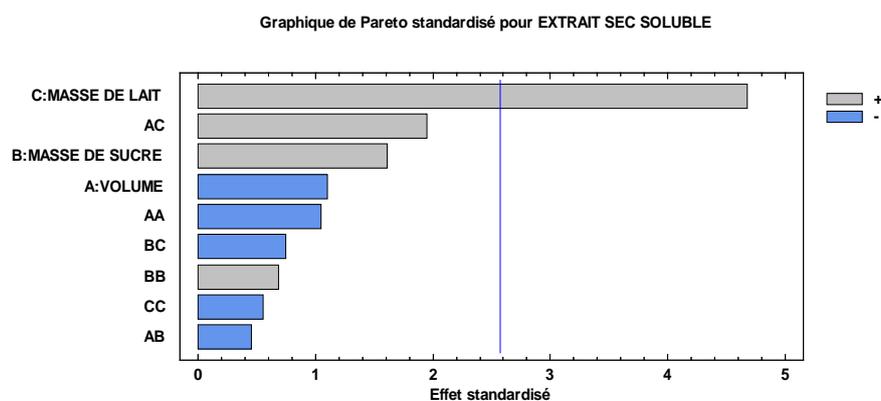
### 6) Soluble dry extract

ANOVA indicates an  $R^2$  equal to 86.58% on the variability of soluble solids content. Its adjusted  $R^2$  is 62.38%. The various yogurt samples have dry soluble extract contents ranging between 11.90% to 23.55%. However, the milk powder

mass remains the only factor with a significant ( $p < 0.05$ ) positive influence on the dry soluble extract content. Nevertheless, the linear factor of the sugar mass and the interaction between the volume and the mass of powdered milk and the concentration of the tiger nut milk have less significant ( $p > 0.05$ ) but positive effects. A visual illustration of the relationship is shown in **Figure 5** and its mathematical relationship expressed in Equation (4) is established in **Table 4**. The high value of soluble dry extract is found in formulation (3.5 l; 75 g; 120 g) and the lowest value in formulation (5 l; 62.5 g; 20 g). The higher the milk powder mass, the higher the soluble solids content. Tiger nut milk dry soluble extract would be important the more the volume is important because of the insolubility of certain compounds. If the milk is highly concentrated, it cannot extract the maximum of soluble compounds and the process can increase the amount of insoluble compounds [18]. From the foregoing, it is apparent that the amount of dry soluble extract of a sample is correlated with the quantities of milk powder, sugar and the concentration of milk (**Figure 6**). These results would be due to as much to the milk used to bacteriological activities. The milk used during the formulation is instant milk powder, thus containing a high content of soluble solids. As for lactic acid bacteria, the lactic acid they produce during fermentation is an organic acid that preserves and concentrates the dry matter of milk.



**Figure 5.** Soluble dry extract response surface plot.



**Figure 6.** Pareto graph of the effects on soluble dry extract.

## 7) pH

Using a multiple regression analysis of the experimental results, the pH is obtained by the second-order polynomial Equation (5) (Table 4) which describes the interaction between the three independent factors. The analysis of variance of the coefficient of determination  $R^2$  indicates that the model accounts for 89.47% of the pH variability. The adjusted R-squared is 70.51%. A visual illustration of pH relationship with independent variables is shown in Figure 7 and its mathematical relationship expressed in Equation (5) is established in Table 4. The effects of the quadratic factors of the concentration of the tiger nut milk and the interaction factors of the concentration of the tiger nut milk and the milk powder mass positively and significantly ( $p < 0.05$ ) affect the yogurt pH (Figure 8). The effect of the sugar mass was not significant ( $P > 0.05$ ) on the pH value (Figure 8). The pH of the various tiger nut yogurts varies from 4.39 to 4.97. The highest value was recorded in formulation (2 l; 75 g; 70 g) while the formulation (3.5 l; 75 g; 20 g) had the lowest pH. The tiger nut milk is very little acid but during fermentable combined with milk powder, the production of lactic acid intensifies, the medium acidifies where the pH drop to values around 4. These results are on the other hand below those of [23], for yoghurt based on

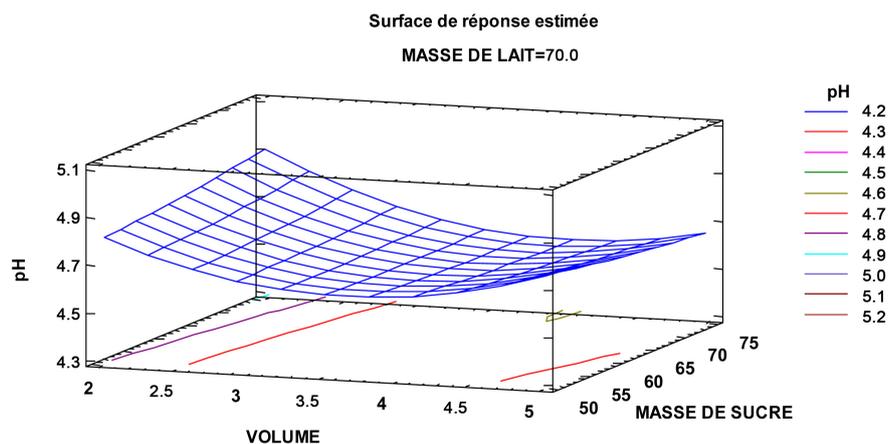


Figure 7. pH response surface.

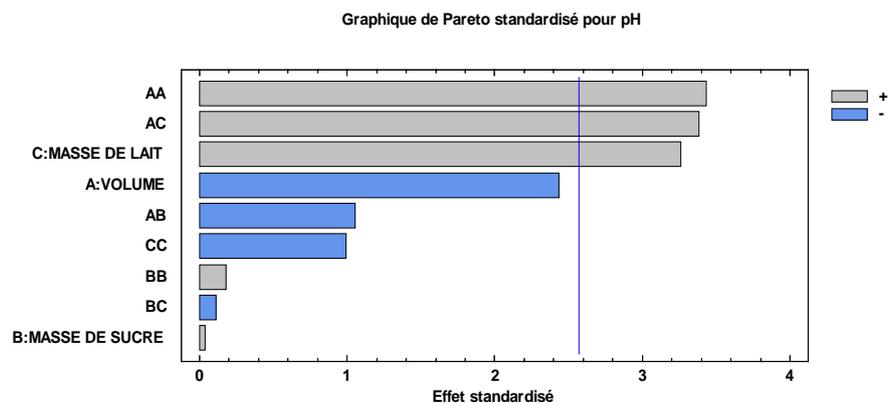


Figure 8. Pareto graph of the effects on the pH.

pure yellow tiger nut with a pH equal to 6.10, yoghurt mixed yam and soybean for a pH 6.32 and yoghurt mixed soya and coconut of pH 6.58. These reported pH values are dependent on tiger nut milk which has a pH of plus or minus 6.6 [22]. The pH is an essential factor for dairy products such as yoghurt; it plays a large role in consumer acceptance of the product and good microbiological quality. A non-standard pH would result in a finished product unfit for consumption. The values reported during this experiment are therefore acceptable and thus demonstrate compliance with the various manufacturing steps: pasteurization, incubation time and sufficient lactic ferments, to reach the appropriate pH.

### 8) Ash

Analysis of variance (ANOVA) of the  $R^2$  indicates that the model accounts for 96.95% (Table 3) of the ash rate variability. The results of analysis carried out, reveal that the total ash content of yoghurt ranges from 0.06% to 1.37%. The highest value of 1.37% was found in formulation (3.5 l; 75 g; 120 g) while the lowest value was found in formulation (5 l; 62.5 g; 20 g) (Table 2). These values are moderately higher than those found by [24] Chukwuma *et al.* (2016), for yoghurts made from pure sapling (0.43%), coconut (0.36%), cow's milk (0.73%) and coconut mix and tiger nut (0.68%). The interaction of the variables of the formulation is described in the following three-dimensional response surface (Figure 9) and the mathematical function (Equation (6)) was the best to describe the relationship between the ashes and the factors and (Table 4). The results show that milk mass ( $p < 0.05$ ), volume ( $p < 0.05$ ) and the effect of quadratic factors of volume ( $p < 0.05$ ) are significant variables (Figure 10). The effect of the sugar mass is not significant ( $p > 0.05$ ) (Figure 10) on the ash content. The ash rate could be important when the tiger nut is concentrated in addition to the milk supply. Factors with a significant influence ( $p < 0.05$ ) on the ash content of the samples are the volume of tiger nut milk, which has a negative impact, inversely proportional to the ash content, as well as its interactions and milk powder mass, which has a positive impact. It has been found that the mass of ash increases with the elevation of the mass of milk.

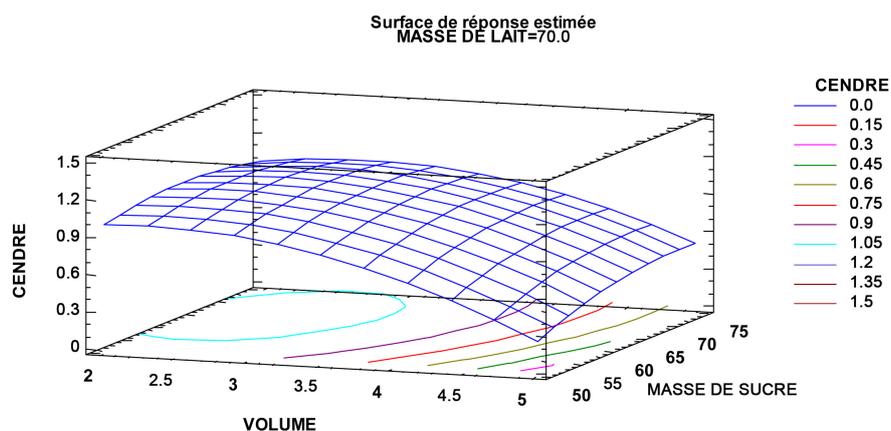
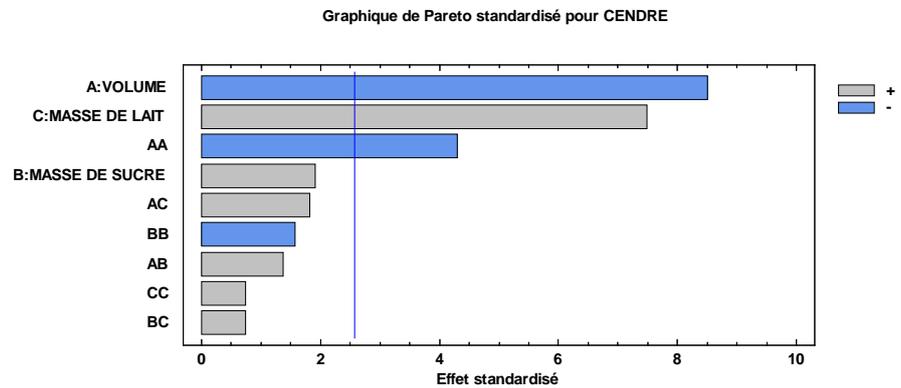


Figure 9. Ash content response surface.



**Figure 10.** Pareto graph of the effects on the ash content.

### 9) Viscosity

The  $R^2$  statistic analysis indicates that the model explains 51.6314% of the variability of the viscosity. Values reported for viscosity range from 296.04 to 30.21 Cp. A visual illustration of the relationship between the viscosity and independent variables is shown in **Figure 11** and its mathematical relationship expressed in Equation (7) is established in **Table 4**. These are influenced mainly by the sugar mass and milk powder interactions, although not significant ( $p > 0.05$ ) (**Figure 12**). The mass of sugar has a negative influence on the viscosity of yogurts. The more it increases and the less yoghurt is viscous. Indeed, lactic acid bacteria proliferate at 45°C and metabolize sugars to acids. This increased acidity causes denaturation of the proteins and forms a gel resulting from the viscous nature characteristic of yogurts. However, a high sugar intake in yogurt would contribute to the proliferation of acidic compounds and thus to a drop in viscosity. The low protein yogurts tend to have low viscosity because of the low water retention capacity of coagulum. These variations in viscosity could be explained by the protein rearrangements that take place where the protein-protein interactions are favored by the low energy bonds, which would lead to an increase in viscosity [25]. As a result, the gel network is even more important that the proportion of protein in yogurt is important. Also, the degradation of lactose into lactic acid leads to a drop in pH then gelling the medium with irreversible changes [26]. Similarly, proteolysis of caseins is induced during technological processes by the addition of lactic acid bacteria producing their proteases. This phenomenon brings about changes in the viscosity of the product and in the consistency with the production of amino acids and peptides. However, for some tests, huge differences in values could be observed. These differences could be attributed not only to the imprecise amounts of ferments incorporated during production. Similarly, during heat treatments, a complex between casein and protein is formed, but the degree of formation of this complex depends on the temperature and the treatment time, as well as the amount of whey protein present in the whey protein milk [27]. The formation of this complex has an influence on the hydrophilic and hydration properties of the casein micelle and therefore on the rheological properties of yoghurt, in

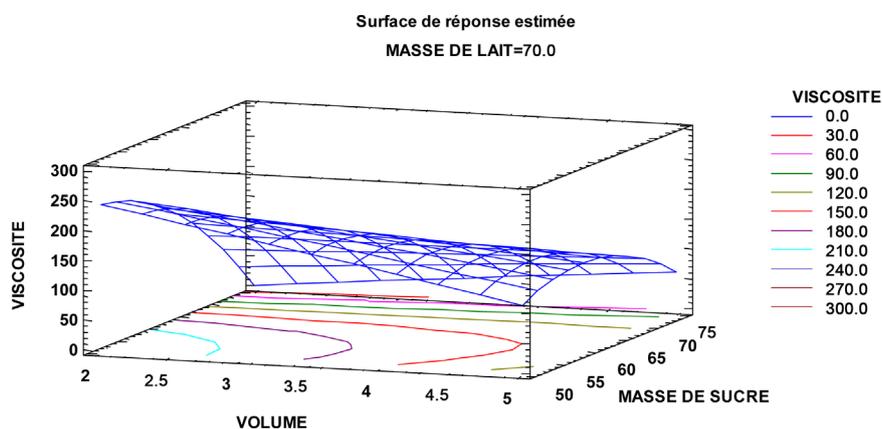


Figure 11. Viscosity response surface.

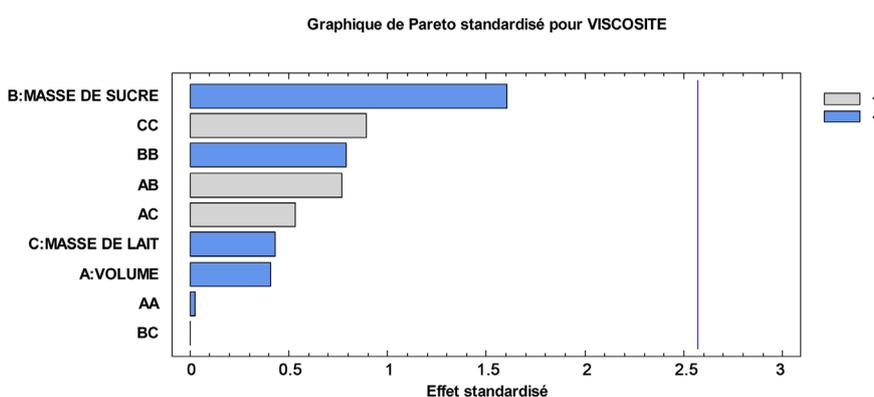


Figure 12. Pareto graph for the viscosity.

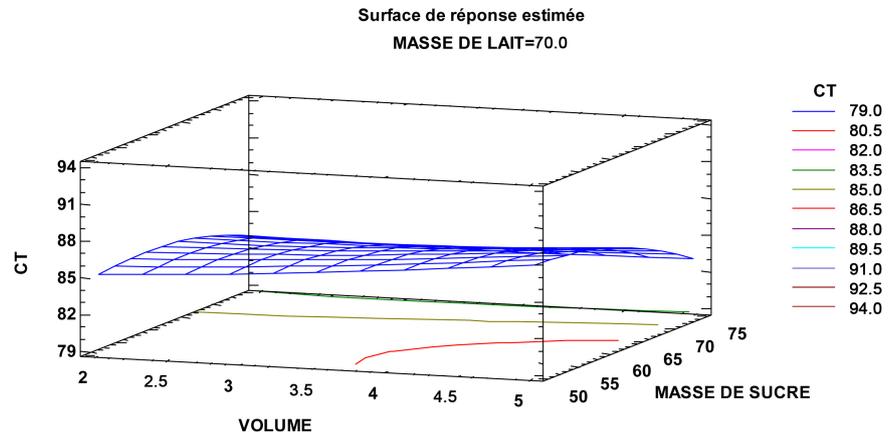
particular its viscosity.

#### 10) Color parameters

In food science, color is one of the main assets of a product that can lead to the purchase by the consumer [28]. It is an important parameter in the making and sometimes tracking of certain products. It can be used as a quality control parameter during storage and its degradation can be attributed to possible reactions. Indeed, different sub-parameters fall into the measure of the color such as the h angle, the chromacy c and the variables  $a^*$  (green to red),  $b^*$  (blue to yellow),  $L^*$  or clarity (black to white) which make it possible to calculate the total color TC and the whiteness index WI.

#### 11) Total color

The analysis results give total color values ranging from 79.5 to 91, 82. These observed variations can be attributed to different opacity levels, related to the level of aggregation of particles. The high value is found in formulation (3.5 l; 50 g; 120 g) and the lowest value in formulation (3.5 l; 75 g; 20 g). The co-efficient of determination predicting the mean score for total color explained up to 42.63% (Table 3) variability in data. The regression Equation (8) predicting the relationship between total color and process variables is established in Table 4 and the response surface is rrepresented by Figure 13. The higher the brightness



**Figure 13.** Surface response for the total color.

values, the higher the opacity, the lower the chromium, which also corresponds to a higher brightness index. No factors had a significant influence on the total yoghurt color ( $p > 0.05$ ) (Table 3). However, the total color of the samples predominate when the mass of milk powder increases and the tiger nut dilution (Figure 14). The results of analysis obtained are in agreement with those of [29], who studied color change as affected by the addition of casein. The addition of proteins significantly affected the yoghurt color. As a result, high milk powder content would mean high milk protein content and thus better protein coagulation. Protein coagulation affects the structure and surface properties of yoghurt and improves the absorption of light [30].

### 12) Hue tint angle

Statistical analysis of the coefficient of determination  $R^2$  indicates that the model accounts for 74.11% of the variability of Hue. The adjusted  $R^2$  is 27.50%. The regression Equation (9) predicted the relationship between Hue value and independent variables is established in Table 4. H value increase with increasing the tiger volume obtain for on kilogram of tuber and decrease with the milk mass (Figure 15). No factor has a significant effect (Figure 16). However, the effects of the linear and interaction factors are much more pronounced ( $p < 0.05$ ) on the H-hue angle but negative. On the other hand, the linear factor of the tiger nut volume and the quadratic factor of the sugar mass have insignificant influence but positive effects on the hue angle.

### 13) Whiteness index (WI)

The whiteness index (WI), which measures the overall whiteness of the food product or its deviation from the white color, ranges from 77.10 to 81.97 (Table 2). Variance analysis of the coefficient of determination  $R^2$  indicates that the model explains 98.18% of the variability of WI however the adjusted  $R^2$  is 94.90%. The regression Equation (10) predicting the relationship between whiteness and process variables is established in Table 4 and the interaction of the variables of the formulation is described by the three-dimensional response surface in Figure 17. The high value of WI is obtained in the formulation (5 l,

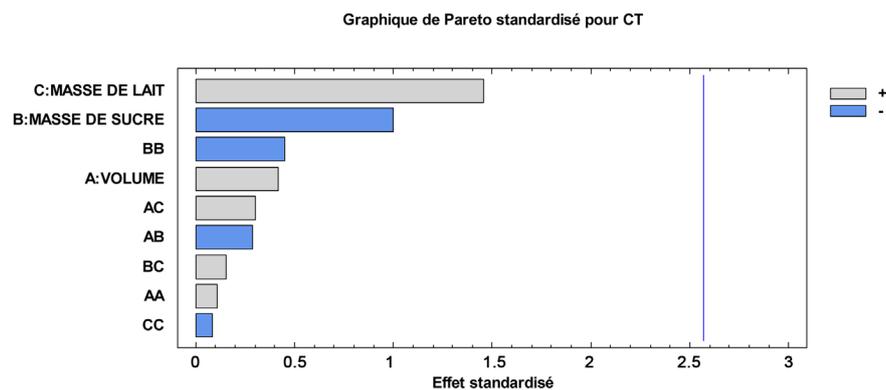


Figure 14. Pareto graph for the total color.

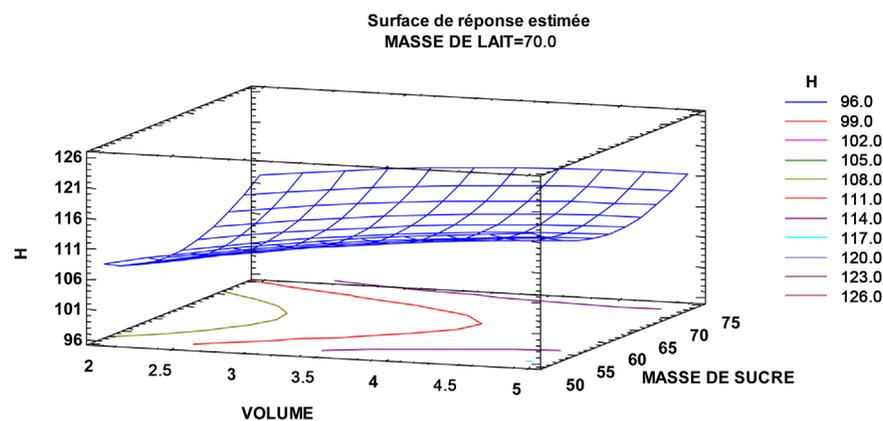


Figure 15. Hue angle response surface.

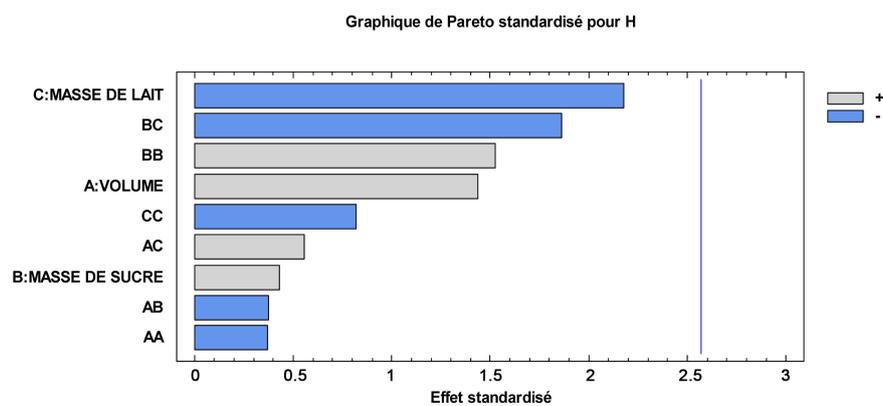


Figure 16. Pareto graph for the hue angle.

62.5 g and 120 g) and the low respectively for tiger nut volume, sugar mass and milk mass. The linear factors of milk mass ( $p < 0.001$ ), and of tiger nut milk volume (concentration) ( $p < 0.0001$ ) are the most factors which have the most significant effect, the interactive factor of sugar and milk masses ( $p < 0.05$ ) and the quadratic factor of the milk mass have significant influences on the whiteness index of yogurts (Figure 18). The increase in volume (concentration) causes a highly significant increase in bleaching index (Figure 17). WI is closely related

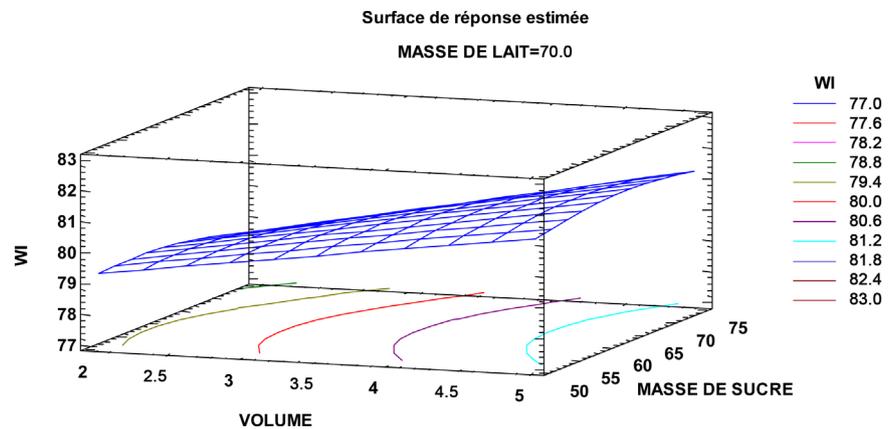


Figure 17. WI response surface.

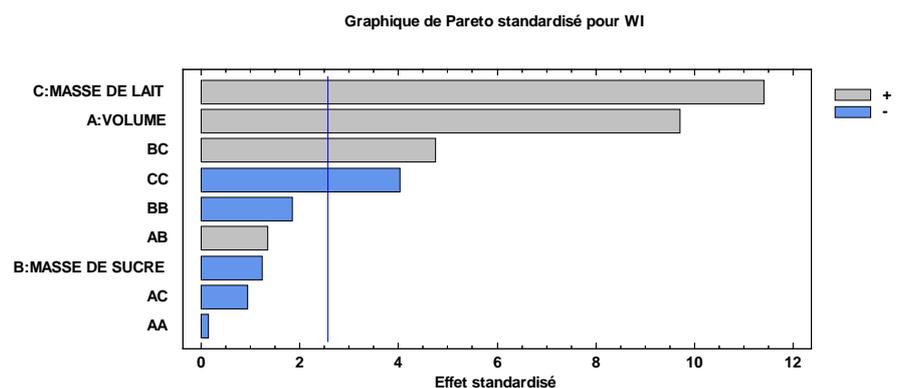


Figure 18. Pareto graph of the index WI.

to brightness  $L$ . A high value of  $L$  in the case of yogurt is a good quality factor however a drop in  $L$  can be attributed to color degradation [31] and can be encouraged by the oil content. The variations in the whiteness index of yogurts are due to the physico-chemical properties of the various components and the pretreatment carried out during production, in particular pasteurization which could cause the gelatinization of the starch contained in the tiger nut milk but also non-enzymatic browning reactions and thus interactions between sugar reducing sugars and milk proteins. It has been found that all values of  $a^*$  are negative thus revealing the presence of green compounds in yoghurt if it is known that tiger nut contains family B vitamins and certain phytochemicals. These findings were made by [32] and attributed these negative values by the presence of tryptophan, tyrosine and riboflavin.

#### 14) Optimization

Multi-response optimization of RSM was applied to determine the optimal combination in yoghurt preparation. The optimum formulation predicted by the regression model is the follows: for tiger nut milk volume (3.77 l), for milk mass (63.84 g) and 75 g for sugar mass. These conditions were tested and compared to the predicted optimal response. Yoghurt optimal sample were prepared using the derived optimum formulation conditions to check the validity of surface

**Table 5.** Optimal values of process variables.

Factors	Optimal value	Responses	Desirability	Prevision	Experimental measures	Inferior limit at 95.0%	Superior limit at 95.0%
		Titration acidity	1	0.80	0.66 ± 0.06	0.596	1.004
Volume (liter)	3.77	Soluble dry extract	0.739	20.03	18 ± 0.1	16.66	23.39
		pH	0.625	4.59	4.5 ± 0.004	4.44	4.74
		Ash	0.662	0.89	1.25	0.726	1.06
Mass of powdered milk (g)	63.84	WI	0.623	79.93	75.24 ± 0.003	79.38	80.47
		H	0.766	117.37	115.27 ± 0.145	107.26	127.49
Mass of sugar (g)	75.0	CT	0.746	82.63	80.41 ± 0.19	75.08	90.17
		Viscosity	0.886	43.17	36.4	-160.87	247.23

**Table 6.** Proximate analysis of optimal yoghurt.

Parameter (/100g)	Content
Proteins %	2.2
Fat %	5.67
Calcium (mg)	160
Magnesium (mg)	20
Potassium (mg)	180

response model. The experimental data were compared with predicted values in order to verify adequacy of final reduced flour (**Table 5**). The overall desirability is 75% and this shows that is fit to explain this study. The results of the proximate composition of the tiger nut yoghurt are presented in **Table 6**. Protein and fat content are respectively 2.5% and 5.67%. Calcium, magnesium and potassium content are respectively are 160 mg, 40 mg and 180 mg in 100 g of the optimal yoghurt.

### 3. Conclusion

The response surface methodology has successfully optimized the tiger nut yoghurt formulation process. The experimental data after optimization and the predicted data are significantly the same overall. The development of yoghurt based on tiger nuts by adding milk powder can constitute on the one hand a real nutritional advantage in view of these many biological properties of nutsedge and other departure its valuation. Optimization has made it possible to minimize milk powder intake. The physicochemical and biochemical analysis confirms the nutritional value of the product compared to those already existing on the market. Further studies on microbial quality and stability would predict the

conditions of optimized product use. Finally, the production, utilization, and consumption of tigernut-based yoghurt should be encouraged because such product will be helpful in providing nutritious, safe and wholesome food for the poor and malnourished populations in developing country.

### Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

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