

# The Exogenous Application of Absciscic Acid Induce Accumulation of Anthocyanins and Phenolic Compounds of the 'Rubi' Grape

Francisco José Domingues Neto<sup>1\*</sup>, Adilson Pimentel Junior<sup>1</sup>, Cristine Vanz Borges<sup>1</sup>,  
Silvia Regina Cunha<sup>1</sup>, Daniel Callili<sup>1</sup>, Giuseppina Pace Pereira Lima<sup>2</sup>, Sergio Ruffo Roberto<sup>3</sup>,  
Sarita Leonel<sup>1</sup>, Marco Antonio Tecchio<sup>1</sup>

<sup>1</sup>School of Agriculture, São Paulo State University (UNESP), Botucatu, Brazil

<sup>2</sup>Institute of Biosciences, São Paulo State University (UNESP), Botucatu, Brazil

<sup>3</sup>Department of Agronomy, Londrina State University (UEL), Londrina, Brazil

Email: \*fjdominguesneto@hotmail.com

**How to cite this paper:** Domingues Neto, F.J., Pimentel Junior, A., Borges, C.V., Cunha, S.R., Callili, D., Lima, G.P.P., Roberto, S.R., Leonel, S. and Tecchio, M.A. (2017) The Exogenous Application of Absciscic Acid Induce Accumulation of Anthocyanins and Phenolic Compounds of the 'Rubi' Grape. *American Journal of Plant Sciences*, 8, 2422-2432.

<https://doi.org/10.4236/ajps.2017.810164>

**Received:** August 9, 2017

**Accepted:** September 12, 2017

**Published:** September 15, 2017

Copyright © 2017 by authors and

Scientific Research Publishing Inc.

This work is licensed under the Creative Commons Attribution International

License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

## Abstract

The absciscic acid (ABA) is related to the formation of certain polyphenols, such as anthocyanin, improving the nutritional and commercial quality of the red grapes. In this study, we verified whether the exogenous application of *S*-ABA favors the accumulation of anthocyanin, the total phenolic compounds and the antioxidant activity of the 'Rubi' grape. The experiment was performed during the cycle 2014/2015 with the 'Rubi' vine (*Vitis vinifera* L.) grafted in the rootstock 420-A. The experimental design was composed of randomized blocks, with four treatments and six blocks, in a total of 24 experimental parcels, constituted of one plant each. We used the *S*-ABA isomer in different concentrations and times of application, 400 mg·L<sup>-1</sup> *S*-ABA in the beginning of the ripening (BR); 400 mg·L<sup>-1</sup> at BR + 200 mg·L<sup>-1</sup> at 25 days after first application (DAFA); 400 mg·L<sup>-1</sup> at BR + 400 mg·L<sup>-1</sup> at 25 DAFA; besides the control (0 mg·L<sup>-1</sup>). After 40 days of the first application, the samples were harvested and we verified that the *S*-ABA induced the accumulation of anthocyanins and total phenolic compounds in the 'Rubi' grape. From these results we can recommend two applications of *S*-ABA, 400 mg·L<sup>-1</sup> at BR + 200 mg·L<sup>-1</sup> at 25 DAFA.

## Keywords

*Vitis vinifera*, *S*-ABA, Table Grape, Bioactives Compounds

## 1. Introduction

The 'Rubi' grape (*Vitis vinifera* L.) is a fine table cultivar and occurred from a

mutation from the 'Italia' grape, distinguishing only by the pink color of the berry [1]. However, in some regions of tropical climate, the colored grapes show an undesirable color in the moment of the harvest and this is mainly due to the low thermal amplitude and high rainfall. This lack of color can be supplied by the use of growth regulators, as the abscisic acid (ABA) that promotes the synthesis and accumulation of anthocyanins, which are pigments that provide color to the fruits, flowers and vegetables [2].

The fruit color has a central role in the direct consumer attraction and for this reason, is one of the most important ampelographic traces and also is one of the main characteristics to be considered in order to evaluate the quality by phenotyping of a potential genotype [3]. The proportion of pigments such as chlorophylls, carotenoids and anthocyanins determines the external color and appearance of the fruit. Some of the phytochemicals of the polyphenols class, such as anthocyanins, are considered as important markers of the commercial values of grapes [4], besides acting as antioxidants in the human organism, preventing a series of generative diseases [5].

The concentration of phenolic compounds in fruits and vegetables is regulated by genetic, environmental and physiological factors, such as temperature, light, rainfall, soil, chemical products and growth regulators. Many agronomic strategies as alteration of the environmental conditions, water availability, grafting and stimulating agents have been employed to increase the biosynthesis of phenolic compounds in fruits and vegetables [6]. The exogenous application of abscisic acid (ABA) in the beginning of the berries ripening is a way of increasing the synthesis of secondary metabolism compounds, including (poly) phenols. This growth regulator is involved in various biochemical and physiological processes, promoting the synthesis and accumulation of anthocyanins, which directly influence the color development. The expression of this pigment depends on internal factors as the ABA, which induces the transcription factor MYB1A, protein responsible for regulating gene transcription genes that compose the anthocyanins biosynthetic pathway in colored grapes [7].

The efficiency of exogenous application of ABA has been demonstrated in the synthesis and accumulation of anthocyanins (polyphenols) in the peel of many grape cultivars [8] [9] [10] [11], as well as it provides uniformity of color, improves the quality of the treated grapes [8] [12], and increase the phenolic compounds contents [11] [13]. Anthocyanin performs an important role in the color and in the sensorial characteristics of fruits and vegetables, beyond its protective effects against certain types of cancer and cardiovascular diseases [14] [15]. The accumulation of these compounds adds commercial quality to the product, as well as attracts consumers that, increasingly, search for fruits with better nutritional quality as a natural fount of antioxidants.

In this way, the aim of this study was the effects of the (*S*)-cis-abscisic acid (*S*-ABA) exogenous application in the anthocyanins content, phenolic compounds and in the antioxidant capacity of the 'Rubi' grape.

## 2. Material and Methods

### 2.1. Location and Plant Material

The experiment was performed during the cycle 2014/2015 of the 'Rubi' vine (*Vitis vinifera* L.) grafted in the rootstock '420-A' spaced out  $4 \times 2$  m, in the sixth production year, supported in the pergola system, located in São Miguel Arcanjo, state of São Paulo (23°31'S, 47°35'W and altitude 660 m), Brazil. The soil of the region was classified as Dystrophic Red Latosol [16] and the climate region, according to Köppen classification, as *Cwa*, with average annual rainfall of 1.396 mm, temperature average of 20.4°C and 70.6% relative humidity.

The grapevines were pruned at August 2014, leaving six buds for each productive branch and subsequently, 2% hydrogen cyanamide was applied aiming the induction and standardizes prouting. In the beginning of the gems sprouting, the vegetative branches were eliminated and 2 to 3 sprouts with cluster were kept by branch. The other cultural treatments, as shoot and branch thinning, branch training, fertilization, weed, pests and diseases control were performed according to the techniques employed in the cultivation region.

### 2.2. Experimental Design and Treatments

The experimental design was composed of randomized blocks, with four treatments and six blocks, in a total of 24 experimental parcels, constituted of one plant each. The effect of the isomer *S*-ABA supplied by Valent BioScience Corporation (Libertyville, IL, USA) (100 g·L<sup>-1</sup> of active ingredient) was evaluated in different concentrations, 400 mg·L<sup>-1</sup> *S*-ABA in the beginning of the ripening (BR); 400 mg·L<sup>-1</sup> at BR + 200 mg·L<sup>-1</sup> at 25 days after first application (DAFA); 400 mg·L<sup>-1</sup> at BR + 400 mg·L<sup>-1</sup> at 25 DAFA; beyond the control (no *S*-ABA application), in a total of 4 treatments. It was added to all treatments 0.3 mL of BreakThru® (Evonik Industries, Germany) non-ionic surfactant. It was considered as BR for the first application softening berries and color change, which presented soluble solids values of 9° Brix and acidity of 0.9% of tartaric acid. The clusters were sprayed in the morning using costal sprayer (40 kg f<sup>-1</sup> pressure) with JA1 hollow cone nozzle tips to provide complete and uniform coverage, employing 900 L·ha<sup>-1</sup>.

### 2.3. Harvest and Characteristics Evaluated

The harvest was performed 40 days after the first *S*-ABA application, were harvested five representative clusters in each experimental parcel and sampling 10 berries per cluster for the analyses of the anthocyanins content, total phenolic compounds, total flavonoids, total chlorophyll, total carotenoids and total antioxidant activity. The berries were cut in half to remove the seeds and the peel and pulp were pulverized in liquid nitrogen and stored at -80°C until the moment of the analyses.

Chlorophyll, anthocyanins and carotenoids were determined according to Sims and Gamon [17]. The extraction was performed in cold acetone/Tris-HCl

buffer solution (80:20 vol:vol, pH 7.8), in ambient protected from light. The readings were made in spectrophotometer BEL Photonics®, SP 2000 UV/vis. The absorbance was converted in mg 100 g<sup>-1</sup> of fresh weight, through the following equations:

$$\text{Anthocyanin} = 0.08173A_{537} - 0.00697A_{647} - 0.002228A_{663} \quad (1)$$

$$\text{Chlorophyll } a = 0.01373A_{663} - 0.000897A_{537} - 0.003046A_{647} \quad (2)$$

$$\text{Chlorophyll } b = 0.02405A_{647} - 0.004305A_{537} - 0.005507A_{663} \quad (3)$$

$$\text{Carotenoids} = \left[ A_{470} - (17.1 \times (\text{Chl } a + \text{Chl } b) - 9.479 \times \text{Anthocyanin}) \right] / 11926 \quad (4)$$

The total chlorophyll content was obtained from the sum of Equation (2) and Equation (3).

Total phenolic content (TPC) was determined using Folin-Ciocalteu method [18]. The absorbance was measured at 725 nm using a UV-VIS spectrophotometer (BEL Photonics®, SP 2000). TPC was expressed as mg gallic acid equivalent 100 g<sup>-1</sup> of fresh weight.

Total flavonoids were determined according to the method described by Popova *et al.* [19], with adjustments [20]. The extraction was performed in 70% acidified methanol and the absorbance was measured at 425 nm in spectrophotometer (BEL Photonics®, SP 2000 UV/VIS). The total flavonoids were expressed in mg quercetin equivalent per 100 g of fresh weight.

The antioxidant activity was performed according to the methodology of Brand-Williams *et al.* [21], altered by Rossetto *et al.* [22]. The DPPH (2,2-diphenyl, 1-picrylhydrazyl hydrate) (Sigma Aldrich Brazil) in 80% methanol and the absorbance was measured at 517 nm and converted in percentage of the antioxidant capacity through the equation:

$$\% \text{ of DPPH reduction} = \frac{\text{Abs}_{\text{control}} - \text{Abs}_{\text{sample}}}{\text{Abs}_{\text{control}}} \times 100$$

The calibration curve was prepared with Trolo × at 5, 10, 15, 20 and 25 µg and the results were expressed in µg TEAC equivalent g<sup>-1</sup>.





## 2.4. Statistical Analysis

The data were submitted to the variance analysis (F test) and the means were compared using Tukey test at 5% of probability. The correlation analysis was performed between the contents of anthocyanins, chlorophylls, carotenoids, phenolic compounds, flavonoids and antioxidant activity.

## 3. Results and Discussion

The application of S-ABA provided a raise in the intensity of the color in 'Rubi' grape berries (analyzed via anthocyanins) in all the treatments, when compared to control (Table 1). The color of the grape berries is resulting of the accumulation of phenolic compounds (e.g., anthocyanins) in the beginning of the ripening. Generally, it is found a high curvilinear relation between the anthocyanins content and parameters of color measures (e.g.,  $L^*$  e  $h^*$ ) [12]. With two S-ABA

**Table 1.** Anthocyanins (Ant), total phenolic compounds (TPC), flavonoids (Flav) and carotenoids (Car) of 'Rubi' grape submitted to the application of abscisic acid (*S*-ABA).

	Treatments ( <i>S</i> -ABA mg·L <sup>-1</sup> )	Ant (mg 100 g <sup>-1</sup> )	TPC (mg 100 g <sup>-1</sup> )	Flav <sup>a</sup>	Car (µg 100 g <sup>-1</sup> )
	Control (0)	0.16 ± 0.03* c	61.14 ± 5.02* c	0.84 ± 0.04* b	73.42 ± 8.26* a
	400 (BR)	0.22 ± 0.03 b	72.41 ± 4.34 b	0.89 ± 0.04 b	62.83 ± 4.21 b
	400 (BR) + 200 (25 DAFA)	0.30 ± 0.03 a	82.53 ± 4.96 a	0.98 ± 0.13 ab	74.74 ± 2.65 a
	400 (BR) + 400 (25 DAFA)	0.25 ± 0.02 b	78.99 ± 2.81 ab	1.16 ± 0.16 a	77.76 ± 6.68 a
	CV (%)	11.38	5.65	12.06	8.36

\*Means followed by the same letter in the column do not show any significant difference by the Tukey's test at 5% probability. Note: BR = Beginning of Ripening; 25 DAFA = 25 days after the first application. <sup>a</sup>Expressed in mg quercetin equivalente per 100 g<sup>-1</sup> of fresh sample.

applications (400 mg·L<sup>-1</sup> at BR + 200 mg·L<sup>-1</sup> at 25 DAFA) there was an induction of anthocyanins synthesis and accumulation (0.30 mg 100 g<sup>-1</sup>), 46.66 % higher than control (0.16 mg 100 g<sup>-1</sup>) (Table 1). The use of this regulator applied twice (400 mg·L<sup>-1</sup> 7 days after the beginning of the ripening + 400 mg·L<sup>-1</sup> at 15 days before the harvest) improved the 'Benitaka' grape color, besides improving the physical characteristic of the clusters, when compared to one application (400 mg·L<sup>-1</sup> 7 days after the beginning of the ripening) [8].

The synthesis of anthocyanins can occur from the beginning of the ripening until the harvest, as well as during the first half of the ripening [23], which would justify the positive effect of the *S*-ABA application in two phases of the grapes ripening. Many studies have showed that the *S*-ABA exogenous increases the acnthocyanins contents in grapes [10] [11] [24] [25] and can also improve the color and quality of the fruit [8] [12], by increasing other classes of polyphenols (e.g., flavonoids and ellagic acid) [13].

Independently on the concentration, treatments with two *S*-ABA applications were efficient in the induction of total phenolic compounds (TPC) and flavonoids in 'Rubi' grape (Table 1). For total flavonoids, there was a high concentration in the grapes treated with 400 mg·L<sup>-1</sup> BR + 400 mg·L<sup>-1</sup> 25 DAFA distinguishing from the control (Table 1). The variations in the flavonoid content can be attributed to genetic and climatic factors, management of the vineyard, degree of ripeness, among others [26]. In this study, all treatments received the same management and the same growing conditions, thus, the grapes ripening

degree promoted the synthesis and accumulation of flavonoids, which is evidenced by the intense pigmentation of the berries, provided after *S*-ABA application.

With two applications of 400 mg·L<sup>-1</sup> of *S*-ABA, the grapes showed 1.16 mg 100 g<sup>-1</sup> of flavonoids, 27.58% superior when compared to the control (0.84 mg 100 g<sup>-1</sup>). The anthocyanins and flavonoids are products of the phenylalanine ammonia lyase (PAL - EC 4.3.1.5) and of the cinnamate 4-hydroxylase (C4H - EC 1.14.13.11) [27] activities. In this way, the color of the 'Rubi' grape using 400 mg·L<sup>-1</sup> BR + 200 mg·L<sup>-1</sup> at 25 DAFA, may be a consequence of the induction of the enzymes activities.

There was a significant effect of the treatments in the total chlorophyll levels and total carotenoids of 'Rubi' grapes (Table 2). Grapes that were not treated with *S*-ABA showed the highest content (104.22 µg 100 g<sup>-1</sup>), not differing statistically from the grapes treated with two applications of *S*-ABA (400 mg·L<sup>-1</sup> BR + 200 mg·L<sup>-1</sup> at 25 DAFA), which showed a content of 102.16 µg 100 g<sup>-1</sup> (Table 2). The result obtained with the grapes that received application of *S*-ABA showed chlorophyll content similar to the control, what can be explained by the analyses in the whole berries (peel and pulp), because the 'Rubi' grape shows white pulp.

Grapes treated with two *S*-ABA application showed the highest of total carotenoids content (74.74 and 77.76 µg 100 g<sup>-1</sup>), distinguishing from the ones that received only one application (62.83 µg 100 g<sup>-1</sup>), however, they did not differ from the control treatment (73.42 µg 100 g<sup>-1</sup>) (Table 1). Some studies that quantify the chlorophyll and carotenoids in the berries of colored grapes are scarce, mainly the ones submitted to the application of abscisic acid. According to Rusconi *et al.* [28] these compounds perform an important role in the grapes pigmentation, but especially in white grapes, when the anthocyanins are not present. In the grapes, as well as in its derivatives, the content and profile of the carotenoids influence in the fruit bouquet and are considered determining factors for the aroma, beyond its importance to the human health (pro-vitamin A and antioxidant activity) [29].

Even though the analyzed compounds with antioxidant potential have been affected by the *S*-ABA application, there was no variation in the total antioxidant

**Table 2.** Total chlorophyll and total antioxidant activity of the 'Rubi' grape, submitted to the application of abscisic acid (*S*-ABA).

Treatments ( <i>S</i> -ABA mg·L <sup>-1</sup> )	Chlorophyll total (µg 100 g <sup>-1</sup> )	Antioxidant activity <sup>aNS**</sup>
Control (0)	104.22 ± 7.34* a	341.35 ± 28.93*
400 (BR)	90.71 ± 7.38 b	322.96 ± 18.19
400 (BR) + 200 (25 DAFA)	91.28 ± 8.22 b	341.16 ± 24.34
400 (BR) + 400 (25 DAFA)	102.16 ± 13.19 ab	346.96 ± 29.03
CV (%)	7.94	7.62

\*Means followed by the same letter in the column do not show any significant difference by the Tukey's test at 5% probability. Note: BR = Beginning of Ripening; 25 DAFA = 25 days after the first application. <sup>a</sup>Expressed in µg Trolox equivalent g<sup>-1</sup> of fresh sample. <sup>NS\*\*</sup>Non-significant.

activity measured via DPPH (**Table 2**). The antioxidant capacity in grapes treated with abscisic acid is variable in grapes cultivars. Sandhu *et al.* [13] studied the effects of the ABA exogenous in grapes ‘Alachua’ (table grape) and ‘Noble’ (wine grape) and verified that the ABA increased the antioxidant capacity of the ‘Noble’ grape in two times (8 and 10 days after the first and second application, respectively), while that for ‘Alachua’ no differences were found in no sampling period. In the juice of the ‘Isabel’ grape, Yamamoto *et al.* [10] found no correlation between the content of phenolic compounds and antioxidant activity.

In our study, the antioxidant activity measured via DPPH can be attributed mainly to the phenolic compounds, however, other molecules as carotenoids and chlorophylls may have influenced the response, due to the properties of scavenger reactive oxygen species (ROS) [30] [31]. Other analytic methods have been used to determine the antioxidant activity in grapes (ABTS, DPPH, FRAP, ORAC, among others), increasing the difficult in the data comparison. Other studies showed the difficulty in obtaining similar data for the total antioxidant activity, making it difficult to compare the obtained data [32] [33], which can be attributed to the method utilized, or by the expression of results, or even by the analyzed material (peel, pulp, seeds or full berries).

The anthocyanins showed a positive and high correlation with the concentration of total phenolic compounds ( $r = 0.7852^{**}$ ) (**Table 3**), evidencing that the increase of anthocyanins is related to the increase of total phenolic compounds of the ‘Rubi’ grape. These compounds are directly or indirectly involved with the grapes color and red grape juice color, beyond important characteristics of the fruit quality [10].

There was a significant correlation between the total phenolic compounds and flavonoids, evidencing that the increase of the flavonoids contribute to the TPC. Nevertheless, this was already expected, since the flavonoids, as well as the anthocyanins, belong to the polyphenols class. We found that the total chlorophyll and carotenoids contributing to the increase of the flavonoids content (**Table 3**). According to Han *et al.* [34], the carotenoids have been described for maintaining the cell wall integrity and the efficiency of this pigment as an antioxidant

**Table 3.** Correlation between total chlorophyll (Cl *tot*), anthocyanins (Ant), carotenoids (Car), flavonoids (Flav), total phenolic compounds (TPC) and antioxidant activity (AA) of the ‘Rubi’ grape, submitted to the application of abscisic acid (S-ABA).

	Ant	Car	Flav	TPC	AA
Cl <i>tot</i>	-0.2326 <sup>ns</sup>	-0.0687 <sup>ns</sup>	0.5214 <sup>**</sup>	0.0344 <sup>ns</sup>	0.1005 <sup>ns</sup>
Ant		-0.0687 <sup>ns</sup>	0.2715 <sup>ns</sup>	0.7852 <sup>**</sup>	0.1159 <sup>ns</sup>
Car			0.5214 <sup>**</sup>	-0.0465 <sup>ns</sup>	0.2170 <sup>ns</sup>
Flav				0.4465 <sup>*</sup>	0.1005 <sup>ns</sup>
TPC					0.0344 <sup>ns</sup>

ns: non-significant; \*Significant 5%; \*\*Significant 1%, by Tukey’s test ( $P < 0.05$ ).

against injuries promoted by ROS depends on the synergistic interaction with flavonoids and with other carotenoids. In this way, the positive correlation found between carotenoids and flavonoids can be attributed to the synergic action and antioxidant of both phytochemicals.

It is known that well-colored grapes are more acceptable in the market of fresh fruits, thus growers should consider the application costs and the grape price to evaluate the benefits of the *S*-ABA application to increase profits. Thus, from the data, we noticed that *S*-ABA promotes the synthesis and accumulation of anthocyanins, phenolic compounds and total flavonoids of the 'Rubi' grape, mainly when applied twice, adding commercial value to this grape cultivar, since the consumers are attracted by fruits with antioxidant potential. It is also important to state that none of *S*-ABA treatments changed the berry taste, what indicates that the plant growth regulator has no influence on ripening evolution.

## 4. Conclusion

The application of *S*-ABA in two phases in the clusters development (beginning of ripening and after 25 days) is efficient to improve the berries color, as well as for increase of anthocyanins and phenolic compounds content of the 'Rubi' grape. The dose of 400 mg·L<sup>-1</sup> BR + 200 mg·L<sup>-1</sup> DAFA is enough to increase the anthocyanins, which are highly correlated to the berries color, increasing the antioxidants content of the fruit.

## Acknowledgements

We would like to thank the "Coordenação de Aperfeiçoamento de Pessoal de Nível Superior" (CAPES), the "Conselho Nacional de Desenvolvimento Científico e Tecnológico" (CNPq) (Process 301577/2015-0) and the "Fundação de Amparo à Pesquisa do Estado de São Paulo" (FAPESP) (2015/16440-5) for their financial support for this project.

## References

- [1] Terra, M.M., Pires, E.J.P. and Nogueira, N.A.M. (1998) Tecnologia para produção de uva Itália na região noroeste do estado de São Paulo. Coordenadoria de Assistência Técnica Integral, Campinas, 81 p.
- [2] Ribichaud, J.L. and Noble, A.C. (1990) Astringency and Bitterness of Selected Phenolic in Wines. *Journal of the Science of Food and Agriculture*, **53**, 343-353. <https://doi.org/10.1002/jsfa.2740530307>
- [3] Rustioni, L., Basilico, R., Fiori, S., Leoni, A., Maghradze, D. and Failla, O. (2013) Grape Colour Phenotyping: Development of a Method Based on the Reflectance Spectrum. *Phytochemical Analysis*, **24**, 453-459. <https://doi.org/10.1002/pca.2434>
- [4] Rustioni, L., Rocchi, L. and Failla, O. (2015) Effect of Anthocyanin Absence on White Berry Grape (*Vitis vinifera* L.). *Vitis*, **54**, 239-242.
- [5] Nixdorf, S.L. and Hermosín-Gutiérrez, I. (2010) Brazilian Red Wines Made from the Hybrid Grape Cultivar Isabel: Phenolic Composition and Antioxidant Capacity. *Analytica Chimica Acta*, **659**, 208-215. <https://doi.org/10.1016/j.aca.2009.11.058>

- [6] Treutter, D. (2010) Managing Phenol Contents in Crop Plants by Phytochemical Farming and Breeding-Visions and Constraints. *International Journal of Molecular Sciences*, **11**, 807-857. <https://doi.org/10.3390/ijms11030807>
- [7] Jeong, S.T., Goto-Yamamoto, N., Kobayashi, S. and Esaka, M. (2004) Effects of Plant Hormones and Shading on the Accumulation of Anthocyanin and the Expression of Anthocyanin Biosynthetic Genes in Berry Skins. *Plant Science*, **167**, 247-252. <https://doi.org/10.1016/j.plantsci.2004.03.021>
- [8] Parte superior do formulário Roberto, S.R., Assis, A.M., Yamamoto, L.Y., Miotto, L.C.V., Sato, A.J., Koyama, R. and Genta, W. (2012) Application Timing and Concentration of Abscissic Acid Improve Color of 'Benitaka' Table Grape. *Scientia Horticulturae*, **142**, 44-48. <https://doi.org/10.1016/j.scienta.2012.04.028>
- [9] Leão, P.C.S., Coelho Lima, M.A., Costa, J.P.D. and Trindade, D.C.G. (2014) Abscissic Acid and Ethephon for Improving Red Color and Quality of Crimson Seedless Grapes Grown in a Tropical Region. *American Journal of Enology and Viticulture*, **66**, 37-45. <https://doi.org/10.5344/ajev.2014.14041>
- [10] Yamamoto, L.Y., Assis, A.M., Roberto, S.R., Bovolenta, Y.R., Nixdorf, S.L., García-Romero, E., Gómez-Alonso, S. and Hermosín-Gutiérrez, I. (2015) Application of Abscissic Acid (S-ABA) to cv. Isabel Grapes (*Vitis vinifera* × *Vitis labrusca*) for Color Improvement: Effects on Color, Phenolic Composition and Antioxidant Capacity of Their Grape Juice. *Food Research International*, **77**, 572-583. <https://doi.org/10.1016/j.foodres.2015.10.019>
- [11] Domingues Neto, F.J., Tecchio, M.A., Pimentel Junior, A., Vedoato, B.T.F., Lima, G.P.P. and Roberto, S.R. (2017) Effect of ABA on Colour of Berries and in the Anthocyanin Accumulation and Total Phenolic Compounds of 'Rubi' Table Grape (*Vitis Vinifera*). *Australian Journal of Crop Science*, **11**, 199-205. <https://doi.org/10.21475/ajcs.17.11.02.p269>
- [12] Peppi, M.C., Fidelibus, M.W. and Dokoozlian, N. (2007) Application Timing and Concentration of Abscissic Acid Affect the Quality of 'Redglobe' Grapes. *The Journal of Horticultural Science and Biotechnology*, **82**, 304-310. <https://doi.org/10.1080/14620316.2007.11512233>
- [13] Sandhu, A.K., Gray, D.J., Lu, J. and Gu, L. (2011) Effects of Exogenous Abscissic Acid on Antioxidant Capacities, Anthocyanins, and Flavonol Contents of Muscadine Grape (*Vitis rotundifolia*) Skins. *Food Chemistry*, **126**, 982-988.
- [14] Djousse, L., Arnett, D.K., Coon, H., Province, M.A., Moore, L.L. and Ellison, R.C. (2004) Fruit and Vegetable Consumption and LDL Cholesterol: The National Heart, Lung, and Blood Institute Family Heart Study. *The American Journal of Clinical Nutrition*, **79**, 213-217.
- [15] Arts, I.C.W. and Hollman, P.C.H. (2005) Polyphenols and Disease Risk in Epidemiologic Studies 1-4. *The American Journal of Clinical Nutrition*, **81**, 317-325.
- [16] EMBRAPA Solo (1999) Embrapa. Sistema Brasileiro de Classificação do solo. EMBRAPA, Produção de Informação, Rio de Janeiro, Brasília, 42 p.
- [17] Sims, D.A. and Gamon, J.A. (2002) Relationships between Leaf Pigment Content and Spectral Reflectance across a Wide Range of Species, Leaf Structures and Developmental Stages. *Remote Sensing of Environment*, **81**, 337-354.
- [18] Singleton, V.L. and Rossi Junior, J.A. (1965) Colorimetry of Total Phenolics with Phosphomolybdic-Phosphotungstic Acid Reagents. *American Journal of Enology and Viticulture*, **16**, 144-158.
- [19] Popova, M., Bankova, V., Butovska, D., Petkov, V., Nikolova-Danyanova, B., Sabatini, A.G., Marazzan, G.L. and Bogdanov, S. (2004) Validated Methods for the

- Quantification of Biologically Active Constituents of Poplar-Type Propolis. *Phytochemical Analysis*, **15**, 235-240. <https://doi.org/10.1002/pca.777>
- [20] Silva, M.B., Rodrigues, L.F.O.S., Rossi, T.C., Vieira, M.C.S., Minatel, I.O. and Lima, G.P.P. (2016) Effects of Boiling and Oil or Vinegar on Pickled Jurubeba (*Solanum paniculatum* L.) Fruit. *African Journal of Biotechnology*, **15**, 125-133. <https://doi.org/10.5897/AJB2015.15077>
- [21] Brand-Williams, W., Cuvelier, M.E. and Berset, C. (1995) Use of a Free Radical Method to Evaluate Antioxidant Activity. *Lebensmittel- Wissenschaft & Technologie*, **28**, 25-30.
- [22] Rossetto, M.R.M., Vianello, F., Rocha, S.A. and Lima, G.P.P. (2009) Antioxidant Substances and Pesticide in Parts of Beet Organic and Conventional Manure. *African Journal of Plant Science*, **3**, 245-253.
- [23] Mullins, M.G., Bouquet, A. and Willians, L.E. (1992) Biology of the Grapevine. Cambridge University Press, Cambridge, 252 p.
- [24] Ban, T., Ishimaru, M., Kobayashi, S., Shiozaki, S., Goto-Yamamoto, N. and Horiuchi, S. (2003) Absciscic Acid and 2,4-Dichlorophenoxyacetic Acid Affect the Expression of Anthocyanin Biosynthetic Pathway Genes in 'Kyoho' Grape Berries. *The Journal of Horticultural Science and Biotechnology*, **78**, 586-589. <https://doi.org/10.1080/14620316.2003.11511668>
- [25] Kobayashi, S., Goto-Yamamoto, N. and Hirochika, H. (2004) Retrotransposon Induced Mutations in Grape Skin Color. *Science*, **1**, 982-982. <https://doi.org/10.1126/science.1095011>
- [26] Rockenbach, I.I., Gonzaga, L.V., Rizelio, V.M., Gonçalves, A.E.S.S., Genovese, M.I. and Fett, R. (2011) Phenolic Compounds and Antioxidant Activity of Seed and Skin Extracts of Red Grape (*Vitis vinifera* and *Vitis labrusca*) Pomace from Brazilian Winemaking. *Food Research International*, **44**, 897-901.
- [27] Huang, B., Duan, Y., Yi, B., Sun, L., Lu, B., Yu, X., Sun, H., Zhang, H. and Chen, W. (2008) Characterization and Expression. Profiling of Cinnamate 4-Hydroxylase Gene from *Salvia miltiorrhiza* in Rosmarinic Acid Biosynthesis Pathway. *Russian Journal of Plant Physiology*, **55**, 390-399. <https://doi.org/10.1134/S1021443708030163>
- [28] Rustioni, L., Rocchi, L., Guffanti, E., Cola, G. and Failla, O. (2014) Characterization of Grape (*Vitis vinifera* L.) Berry Sunburn Symptoms by Reflectance. *Journal of Agricultural and Food Chemistry*, **62**, 3043-3046. <https://doi.org/10.1021/jf405772f>
- [29] Krinsky, N.I. and Johnson, E.J. (2005) Carotenoid Actions and Their Relation to Health and Disease. *Molecular Aspects of Medicine*, **26**, 459-516.
- [30] Van Den Berg, R., Haenen, G.R.M.M. and Van Den Berg, H. (1999) Applicability of an Improved Trolox Equivalent Antioxidant Capacity Assay for Evaluation of Antioxidant Capacity Measurements of Mixtures. *Food Chemistry*, **66**, 511-517.
- [31] Ferruzzi, M.G., Bohm, V., Courtney, P. and Schwartz, S.J. (2002) Antioxidant and Antimutagenic Activity of Dietary Chlorophyll Derivatives Determined by Radical Scavenging and Bacterial Reverse Mutagenesis Assays. *Journal of Food Science*, **67**, 2589-2595. <https://doi.org/10.1111/j.1365-2621.2002.tb08782.x>
- [32] Lago-Vanzela, E.S., Silva, R., Gomes, E., García-Romero, E. and Hermosín-Gutiérrez, I. (2011) Phenolic Composition of the Edible Parts (Flesh and Skin) of Bordô Grape (*Vitis labrusca*) using HPLC-DAD-ESI-MS/MS. *Journal of Agricultural and Food Chemistry*, **59**, 13136-13146. <https://doi.org/10.1021/jf203679n>

- [33] González-Caetano, M.R., Jourdes, M., Femenia, A., Simal, S., Rosselló, C. and Teissedre, P.L. (2013) Characterization of Polyphenols and Antioxidant Potential of White Grape Pomace Byproducts (*Vitis vinifera* L.). *Journal of Agricultural and Food Chemistry*, **61**, 11579-11587. <https://doi.org/10.1021/jf403168k>
- [34] Han, S.H., Sakuraba, Y., Koh, H.J. and Paek, N.C. (2012) Leaf Variegation in the Rice zebra2 Mutant Is Caused by Photoperiodic Accumulation of Tetra-Cis-Lycopene and Singlet Oxygen. *Molecular Cell*, **33**, 87-97. <https://doi.org/10.1007/s10059-012-2218-0>



Scientific Research Publishing

**Submit or recommend next manuscript to SCIRP and we will provide best service for you:**

Accepting pre-submission inquiries through Email, Facebook, LinkedIn, Twitter, etc.

A wide selection of journals (inclusive of 9 subjects, more than 200 journals)

Providing 24-hour high-quality service

User-friendly online submission system

Fair and swift peer-review system

Efficient typesetting and proofreading procedure

Display of the result of downloads and visits, as well as the number of cited articles

Maximum dissemination of your research work

Submit your manuscript at: <http://papersubmission.scirp.org/>

Or contact [ajps@scirp.org](mailto:ajps@scirp.org)