

Effect of Hoechunyangkyeok-San Extract on Melanogenesis

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Received 14 March 2016; accepted 23 May 2016; published 26 May 2016

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Abstract

Forsythia fructus has been shown to have antioxidative, anti-inflammatory, antibacterial, antiaging and whitening effects. Hoechunyangkyeok-san (Forsythia viridissima-prescription) is a traditional herbal medicine, which has been clinically used for treating febrile and inflammatory disorders. This work was carried out to investigate the skin whitening effects of Forsythia viridissima-prescription extract (a hydrolyzed extract of Hoechunyangkyeok-san: SID White HYC) on skin. The effects of SID White HYC were assessed the melanin contents in B161 melanoma cells and the pigmented equivalent with HMB45 and Fontana Masson staining in 3D skin model. Then, we examined the expression of major pigment enzymes regulating melanin synthesis and melanosome transport related proteins in B16F1 cells. SID White HYC significantly inhibited the melanin synthesis (56.7% and 30.6% inhibition at 100 µg/mL, intracellular and secreted, respectively) in B16F1 cells and 3D skin model. In addition, western blotting analysis showed that SID White HYC reduced the expression of melanin synthesis and melanosome transport related proteins in B16F1 cells. In clinical trials, the cream containing 0.05% SID White HYC showed skin depigmentation effect without any irritation. These results suggest that SID White HYC may be useful inhibition of melanogenesis and melanosome transport. Therefore, SID White HYC may have potential as a skin-whitening ingredient in cosmetics.

Keywords

Hoechunyangkyeok-San (Formula Extract Containing Forsythia viridissima Fruit, Platycodon grandiflorum Root, Glycyrrhiza glabra Root, Paeonia japonica Root, Angelica gigas Root, Scutellaria baicalensis Root, Gardenia jasminoides Fruit, Coptis chinensis Root, Mentha arvensis Leaf, Rehmannia glutinosa Root, Poncirus trifoliate Fruit), Melanogenesis, Melanosome Transport, Melanin, Whitening

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How to cite this paper: Kim, M.J., Jung, T.K., Park, H.-C. and Yoon, K.-S. (2016) Effect of Hoechunyangkyeok-San Extract on Melanogenesis. *Journal of Cosmetics, Dermatological Sciences and Applications*, **6**, 85-95. http://dx.doi.org/10.4236/jcdsa.2016.63011

1. Introduction

In mammalian hair and skin, melanogenesis is occurred within melanosomes, the specialized lysosome-related vesicles of melanocytes located in the basal layer of the epidermis [1].

Melanosomes contain three enzymes that are absolutely necessary for the synthesis of both types of melanin (pheomelanin and eumelanin). The first enzyme is tyrosinase, which is responsible for the critical and limiting step in melanogenesis, the hydroxylation of tyrosine to form 3,4-dihydroxyphenylalanine (L-DOPA) followed by oxidation of L-DOPA to form dopaquinone. The second and third enzymes are Tyrosinase-Related Protein-1 (TRP-1) and Dopachrome Tautomerase (DCT), which are more concerned with the generation of different types of melanin [2] [3].

In humans, pigmentation results from the synthesis and distribution of melanin in skin, hair bulbs and eyes [4]. The distribution of melanin in the skin is completed by melanosome transport to keratinocytes. In melanosome transport, there are three kinds of the key player proteins. At the cell dendrite tips, the Rab27a-melanophilin complex recruits the actin-dependent motor protein, myosinVa (Myo5a), allowing melanosomes to move on the actin 4 network. Rab27a, melanophilin (Mlph) and Myo5a make a complex to link melanosomes to the F-actin based motors [5]. Myo5a is a molecular motor that binds to actin filaments through its N-terminal head domain and to the cargo through its C-terminal globular tail [6]. Interaction of Myo5a with melanosomes is mediated by a small Ras-like GTPase Rab27a and Mlph. The GTP bound Rab27a interacts with Mlph that in turn recruits Myo5a to form a tripartite complex [7].

According to a recent study, Rab27a-Mlph and Mlph-Myo5a interactions are very important for melanosome transport. For example, hesperidin blocks the Rab27a-Mlph interaction and therefore causes melanosome aggregation in melanocytes [8]. Also, manassantin B causes melanosome aggregation in melanocytes by blocking the Mlph-Myo5a interaction [9]. From these studies, we know that melanosome transport is influenced by inhibiting interaction of tripartite complex as well as loss of any one of the components of the tripartite complex.

Hoechunyangkyeok-san is a traditional herbal prescription for treating febrile and inflammatory diseases, which recorded in Manbyeonghoechun (which means "Restoration of all illnesses", 1587 at first) and Bangyak-happyeon, clinical herbal prescription book [10] [11]. It consists of *Forsythia viridissima* fruit, *Platycodon grandiflorum* root, *Glycyrrhiza glabra* root, *Paeonia japonica* root, *Angelica gigas* root, *Scutellaria baicalensis* root, *Gardenia jasminoides* fruit, *Coptis chinensis* root, *Mentha arvensis* leaf, *Rehmannia glutinosa* root, and *Poncirus trifoliata* fruit. Hoechunyangkyeok-san has been reported to be a useful treatment for diabetes, atherosclerosis and hyperlipidemia in the type 1 diabetic model, and inhibit melanin content in B16F1 cells [12] [13]. However, it has been rarely conducted to evaluate the skin whitening activities. To search for new skin-lightening agents, we investigated the effect of Hoechunyangkyeok-san on depigmenting mechanisms.

2. Materials and Methods

2.1. Chemicals and Reagents

 α -MSH, 3-(4,5-dimetylyhiazol-2-yl)-2,5-diphenyltetrazoliumbromide (MTT) and dimethylsulfoxide (DMSO) were purchased from Sigma-Aldrich (USA). Penicillin/streptomycin, phosphate buffered saline (PBS), fetal bovine serum (FBS) and tryps in were purchased from Gibco (USA). Dulbecco's modified Eagle's medium (DMEM) was obtained from HyClone (USA). Antibodies recognizing tyrosinase, TRP-1, -2, MITF and Rab27a were purchased from Santa Cruz Biotechnology (USA), β -actin and Myo5a were purchased from Cell Signaling Technology (USA). Anti-Mlph was bought from Proteintech (USA).

2.2. Preparation of SID White HYC

Herbs which constitute Hoechunyangkyeok-san were purchased from a Korean herbal medicine dealer in Gumediherb, Korea and composed of 11 herbs shown as **Table 1** [11].

Hoechunyangkyeok-san (50 g, in the ratio of **Table 1**) was extracted twice with 75% aqueous ethanol under reflux at 60° C - 90° C for 4 hours, filtered, and evaporated under reduced pressure. The 75% aqueous ethanol extract was redissolved in 10% aqueous ethanol and hydrolyzed by enzymes, Pectinex (Novozyme, Denmark) at 50° C - 60° C for 30 hours and fractionated by ethyl acetate. The fraction was cooled, filtered, and evaporated under reduced pressure. Finally we obtained the SID White HYC.

| Table 1. The composition of Hoechunyangkyeok-san. | |
|---|------------|
| Herbal name | Amount (%) |
| Forsythia viridissima fruit | 14.70 |
| Platycodon grandiflorum root | 8.53 |
| Glycyrrhiza glabra root | 8.53 |
| Paeonia japonica root | 8.53 |
| Angelica gigas root | 8.53 |
| Scutellaria baicalensis root | 8.53 |
| Gardenia jasminoides fruit | 8.53 |
| Coptis chinensis root | 8.53 |
| Mentha arvensis leaf | 8.53 |
| Rehmannia glutinosa root | 8.53 |
| Poncirus trifoliata fruit | 8.53 |

2.3. Cell Culture

B16F1 murine melanoma cells were obtained from the American Type Culture Collection (ATCC, USA) and cultured in DMEM supplemented with penicillin (100 U/mL), streptomycin (100 μ g/mL) and 10% FBS. Cells were incubated at 37°C in a humidified atmosphere containing 5% CO₂.

2.4. Cytotoxicity (MTT) Assay

Cytotoxicity was evaluated using the MTT assay. The B16F1 cells were seeded at a density of 5×10^3 cells/well in a 96-well plate, and were cultured at 37° C with 5% CO₂. The cells were then treated with SID White HYC at various concentrations (1, 20, 50, 100 µg/mL) for 72 hours. Following incubation, the cells were treated with the MTT solution for 4 hours at 37° C. The supernatants were aspirated, and DMSO was added to each well. After incubation for 20 minutes, the absorbance was measured at 540 nm using a microplate reader (Synergy, BioTek, USA). Cytotoxicity is presented as a percentage of the optical density of the vehicle group.

2.5. Measurement of Intracellular and Secreted Melanin Contents

The intracellular and secreted melanin content was measured using a slight modification of a previously reported method [14] [15]. Briefly, B16F1 cells were incubated at a density of 5×10^4 cells/well in a 6-well plate overnight. B16F1 cells, pretreated with the indicated concentrations of SID White HYC for 1 hour, were treated for 72 hours with α -MSH (200 nM). In order to measure the secreted melanin, 200 µL aliquots of media were placed in 96-well plates and absorbance was measured at 405 nm using a microplate reader. Cells were detached by incubation in trypsin/EDTA. After precipitation, the color of the cell pellets was evaluated visually, and pellets were solubilized in 1 N NaOH containing 10% DMSO. Spectrophotometric analysis of cellular melanin content was performed by quantifying absorbance at 405 nm. Three independent experiments were performed in triplicate.

2.6. Cellular Tyrosinase Assay

Tyrosinase activity in B16F1 cells was assayed as described previously with modifications [16] [17]. Cells were washed twice with ice-cold phosphate buffered saline (PBS) and lysed with cell lysis buffer (0.1% Triton-X 100 in 0.1 M phosphate buffer, pH 6.8). The reaction mixture containing 40 μ L of 10 mM L-tyrosine, 140 μ L of enzyme solution (lysate 40 μ g in 0.1 M phosphate buffer, pH 6.8) and 20 μ L of indicated concentration SID White HYC (in PBS) was incubated at 37°C for 10 minutes. The absorbance of samples was measured at 475 nm. Enzyme activities were expressed as a percentage of vehicle.

2.7. Western Blot Analysis

Cells were rinsed twice with PBS and scraped into lysis buffer. Lysates were prepared with lysis buffer according to the manufacturer's instructions (Thermo, USA; iNtRON, Korea). The protein concentration was determined using a BCA Assay Reagent Kit (Thermo Fisher Scientific Inc., USA). Proteins present in the cell lysates were separated by Sodium Dodecyl Sulfate-Polyacrylamide Gel Electrophoresis (SDS-PAGE) using a 4% - 20% gel, transferred onto polyvinylidene fluoride membranes (GE Healthcare, UK), blocked with 5% skim milk, and treated with primary antibodies for 2 hoursat room temperature. After incubation with horseradish peroxidaseconjugated secondary antibodies (Santa Cruz Biotechnology, USA) at room temperature, immunoreactive proteins were detected with a chemiluminescent ECL assay kit (GE Healthcare, UK) according to the manufacturer's instructions. Bands were visualized using a ChemiDoc image analyzer (BIO-RAD, USA).

2.8. Pigmented Reconstructed Skin Model

Reconstructed human epidermis MelaSkinTM (MCTT, Korea) consisted of normal human derived epidermal keratinocytes and melanocytes that had been cultured to form a multilayered, highly differentiated model. Reconstructed epidermis was placed in 6-well plates and incubated at 37°C in a humidified atmosphere of 5% CO₂ in air. The tissue was incubated for 24 hours with the indicated concentration of SID White HYC.

2.9. Histology and Immunohistochemistry

After 24 hours, tissue were fixed in 10% formaldehyde for 1 day and processed for conventional paraffin embedment. 4 µm thickness sections were then prepared. Sections were stained with hematoxylin and eosin (H&E), HMB-45 (Santa Cruz Biotechnology, USA) and Fontana-Masson for light microscopic observation.

2.10. Clinical Evaluation

A randomized, double-blind, placebo-test (0.05% SID White HYC containing cream), split-face comparison study was conducted. All the participants provided informed written consent prior to their participation. The participants were twenty-two healthy female volunteers aged 37 - 54 years old (average 48.05 ± 4.40 years).

2.10.1. Clinical Assessment

Visual assessment was done at before, 4 and 8 weeks after treatment. Two investigator clinically graded skin color base on 10 points scale at each visit (0 point, clear and transparent; 10 point, dark and unclear). The mean value of the data obtained from each investigator was then calculated and compared.

2.10.2. Objective Evaluation

Objective measurement was done at before, 4 and 8 weeks after treatment. Objective skin color measurements were performed using a Mexameter (MX-18, Courage & Khazaka Electronic GmbH, Cologne, Germany) and Spectrophotometer (CM-2500d, Minolta, Japan) during each visit. Three successive measurements of the Mexameter and Spectrophotometer were made on the darkest portion of the test and placebo cream treated skin. The mean value of the data obtained from each exam was then calculated and compared. Melanin index was measured with Mexameter. High melanin index score indicates dark skin color. The *L*, *a* and *b* values were recorded with Spectrophotometer. (L^* : Luminance parameters (brightness), a^* : Chrominance parameters (green-to-red), b^* : Chrominance parameters (blue-to-yellow)). All the experimental data were expressed as means \pm SEM. Individual typology angle (ITA°) value was achieved by the formulation. Low ITA° value indicates dark skin color.

ITAb value =
$$\left[\operatorname{Arc Tangent}\left\{\left(L^* - 50\right)/b^*\right\}\right] \times 180/3.14159$$

2.10.3. Statistical Analysis

For statistical analysis, an RM ANOVA method was used to compare the change in the visual assessment score, melanin index score and ITA° value resulting from treatment between test and placebo treated skin. The data was analyzed using SPSS Package Program 11.5 (IBM, USA). *P*-values < 0.05 were considered statistically significant.

2.11. Statistical Analysis

Data were analyzed using Sigma Plot. Results were expressed as the mean \pm S.D. of three independent experiments. Comparisons were performed using a one-way ANOVA followed by Duncan's multiple range test. A *P*-value < 0.05 was considered statistically significant.

3. Results

3.1. Effect of SID White HYC on Cell Viability and Melanin Biosynthesis

B16F1 cells were exposed to various concentration of SID White HYC, and cell viability was measured by MTT assay. As shown in **Figure 1**, cell viability of SID White HYC was above 80% up to 100 μ g/mL (**Figure 1**).

To investigate the inhibitory effect of SID White HYC on melanogenesis, the melanin contents of the B16F1 cells were measured after treatment with SID White HYC. Cells were exposed to SID White HYC (10, 50, 100 μ g/mL) for 72 hours with α -MSH (200 nM), and then the amounts of melanin intracellular spaces and secreted into the medium were measured individually. As **Figure 2** shows, SID White HYC decreased the amount of melanin intracellular spaces and secreted into the media (at 100 μ g/mL, 56.72% and 30.64% inhibition, respectively). These results indicate that SID White HYC might have inhibited the synthesis of melanin.

3.2. Inhibitory Effect of SID White HYC on Tyrosinase Activity

SID White HYC inhibited cellular tyrosinase activity in a dose dependent manner but the effect did not inhibit a mushroom tyrosinase activity (data not shown). SID White HYC at 50 μ g/mL reduced tyrosinase activity by 35% (Figure 3).

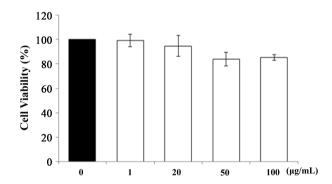


Figure 1. Cell viability of SID White HYC in B16F1 cells. The cells were treated with various concentration of samples for 72 hours. Data are the mean \pm S.D from three independent experiments. **P* < 0.05: statistically significant vs. control group (vehicle).

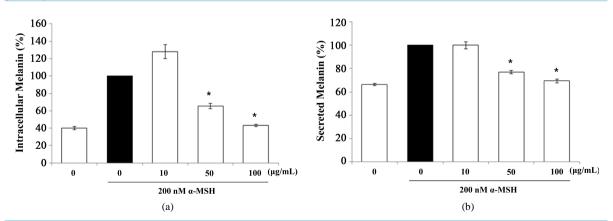


Figure 2. The effect of SID White HYC on intracellular (a) and secreted melanin (b) inmurine B16F1 melanoma cells treated with α -MSH. Thecells were treated with α -MSH and SID White HYC at indicated concentration for 72 hours. Data are the mean \pm S.D from three independent experiments. **P* < 0.05 vs. control group (vehicle).

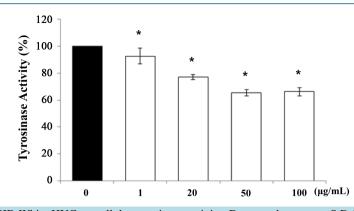


Figure 3. The effect of SID White HYC on cellular tyrosinase activity. Data are the mean \pm S.D from three independent experiments. **P* < 0.05 vs. control group (vehicle).

3.3. Inhibitory Effect of SID White HYC on Melanogenesis-Related Proteins Expression

The expression levels of melanogenesis-related proteins were determined by western blot in α -MSH stimulated B16F1 cells. Protein levels of tyrosinase and TRP-1 were significantly decreased but not TRP-2 after treatment with SID White HYC (**Figure 4**). These results suggest that SID White HYC decease melanin synthesis by down-regulating of melanogenesis-related proteins.

3.4. Inhibitory Effect of SID White HYC on Melanosome Transport-Related Proteins Expression

The function of melanosomes in skin pigmentation requires not only melaninformation, but also transport within the cell. Melanosomes are transported along microtubules towards actin near the plasma membrane and in the tips of dendrites, from where they are transferred to keratinocytes [18]. In melanocytes, the transport of melanosome is carried out by a complex of three proteins: Rab27a, Mlph and Myo5a.

To know the effects of SID White HYC on triplicate protein involved in melanosome transport, we checked expression level of Rab27a, Mlph and Myo5a after treatment with SID White HYC for 72 hours on B16F1 cells. There was dose-dependent reduction of Rab27a, Mlph and Myo5a expression with SID White HYC treatment compared with control (**Figure 5**). These results demonstrate that SID White HYC decease melanin secretion by down-regulating of melanosome transport-related proteins.

3.5. Effect of SID White HYC on Melanogenesis in Cultured Human Skin Model

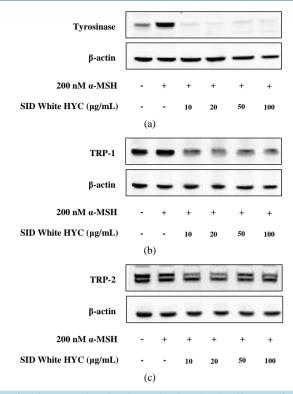
To identify the effects of SID White HYC on skin pigmentation, we used three-dimensional cultured human skin model (MelaSkinTM). The MelaSkin exhibits *in vivo*-like morphological and ultrastructural characteristics. When MelaSkin was cultured in media, melanogenesis was induced naturally. Thus the model provides a useful *in vi-tro* means to evaluate cosmetic and pharmaceutical agents designed to modulate skin pigmentation. Different concentration of SID White HYC and the vehicle were applied on the reconstructed skin for 24 hours.

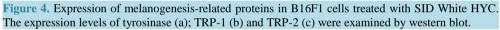
Distribution of melanin in MelaSkin was shown by Fontana-Masson and HMB45 staining. Melanin is stained strongly by Fontana-Masson stain in all layers of the epidermis. One of the most frequently used melanocytic markers in clinical practice is HMB45. HMB-45 is a monoclonal antibody that reacts against an antigen present in melanocytic tumors such as melanomas but not other tumors, thus demonstrating specificity and sensitivity [19].

Figure 6 showed decreased deposition of pigment granule in MelaSkin equivalents following SID White HYC treatment. Fontana-Masson and HMB45 staining revealed the hypopigmentation activities of SID White HYC.

3.6. Clinical Evaluation

Figure 7 displays the decrease of the clinical visual assessment score over time compared with before. A decrease in the visual assessment score over time was observed for both test and placebo treated skin. The visual assessment score of the test group was significantly lower than that of the placebo group (P < 0.05). The visual





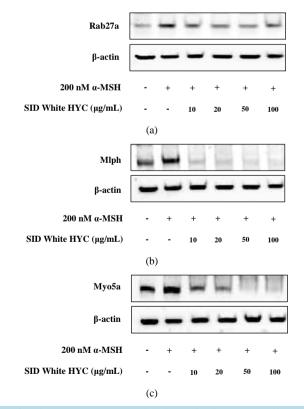


Figure 5. Expression of melanosome transport-related proteins in B16F1 cells treated with SID White HYC. The expression levels of Rab27a (a); Mlph (b) and Myo5a (c) were examined by western blot.

assessment score change at the end of the study for the test and placebo treated skin were -2.5% and -0.43% respectively, with a significant different after 8 weeks (P < 0.05) (Figure 7(a)).

A significant decrease in the melanin index score was observed from 4 week through the end of study at both group (Figure 7(b)). The mean melanin index score change at the end of the study for the test and placebo treated skin were -5.08% and -2.48% respectively, with a statistically significant difference (P < 0.05).

At the Spectrophotometer analysis, the test group also showed significant increase in ITA° value after 4 week of treatment, which lasted until the end of the study. The ITA° value change at the end of the study for the test and the placebo treated skin were +11.72% and +7.82% respectively, with a statistically significant difference (P < 0.05) (Figure 7(c)).

4. Discussion

Depigmentation can be accomplished by specific interaction of an active: before melanin synthesis; during melanin synthesis; after melanin synthesis [20].

Skin pigmentation results from melanin synthesis by melanocytes, which is induced by exposure to UV radiation. In mammals, melanin synthesis is stimulated by a large number of effectors, including UV-B radiation and cAMP-elevating agents (forskolin, 3-isobutyl-1-methylxanthine, α -melanocytestimulating hormone (α -MSH), glycyrrhizin) [21]-[25].

Melanin synthesis takes place within specialized intracellular organelles named melanosome, where the enzymes involved in melanogenesis, including tyrosinase, are located. Mature melanin-filled melanosomes move from the perinuclear region to the dendrite tips of melanocytes and are transferred to neighboring keratinocytes by a mechanism that is still now well characterized. The result of this process is a uniform distribution of melanin pigments in the epidermis that is responsible for skin and hair color in humans and other animals. The ability to inhibit melanin synthesis effectively is very important in regulating pigmentation and skin whitening [26] [27].

With regard to the inhibitory effect of SID White HYC on melanogenesis, SID White HYC reduced the synthesis of melanin in α -MSH stimulated B16F1 cells (**Figure 2**). According to the HMB45 immunostaining assay, in which HMB45 was found to react with the neuraminidase-sensitive sialylated glycol-conjugate present in early-stage melanosomes, the decreased presence of HMB45 indicated that SID White HYC inhibited the formation of melanosome (**Figure 6**). The results of Fontana-Masson staining were in complete agreement with this observation, confirming that SID White HYC influenced active melanosome formation and thus melanocytic differentiation. SID White HYC reduced the activity of cellular tyrosinase but the effect was not a direct inhibition of tyrosinase, since it had no effect on mushroom tyrosinase (**Figure 3**). These data suggest that SID White HYC has an influence on the expression of melanogenic proteins such as tyrosinase, TRP-1 and TRP-2. The changes in the level of melanogenic proteins expression analyzed by immunoblot indicate that SID White HYC decreased melanogenic proteins level (**Figure 4**).

In melanosome transport, there are three kinds of the key player proteins. Rab27a, Mlph and Myo5a form a tripartite complex that allows interaction of melanosomes with the actin network [28] [29]. When the three constituents meet each other to form a complex, it is possible to normally transport melanosome to the periphery. If any of the three constituents is abnormal, it is observed that a problem arises in the transport of melanosomes without causing any problem in the production of melanin, so that the color of the skin becomes white.

In regard to the inhibitory effect of SID White HYC on melanogenesis, SID White HYC reduced the secreted melanin in α -MSH stimulated B16F1 cells (**Figure 2(b)**). It seems that SID White HYC has an influence on melanosome transport. The changes in the level of transport-related proteins expression analyzed by immunoblot indicate that SID White HYC decreased transport-related proteins level (**Figure 5**). These results has led to development of effective skin-whitening ingredients with blocking the before and after melanin synthesis.

The levels of skin pigmentation of the test and placebo groups were measured at 4 and 8 weeks using the Mexameter and Spectrophotometer, melanin index and ITA°, respectively. The test group had significantly less pigmentation throughout the study compared with before (**Figure 7**). The ITA° value is believed to provide a quantitative means for classifying individual skin color [30]. It indicates the melanotic status of the skin at the time of the testing to be taken into account [31] [32]. None of adverse reactions such as erythema, burning sensation and pruritus were observed during 8 weeks of study period. These results suggest that SID White HYC ameliorates darkness of the skin.

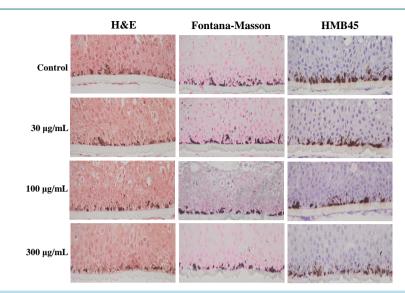


Figure 6. Histological analysis of melanin deposition in cultured human skin model treated with SID White HYC using Fontana-Masson and HMB 45 stains.

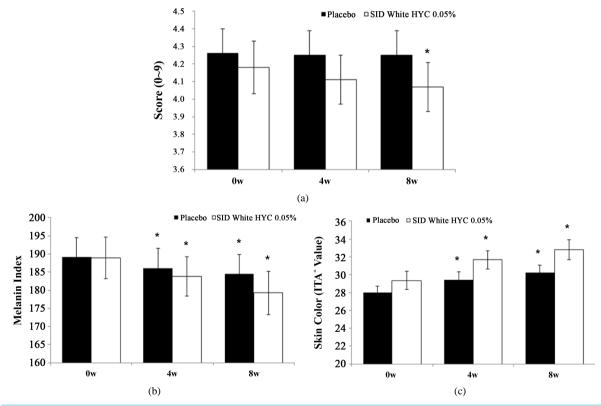


Figure 7. The effects of SID White HYC containing cream on the visual assessment score (a); melanin index (b) and ITA° value (c) compared with placebo. Mean \pm SEM, *P < 0.05 vs. before treatment.

5. Conclusion

Taken together, SID White HYC could be expected to exert skin whitening effect by decrease of the melanogenesis-related proteins and melanosome transport-related proteins expression without any adverse effects. In conclusion, these studies demonstrate that SID White HYC may be useful inhibitor of melanogeneiss and that SID White HYC may be an effective component in cosmetics designed to lighten skin.

Acknowledgements

This work was supported by Chungcheong Institute for Regional Program Evaluation Promotion Project (R0002893) of the Ministry of Trade, Industry and Energy Republic of Korea.

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