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A Paraquat Tolerance Mutant in Rice (*Oryza sativa* L.) Is Controlled by Maternal Inheritance

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Abstract

Paraquat (1,1'-dimethyl-4,4'-bipyridinium) tolerance is an important trait in the weed control during crop production. The paraquat tolerant (Pq72) and susceptible (Pq1192) mutants are pure lines derived from the mutation pool of rice cultivar TNG67. Two reciprocal crosses, Pq72/Pq1192 and Pq1192/Pq72, were conducted between Pq72 and Pq1192 mutant lines for studying the genetic of paraquat tolerance by investigations of physiological characteristics related to paraquat tolerance including leaf injury index, leaf chlorophyll fluorescence (Fv/Fm) and electrolyte leakage in the F_2 populations of two reciprocal crosses after paraquat treatment. The results suggested that a maternal inheritance of paraquat tolerance is existed in these mutants. Further analysis found that the F_2 population of Pq72/Pq1192 segregated 3:1 (tolerant to susceptible) in both Fv/Fm and electrolyte leakage, respectively. This result implies that the paraquat tolerance of the Pq72 mutant is controlled by a single dominant gene.

Keywords

Paraquat, Tolerance, Leaf Chlorophyll Fluorescence, Electrolyte Leakage, Maternal Effect

1. Introduction

Weed control has become a problem for farmers in most rice production areas. Paraquat (PQ, 1,1'-dimethyl-4,4'-bipyridinium; methyl viologen) is a rapid acting and non-selective herbicide for controlling weeds in cultivated as well as non-cultivated land throughout Taiwan [1]. PQ can cause membrane damage,

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DNA denaturation, protein reduction by transferring electron from photosystem I (PSI) to oxygen producing toxic reactive oxygen species (ROSs), efficiently induce the death of plant cells [2] [3] [4] [5]. Therefore, paraquat can kill a wide range of annual grasses, broad-leaved and perennial weeds upon contact, but it becomes biologically inactive and less toxic toward roots as well as rhizomes when entering the soil environment [3] [6]. Up to now, more than 30 paraquat-resistant weed species, *i.e. Conyza bonariensis* (L.) Corng. *Lolium rigidum, Arctotheca calendula, Rehmannia glutinosa* and *Mazus pumilus* have been reported in the world (http://www.weedscience.org/; [7]). In Arabidopsis, several paraquat resistant mutants including photoautotrophic salt tolerance 1 (*pst*1), radical-induced cell death1 (*rcd*1), paraquat resistant 1 (*par*1), paraquat resistant 2 (*par*2), pleiotropic drug resistance 11 (*atpdr*11) and resistant to methyl viologen 1 (*rvm*1) [8]-[13] have been identified and characterized.

Fuerst and Vaughn [14] proposed mechanisms of paraquat resistance as follows: reducing quantities of paraquat absorbed by leaf surface; reducing the efficiency of electron acceptation of paraquat, *i.e.* modification of photosystem I; increasing paraquat metabolic rate; increasing the enzymatic activities for free radical scavenging; and sequestration of paraquat. The first three mechanisms still need to be defined experimentally, however, the increase of enzymatic activities for free radical scavenging and sequestration of paraquat have been considered as the critical mechanisms for paraquat resistance [15].

A mutant pool of rice variety TNG67 contains more than 3000 mutants with many desirable traits were developed by sodium azide mutagenesis at the Taiwan Agricultural Research Institute [16]. In our previous study, two pure mutant lines, the paraquat tolerant 72 (T, Pq72) and susceptible 1192 (S, Pq1192), have been screened from this mutant pool [17], and the physiological mechanism study on the paraquat tolerance mutants revealed no difference in uptake, translocation, and metabolism of paraquat between T and S lines (C. S. Wang, unpublished data). However, the induction of enzymatic activities for free radical scavenging played a critical role in paraquat tolerance [15]. Therefore, it is interesting to explore the expression of genes encoding anti-oxidation related enzymes and the mutations induced by sodium azide. In this study, the inheritance of paraquat tolerance in rice mutant was studied based on the tolerance expression in F₂ populations from the reciprocal crosses of Pq72 (T) and Pq1192 (S) pure lines.

2. Materials and Methods

2.1. Rice Materials

Six paraquat-susceptible (S) lines (No. 81, 802, 875, 948, 1081 and 1192) and 7 tolerant (T) lines (No. 72, 713, 881, 882, 883, 986 and 1067) have been selected from 1343 pure mutants at M7 generation from the mutant pool of TNG67 rice variety [17]. After further identification of paraquat tolerance was conducted for three selfed generations, two most stable pure lines, the tolerant 72 (T, Pq72)

and susceptible 1192 (S, Pq1192), were confirmed in the M10 generation and selected as the materials for studying the inheritance of paraquat tolerance. The rice plants of parental and F_1 generations used for inheritance analysis were transplanted into pots at the three-leaf (V3) stage and grown to 21 days in greenhouse for tolerance test. All rice plants of F_2 populations used for inheritance analysis were grown in the paddy field according to traditional management

2.2. Paraquat Treatment

Paraquat (1,1'-dimethyl-4,4'-bipyridinium) treatment was performed as previous described [17]. To acquire a homogenized spraying effect on the tested pot-grown plants, an auto sprayer system consisting of automobile orbital sprayer with a speed of $187 \text{ cm} \cdot \text{min}^{-1}$, high-pressure compressor providing 0.2 MPa with a flow rate of $1 \text{ m} \cdot \text{sec}^{-1}$, and three nozzles at 45 cm above plants producing droplet of 30 μ m diameter, were applied in this work in our herbicide spraying facility in the greenhouse at the NCHU main campus.

2.3. Genetic Crossing

Two paraquat mutants, Pq72 (T) and Pq1192 (S), were selected as parents to make reciprocal crosses to generate the offspring for the genetic analysis of paraquat tolerance. The F_1 plants from the reciprocal crosses between Pq72 and Pq1192 were analyzed by at least two polymorphic SSR markers (data not shown) to identify the true hybrid and then selfed to obtain the F_2 generation. Responses of these plants to paraquat were determined (Table S1).

Duration and location. All the experiments were conducted during 2010 to 2013 either in greenhouse at the main campus of NCHU or in the paddy field at the experimental farm of Agronomy Department of NCHU at Wu-Feng, Taichung, Taiwan according to the experimental requirement.

2.4. Evaluation of Plant Response to Paraquat

The parameters of paraquat tolerance were analyzed by investigations of injury index, chlorophyll fluorescence [18] and electrolyte leakage [19]. At tillering stage, nine completely expanded leaves for each rice plant were harvested and the 1.5 cm-leaf segments were excised 5 cm distance from leaf tip for the following measurements.

Injury index. The paraquat treatment in this injury analysis was performed as previous method [17]. An aliquot of 15 mL 100 μ M paraquat was applied to a Petri dish (diameter 15 cm) lined with filter paper and then the leaf segments of rice plants were placed on the paper. These samples were placed under a light intensity of 200 μ ·mole·m⁻²·s⁻¹ for 9 hrs and the injury indices were determined. The injury index could be classified from 0 to 7, where 0 indicates healthy leaf segment without any herbicidal damage and 7 indicates the completely etiolated leaf segment caused by paraquat (**Figure S1**).

Chlorophyll fluorescence. The paraquat-treated leaves were blotted with tissue paper for drying and then fixed to the sensor clip of portable Chlorophyll Fluorimeter (OS1-FL, Opti-Sciences, and Tyngsboro, USA) for 10 min in the dark before analyzing the maximal efficiency of PSII photochemistry (Fv/Fm), and a sensitive physiological trait reflecting rapid damages to photosystem. Fv/Fm = (Fm-Fo)/Fm, where Fo and Fm indicate the minimal and maximal fluorescence after dark acclimation, respectively [20].

Electrolyte leakage. The paraquat-treated leaves were placed in a tube containing 8 mL deionized water and shaken gently for 24 hrs. The electric conductivity of the solution was determined by a conductivity meter (SC-170, SUNTEX INSTRUMENTS CO., Taipei, Taiwan). Subsequently, samples were heated in a boiling water bath for 15 min and then cooled for the second assay of conductivity. The electrolyte leakage was calculated as the ratio of conductivity prior to and after boiling.

2.5. Data Analysis

A completely randomized design (CRD) with three replications was conducted in this work. Data was subjected to ANOVA, and the difference among treatments was compared statistically based on the least significant difference at 5% level (LSD_{0.05}). In genetic analysis, χ^2 test was applied to analyze the fitness of segregation ratio for paraquat tolerance of F_2 populations from two crosses, Pq72/Pq1192 and Pq1192/Pq72.

3. Results and Discussion

3.1. Responses of the Pq1192 and Pq72 Mutants to Paraguat

Rice mutants Pq1192 (S) and Pq72 (T) have been selected from the M7 generation of TNG67 mutation pool by 100 μ M paraquat treatment [17] and continued selfing to obtain the M10 pure lines. In order to understand the response against paraquat, the rice plants, TNG67, Pq1192 and Pq72, at tillering stage were treated with 100 μ M paraquat. At 3 days after treatment, Pq72 displayed an almost unaffected appearance in contrast with Pq1192 showed a serious injurious appearance (Figure 1(a)). Besides, some significant water soaking injured spots were observed on the leaves of TNG67 after treatment. These results indicate that Pq72 has a better paraquat tolerance than the other two lines.

The morphology of paraquat-treated leaves was investigated at 0, 3, 6, 9, 12, 24 and 48 h after treatment to reveal the progress of paraquat damage of these lines (**Figure 1(b)**). Some water soaking injured spots were observed on the leaves of Pq1192 at 3 h after treatment. The necrosis spots appeared on all leaves of TNG67, Pq1192 and Pq72 after 6 h of treatment, and the Pq1192 leaves showed the most serious injury. At the 48 h after treatment, the severe desiccation occurred on the leaf tissues of TNG67 and Pq1192 and large necrosis lesion were observed on the leaves. The Pq72 leaves showed a less injury than TNG67 and Pq1192 after paraquat treatment. These results indicate that Pq72 effectively

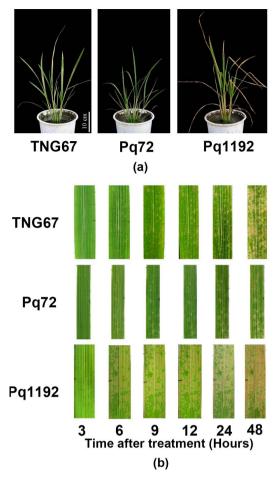


Figure 1. Responses of three rice plants to paraquat, injury of paraquat-susceptible mutant appeared 21 days after 100 μM paraquat treatment (a). Injury of leaf segments excised from 21-days-old rice TNG67, Pq72(T) and Pq1192(S), respectively, caused by 100 μM paraquat (b).

delayed the formation of water soaking injured spots and show tolerance to paraquat treatment.

The maximal efficiency of photosystem II (PSII) photochemistry (Fv/Fm) can be applied as an indicator to distinguish the injury level of plants under stress [21]. For example, the Fv/Fm values of health leaves were distributed in the range 0.72 - 0.85 and the less Fv/Fm denoted that the PSII has been damaged. In general, the Fv/Fm < 0.6 denoted the plants is injured but could be recovered; otherwise, the Fv/Fm < 0.3 denoted the plants is injured severely and could not be recovered [22]. In order to understand the injury level of TNG67, Pq72 and Pq1192 under paraquat treatment, Fv/Fm values of rice plants at tillering stage were determined (Table 1). No significant difference in Fv/Fm of leaf segments excised from TNG67, Pq1192 and Pq72 before treatment, all the Fv/Fm values were about 0.8. However, the Fv/Fm of Pq1192(S) line was reduced to 0.596 (74.1%) which is lower than TNG67 (0.662; 81.3%) and Pq72 (0.687; 85.0%) at 3 h after treatment. At the same time, significant water soaking spots were also

Table 1. Photosystem II inhibition of leaf segments, excised from 21-day-old rice plants, after treatment with 100 μ Mparaqua. Rice cv. TNG67, and both tolerant (Pq72) and susceptible (Pq1192) mutants were compared for the evaluation of paraquat tolerance.

Line				Fv/Fm					
	Time after treatment (Hours)								
	0	3	6	9	12	24	48		
TNG67	$0.814 \pm 0.005^{a*}$	0.662 ± 0.028^a	0.554 ± 0.034^{a}	0.521 ± 0.045 ^b	0.555 ± 0.043^{b}	0.500 ± 0.043^{b}	0.442 ± 0.076^{ab}		
Pq72	0.808 ± 0.008^{a}	0.687 ± 0.019^a	0.621 ± 0.014^{a}	0.645 ± 0.035^a	0.677 ± 0.029^a	0.635 ± 0.041^{a}	0.587 ± 0.060^{a}		
Pq1192	0.805 ± 0.007^{a}	0.596 ± 0.045^{b}	0.447 ± 0.056^{b}	0.463 ± 0.044^{b}	0.503 ± 0.044^{b}	0.437 ± 0.041^{b}	0.365 ± 0.094^{b}		

^{*}Values are mean of Fv/Fm \pm SD. Values with the same letter are not significantly at P < 0.05.

observed on the leaves of Pq1192. At 6 h after treatment, the Fv/Fm values of all lines were significantly reduced and the injury spots were observed on the leaves. However, the injury of Pq72 delayed from 6 to 24 h and Fv/Fm ratio maintained above 0.6 (Table 1).

At any time during treatment, the Fv/Fm values of Pq72 were almost higher than the ones of TNG67 and Pq1192. Even at 48 h, the Fv/Fm ration of Pq72 was 0.587 (72.6%) still significantly higher than those of TNG67 (0.442; 54.3%) and Pq1192 (0.365; 45.4%). These results revealed that Pq72 was also injured slightly by paraquat in the initial stage but the injury development was significantly delayed. These results are also consistent with the suggestion that the Pq72 is more tolerant than the other lines, and Pq72 efficiently delays the injury of paraquat [17].

3.2. Inheritance of Paraquat Tolerance

In order to determine the inheritance of paraquat tolerance, reciprocal crosses were made between Pq1192 and Pq72 and the F_1 progeny was confirmed by SSR (simple sequences repeat) markers (data not shown). There were 36 and 42 of true bred F_1 individuals selected from Pq72/Pq1192 and Pq1192/Pq72, respectively. After paraquat treatment, the frequency distribution of injury index of F_1 populations was shown in **Figure 2**. The F_1 plants of Pq72/Pq1192 show significant less injury index than that of Pq1192/Pq72 cross. The significant differences in injury index of two reciprocal crosses suggest the maternal effect on paraquat tolerance.

In the F_2 population, a total of 120 individuals were planted in the field and 40 individuals were selected randomly for determination of chlorophyll fluorescence and electrolyte leakage after paraquat treatment. In chlorophyll fluorescence analysis, the Fv/Fm values of TNG67, Pq72 and Pq1192 were 0.665, 0.724 and 0.508, respectively. The Fv/Fm values of the F_2 populations from Pq72/Pq1192 and Pq1192/Pq72 distributed among 0.354 - 0.732 and 0.254-0.602, respectively (**Figure 3**). The mean Fv/Fm values showed significant differences from the F_2 populations of Pq72/Pq1192 and Pq1192/Pq72 were 0.574 and 0.457, respectively.

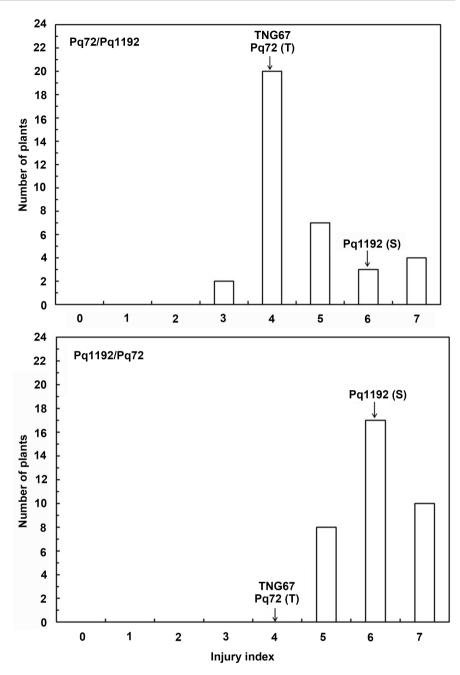


Figure 2. Frequency distribution of injury index of the leaf segments excised from the individuals of the reciprocal crosses, Pq72/Pq1192 and Pq1192/Pq72, in F_1 generation.

The responses of rice in electrolyte leakage assay, the percentages of electrolyte leakage (PELs) from three rice lines TNG67, Pq72 and Pq1192 were 30.2%, 20.4% and 44.6%, respectively. The frequency distributions of PELs of F_2 populations, Pq72/Pq1192 and Pq1192/Pq72, were shown in **Figure 4**. The PELs of the F_2 populations from Pq72/Pq1192 and Pq1192/Pq72 distributed among 16.4% - 50.5% and 27.3% - 58.7%, respectively. The mean PELs of the F_2 populations from Pq72/Pq1192 and Pq1192/Pq72 were 32.3% and 44.6%, respectively. These results also display a significant difference in the results of reciprocal

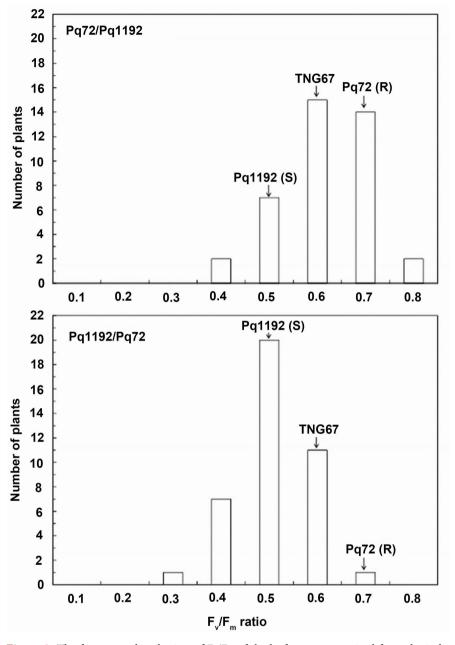


Figure 3. The frequency distribution of F_v/F_m of the leaf segments excised from the individuals of the two reciprocal crosses, Pq72/Pq1192 and Pq1192/Pq72, in F_2 generation.

crosses and are similar to the results in the frequency distributions of injury index of F_1 populations. These findings suggested that the influence of maternal effect on paraquat tolerance is present.

Furthermore, the individuals with Fv/Fm value less than 0.5 were defined as susceptibility; otherwise the ones higher than 0.5 were defined as tolerance. At the same time, the individuals with PLEs higher than 35% were defined as susceptibility; otherwise the ones lower than 35% were defined as tolerance. The Chi-square test fits the expected ratio in F_2 population (**Table 2**). The results suggested that the F_2 population of Pq72/Pq1192 segregated to 3:1 (T:S)

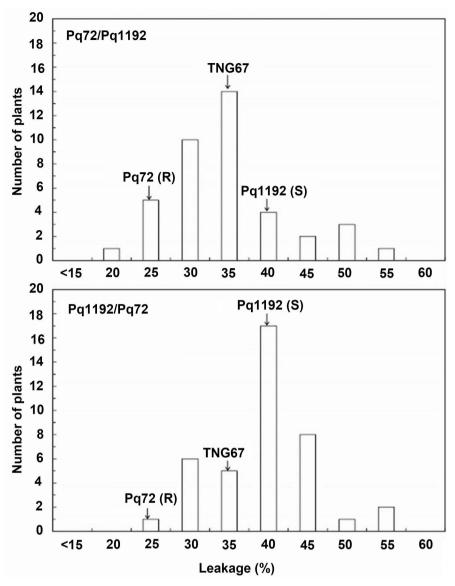


Figure 4. Frequency distribution of electrolyte leakage percentages of the leaf segments excised from the individuals of the two reciprocal crosses, Pq72/Pq1192 and Pq1192/Pq72, in F_2 generation.

according to Fv/Fm and leakage investigation. However, the F_2 population of reciprocal cross, Pq72/Pq1192, did not segregate to theoretical ratio, 1:3. It implies that the paraquat tolerance of Pq72 is maternally controlled by a single dominant gene.

Most studies displayed that genetic characteristic of paraquat resistance was controlled by the nuclear genes, such as *Conyza bonariensis* [23], *Erigeron philadelphicus* [24], *Erigeron Canadensis* [25], *Hordeum glaucum* [26], *Hordeum lepornum* [27], *Arctotheca calendula* [27] and *Lolium rigidum* [28]. However, the paraquat tolerance inheritance of Pq72 is influenced by the maternal effect. This result is similar to the findings that the inheritances of triazine resistance of some weeds were the examples of cytoplasmic inheritance [29] [30] [31].

Table 2. Segregation for paraquat tolerances of F_2 populations from Pq72/Pq1192 and Pq1192/Pq72.

Dhysiala sigal tuaita	F	No	of F ₂ pla	nts	Genetic	- 2	
Physiological traits	F ₂ generation	T	S	Total	ratio tested	χ^2	
E /El	Pq72/Pq1192	31	9	40	3:1	0.133 ^{ns}	
F _v /F _m value	Pq1192/Pq72	12	28	40	1:3	0.533 ^{ns}	
Tl	Pq72/Pq1192	30	10	40	3:1	$0^{\rm ns}$	
Leakage	Pq1192/Pq72	12	28	40	1:3	0.533 ^{ns}	

Critical value: χ^2 (1, 0.05) = 3.84; ^{ns} means that the probability is not significant.

The mechanism of triazines resistance was postulated that the electron transfer in photosystem II is hindered by this kind of herbicide, and a G264S mutation of D1 protein in photosystem II causes the triazine resistance of *Brassica napus* L. [32]. On the other hand, the action mechanism of paraquat was proposed that the electron transfer in photosystem I of plants was interfered by paraquat [2] [3] [4] [5]. The genetic analysis also displays that the paraquat tolerance of Pq72 is controlled by a single dominant gene. According to these findings, we propose that the paraquat tolerance gene of Pq72 might participate in the electron transfer in the chloroplastic thylakoid.

It has been found that the paraquat resistant mutant (parl) of Arabidopsis is contributed by a strong transporter protein localized to the Golgi apparatus due to an amino acid mutation caused the reduction of intracellular transportation of paraquat and accumulation into chloroplast [11]. Transgenic rice plants carry the RNA interference of the PAR1-like gene, OsPAR1, showed significant paraquat resistance by reduction of paraquat transportation and accumulation less into chloroplasts but no difference in paraquat uptake was observed [11]. Same results were reported in our previous report that no differences were detected in the uptake and transportation of paraquat after application in the three tested rice lines and the tolerance is due to the delay of cellular damage in the tolerance mutant [17]. These results supported that the organelles or cytoplasmic factors [7] [33] [34] of the tolerance lines may contribute to paraguat tolerance. Therefore, our results showed significant maternal effects in all the physiological traits detected. However, it is difficult to map the paraquat tolerance gene of Pq72, because of cytoplasmic inheritance of Pq72 paraquat tolerance. Our ongoing molecular genetic study on the paraquat T/S mutants will provide a clear information for the mechanism of paraguat tolerance in mutants.

In this study, the properties and inheritance of paraquat tolerance in mutant Pq72 was characterized. These results demonstrate that Pq72 is a useful germplasm for studying the mechanism of paraquat tolerance in rice improvement. The increasing activities of anti-oxidation enzymes were proposed to explain to be responsible for paraquat tolerance [17]. Therefore, the expression profiles of the genes involved in the anti-oxidation pathway of Pq72 will be de-

termined through transcriptomics analysis and molecular genetics to clarify the mechanism of paraquat tolerance in Pq72.

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Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] TACTRI/COA (2012) Plant Protection Manual: Agricultural Chemicals and Toxic Substances Research Institute, Council of Agriculture, Executive Yuan, Taiwan.
- [2] Babbs, C.F., Pham, J.A. and Coolbaugh, R.C. (1989) Lethal Hydroxyl Radical Production in Paraquat-Treated Plants. *Plant Physiology*, 90, 1267-1270. https://doi.org/10.1104/pp.90.4.1267
- [3] Dodge, A.D. (1971) The Mode of Action of the Bipyridylium Herbicides, Paraquat and Diquat. *Endeavour*, **30**, 130-135. https://doi.org/10.1016/0160-9327(71)90039-1
- [4] Haley, T.J. (1979) Review of the Toxicology of Paraquat (1,1'-dimethyl-4,4'-bipyridinium chloride). Clinical Toxicology, 14, 1-46. https://doi.org/10.3109/15563657909030112
- [5] Suntres, Z.E. (2002) Role of Antioxidants in Paraquat Toxicity. *Toxicology*, **180**, 65-77. https://doi.org/10.1016/S0300-483X(02)00382-7
- [6] Bromilow, R.H. (2004) Paraquat and Sustainable Agriculture. *Pest Management Science*, **60**, 340-349. https://doi.org/10.1002/ps.823
- [7] Tsuji, K.H.M., Morita, S., Miura, R. and Tominaga, T. (2013) Resistance to Paraquat in *Mazus pumilus. Weed Research*, **53**, 176-182. https://doi.org/10.1111/wre.12012
- [8] Ahlfors, R., et al. (2004) Arabidopsis RADICAL-INDUCED CELL DEATH1 Belongs to the WWE Protein-Protein Interaction Domain Protein Family and Modulates Abscisic Acid, Ethylene, and Methyl Jasmonate Responses. Plant Cell, 16, 1925-1937. https://doi.org/10.1105/tpc.021832
- [9] Chen, R., et al. (2009) The Arabidopsis PARAQUAT RESISTANT2 Gene Encodes an S-Nitrosoglutathione Reductase That Is a Key Regulator of Cell Death. Cell Research, 19, 1377-1387. https://doi.org/10.1038/cr.2009.117
- [10] Fujita, M., et al. (2012) Natural Variation in a Polyamine Transporter Determines Paraquat Tolerance in Arabidopsis. Proceedings of the National Academy of Sciences of the United States of America, 109, 6343-6347. https://doi.org/10.1073/pnas.1121406109
- [11] Li, J., *et al.* (2013) PARAQUAT RESISTANT 1, a Golgi-Localized Putative Transporter Protein, Is Involved in Intracellular Transport of Paraquat. *Plant Physiology*, **162**, 470-483. https://doi.org/10.1104/pp.113.213892
- [12] Tsugane, K., et al. (1999) A Recessive Arabidopsis Mutant That Grows Photoautotrophically under Salt Stress Shows Enhanced Active Oxygen Detoxification. Plant

- Cell, 11, 1195-1206. https://doi.org/10.1105/tpc.11.7.1195
- [13] Xi, J., Xu, P. and Xiang, C.B. (2012) Loss of AtPDR11, a Plasma Membrane-Localized ABC Transporter, Confers Paraquat Tolerance in Arabidopsis Thaliana. *The Plant Journal*, **69**, 782-791. https://doi.org/10.1111/j.1365-313X.2011.04830.x
- [14] Fuerst, E.P. and Vaughn, K.C. (1990) Mechanisms of Paraquat Resistance. *Weed Technology*, **4**, 150-156. https://doi.org/10.1017/S0890037X0002515X
- [15] Hart, J.J. and Di Tomaso, J.M. (1994) Sequestration and Oxygen Radical Detoxification as Mechanisms of Paraquat Resistance. *Weed Science*, **42**, 277-284.
- [16] Wang, C.S., Tseng, T.H. and Lin, C.Y. (2003) Rice Biotech Research at the Taiwan Agricultural Research Institute. In: Yang, H., Yu, J., Ramachandran, S. and Pan, S., Eds., Rice Biotech Research at the Taiwan Agricultural Research Institute in Rice Genomics (I): A Holistic Approach to Rice Research and Genetic Engineering, World Scientific Publishing Co., Singapore, 111-128. https://doi.org/10.1142/9789812791344_0009
- [17] Tseng, T.Y., Ou, J.F. and Wang, C.Y. (2013) Role of Ascorbate-Glutathione Cycle in Paraquat Tolerance of Rice (*Oryza sativa*). Weed Science, 61, 361-373. https://doi.org/10.1614/WS-D-12-00133.1
- [18] Harbinson, J., Genty, B. and Baker, N.R. (1989) Relationship between the Quantum Efficiencies of Photosystems I and II in Pea Leaves. *Plant Physiology*, **90**, 1029-1034. https://doi.org/10.1104/pp.90.3.1029
- [19] Lu, S., Guo, Z. and Peng, X. (2003) Effects of ABA and S-3307 on Drought Resistance and Antioxidative Enzyme Activity of Turfgrass. *The Journal of Horticultural Science and Biotechnology*, 78, 663-666. https://doi.org/10.1080/14620316.2003.11511681
- [20] Genty, B., Briantais, J.-M. and Baker, N.R. (1989) The Relationship between the Quantum Yield of Photosynthetic Electron Transport and Quenching of Chlorophyll Fluorescence. *Biochimica et Biophysica Acta*, 990, 87-92. https://doi.org/10.1016/S0304-4165(89)80016-9
- [21] Kooten, O. and Snel, J.F.H. (1990) The Use of Chlorophyll Fluorescence Nomenclature in Plant Stress Physiology. *Photosynthesis Research*, 25, 147-150. https://doi.org/10.1007/BF00033156
- [22] Bolhár-Nordenkampf, H.R. and Götzl, M. (1992) Chlorophyllfluoreszenz als Indikator der mit Seehöhe zunehmenden Streßbelastung von Fichtennadeln. *FBVA Berichte. Schriftenreihed. Forstl. Bundesveranst.*, **67**, 119-131.
- [23] Shaaltiel, Y., *et al.* (1988) Dominant Pleiotropy Controls Enzymes Co-Segregating with Paraquat Resistance in *Conyza bonariensis. Theoretical and Applied Genetics*, **75**, 850-856.
- [24] Itoh, K. and Miyahara, M. (1984) Inheritance of Paraquat Resistance in *Erigeron philadelphicus* L. *Weed Research*, **29**, 301-307.
- [25] Yamasue, Y., et al. (1992) Paraquat Resistance and Its Inheritance in Seed Germination of the Foliar-Resistant Biotypes of Erigeron canadensis L. and E. sumatrensis Retz. Pesticide Biochemistry and Physiology, 44, 21-27. https://doi.org/10.1016/0048-3575(92)90005-K
- [26] Islam, A.K.M.R. and Powles, S.B. (1988) Inheritance of Resistance to Paraquat in Barley Grass *Hordeum glaucum* Steud. *Weed Research*, **28**, 393-397. https://doi.org/10.1111/j.1365-3180.1988.tb00819.x
- [27] Purba, E., Preston, C. and Powles, S.B. (1993) Inheritance of Bipyridyl Herbicide Resistance in *Arctotheca calendula* and *Hordeum leporinum. Theoretical and Applied Genetics*, **87**, 598-602.

- [28] Yu, Q., *et al.* (2009) Paraquat Resistance in a *Lolium rigidum* Population Is Governed by One Major Nuclear Gene. *Theoretical and Applied Genetics*, **118**, 1601-1608. https://doi.org/10.1007/s00122-009-1008-3
- [29] Darmency, H. and Pernes, J. (1985) Use of Wild *Setaria viridis* (L.) Beauv. to Improve Triazine Resistance in Cultivated *S. italica* (L.) by Hybridization. *Weed Research*. **25**, 175-179.
- [30] Scott, K.R. and Putwain, P.D. (1981) Maternal Inheritance of Simazine Resistance in a Population of *Senecio vulgaris. Weed Research*, **21**, 137-140. https://doi.org/10.1111/j.1365-3180.1981.tb00108.x
- [31] Souza Machado, V., *et al.* (1978) Uniparental Inheritance of Chloroplast Atrazine Tolerance in *Brassica campestris. Canadian Journal of Plant Science*, **58**, 977-981. https://doi.org/10.4141/cjps78-150
- [32] Sundby, C., Chow, W.S. and Anderson, J.M. (1993) Effects on Photosystem II Function, Photoinhibition, and Plant Performance of the Spontaneous Mutation of Serine-264 in the Photosystem II Reaction Center D1 Protein in Triazine-Resistant *Brassica napus* L. *Plant Physiology*, 103, 105-113. https://doi.org/10.1104/pp.103.1.105
- [33] Han, H.J., Zhu, B., Fu, X.Y., Zhao, W., Shi, B. and Yao, Q.H. (2014) Gene Expression Profiles of Arabidopsis under the Stress of Methyl Viologen: A Microarray Analysis. *Molecular Biology Reports*, 41, 7089-7102. https://doi.org/10.1007/s11033-014-3396-y
- [34] Kim, J.-G. and Jung, S. (2013) Differential Antioxidant Mechanisms of Rice Plants in Response to Oxyfluorfen and Paraquat. *Weed & Turfgrass Science*, **2**, 254-259. https://doi.org/10.5660/WTS.2013.2.3.254

Supplemental

Table S1. The 21 polymorphic SSR markers between TNG67, Pq72 (T) and Pq1192 (S).

Marker	CI.	Position	PCR product (bp) in		
	Chromosome	(cM)	TNG67 or Pq72(T)	Pq1192(S)	
RM495	1	2.8	150	136	
RM1	1	29.7	82	97	
RM259	1	54.2	163	146	
RM307	4	0	119	113	
RM507	5	0	256	247	
RM413	5	26.7	63	69	
RM161	5	96.9	168	152	
RM178	5	118.8	105	107	
RM334	5	141.8	132	173	
RM133	6	0	111	109	
RM125	7	24.8	117	114	
RM11	7	47	116	131	
RM455	7	65.7	122	119	
RM118	7	96.9	148	151	
RM408	8	0	121	115	
RM105	9	32.1	94	101	
RM215	9	99.4	147	143	
RM271	10	59.4	86	99	
RM287	11	68.6	212	250	
RM19	12	20.9	104	115	
RM277	12	57.2	115	119	

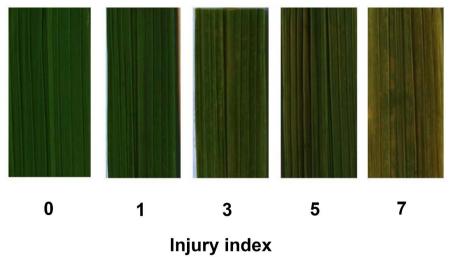


Figure S1. The injury index used in this study. It was classified from 0 to 7, where 0 indicates healthy leaf segment without any herbicidal damage and 7 indicates the completely etiolated leaf segment caused by paraquat.