

Persistence of *Venturia inaequalis* Populations Resistant to Strobilurins in the Field and in the Glasshouse

Riccardo Fiaccadori

Department of Agricultural Sciences (DIPSA), University of Bologna, Bologna, Italy

Email: riccardo.fiaccadori@unibo.it

How to cite this paper: Fiaccadori, R. (2018) Persistence of *Venturia inaequalis* Populations Resistant to Strobilurins in the Field and in the Glasshouse. *American Journal of Plant Sciences*, 9, 552-560. <https://doi.org/10.4236/ajps.2018.94042>

Received: November 18, 2017

Accepted: March 4, 2018

Published: March 7, 2018

Copyright © 2018 by author and Scientific Research Publishing Inc. This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

The research was focused on the persistence of resistance of *V. inaequalis* to strobilurins from resistant populations occurring in the field. It was studied with two types of experiments: the first in apple orchards where resistance occurred in 2005 and employ of these fungicides was suspended from 2006 to 2011, realizing every year sensitivity/resistance assays on strobilurins. In the second was verified the sensitivity/resistance of the same 2015 populations, repeatedly multiplied (ten inoculations) in glasshouse on apple seedlings. Results showed that the resistance survived in orchards all the years of tests (six years), despite sensitivity fluctuations occurred on infected leaves due to sexual crosses, competition with sensible strains and soil management. In glasshouse, the level of resistance of all populations increased quickly in first inoculations and values remained high ($EC_{50} > 10 \text{ mg}\cdot\text{L}^{-1}$) in all the following inoculations, until the last, the 10th. *V. inaequalis* resistant strains seemed to show high fitness: relevant persistence in the field, and a capacity of prevail on sensible ones in absence of factors of field variability.

Keywords

Venturia inaequalis, Persistence, Resistance, Strobilurins, Scab Management

1. Introduction

Venturia inaequalis (Cke) Wint. is the causal agent of one of the most important diseases of apple cultivation in over 65 countries in the world [1]. The control of this disease requires many treatments with several chemical groups of fungicides and possibly an expert technical organization. Strobilurins fungicides, named also QoI (Inhibitors of Quinone outside), were introduced at the end of 90 years.

They showed excellent activity against apple scab even when applied under high concentration of inoculum and favorable epidemiological conditions. However, cases of practical resistance of *V. inaequalis* were reported in the early 2000s [2] [3] and were confirmed by further surveys [4] [5].

In particular, the survey conducted in Italy showed that the presence and frequency of resistance to strobilurins differed among apple areas [6] and in those more interested by the problem the use of these QoI fungicides was generally interrupted. The knowledge of the fitness of these populations could be interesting in these situations, especially to evaluate the persistence and the possibility to the back employ of these fungicides where the problem occurred. There is currently a lack of information about the persistence of resistant populations to different fungicides groups: few reports deal on SBI and dicarboximides, while a higher number of researches are available for benzimidazoles. Concerning the last group of fungicides, resistant strains have shown a high persistence of resistance, 10 years in *Tapesia* sp. [7] and over 4 years in *Venturia pirina* [8]. A persistence of resistance of 10 years was also detected for ethirimol (chemical group IBS) in *Spaeroteca fuliginea* [9], while resistance to dicarboximides showed a rather high decrease after two years of no use on *Botrytis cinerea* [10] and *Monilia fructicola* [11].

The aim of this research was to verify the evolutions of *V. inaequalis* populations that were found to be resistant to strobilurins in Italian commercial orchards in 2005. Experiments were realized following two lines: 1) to verify the persistence of resistance of these 2005 field populations in the same orchards, from 2006 to 2011, in which strobilurins were no more used; 2) to assay persistence of resistance and fitness of these 2005 resistant populations in glasshouse, in absence of perturbative agents on these phenotypes (gamic cross, climatic and epidemiologic situations, fungicides etc.).

2. Materials and Methods

2.1. Population's Collection

Populations of *V. inaequalis* were collected from four apple orchards (N. 143, 144, 148 and 150, comprehending apples cv Imperatore Dallago, from five to eight years old) of Emilia-Romagna Region (Northern Italy) where strobilurin fungicides (kresoxim-methyl firstly and trifloxystrobin subsequently) were applied for several years, and where resistance to strobilurins in 2005 was detected [3].

So scabbed leaves (30 - 50 in number) of the field for each population were randomly collected and 50 - 60 scab spots were added with a drop of sterile water, successively sucked, obtaining a dispersion with conidia. It was filtered and adjusted to a conidia concentration of $1 - 2 \times 10^5 \text{ mL}^{-1}$. The dispersion was inoculated on leaves of apple seedlings in glasshouse and after about 22 days the leaves with sporulated conidia were detached and conserved in silica gel until the use in the two kinds of test.

2.2. Persistence of Resistance: Assays in the Field

In the tested orchards, where strobilurins were used and resistance was detected in 2005, strobilurins were not applied from 2006 to 2011, and apple scab control was carried out with other kind of fungicides such as aniline-pyrimidines, sterol biosynthesis inhibitors and dithianon.

Every year, samples of leaves were collected in each orchard, at the end of primary or at the beginning of secondary infective cycle, choosing scabbed leaves, on which was realized the multiplication of conidia, as explained before.

2.2.1. Preparation of Fungicide

The strobilurin named trifloxystrobin was used for tests, because it was the mostly employed in considered orchards and in apple areas. The fungicide was used as active material (Sigma-Aldrich, St. Louis, USA). It was dissolved in acetone at final concentration $< 0.01 \text{ L}^{-1}$ and added to water-agar (2%, Agar Grade A, Becton, Dickinson and Company) to achieve the following fungicide concentrations: 0, 0.001, 0.01, 0.1 and $2 \text{ mg}\cdot\text{L}^{-1}$. The antibiotic streptomycin sulphate was added to water agar at a final concentration of $200 \text{ mg}\cdot\text{L}^{-1}$.

2.2.2. Preparation of Conidia

Pieces (0.01 g) of scabbed leaves were randomly collected from each multiplied population and introduced in micro tubes containing 1 mL of sterile water. After shaking, the conidial concentration was adjusted to $1 - 3 \times 10^5 \text{ conidium L}^{-1}$. Two drops of $20 \mu\text{L}$ of spore suspension for every fungicide concentration were placed on agar plates and incubated for 24 hours at 20°C .

2.2.3. In Vitro Assays

300 conidia per concentration were visually assessed at the microscope, counting those germinated. Tests showing less than 50% conidia germinated in the control were not considered. On each population the assays were repeated three times, which permitted to get the mean (geometric) EC_{50} values (fungicide Efficacy Concentration reducing 50% the percentage of germination of conidia) by probits analysis and $\text{EC}_{50 \text{ max}}$ (maximum value of EC_{50} detected among the several *in vitro* tests on every population). EC_{50} values were calculated with probits using Finney & Stevens table [12]. The software of the probits program was Excel.

The classification of a population as sensible or resistant was based on previous researches on different types of population (wild type, well and poorly controlled ones) that showed specific sensitivities [3] [6].

The used parameter was the $\text{EC}_{50 \text{ max}}$ value because had a practical use, identifying the lowest value of sensitivity in population and consequently in the orchard. Sensible populations showed all the EC_{50} values inferior to 0.063 mg/L . When at least one EC_{50} value was higher than $0.169 \text{ mg}\cdot\text{L}^{-1}$, the population was considered resistant. Values in the very short intermediate range from 0.063 and 0.169 mg/L are considered in an intermediate situation, difficultly to evaluate as sensitivity. Wild-types populations showed always EC_{50} values inferior to 0.031

mg/L.

A classification of populations was also realized as mean EC_{50} values, where wild-type populations have EC_{50} mean inferior to 0.013 mg/L, those sensible are inferior to 0.049 mg/L, while that resistant are superior to 0.049 mg/L. This method presents values not highly different from that based on EC_{50} max, but this is more reliable for a practical evaluation of the presence of resistance in the orchards. However the EC_{50} value presents the advantage to be scientifically more available for evaluations of sensitivity of populations and can be better utilized in statistic elaborations.

2.3. Persistence of Resistance: Assays in Glasshouse

The conidia from orchards with resistance to strobilurins, after the propagation (see Chapter 2.2), were repeatedly inoculated on apple seedlings (10 times) in glasshouse, using always the conidia of the previous inoculation (Table 4). The inoculations were realized with little mechanical sprayers containing from 20 to 30 mL. After two-four inoculations, the conidia of each sample were subjected to an *in vitro* evaluation on strobilurins sensitivity. Each inoculation was realized on 8 - 10 apple seedlings (cv. Golden), 4 - 5 weeks old, with 5 - 7 formed leaves.

Inoculation of *V. inaequalis* populations was performed with a suspension of conidia at the concentration of $1 - 2 \times 10^5$ mL of sterile water that was uniformly sprayed on both leaf surfaces (1.2 mL each plant). The seedlings were then put in a plastic bag, closed to maintain wetness on leaves for 48 hours at 18°C. Subsequently the plants were incubated for further 18 days with light at an intensity of 25μ Einstein. $m^{-2} \cdot s^{-2}$, a photoperiod of 12 hours a day, mean temperature about 18°C and 60% - 80% relative humidity. Then the scabbed leaves were collected and dried with paper and silica gel until the following inoculation.

2.3.1. Sensitivity Assays *In Vivo*

To evaluate the persistence of resistance, *in vitro* tests (sensitivity as EC_{50} values) were realized with the same methodology used for field and glasshouse populations. Three assays were realized, utilizing for each one a subpopulation with a part of scabbed leaves. The methodology used was previously described in the paper.

2.3.2. Statistic Analysis

ANOVA factorial analysis was preliminarily realized on the three populations with the higher number of data based on the most important parameter: mean EC_{50} value. Successively a non-parametric analysis with ez-Perm function (Ez package in R) was performed [Stat.Soft.inc. (2013), STATISTICA (version 12)], P: 0.05.

3. Results

3.1. Assays from Orchards

The evolution of sensitivity to strobilurins from 2005 to 2011 in the four orc-

hards, expressed as concentrations of fungicide (mean and max EC₅₀ values) is showed in **Table 1** and data are evaluated with parametric and non parametric analysis. EC₅₀ values in the first year presented a rather high range of values. The populations N. 143 presented high mean and max EC₅₀ values in 2005, and showed a light fluctuation of sensitivity in the following years, without employ of strobilurins, always in the range of resistant values, returning in 2011 to EC₅₀ values similar to 2005.

Population N. 144 presented lower values (as EC₅₀) in 2005, but in the following years, the EC₅₀'s showed no reduction, but on the contrary, there is an increase until 2011. Even in this case the EC₅₀ values are always superior to the minimum values for resistance.

In addition, population N. 150 showed a variability of EC₅₀ values (both considering mean and max), with fluctuations in the years, from 2005 to 2009, that, however, permitted to population to remain always in a situation of resistance.

Samples of population N. 148 were collected only in 2005, 2009 and 2011 and showed, quite always, very low values of sensitivity as EC₅₀ values, with only a case in 2011, of moderate increase of sensitivity (EC₅₀ mean) that remained, however, in a situation of resistance.

The statistic elaboration used, with ANOVA factorial analysis (**Table 2**), showed no differences about mean sensitivity data among the populations and the years; but it presents a poor homogeneity in variance.

As data are not suitable to be examined in a parametric method, anon-parametric analysis with ez-Perm function was realized (**Table 3**). Also this elaboration evidenced a non significative interaction between years and populations in EC₅₀ means; so all the sensitivity values, in all considered years, were caused only by the occurrence and the persistence of resistance.

Table 1. Effective EC₅₀ values of trifloxystrobin on *V. inaequalis* resistant populations in commercial orchards from 2005 to 2011.

N° population	2005		2006		2007		2008		2009		2010		2011	
	EC ₅₀ mean	EC ₅₀ max												
143	>10	>10	3.3	5.6	2.11	4.60	1.81	>10	7.35	>10	1	1	3.7	>10
144	0.32	4.11	0.29	3.19	1.47	1.92	1.2	6.6	3.68	>10	>10	>10	7.6	<10
150	1.11	6.47	>10	>10	1.25	5.12	2.11	4.8	3.42	<10		2		
148	>10	>10	1	1	1	1	1	1	>10	>10	1	1	1.48	>10

1: sample not available; 2: estirpate orchard.

Table 2. Parametric variance analysis with ANOVA test.

Effect	Sum of squares	Degree of freedom	Variance	Variance error	F-value	P-value
Year	132.72	5	26.45	29.98	0.88	0.53
Population	4.25	2	2.12	29.98	0.07	0.93
Year × population	269.90	9	29.98	27.95	1.07	0.40

Table 3. Non-parametric analysis of variance with EZ-Perm.

Effect	Effect	SS	Degree	MS	Den.Syn.	Den.Syn.	F-value	P-value
	(F/R)		Freedom		Error df	Error MS		
Intercept	Fixed	183.38	1	183.38	5.04	26.46	6.92	0.04
Year	Random	132.27	5	26.45	9.00	29.98	0.88	0.53
Popul/site	Fixed	4.25	2	2.12	9.00	29.98	0.07	0.93
Year × popul/site	Random	269.90	9	29.98	34.00	27.95	1.07	0.40
Error		950.32	34	27.95				

3.2. Assays from Glasshouse

Three of the four *V. inaequalis* populations, which occurred as resistant to strobilurins in orchards in 2005 year, were used also for glasshouse studies. These populations were repeatedly inoculated in glasshouse as reported in **Table 4**. The first examined population, N. 143, showed very high EC_{50} values in the first assessment (after the second inoculation), similar to that of field sensitivity in 2005. This high reduction of sensitivity (mean and max EC_{50} values) remained constantly high also after the 4th, 8th and 10th inoculation.

Also the population N. 144 showed very high EC_{50} values already after the first assessment (2nd inoculation) with mean and max EC_{50} values that are sensibly higher compared to the initial sensitivity in 2005. This population maintained substantially the same level of very low sensitivity after the other three inoculations.

Likewise the population N. 150 presented a very low sensitivity already after the first glasshouse assessment, showing higher EC_{50} (mean and max) compared to the initial sensitivity values from the orchard. The population maintains the same high degree of resistance also after the three cited assays, subsequent to the 4th, 8th and 10th inoculation.

4. Discussion

The research was carried out examining *V. inaequalis* populations from four Italian orchards that were assayed and classified as resistant to strobilurins in 2005 (characterized by poor field control and high EC_{50} values for the prevalence of resistant spores respecting sensible ones). The evolution and persistence of resistant populations were studied in that orchards, assaying sensitivity to strobilurins in the following six years), and in glasshouse, where the populations coming from these orchards were repeatedly inoculated (ten times).

The assessment of sensitivity of *V. inaequalis* populations permits to evidence that all tested orchards can be considered resistant during all the six years, despite fluctuations in sensitivity due to several causes.

Indeed all fungi populations, comprehending also those that are a mix between sensible and resistant spores, present sexual crosses between antheridia and oogonia in parasitic phase of the fungus in winter that can modify characteristics of strains and populations, also regarding sensitivity. Moreover, during

Table 4. Mean EC₅₀ values of trifloxystrobin on *V. inaequalis* resistant populations, repeatedly inoculated* on seedlings in glasshouse.

N° population	Initial sensitivity of populations (2005)		Results of sensitivity (<i>in vitro</i> test) on spores after the sporulation of:								
			2 nd inoculation		4 th inoculation		8 th inoculation		10 th inoculation		
	Mean EC ₅₀	Max EC ₅₀	Mean EC ₅₀	Max EC ₅₀	Mean EC ₅₀	Max EC ₅₀	Mean EC ₅₀	Max EC ₅₀	Mean EC ₅₀	Max EC ₅₀	
143	>10	>10	8.6	>10	>10	>10	>10	>10	>10	>10	>10
144	0.32	0.39	>10	>10	>10	>10	>10	>10	>10	>10	>10
150	0.18	6.47	>10	>10	>10	>10	>10	>10	>10	>10	>10

*Inoculations: 1st 08.05.09; 2nd 10.11.09; 3rd 29.12.09; 4th 02.02.10; 5th 31.03.10; 6th 12.05.10; 7th 17.06.10; 8th 12.07.10; 9th 07.09.11; 10th 08.01.11.

the vegetative phase, the competition between resistant and sensible strains during repeated agamic cycles of the fungus can modify the proportion of resistant and sensible spores. In addition, the soil management could have influence, as lawn permits a better conservation of the leaves and of the populations of *V. inaequalis*, while ploughings and shredding could cause reductions, selections and modifications of populations. However, I have observed that these modifications were not able to modify the status of populations which have a majority of spores resistant to strobilurins. Concluding, the presented data emphasize that the resistance of *V. inaequalis* to strobilurins showed characteristics of a long persistence in the orchards, at least 6 years.

The assays in glasshouse evidenced that repeated inocula increased quickly the levels of resistance until the maximum admissible values (EC₅₀ >10 mg/L). It must be pointed out that the conidia were used in glasshouse, only with agamic multiplications, while the conservation occurred at -20°C, in lyophilized status. In these situations, these conidia appear to undergo an inferior variability respecting those in field conditions, previously examined.

As presented in introduction, the knowledges about the persistence of fungicide resistance of strains of populations are very low on less recent chemical groups (IBS, dicarbossimides, benzimidazoles) and practically absent on more recent others, as strobilurins, and anilino-pyrimidines. Now we are continuing researches on persistence to QoI in the same orchards and are working on persistence of resistance to difenoconazole, but I do not know other research groups that are involved on the same arguments.

The persistence of resistance in fungi populations is an aspect of the much wider argument of the fitness of resistance strains.

About strains resistant to strobilurins, researches on *Erisiphe graminis* showed no differences in aggressiveness between the two types of strains (sensible and resistant) [13], while in several other cases a reduced fitness of isolates resistant to strobilurins is reported, but it must be pointed out that it generally concerns artificial mutants with the introduction of G143A substitution, as on *Botrytis cinerea* [14]. However it is not known the field behavior of these resistant

strains.

5. Conclusions

In conclusion, the aspects of fitness studied that spores resistant to strobilurins, generally related to G143A substitution, do not present a lower fitness respecting sensible ones, as demonstrated on animal and vegetal types of organisms [15]. Our research on *Venturia inaequalis*, especially in glasshouse, supports the theory that the resistant strains to strobilurins present characteristics of fitness surely not inferior to that of sensible strains. I also demonstrated the maintenance of resistance in orchards for several years after its appearance.

Moreover, in controlled conditions, where genetic, environmental or agronomic factors cannot influence these phenotypes and genotypes, the fitness of strains resistant to strobilurins seemed to appear superior to that of sensible strains. However very few data are available about fitness of other resistant fungi populations respecting sensible ones, with referring to all recent and actually used fungicides.

Acknowledgements

Thanks to Enrico Muzzi for advising in statistic analysis, Angela Finestrelli and Elena Cicognani for laboratory help, A. Gilli for field supervision, Cesare Gessler for advices and Marina Collina for logistic help.

Fund

Research realized with fund by Regione Emilia-Romagna (Italy) Servizio Fitosanitario Regionale.

References

- [1] MacHardy, W.E. (1996) Apple Scab Biology, Epidemiology and Management. APS Press, St. Paul, Minnesota, 3.
- [2] Kuck, K.H. and Mehl, A. (2003) Trifloxystrobin: Resistance Risk and Resistance Management. *Pflanzenschutz Nachrichten Bayer*, **56**, 313-325.
- [3] Fiaccadori, R., Cicognani, E., Abbatecola, M., Collina, M. and Brunelli, A. (2005) Sensitivity of *Venturia inaequalis* to Strobilurin Fungicides in Italy. *Communications in Agricultural and Applied Biological Sciences*, **70**, 73-78.
- [4] Sallato, B.V., Latorre, B.A. and Aylwin, G. (2006) First Report on Practical Resistance to QoI Fungicides in *Venturia inaequalis* (Apple Scab) in Chile. *Plant Disease*, **90**, 375. <https://doi.org/10.1094/PD-90-0375A>
- [5] Lesniak, K., Proffer, T., Beckerman, J. and Sundin, G. (2011) Occurrence of QoI Resistance and Detection of the G143A Mutation in Michigan Populations of *Venturia inaequalis*. *Plant Disease*, **95**, 927-934. <https://doi.org/10.1094/PDIS-12-10-0898>
- [6] Fiaccadori, R., Collina, M. and Brunelli, A. (2013) Reduced Sensitivity of *Venturia inaequalis* to Strobilurins and Anilinopyrimidines in Italy. *Proceedings "8th International Conference on Integrated Fruit Production"*, *IOBC-WPRS Bulletin*, **91**, 345-350.
- [7] Leroux, M. and Gredt, E. (1997) Evolution of Fungicide Resistance in the Cereal

- Eyespot Fungi, *Tapesia yallundae* and *Tapesia acuformis* in France. *Pesticide Science*, **51**, 312.
[https://doi.org/10.1002/\(SICI\)1096-9063\(199711\)51:3<321::AID-PS639>3.0.CO;2-U](https://doi.org/10.1002/(SICI)1096-9063(199711)51:3<321::AID-PS639>3.0.CO;2-U)
- [8] Shabi, E. and Kaian, I. (1980) Fitness of *Venturiapirina* Isolates Resistant to Benzimidazoles Fungicides. *Phytopathology*, **70**, 1172-1174.
<https://doi.org/10.1094/Phyto-70-1172>
- [9] Schepers, H.T.A.M. (1984) Persistence of Resistance to Fungicides in *Sphaeroteca fuliginea*. *Netherlands Journal of Plant Pathology*, **90**, 165-171.
<https://doi.org/10.1007/BF02006480>
- [10] Northover, J. (1988) Persistence of Dicarboximide-Resistant *Botrytis cinerea* in Ontario vineyards. *Canadian Journal of Plant Pathology*, **10**, 123-132.
<https://doi.org/10.1080/07060668809501744>
- [11] Sanomuang, N. and Gaunt, R.E. (1995) Persistence and Fitness of Carbendazim- and Dicarboximide-Resistant Isolates of *Monilia fructicola* (Wint.) Honey in Flowers, Shoots and Fruits of Stone Fruits. *Plant Pathology*, **44**, 448-457.
<https://doi.org/10.1111/j.1365-3059.1995.tb01667.x>
- [12] Finney, D.J. and Stevens, W.L. (1948) A Table for the Calculation of Working Probits and Weights in Probit Analysis. *Biometrika*, **35**, 191-201.
<https://doi.org/10.1093/biomet/35.1-2.191>
- [13] Chin, K.M., Chavaillaz, D., Kaebuhrer, M., Staub, T. and Felsenstein, F.G. (2001) Characterizing Resistance Risk of *Erisiphegraminis* sp. *tritici* to Strobilurins. *Crop Protection*, **20**, 87-96. [https://doi.org/10.1016/S0261-2194\(00\)00059-4](https://doi.org/10.1016/S0261-2194(00)00059-4)
- [14] Markoglou, A.N., Malandrakis, A.A., Vitoratos, A.G. and Ziogas, B.N. (2006) Characterization of Laboratory Mutants of *Botrytis cinerea* Resistant to QoI Fungicides. *European Journal of Plant Pathology*, **115**, 149-162.
<https://doi.org/10.1007/s10658-006-0008-2>
- [15] Brasseur, G., Saribas, S.A. and Daldal, F. (1996) A Compilation of Mutations Located in the *Cytochrome b* subunit of the Bacterial and Mitochondrial *bc₁* Complex. *Biochimica et Biophysica Acta*, **1275**, 61-69.
[https://doi.org/10.1016/0005-2728\(96\)00051-5](https://doi.org/10.1016/0005-2728(96)00051-5)