

Clinical Characteristics of *Haemophilus influenzae* at General Hospital in the Central Region of Japan

Masaaki Minami^{1*}, Ryoko Sakakibara², Taichi Imura², Mika Watanabe², Hideo Morita², Naoto Kanemaki³, Michio Ohta⁴

¹Department of Bacteriology, Graduate School of Medical Sciences, Nagoya City University, Nagoya, Japan

²Department of Clinical Investigation, Daido Hospital, Nagoya, Japan

³Department of Gastroenterology, Daido Hospital, Nagoya, Japan

⁴School of Nursing, Sugiyama Jyogakuen University, Nagoya, Japan

Email: ^{*}minami@med.nagoya-cu.ac.jp

Received 14 April 2016; accepted 19 May 2016; published 24 May 2016

Abstract

Haemophilus influenzae is an important pathogen that caused several infection diseases, such as sinusitis, otitis media, sepsis, and meningitis. This study was conducted to find out the prevalence and antimicrobial susceptibility pattern of Haemophilus influenzae isolates at general hospital in the central region of Japan from December 2015 to January 2016. Haemophilus influenzae was identified by standard laboratory procedure. Antimicrobial susceptibility testing was performed by micro dilution assay according to CLSI recommendation. One hundred ninety-one Haemophilus influenzae were isolated, among which 95 (49.7%) were from male and 96 (50.3%) were from female. The age incidence of (0) years, (≤ 2) years, (≤ 5) years, and ($6 \leq$) years groups were 22(11.5%), 92(48.2%), 61(31.9%), and 16(8.4%), respectively. Positive samples were received mostly from the nasal discharge (177/92.7%), sputum (6/3.1%), tonsillar (6/3.1%), and pharynx (2/1.0%). Ceftriaxone was the most active antibiotics with 100% susceptible rates, followed by ciprofloxacin (99.5%) and minocycline (99%) in our study. Furthermore, we categorized four patterns: beta lactamase-negative ampicillin-sensitive strain (BLNAS), beta lactamase-negative ampicillin-resistant strain (BLNAR), beta lactamase-positive ampicillin resistant strain (BLPAR), and beta lactamase-positive amoxicillin-clavulanic acid-resistant strain (BLPACR) from those ampicillin susceptible results. The numbers of female were significant greater than those of male in BLPAR (p =0.0336). With respect to antimicrobial susceptible pattern, there was no minocycline and piperacillin resistant strain in both BLNAS and BLNAR (p < 0.0001). Haemophilus influenzae infection spreads worldwide and inadequate use of antibiotics contributes to uptake of their new antimicrobial resistance. Continuous antimicrobial surveys are need for control the emergence and spread of antimicrobial resistance to reduce the morbidity and mortality.

Keywords

Haemophilus influenzae, Susceptibility, Antimicrobial Resistance

^{*}Corresponding author.

How to cite this paper: Minami, M., Sakakibara, R., Imura, T., Watanabe, M., Morita, H., Kanemaki, N. and Ohta, M. (2016) Clinical Characteristics of *Haemophilus influenzae* at General Hospital in theCentral Region of Japan. *Journal of Biosciences and Medicines*, **4**, 18-23. <u>http://dx.doi.org/10.4236/jbm.2016.46003</u>

1. Introduction

Haemophilus influenzae is a Gram-negative bacterium which is isolated from the upper respiratory tract of certain normal humans. It is a major cause of bacterial meningitis in children aged under 5 years old and is also a significant agent of respiratory tract infections, including acute otitis media, sinusitis, pneumonia, and other serious invasive infections [1]. In elderly individuals, particularly those with underlying lung disease, Haemophilus influenzae can cause severe pneumonia. Of all six serotypes (a - f) of Haemophilus influenzae, serotype b caused the most invasive diseases prior to the introduction of the Haemophilus influenzae type b (Hib) vaccine [2] [3]. When the conjugate vaccine was introduced, the incidence of invasive Hib disease has been reduced significantly [4]. However, Hib immunization has vet to be included in routine childhood vaccinations in many countries, and Haemophilus influenzae remains one of the most important causes of community-acquired pneumonia [5]. In the past, ampicillin had been recommended as the drug of first choice for Haemophilus influenzae infection. However, the first ampicillin-resistant Haemophilus influenzae was reported in 1974 in several countries [6]. The major mechanism of this resistance was the production of beta-lactamases [7]. Non-beta-lactamase-mediated resistance to ampicillin in *Haemophilus influenzae* was first reported in the early 1980s [8]. This beta-lactamase-negative ampicillin-resistant (BLNAR) determinant was associated with the alteration of penicillin-binding proteins (PBPs) as a result of ftsI gene mutation [9] [10]. Prevalence of BLNAR among Haemophilus influenzae has been increasing in various countries in Europe and Asia [11] [12].

The present study was conducted to find out the recent clinical characteristics of *Haemophilus influenzae* isolates at general hospital in the central of Japan.

2. Materials and Methods

2.1. Strains and Clinical Data Collection

A total of 191 *Haemophilus influenzae* was obtained from various clinical specimens at Daido Hospital from December 2015 to January 2016. Daido Hospital is a 404-bed private general hospital in the central region of Japan. We used medical records appended to clinical species for the analysis of clinical feature at Daido Hospital. We considered several isolates from the same region of the same patient as one isolate per one patient for the analysis in this study. All *Haemophilus influenzae* isolates were identified by standard conventional biochemical methods or the VITEK2 system (bioMérieux, Durham, NC, USA). Our experimental design was approved by the ethics committee at Daido hospital.

2.2. Antimicrobial Susceptibility Analysis

Haemophilus influenzae isolates were examined for seven antimicrobial susceptibilities as follows; Ampicillin (ABPC), subactum/Ampicillin (SBT/ABPC), Clarithromycin (CAM), Ceftriaxone (CTX), Ciprofloxacin (CPFX), Minocycline (MINO), and Piperacillin (PIPC). Minimal inhibitory concentrations (MICs) were determined at clinical laboratory in Daido Hospital using broth micro dilution methodology with the VITEK2 system. Evaluation of susceptibilities was calculated based on Clinical Laboratory Standard Institute (CLSI) break point [13]. We defined the ampicillin-resistant *Haemophilus influenzae* as follows; beta lactamase-negative ampicillin-sensitive strain (BLNAS): the MIC of ABPC ≤ 1 , beta lactamase-negative ampicillin-resistant strain (BLNAR): the MIC of SBT/ABPC ≤ 1 , beta lactamase-positive amoxicillin-clavulanic acid-resistant strain (BLPACR): the MIC of ABPC ≥ 4 and the MIC of SBT/ABPC ≤ 2 [14].

2.3. Statistical Analysis of the Data

We conducted the statistical analysis with the chi-squared test or Fisher's exact test when appropriate. Differences were considered significant when p was <0.05.

3. Results

First of all, we screened the ampicillin susceptibility of all *Haemophilus influenzae* isolates. From those ampicillin susceptible results, we categorized four patterns: BLNAS, BLNAR, BLPAR, and BLPACR (**Table 1**).

Characteristics and description	Total number of isolates	Number of isolates				
		BLNAS	BLNAR	BLPAR	BLPACR	– <i>p</i> value
Gender						
Male	95	26	22	3	44	0.0336
Female	96	36	16	11	33	
Age						
0	22	4	4	2	12	0.3990
≤2	92	28	19	9	36	0.6162
≤5	61	21	13	3	24	0.9203
6≤	16	9	2	0	5	0.160
Speciemen type						
Nasal discharge	177	54	37	14	72	0.153
Sputum	6	4	1	0	1	0.310
Tonsillar	6	2	0	0	4	0.431
Pharynx	2	2	0	0	0	0.240
Total	191	62	38	14	77	

Table 1. Clinical characteristic of *Haemophilus influenzae* isolates.

Next, we compared the differences of clinical characteristics and antimicrobial susceptible pattern except ampicillin among ampicillin-resistant *Haemophilus influenzae* in this study. One hundred ninety-one *Haemophilus influenzae* were isolated among which 95(55.6%) were from male and 96(44.4%) were from female (**Table 1**). According to ampicillin susceptibility, the numbers of female were significant greater than those of male in BLPAR (p = 0.0336). The age incidence among 0 years age group was 22 (11.5%), ≤ 2 years age group, 92(48.2%), ≤ 5 years age group, 61(31.9%), and $6\leq$ years it was 16(8.4%) (**Table 1**). According to ampicillin susceptibility, there was no significant difference in age group. Most of the *Haemophilus influenzae* isolates were from nasal discharge (177/92.7%), followed by sputum (6/3.1%), tonsillar (6/3.1%) and pharynx (2/1.0%) (**Table 1**). According to ampicillin susceptibility, there was no significant difference in specimen group.

The results of antimicrobial susceptible patterns of *Haemophilus influenzae* isolates to various antibiotics tested in this study were shown in **Table 2**. Ceftriaxone was the most active antibiotics with 100% susceptible rates, followed by ciprofloxacin (99.5%) and minocycline (99%). According to ampicillin susceptibility, there was no minocycline and piperacillin resistant strain in both BLNAS and BLNAR.

4. Discussion

In this study, we described the characteristics of *Haemophilus influenzae* isolates from July 2014 to June 2015 at general hospital in the central region of Japan. With respect to gender group, our study showed the male to female ratio was about 0.99 and there were no significant differences among gender. We clarified *Haemophilus influenzae* with age distribution. The present study reveals the about half of *Haemophilus influenzae* were isolated from under 2 years age patients. In the analysis of specimen's type, we found that specimens where most patients with *Haemophilus influenzae* were detected were nasal discharge. In the analysis of antimicrobial susceptibility, we did not find any ceftriaxone-resistant *Haemophilus influenzae* in our study. Furthermore, in our study, ciprofloxacin, minocycline, and piperacillin were also effective antibacterial agent against *Haemophilus influenzae*, especially BLNAS and BLNAR. However, about 23% of *Hemophilus influenzae* were non-susceptible to clarithromycin. As ampicillin-resistant *Haemophilus influenzae* spreads widely, several investigators reported antimicrobial susceptible patterns of *Haemophilus influenzae* in Asian country.

	Number of isolates				
Antimicrobial agent non-susceptibility	BLNAS	BLNAR BLPAR		BLPACR	– <i>p</i> value
Clarithromycin intermediate	12	5	1	10	0.5796
Clarithromycin resistant	3	4	1	8	0.6427
Ceftriaxione resistant	0	0	0	0	N.D.
Ciprofloxacin resistant	1	0	0	0	0.5536
Minocycline resistant	0	0	1	1	0.1056
Piperacillin resistant	0	0	12	9	< 0.000

Table 2. Antimicrobial non-susceptible patterns of Haemophilus influenzae isolates.

In Japan, the prevalence of BLNAR was 63.5% and the rate of BLPAR also ranged from 6.4% to 8.7% from acute otitis media patient [15]. Another Japanese report from acute otitis media patients showed that the rate of BLNAR and BLPAR were 60.5% and 8.3% [16]. Furthermore, the prevalence of BLNAR, BLPAR, and BLPACR were 62.4, 6.1, and 4.1% in other Japanese study [17]. The rate of levofloxacin-resistant was 24.3% in another Japanese report [18]. In South Korea, the rate of BLNAR, BLPAR, and BLPACR were 40.2, 9, and 24.6 [19].

In China, the rate of beta-lactamase positive was 35.8% [20]. In another Chinese report, the rate of beta-lactamase positive was 61% and the rate of tetracycline resistant was 94.5% [21]. In Thailand, the rate of ampicillin resistant, tetracycline resistant, clarithromycin resistant, and ciprofloxacin resistant bacteria were 50, 31.9, 5.7, and 0%, respectively [22]. In Turkey, the rate of ampicillin/clavulanic acid resistant, tetracycline resistant, clarithromycin resistant bacteria were 0, 15, 4, and 0%, respectively [23]. In our case, the prevalence of BLNAR, BLPAR, and BLPACR were 19.9, 7.3, and 40.3% and the rate of beta-lactamase positive was 47.6%. Our results are similar to the result in South Korea, not in Japan. For the other antibiotic-resistance except ampicillin, our result showed that the CPFX-, MINO, and CAM-resistant were 0.5, 1, and 8.4%. The antibiotic resistant rates in our studies were relatively low compared to other country. Although *Haemophilus influenzae* spreads widely, the susceptibility patterns of *Haemophilus influenzae* in Asia have diversified by region [24]. Further analysis is needed for the clarification of epidemiology of *Haemophilus influenzae* in Asian countries.

5. Conclusion

Incidence of *Haemophilus influenzae* infection is increasing worldwide and may lead to severe invasive infection. Our investigation aims to guide medical staffs on appropriate use of antibiotics. Furthermore, our aim is not only to reduce the morbidity and mortality in the patients but also to control the emergence and spread of antimicrobial resistance among *Haemophilus influenzae*. Our results strongly emphasize the need for continuous epidemiological monitoring of antibiotic resistant *Haemophilus influenzae*.

Acknowledgements

We thank Mr. Masashi Ishihara and Ms. Miwako Fujimura for special encouragement. This study was supported by a grant-in-aid for research from the Nagoya City University, Japan.

References

- Ladhani, S., Slack, M.P., Heath, P.T., von Gottberg, A., Chandra, M., Ramsay, M.E., and European Union Invasive Bacterial Infection Surveillance Participants (2010) Invasive *Haemophilus influenzae* Disease, Europe, 1996-2006. *Emerg. Infect. Dis.*, 16, 455-463. <u>http://dx.doi.org/10.3201/eid1603.090290</u>
- [2] Watt, J.P., Wolfson, L.J., O'Brien, K.L., Henkle, E., Deloria-Knoll, M., McCall, N., et al. (2009) Burden of Disease Caused by *Haemophilus influenzae* Type b in Children Younger than 5 Years: Global Estimates. *Lancet*, **374**, 903-911. http://dx.doi.org/10.1016/S0140-6736(09)61203-4
- [3] Peltola, H. (1999) Spectrum and Burden of Severe Haemophilus influenzae Type b Diseases in Asia. Bull. World

Health Organ., 77, 878-887.

- [4] Peltola, H. (1998) Haemophilus influenzae Type b Disease and Vaccination in Europe: Lessons Learned. Pediatr. Infect. Dis. J., 17, S126-S132. <u>http://dx.doi.org/10.1097/00006454-199809001-00007</u>
- [5] Apisarnthanarak, A. and Mundy, L.M. (2005) Etiology of Community-Acquired Pneumonia. *Clin. Chest. Med.*, 26, 47-55. <u>http://dx.doi.org/10.1016/j.ccm.2004.10.016</u>
- [6] Thornsberry, C. and Kieven, L.A. (1974) Antimicrobial Susceptibility of Haemohpilus influenzae. Antimicrob. Agents Chemother., 6, 620-624. <u>http://dx.doi.org/10.1128/AAC.6.5.620</u>
- [7] Medeiros, A.A. and O'Brien, T.F. (1975) Ampicillin-Resistant Haemophilus influenzae Type b Possessing a TEM-Type Beta-Lactamase but Little Permeability Barrier to Ampicillin. Lancet, 1, 716-719. http://dx.doi.org/10.1016/S0140-6736(75)91630-X
- [8] Markowitz, S.M. (1980) Isolation of an Ampicillin-Resistant, Non-Beta-Lactamase-Producing Strain of Haemophilus influenzae. Antimicrob. Agents. Chemother., 17, 80-83. <u>http://dx.doi.org/10.1128/AAC.17.1.80</u>
- [9] Parr Jr., T.R., and Bryan, L.E. (1984) Mechanism of Resistance of an Ampicillin-Resistant, Beta-Lactamase-Negative Clinical Isolate of *Haemophilus influenzae* Type b to Beta-Lactam Antibiotics. *Antimicrob. Agents. Chemother.*, 25, 747-753. <u>http://dx.doi.org/10.1128/AAC.25.6.747</u>
- [10] Ubukata, K., Shibasaki, Y., Yamamoto, K., Chiba, N., Hasegawa, K., Takeuchi, Y., et al. (2001) Association of Amino Acid Substitutions in Penicillin-Binding Protein 3 with Beta-Lactam Resistance in Beta-Lactamase-Negative Ampicillin Resistant Haemophilus influenzae. Antimicrob. Agents. Chemother., 45, 1693-1699. http://dx.doi.org/10.1128/AAC.45.6.1693-1699.2001
- [11] Fluit, A.C., Florijn, A., Verhoef, J., and Milatovic, D. (2005) Susceptibility of European Beta-Lactamase-Positive and -Negative *Haemophilus influenzae* Isolates from the Periods 1997/1998 and 2002/2003. J. Antimicrob. Chemother., 56, 133-138. <u>http://dx.doi.org/10.1093/jac/dki167</u>
- [12] Hasegawa, K., Yamamoto, K., Chiba, N., Kobayashi, R., Nagai, K., Jacobs, M.R., et al. (2003) Diversity of Ampicillin-Resistance Genes in *Haemophilus influenzae* in Japan and the United States. *Microb. Drug Resist.*, 9, 39-46. http://dx.doi.org/10.1089/107662903764736337
- [13] Clinical and Laboratory Standards Institute (CLSI) (2014) Performance Standards for Antimicrobial Susceptibility Testing: 24th Informational Supplement. Clinical and Laboratory Standards Institute M100-S24, Wayne.
- [14] Tristram, S., Jacobs, M.R. and Appelbaum, P.C. (2007) Antimicrobial Resistance in *Haemophilus influenzae*. Clin Microbiol Rev, 20, 368-389. <u>http://dx.doi.org/10.1128/CMR.00040-06</u>
- [15] Shiro, H., Sato, Y., Toyonaga, Y., Hanaki, H. and Sunakawa, K. (2015) Nationwide Survey of the Development of Drug Resistance in the Pediatric Field in 2000-2001, 2004, 2007, 2010, and 2012: Evaluation of the Changes in Drug Sensitivity of *Haemophilus influenzae* and Patients' Background Factors. J. Infect. Chemother, 21, 247-256. <u>http://dx.doi.org/10.1016/j.jiac.2014.11.012</u>
- [16] Kakuta, R., Yano, H., Hidaka, H., Kanamori, H., Endo, S., Ichimura, S., et al. (2016) Molecular Epidemiology of Ampicillin-Resistant Haemophilus influenzae Causing acute Otitis Media in Japanese Infants and Young Children. Pediatr. Infect. Dis. J. http://dx.doi.org/10.1097/inf.000000000001066
- [17] Takakura, M., Fukuda, Y., Nomura, N., Mitsuyama, J., Yamaoka, K., Asano, Y., et al. (2012) Antibacterial Susceptibility Surveillance of *Haemophilus influenzae* Isolated from Pediatric Patients in Gifu and Aichi Prefectures (2009-2010). Jpn. J. Antibiot, 65, 305-321.
- [18] Kuo, S.C., Chen, P.C., Shiau, Y.R., Wang, H.Y., Lai, J.F., Huang, W., et al. (2014) Levofloxacin-Resistant Haemophilus influenzae, Taiwan, 2004-2010. Emerg. Infect. Dis, 20, 1386-1390. <u>http://dx.doi.org/10.3201/eid2008.140341</u>
- [19] Park, C., Kim, K.H., Shin, N.Y., Byun, J.H., Kwon, E.Y., Lee, J.W., *et al.* (2013) Genetic Diversity of the ftsI Gene in β-Lactamase-Nonproducing Ampicillin-Resistant and β-Lactamase-Producing Amoxicillin-/Clavulanic Acid-Resistant Nasopharyngeal *Haemophilus influenzae* Strains Isolated from Children in South Korea. *Microb Drug Resist*, **19**, 224-230. http://dx.doi.org/10.1089/mdr.2012.0116
- [20] Luo, C., Xia, Y., Liu, Q., Chu, L., Fu, X., Jing, C., et al. (2012) Antibiotic Resistance and Molecular Epidemiology of the Beta-Lactamase-Producing Haemophilus influenzae Isolated in Chongqing, China. APMIS, 120, 926-934. http://dx.doi.org/10.1111/j.1600-0463.2012.02921.x
- [21] Zhu, H., Wang, A., Tong, J., Yuan, L., Gao, W., Shi, W., et al. (2015) Nasopharyngeal Carriage and Antimicrobial Susceptibility of *Haemophilus influenzae* among Children Younger than 5 Years of Age in Beijing, China. BMC Microbiol, 15, 6. <u>http://dx.doi.org/10.1186/s12866-015-0350-7</u>
- [22] Lulitanond, A., Chanawong, A., Pienthaweechai, K., Sribenjalux, P., Tavichakorntrakool, R., Wilailuckana, C., *et al.* (2012) Prevalence of β-Lactamase-Negative Ampicillin-Resistant *Haemophilus influenzae* Isolated from Patients of a Teaching Hospital in Thailand. *Jpn. J. Infect. Dis*, **65**, 122-125.

- [23] Kuvat, N., Nazik, H., Berkiten, R. and Öngen, B. (2015) TEM-1 and ROB-1 Presence and Antimicrobial Resistance in Haemophilus influenzae Strains, Istanbul, Turkey. Southeast Asian J. Trop. Med. Pub. Health, 46, 254-261.
- [24] Peto, L., Nadjm, B., Horby, P., Ngan, T.T. and van Doorn, R. (2014) The Bacterial Aetiology of Adult Community-Acquired Pneumonia in Asia: A Systematic Review. *Trans. R. Soc. Trop. Med. Hyg*, **108**, 326-337. <u>http://dx.doi.org/10.1093/trstmh/tru058</u>