

# Optimization of Parameters for the Production of Lipase from *Pseudomonas* sp. BUP6 by Solid State Fermentation

Panichikkal Abdul Faisal, Erandapurthukadumana Sreedharan Hareesh, Prakasan Priji, Kizhakkepowathial Nair Unni, Sreedharan Sajith, Sasidharan Sreedevi, Moolakkariyil Sarath Josh, Sailas Benjamin\*

Enzyme Technology Laboratory, Biotechnology Division, Department of Botany, University of Calicut, Kerala, India

Email: \*[benjamin@uoc.ac.in](mailto:benjamin@uoc.ac.in), \*[sailasben@yahoo.co.in](mailto:sailasben@yahoo.co.in)

Received 17 September 2014; revised 23 October 2014; accepted 6 November 2014

Academic Editor: Sagar Naskar, Bengal College of Pharmaceutical Sciences and Research, India

Copyright © 2014 by authors and Scientific Research Publishing Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

---

## Abstract

Solid-state fermentation (SSF) holds tremendous potentials for the production of industrially significant enzymes. The present study describes the production of lipase by a novel rumen bacterium, *Pseudomonas* sp. strain BUP6 on agro-industrial residues. *Pseudomonas* sp. strain BUP6 showed higher lipase production when grown in Basal salt medium (BSM) supplemented with oil cakes. Initially, five different oil cakes (obtained after extracting oil from coconut, groundnut, cotton seed, gingelly or soybean) were screened to find out the most suitable substrate-cum-inducer for the production of lipase. Among them, groundnut cake supported the maximum production of lipase (107.44 U/gds). Box-Behnken Design (BBD), followed by response surface methodology (RSM) was employed to optimize the culture parameters for maximizing the production of lipase. Using the software Minitab 14, four different parameters like temperature, pH, moisture content and incubation time were selected for the statistical optimization, which resulted in 0.7 fold increase (*i.e.*, 180.75 U/gds) in production of lipase under the optimum culture conditions (temperature 28°C, pH 5.9, moisture 33% and incubation 2 d). Thus, this study signifies the importance of SSF for the production of industrially-significant lipase using agro-industrial residues as solid support.

---

\*Corresponding author.

## Keywords

**Lipase, Solid-State Fermentation, Basal Salt Medium, Oil Cakes, Response Surface Methodology**

## 1. Introduction

Lipases (triacylglycerol acyl hydrolase EC 3.1.1.3) are a cluster of enzymes catalyzing the hydrolysis of triacylglycerol to form glycerol and free fatty acids. In contrast to esterase, lipases are activated only at an oil-water interface. Many lipases are catalyzing a number of useful reactions including hydrolysis, esterification, transesterification, acidolysis, alcoholysis, and synthesis of peptides [1]. The extensive utilization of lipase has wide range of applications in synthesis of detergents, biosurfactants; organic, oleo-chemical, leather, cosmetic, perfume, dairy and agrochemical industries; environmental management; biosensors, etc.

Microorganisms with potentials for producing lipases can be found in different habitats, including wastes of vegetable oils and dairy industries, soils contaminated with oils, seeds, and deteriorated food [2]. Since, lipase is among the most widely used class of enzymes in biotechnological applications and organic chemistry [3], utilization of agro-industrial wastes as alternative sources of substrates would help solving pollution problems. The nature of the substrate is the most important factor affecting fermentative processes. The choice of the substrate depends upon several factors; mainly related to cost and availability.

Lipases are produced by several microorganisms, *viz.*, bacteria, fungi, yeast, actinomycetes, archaea, eucarya, *etc.* Microbial genera involved in the commercial production of lipases include: *Candida*, *Mucor*, *Rhizopus*, *Aspergillus*, *Penicillium*, *Geotrichum*, *Rhizomucor*, *Bacillus*, *Pseudomonas* and *Staphylococcus* [4]. Since they perform at wide range of pH and temperature, and they can be produced by solid-state fermentation (SSF) as well as submerged fermentation (SmF), microbial lipases are considered as highly robust. By dint of low production cost, greater stability, more simplicity and wider availability than SmF, SSF is considered as a novel strategy with higher physiological significance and industrial potentials [5]. Lipases are inducible enzymes; hence, different natural agro-industrial residues such as brans of wheat and rice, sugar cane bagasse, wastes from vegetable oil-refining, etc. can effectively be used as inducers or substrates for lipase production employing SSF strategy [4]-[8]. Deoiled kernels (*i.e.*, cakes) from coconut, olive, gingelly, cotton, *Jatropha*, *etc.*, obtained after extracting oil have been utilized as solid substrate for the fermentative production of lipases and other industrial enzymes. This is because residual oil and other ingredients contained in it serve as inducers for lipase production [15].

Optimization of environmental parameters as well as the culture parameters may enhance the production of value-added products of commercial interest to many folds. Usually, “one parameter at a time” strategy was used for the optimization; but, this method is time consuming and hectic, and that the utilization of statistical tools makes the process easier. Response surface methodology (RSM) is such a kind of statistical tool being applied widely for the optimization, modeling and analysis of problems related to the production of biomolecules [6]. However, the use of different substrates as well as cultivation strategies for the production of lipase still remains an emerging area of research. In this context, the present study focused on 1) screening of different oil cakes as solid substrate for the production of lipase by a novel rumen bacterium *Pseudomonas* sp. strain BUP6, and 2) statistical optimization of parameters for lipase production employing RSM technique.

## 2. Materials and Methods

### 2.1. Materials

Analytical and bacteriological-grade chemicals from Himedia (India) and Merck India Ltd. were used for the present study.

### 2.2. Culture Medium

The pure bacterial culture *Pseudomonas* sp. strain BUP6 (Genbank Accession No. KF 550910), isolated from the rumen of Malabari goat, was used for this study [9]. The bacterium was cultured on basal salt medium

(BSM), supplemented with 0.3% of vegetable oil, and incubated at 37°C for 24 h. The stock culture was maintained on BSM agar slants, which was sub-cultured in an interval of 2 weeks. The BSM contained the following ingredients (%): 0.5 NH<sub>4</sub>NO<sub>3</sub>; 0.4(NH<sub>4</sub>)<sub>2</sub>SO<sub>4</sub>; 0.3 yeast extract; 0.2 K<sub>2</sub>HPO<sub>4</sub>; 0.2 NaCl; 0.01 MgSO<sub>4</sub>. 7H<sub>2</sub>O and 0.01 CaCl<sub>2</sub> in double distilled water.

### 2.3. SSF Using Agro-Industrial Residues

Deoiled cakes of groundnut (GNC), gingelly (GOC), coconut (COC), soybean (SOC), and cotton seed (CSC) procured from the local market were used as solid substrate-cum-inducer for the production of lipase by SSF. Fermentation was carried out in 100 mL of conical flasks. Five grams of substrate (oil cake) were transferred into 100 mL conical flasks, and then moistened with 10 mL of BSM (50%). In order to check the effect of pH on lipase production, the initial pH of the medium was set at 5, 7 and 9. All preparations in the flask were autoclaved at 121°C for 15 min, and inoculated with 0.1 mL of inoculum under aseptic condition. Production of lipase was assayed at regular intervals of 24 h for 5 days.

### 2.4. Extraction of Crude Enzyme

After incubation, the fermented matter in the whole flask was used for lipase assay at regular intervals of 24 h; for the extraction of crude lipase, 10 mL of 0.1 M Tris-HCl buffer was added to the flask and stirred for 10 min. Then the contents of the flask were centrifuged at 9400× g for 15 min at 4°C, the supernatant was used as crude lipase for the activity assay.

### 2.5. Lipase Activity Assay

Production of lipase was qualitatively assayed by the method of by Priji *et al.* [9]. *Para*-nitro phenyl palmitate (*p*-NPP) was used as substrate for lipase assay. The assay mixture containing 1.8 ml of 0.1 M Tris-HCl buffer, 0.15 M NaCl and 0.5% Triton X-100 was pre-incubated with 200 µl of cell-free culture supernatant at 37°C for 10 min. Subsequently, 20 µl of substrate (50 mM *p*-NPP in acetonitrile) was added to the reaction mixture and incubated at 37°C for 30 min. The quantity of *p*-NP liberated was measured spectrophotometrically at λ<sub>405</sub>. One unit of lipase corresponds to 1 µmol of *p*-NP liberated per minute under the standard assay conditions. The lipase activity was calculated as following Equation (1).

$$\text{Lipase activity (U/gds)} = \frac{\Delta E \times V_f \times V_s}{\Delta t \times \epsilon \times \text{gds}} \quad (1)$$

where  $\Delta E$ —absorbance at 405 nm;  $V_f$ —final volume;  $V_s$ —volume of lipase used;  $\Delta t$ —time of hydrolysis;  $\epsilon$ —extinction co-efficient (0.017); gds—dry weight in grams.

## 3. Statistical Optimization of Lipase Production

Box-Behnken Design (BBD), followed by RSM was employed to develop a mathematical correlation between different independent variables such as temperature, pH, moisture, and incubation time on the production of lipase. The software Minitab version 14 (Minitab USA) was used to generate data and to analyze the experimental design of BBD and RSM.

### 3.1. Box-Behnken Design (BBD)

The culture parameters like temperature (28°C to 40°C), pH (5 to 9), moisture (20% to 60%), and incubation time (1 to 5 d) were selected for BBD. BBD at three levels (+1, 0, and -1)—designated as high, medium and low—was used for this study. Based on this design, a set of 27 experimental trials were suggested by the software. All the trials were carried out in duplicates and the results were analyzed by fitting to a second-order polynomial Equation (2). Each experimental trial was set up and lipase was harvested at proper intervals to measure lipase activity as per the design.

$$Y = \beta_0 + \sum_{i=1}^k \beta_i X_i + \sum_{i=1}^k \beta_{ii} X_i X_i + \sum_{i=1}^k \sum_{j=1}^k \beta_{ij} X_i X_j, \quad i < j \quad (2)$$

where  $Y$  represents the response variable;  $\beta_0$  is the interception coefficient;  $\beta_i$  is the coefficient of the linear effect;  $\beta_{ii}$  is the coefficient of quadratic effect;  $\beta_{ij}$  is the coefficient of interaction effect when  $i < j$ ; and  $k$  is the numbers of involved variables.

### 3.2. Validation Experiment

To check the validity of quadratic model, 4 experiments as predicted by point prediction software Minitab 14 were performed. Lipase activity was estimated and compared with predicted values.

## 4. Results

### 4.1. Effects of Different Substrates on Lipase Activity

Five different oil seed cakes (GNC, GOC, COC, SOC and CSC) were used as solid substrate-cum-inducer for the production of lipase by *Pseudomonas* sp. strain BUP6. GNC, GOC and COC supported the maximum production of lipase (107.44 U/gds, 86.21 U/gds and 9.97 U/gds, respectively) on Day 3 of incubation, subsequently the lipase activity was decreased sharply; while SOC as well as CSC supported the maximum production of lipase after 24 h of incubation, but in lesser amounts (95.74 U/gds and 5.45 U/gds, respectively) (Figure 1). Of them, GOC supported the highest lipase production (107.44 U/gds) (Figure 1). Effects of SOC (95.74 U/gds) and GOC (86.21 U/gds) were comparable to that of GNC (107.44 U/gds), but the other two oil cakes (COC and CSC) showed much lesser activity, *i.e.*, 9.97 and 5.45 U/gds (Table 1).

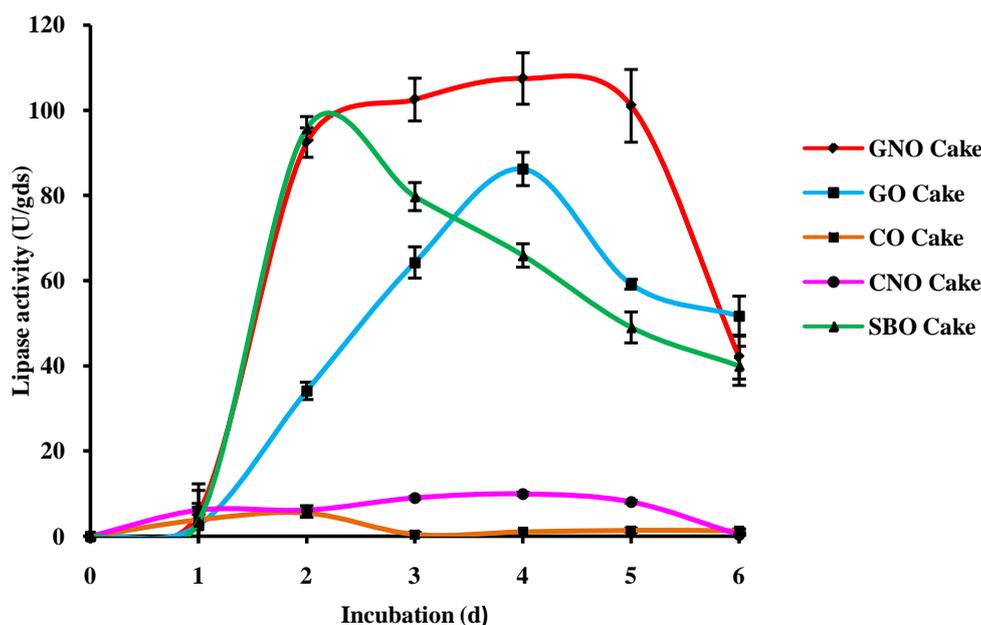


Figure 1. Lipase production by *Pseudomonas* sp. strain BUP6 on different oil cakes.

Table 1. Maximum lipase activity in different substrates.

Substrates	Maximum lipase activity (U/gds)	Incubation (d)
Groundnut oil cake	107.44	3
Soya bean oil cake	95.74	1
Gingelly oil cake	86.21	3
Coconut oil cake	9.97	3
Cotton cake	5.45	1

## 4.2. Statistical Optimization of Lipase Production

GNC which supported the maximum production of lipase was selected as substrate for the optimization studies. Four parameters (temperature, pH, moisture, and incubation) were considered for BBD analysis, followed by RSM to find out the optimum conditions for maximizing the production of lipase. A set of 27 experiments was conducted according to BBD, and the results showed that the predicted and experimental values for lipase activities did not show significant difference (Table 2), i.e., the  $R^2$  value was 0.95-close to unity. A second order polynomial equation was fitted to the experimental lipase activity, which resulted in the following regression Equation (3).

$$Y = 781.660 - 41.872X_1 + 7.275X_2 + 4.549X_3 + 10.584X_4 + 0.555X_1^2 - 6.001X_2^2 - 0.186X_3^2 - 9.213X_4^2 + 0.592X_1X_2 - 0.010X_1X_3 - 0.480X_1X_4 + 1.230X_2X_3 + 4.056X_2X_4 + 0.454X_3X_4 \quad \text{Equation (3)}$$

where  $X_1$ —pH,  $X_2$ —Temperature,  $X_3$ —Moisture,  $X_4$ —Incubation.

The results were analyzed by standard analysis of ANOVA (Table 3). Based on these results, the model was utilized to generate response surfaces for the analysis of the variable effect on the production of lipase. The response surface plots obtained using Equation (3) is depicted in Figure 2.

**Table 2.** Experimental trials according to BBD model for the optimization of lipase production.

Run order	Temperature (°C)	pH	Moisture (%)	Incubation (d)	Observed (U/gds)	Predicted (U/gds)
1	28	5	40	3	137.86	155.597
2	40	5	40	3	112.35	119.362
3	28	9	40	3	155.98	160.407
4	40	9	40	3	158.89	152.592
5	34	7	20	1	97.68	101.597
6	34	7	60	1	26.37	35.46
7	34	7	20	5	5.08	7.429
8	34	7	60	5	6.43	13.952
9	28	7	20	3	129.66	121.105
10	40	7	20	3	98.97	101.595
11	28	7	60	3	86.95	93.814
12	40	7	60	3	51.23	69.274
13	34	5	40	1	133.52	125.784
14	34	9	40	1	111.3	112.359
15	34	5	40	5	27.07	35.5
16	34	9	40	5	69.74	86.965
17	28	7	40	1	171.25	168.312
18	40	7	40	1	161.2	157.807
19	28	7	40	5	139.53	121.994
20	40	7	40	5	106.44	88.449
21	34	5	20	3	109.47	107.044
22	34	9	20	3	25.59	27.679
23	34	5	60	3	1.87	-21.148
24	34	9	60	3	114.76	96.258
25	34	7	40	3	155.76	151.007
26	34	7	40	3	158	151.007
27	34	7	40	3	139.26	151.007

**Table 3.** Analysis of variance for the second-order polynomial model for optimization of lipase production [Degree of freedom (DF), Sequential sum of square (Seq SS), Adjacent sum of square (Adj SS), Adjacent mean square (Adj MS), Text of statistics (F), Probability (P)].

Source	DF	Seq SS	Adj SS	Adj MS	F	P
Regression	4	72999.4	72999.4	5214.2	19.51	0.000
Linear	4	15241.7	4271.3	1067.8	3.99	0.028
Square	4	45364.5	45364.5	11341.1	42.43	0.000
Interaction	6	12393.1	12393.1	2065.5	7.73	0.001
Residual Error	12	3207.8	3207.8	267.3		
Lack-of-Fit	10	2998.3	2998.3	299.8	2.86	0.287
Pure Error	2	209.5	209.5	104.7		
Total	26	76207.2				

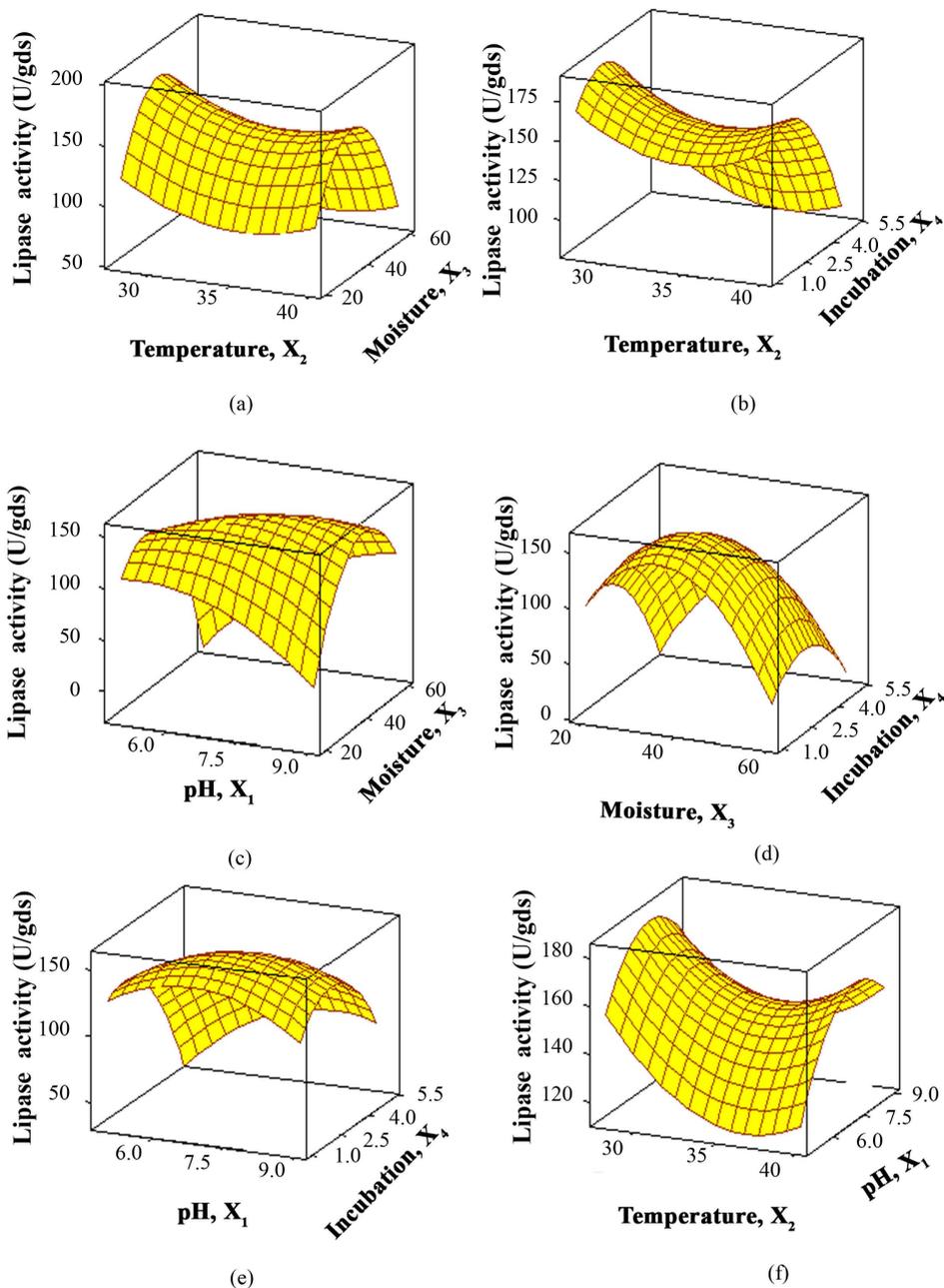
### 4.3. Validation of Lipase Production

Four random experimental conditions were evaluated for the validation of the model. In all these instances, model prediction was in good agreement with the experimental data (considering the experimental error), and correlation coefficient was found to be 0.96 (Table 4).

Correlation coefficient was close to 1.0, suggesting the significance of the model. The optimum production of lipase was found to be 180.75 U/gds (at 28°C, pH 5.9, moisture 33%, and incubation period 2 d). Thus, the statistical optimization resulted in 0.7 fold of lipase activity over the unoptimized condition.

## 5. Discussion

Research on lipase progressed very rapidly during the past few decades, giving much emphasis on the exploitation and recycling of agro-industrial residues. The present study proved that *Psuedomonas* sp. strain BUP6 is an efficient producer of lipase on solid medium (oil cakes). Nowadays, SSF strategy is increasingly employed as a method for the production of lipase on different lipid-bound waste materials, because of the several advantages of SSF such as better yield and easy to control. As a part of this study, different natural substrate such as COC, GNC, SOC, GOC, and CSC were screened to spot out the best solid medium for lipase production. Of them, GNC supported the highest production of lipase (107.44 U/gds at 72 h). The cultivation period varies with the microorganism, *i.e.*, fast growing bacteria were found to secrete lipase within 24 h [10] [11]. Bacteria normally grow in a complex nutrient medium containing carbon (oil, sugars, mixed carbon sources), nitrogen and phosphorous sources and mineral salts. Other significant factors influencing lipase production include: initial pH, growth temperature, incubation period and moisture percentage (water activity). Temperature of the substrate during SSF critically affects the growth of microorganisms, and product formation [5]. Lipase yield by *Pseudomonas* sp. BUP6 appeared to be dependent on moisture content. Results showed that lipase production (expressed as enzyme activity) gradually increased from 20% to 60% moisture content and reached its maximum at 60%, and found that the moisture content was directly proportional to production of lipase; but high moisture content led to more contamination (may be due to poor aeration). The maximum lipase activity (114.75 U/gds) was obtained at 60% moisture content. Like moisture content, incubation time was also an important parameter that influenced the production of lipase. In the present study, 48 h of incubation was found to be optimum for lipase production. Incubation periods ranging from few hours to several days have been found to be best suited for the maximum lipase production by bacteria. For instance, an incubation period of 12 h was found optimum for the lipase production by *Acinetobacter calcoaceticus* and *Bacillus* sp. RSJ1 [10] and 16 h for *B. thermocatenulatus* [11]; while, in the case of *Pseudomonas* sp., *P. fragia* and *P. fluorescens* BW 96CC, the maximum lipase activity was obtained at 72 h and 96 h of incubation, respectively [12] [13]. Employing *Candida rugosa*, Benjamin and Pandey [1] reported the use of mixed-solid substrate containing wheat bran and coconut oil cake for lipase production. Fermentation was carried out for 72 h, and the maximum lipase yield was 118.2 U/g. Similarly, in the present study, the maximum activity of lipase obtained (107.44 U/gds) was at 72 h of incubation using GNC as substrate. It seems that the residual oil in the cake acted as both inducer and additional nutrient.



**Figure 2.** Response surface plots (3D) showing the effects of different parameters ( $X_1$ : pH;  $X_2$ : temperature, °C;  $X_3$ : moisture, %; and  $X_4$ : incubation time, d) on production of lipase by *Pseudomonas* sp. strain BUP6.

**Table 4.** Experimental trails for the validation of the predicted model.

Run Order	Temperature (°C)	pH	Moisture (%)	Incubation (d)	Observed (U/gds)	Predicted (U/gds)
1	28	5.9	33	2	180.75	187.92
2	34	7	50	2	135.15	125.92
3	34	6	33	2	147.66	157.31
4	28	5.9	50	3	141.85	134.07

Therefore, at the later stage of fermentation, the lipase activity was decreased; this might be an indication of the depletion of nutrient in the medium, *i.e.*, lipase production is dependent on the nutrient status.

Few studies reported that oil cakes as the best solid substrate-cum-inducer for the production of lipase by SSF [14] [15]. D'Annibale *et al.* [16] reported that olive mill waste water as a growth medium for lipase production, which showed the highest lipase activity of 9.23 U/ml. Brozzoli *et al.* [17] studied the lipase production in bench-top reactor using the olive mill waste water medium and obtained the maximum production as 20.4 U/ml. Salihi *et al.* [18] used the statistical optimization of nutrient components to enhance lipase production by *C. cylindracea*, and the maximum activity was 20.26 U/ml. Vishnupriya *et al.* [19] assessed the lipase production by *Sterptomyces griseus*, and found the maximum activity as 51.9 U/ml. Compared to all these studies, present study reports higher lipase activity (180.75 U/gds) at optimized condition.

Various physico-chemical parameters evaluated for the maximum production of lipase. Four parameters like temperature, pH, moisture, and incubation time for the lipase production were statistically optimized. Optimization is a complex process, which can be performed in two different ways: conventional and modern methods. The conventional method of optimization defined as the one-at-time strategy, but modern multivariate RSM technique enables optimization of more than one parameter at a time [6], which can be performed for assessing the relationship between environmental and cultural parameters, so as to produce 3D contour and surface plots. This is more effective than conventional methodology, *i.e.*, effective, easier, faster and more economical [6]. Groundnut cake which supported the maximum lipase production was selected for the optimization process. Under optimized conditions (28°C, pH 5.9, moisture 33%, and incubation 2 d), the maximum production of lipase was 180.75 U/gds on GNC, which was 0.7 fold higher than that of unoptimized conditions.  $R^2$  value (0.95) represents the good fixity of experiments with predicted values.

## 6. Conclusion

Briefly, this study investigated the lipase activities of *Pseudomonas* sp. BUP6 on agro-industrial residues as substrate, which showed that groundnut oil cake was the best for enhancing lipase production on solid medium. By statistically optimizing the culture parameters, the lipase production was further enhanced. Thus, this study focuses on the need for the exploitation of agro-industrial residues for the production of industrially-significant and human-friendly biomolecules; moreover, its low cost increases the industrial potentials.

## Acknowledgements

The financial assistance (Grant No. 026/SRSL/2012/CSTE) from Kerala State Council for Science, Technology and Environment (KSCSTE), Government of Kerala, is gratefully acknowledged.

## References

- [1] Benjamin, S. and Pandey, A. (1998) Mixed-Solid Substrate Fermentation—A Novel Process for Enhanced Lipase Production by *Candida rugosa*. *Acta Biotechnologica*, **18**, 315-324. <http://dx.doi.org/10.1002/abio.370180405>
- [2] Sharma, R., Chisti, Y. and Banerjee, U.C. (2001) Production, Purification, Characterization, and Applications of Lipases. *Biotechnology Advances*, **19**, 627-662. [http://dx.doi.org/10.1016/S0734-9750\(01\)00086-6](http://dx.doi.org/10.1016/S0734-9750(01)00086-6)
- [3] Gupta, R., Gupta, N. and Rathi, P. (2004) Bacterial Lipases: An Overview of Production, Purification and Biochemical Properties. *Applied Microbiology and Biotechnology*, **64**, 763-781. <http://dx.doi.org/10.1007/s00253-004-1568-8>
- [4] Aravindan, R., Anbumathi, P. and Viruthagiri, T. (2007) Lipase Applications in Food Industry. *Indian Journal of Biotechnology*, **6**, 141-158.
- [5] Pandey, A., Selvakumar, P., Soccol, C.R. and Nigam, P. (1999) Solid State Fermentation for the Production of Industrial Enzymes. *Current Science*, **77**, 149-162.
- [6] John, R.P., Sukumaran, R.K., Nampoothiri, K.M. and Pandey, A. (2007) Statistical Optimization of Simultaneous Saccharification and l (+)-Lactic Acid Fermentation from Cassava Bagasse Using Mixed Culture of Lactobacilli by Response Surface Methodology. *Biochemical Engineering Journal*, **36**, 262-267. <http://dx.doi.org/10.1016/j.bej.2007.02.028>
- [7] Santis-Navarro, A., Gea, T., Barrena, R. and Sánchez, A. (2011) Production of Lipases by Solid State Fermentation Using Vegetable Oil-Refining Wastes. *Bioresource Technology*, **102**, 10080-10084. <http://dx.doi.org/10.1016/j.biortech.2011.08.062>

- [8] Joshi, C. and Khare, S.K. (2013) Purification and Characterization of *Pseudomonas aeruginosa* Lipase Produced by SSF of Deoiled Jatropha Seed Cake. *Biocatalysis and Agricultural Biotechnology*, **2**, 32-37. <http://dx.doi.org/10.1016/j.bcab.2012.08.006>
- [9] Priji, P., Unni, K.N., Sajith, S., Binod, P. and Benjamin, S. (2014) Production, Optimization and Partial Purification of Lipase from *Pseudomonas* sp. Strain BUP6, a Novel Rumen Bacterium Characterized from Malabari Goat. *Biotechnology and Applied Biochemistry*. <http://dx.doi.org/10.1002/bab.1237>
- [10] Sharma, R., Soni, S.K., Vohra, R.M., Jolly, R.S., Gupta, L.K. and Gupta, J.K. (2002b) Production of Extracellular Alkaline Lipase from a *Bacillus* sp. RSJ1 and Its Application in Ester Hydrolysis. *Indian Journal of Microbiology*, **42**, 49-54.
- [11] Schmidt-Dannert, C., Rua, M.L., Rua, M.L. and Schmid, R.D. (1997) Two Novel Lipases from the Thermophile *Bacillus thermocatenulatus*: Screening, Purification, Cloning, Over Expression and Properties. *Methods in Enzymology*, **284**, 194-219. [http://dx.doi.org/10.1016/S0076-6879\(97\)84013-X](http://dx.doi.org/10.1016/S0076-6879(97)84013-X)
- [12] Pabai, F., Kermasha, S. and Morin, A. (1996) Use of Continuous Culture to Screen for Lipase-Producing Microorganisms and Interesterification of Butterfat by Lipase Isolates. *Canadian Journal of Microbiology*, **42**, 446-452. <http://dx.doi.org/10.1139/m96-061>
- [13] Dong, H., Gao, S., Han, S. and Cao, S. (1999) Purification and Characterization of a *Pseudomonas* sp. Lipase and Its Properties in Non-Aqueous Media. *Applied Microbiology and Biotechnology*, **30**, 251-256.
- [14] Ramachandran, S.S., Singh, S.K., Larroche, C., Soccol, C.R. and Pandey, A. (2007) Oil Cakes and Their Biotechnological Applications—A Review. *Bioresource Technology*, **98**, 2000-2009. <http://dx.doi.org/10.1016/j.biortech.2006.08.002>
- [15] Singhania, R.R., Soccol, C.R. and Pandey, A. (2008) Application of Tropical Agro-Industrial Residues as Substrate for Solid-State Fermentation Processes. In: Pandey, A., Soccol, C.R. and Larroche, C., Eds., *Current Development in Solid-State Fermentation*, Springer, New York, 412-442.
- [16] D'Annibale, A., Sermanni, G.G., Federici, F. and Petruccioli, M. (2006) Olive-Oil Waste Waters: A Promising Substrate for Microbial Lipase Production. *Bioresource Technology*, **97**, 1828-1833. <http://dx.doi.org/10.1016/j.biortech.2005.09.001>
- [17] Brozzoli, V., Crognale, S., Sampedro, I., Federici, F., D'Annibale, A. and Petruccioli, M. (2009) Assessment of Olive-Mill Wastewater as a Growth Medium for Lipase Production by *Candida cylindracea* in Bench-Top Reactor. *Bioresource Technology*, **100**, 3395-3402. <http://dx.doi.org/10.1016/j.biortech.2009.02.022>
- [18] Salihu, A., Alam, M.Z., AbdulKarim, M.I. and Salleh, H.M. (2011) Optimization of Lipase Production by *Candida cylindracea* in Palm Oil Mill Effluent Based Medium Using Statistical Experimental Design. *Journal of Molecular Catalysis B: Enzymatic*, **69**, 66-73. <http://dx.doi.org/10.1016/j.molcatb.2010.12.012>
- [19] Vishnupriya, B., Sundaramoorthi, C., Kalaivani, M. and Selvam, K. (2010) Production of Lipase from *Streptomyces griseus* and Evaluation of Bioparameters. *International Journal of ChemTech Research*, **2**, 1380-1383.

Scientific Research Publishing (SCIRP) is one of the largest Open Access journal publishers. It is currently publishing more than 200 open access, online, peer-reviewed journals covering a wide range of academic disciplines. SCIRP serves the worldwide academic communities and contributes to the progress and application of science with its publication.

Other selected journals from SCIRP are listed as below. Submit your manuscript to us via either [submit@scirp.org](mailto:submit@scirp.org) or [Online Submission Portal](#).

