

Promising Actions of Certain Medicinal Plants of Bangladesh against Diabetes and Oxidative Stress: A Review

Arghya Prosun Sarkar^{1*} , Sozoni Khatun¹, Tonima Enam¹, Mst. Israt Jahan¹, Arjyabrata Sarker², Nilay Saha³, Md. Ariful Islam³, Shantanu Saha Sani⁴

¹Department of Pharmacy, Faculty of Biological Sciences, Islamic University, Kushtia, Bangladesh

²Senior Executive, Strategic Brand Management Department, Beximco Pharmaceuticals Ltd., Dhaka, Bangladesh

³Department of Pharmacy, Faculty of Biological Sciences, Jahangirnagar University, Savar, Dhaka, Bangladesh

⁴Product Management Department, Beacon Pharmaceuticals Ltd., Dhaka, Bangladesh

Email: *arghyapharm7@gmail.com

How to cite this paper: Sarkar, A.P., Khatun, S., Enam, T., Jahan, M.I., Sarker, A., Saha, N., Islam, Md.A. and Sani, S.S. (2022) Promising Actions of Certain Medicinal Plants of Bangladesh against Diabetes and Oxidative Stress: A Review. *Pharmacology & Pharmacy*, 13, 393-416.

<https://doi.org/10.4236/pp.2022.1310030>

Received: August 22, 2022

Accepted: October 25, 2022

Published: October 28, 2022

Copyright © 2022 by author(s) and Scientific Research Publishing Inc.

This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

Diabetes is a metabolic illness defined by hyperglycemia that affects 10% of the world's population. Diabetic complications such as blindness, kidney failure, and heart failure can develop if left untreated and are made worse by oxidative stress. Oxidative stress contributes to the rise of diabetic complications, particularly type-2 diabetes. In the blood vessels of diabetic individuals, it causes endothelial dysfunction. Diabetes is one of the leading causes of death worldwide. Nowadays, it is alarming that the number of diabetic patients is increasing dramatically. There are plenty of anti-diabetic drugs available on the market, but they possess several adverse effects and do not completely cure diabetes. It has now become a financial burden on patients, their families, and society as well. Medicinal plants have gained popularity in developed and developing countries over the last two decades because of their vast natural sources and lack of harmful effects compared to modern allopathic medications. According to the World Health Organization, traditional medicines, which are mostly manufactured from plants, are still used by 80% of the population in developing countries for the management and curing of diseases. This review includes 81 Bangladeshi medicinal plants from 51 different families that can be used to treat diabetes and oxidative stress. Among them, *Psidium guajava* (L.), *Aloe vera*, *Catharanthus roseus*, *Allium sativum*, *Annona squamosa*, *Cinnamom zeylanicum*, *Amaranthus esculentus*, *Eugenia jambolana*, *Azadirachta indica*, *Moringa oleifera*, *Spondias pinnata*, *Coccinia grandis* (L.), *Momordica charantia* L., *Heretiera fomes*, *Trigonella foenum-graecum* were most potent. The fundamental purpose of our study is to find out and highlight certain medicinal plants in Bangladesh that have an-

ti-diabetic and antioxidant capabilities so that the researchers can develop newer anti-diabetic medications with minimal side effects to treat metabolic dysfunction, diabetic complications, and oxidative stress more effectively.

Keywords

Diabetes, Anti-Oxidant, Anti-Diabetic, Oxidative Stress, Medicinal Plants

1. Introduction

Diabetes is becoming the most worrying threat to public health in developed countries as well as in developing countries like Bangladesh [1]. For the very first time, Egyptians introduced the term “Diabetes”, and the Greek Physician Aretaeus cut out the word diabetes mellitus. The word diabetes mellitus comes from Greek word diabetes which means siphon “to pass through” and mellitus comes from a Latin word meaning “honey” or “sweet” [2]. Diabetes mellitus (DM) may be described as a metabolic disorder or a group of metabolic diseases manifested by high blood glucose levels, and oxidative stress either because of insufficient insulin production, insulin action, or both. It disrupts the metabolism of proteins, carbohydrates, and fats, leading to chronic hyperglycemia and abnormal lipid profile resulting in several diabetic complications such as retinopathy, neuropathy, ketosis, polyuria, polydipsia, and cardiovascular complexity as well [3] [4] [5]. Approximately 10% of the world’s population suffers from diabetes. The risk factors that cause diabetes are diet, obesity, sedentary lifestyle, insulin resistance, high family aggregation, and so on [6] [7]. Depending upon characteristics and cause, there are mainly three kinds of diabetes for instance type-1 diabetes, type-2 diabetes, and gestational diabetes [8]. Type-1 diabetes is insulin-dependent diabetes which is also named adolescent diabetes [5] [7]. Approximately 5% - 10% of the people are affected by this type of diabetes. It is caused by cellular-mediated autoimmune destruction of pancreatic beta cells. The most conventional form of diabetes is type-2 diabetes [8]. Nearly 90% to 95% of the people are affected with type-2 diabetes which is commonly known as adult diabetes [5]. It does not depend on insulin and is therefore recognized as non-insulin-dependent diabetes. It is more common in people over 40 years of age [9] and may be occurred due to a lack of insulin action and insulin secretion [8]. The most common cause of this type of diabetes is obesity which causes insulin resistance [5]. Gestational diabetes is seen during pregnancy and is caused by carbohydrate intolerance which leads to hyperglycemia, although the reason for this form is not fully understood [8]. According to International Diabetes Federation (IDF), the number of patients with diabetes mellitus was 366 million in 2011 and that will be 522 million by 2030 [10], where 4.6 million people die every year owing to diabetes [1]. But according to the current survey report, there are 451 million people aged between 18 - 99 years who are suffering from DM worldwide in 2017 and this number will rise to 693 million by 2045 [3].

Diabetes is therefore considered one of the country's top five most serious diseases [11]. Along with cancer, cardiovascular disease, and cerebrovascular illness, diabetes is indeed the world's third-leading cause of death [11]. As a result, DM is gradually turning over the world's most prolific silent killer [8]. Cause it is said to be irremediable [12].

However, there are lots of synthetic anti-diabetic drugs, for instance, biguanides, glucosidase inhibitors, thiazolidinediones, and metformin commercially available to compress hyperglycemia in diabetes mellitus. Although these drugs reduce hyperglycemia, there are several side effects associated with these drugs like liver problems, diarrhea and lactic acidosis. Afterward, it is inappropriate to use them during pregnancy and the price of medicine is much higher. So, overpassing these problems is a challenge to the medical community [4] [11] [13] [14]. Medicinal plants have been used for human well-being since ancient times [15] [16]. In conformity with WHO, approximately 4 billion people, 75% - 80% of the total population, use herbal remedies as first aid that can improve our healthcare and have fewer side effects, and better compatibility with the human body [1] [15] [16] [17]. As a result, the use of herbal medicine is gaining traction in both developed and developing countries [7]. In the United States, about 25% of the prescribed drugs come from natural sources [15]. So it is difficult to separate plants and humans [16].

Bangladesh is known as an agricultural country and is said to be a rich country of biodiversity. More than 500 medicinal plant species are found here that are effective in curing diabetes [13] [16] and contain numerous antioxidant properties like tannin, flavonoids, phenolic compounds, and alkaloids [3] [4] [18]. In diabetes, extemporaneous formation of free radicals has occurred on account of glucose auto-oxidation, and non-enzymatic protein glycation. When the amount of free radicals increases and the activity of antioxidants decreases, lipid peroxidation, enzymes, and cellular organelles are damaged, which also affects the abnormalities of diabetes. Plants having both antioxidant and anti-diabetic properties can prevent diabetes complications more efficiently than the currently available anti-diabetic drugs [4].

In this review, we have focused on Bangladeshi medicinal plants having anti-diabetic properties along with antioxidant substances. We have mainly attempted to show a comparative analysis of different plant parts and their various doses, and we have also tried to include their local names. This article may also be helpful for healthcare professionals, scientists, and scholars working in pharmacology and therapeutics to launch newer forms of medicine for the treatment of diabetes without or with fewer side effects.

2. Methodology

The data for this review was gathered from many research publications and review articles on ayurvedic plants having anti-diabetic and antioxidant potential in Bangladesh. The following databases have been used to collect reference papers:

Google Scholar, Scopemed, Medline, ResearchGate, PubMed, and Science Direct. For searching information, terms such as botanical plants, medicinal plants, traditional plants, anti-hyperglycemic plants, anti-diabetic plants, a survey of anti-diabetic plants, a survey of anti-diabetic active medicinal plants, a review of anti-diabetic medicinal plants, oxidative stress and an ethnomedicinal plant survey in Bangladesh have been used. We have found 81 Bangladeshi medicinal plants from 51 different families that can be used to treat diabetes and oxidative stress.

3. Results

In the present study, 81 Bangladeshi medicinal plants from 51 different families were noted. It was revealed that the plant parts used having anti-diabetic and antioxidant activity included leaves, bulbs, stems, roots, barks, rhizomes, aerial parts, fruits and seeds as well as whole plants (Table 1, Figure 1). The most commonly used species that can be used to treat diabetes and oxidative stress are *Psidium guajava* (L.), *Aloe vera*, *Catharanthus roseus*, *Allium sativum*, *Annona squamosa*, *Cinnamon zeylanicum*, *Amaranthus esculentus*, *Eugenia jambolana*, *Azadirachta indica*, *Moringa oleifera*, *Spondias pinnata*, *Coccinia grandis* (L.), *Momordica charantia* L, *Heretiera fomes*, and *Trigonella foenum-graecum*. The study indicated that the common medicinal plant families were Myrtaceae, Liliaceae, Apocynaceae, Alliaceae, Annonaceae, Lauraceae, Amaranthaceae, Asteraceae, Meliaceae, Moringaceae, Anacardiaceae, Cucurbitaceae, Sterculiaceae, and Fabaceae (Table 1).

Table 1. List of selected Bangladeshi medicinal plants having anti-diabetic and antioxidant activity.

Family	Botanical name	Local name	Plant part	Extract	Active chemical constituents	Dose (mg/kg)	Test model	Anti-diabetic effects	Antioxidant activity	Ref.
Myrtaceae	<i>Psidium guajava</i> (L.)	Peyara	Leaf and Fruit	Aqueous	Terpen, Flavonoid, Strictinin, Isostrictinin, Pedunculagin, and Polysaccharides	50 - 800 mg/kg	Streptozotocin (STZ)-induced diabetic rats	Decreased glucose level, Performed various anti-diabetic effect, and hypoglycemic effect	Showed high antioxidant activity. Fruit extracts exhibited weaker antioxidant effects than did the leaf extracts	[19] [20] [21]
Liliaceae	<i>Allium cepa</i>	Peyaj	Bulb	-	-	200 mg/kg	Alloxen induced diabetic rat	Controlled the activity of glucose 6 phosphate and HMG-CO a reductase	-	[22] [23] [24] [25] [26] [27]
	<i>Asparagus racemosus</i>	Satamuli	Root	Ethanollic	Steroidal, Saponins, Polyphenols (1.69%), Phytosterols (0.79%), Ascorbic acid (0.76%), and Flavonoids (0.48%)	1.25 g/kg	Type1 and type 2diabetic rats	Suppressed postprandial hyperglycemia, reversibly increased unabsorbed sucrose content throughout the gut, enhanced glucose transport and insulin action in 3T3-L1 adipocytes	Showed antioxidant activity	
	<i>Aloe vera</i>	Ghritkumari	Leaf	Ethanollic	Pseudoprototinosaponin AIII, and Prototinosaponins AIII	200 and 300 Mg/kg P.o., 28d	Alloxan induced diabetic mice	Glucose uptake and insulin released against glycogenolysis or gluconeogenesis pathway	Showed high antioxidant activity	
Apocynaceae	<i>Catharanthus roseus</i>	Nayantara	Full plant	-	Alkaloids, Terpenoids, Steroids, Flavonoids	0.5, 0.75 and 1.0 ml/kg	Alloxan-induced diabetic rabbits.	Dose-dependent reduction in blood glucose was observed	Showed high antioxidant activity (81.70%).	[28] [29] [30] [31] [32]
	<i>Alstonia Scholaris</i>	Saptaparni Devil's tree, Chaitan, Chatim	Leaf	Ethanollic	Alkaloids, Saponins, Terpenoids, Flavonoids, Phenolic compounds, Tannins, Steroids, and Glycosides	100, 200, and 400 mg/kg	Streptozotocin-induced diabetic rats	Reduced blood glucose level	Showed antioxidant activity	

Continued

Alliaceae	<i>Allium sativum</i>	Ada	Bulb	Ethanolic	-	0.1, 0.25 and 0.5 g/kg	Streptozotocin-induced diabetic rats.	Garlic alcoholic extract significantly decreased serum glucose	<i>Allium sativum</i> (L.) was rich in antioxidants which helped destroy free radicals' particles that could damage cell membranes and DNA.	[33] [34] [35]	
Solanaceae	<i>Capsicum frutescens</i>	Lal morich	Fruit			40 mg/kg	Type 2 diabetes model of rats	Increased serum insulin concentration in high fat diet	-	[36] [37] [38] [39] [40] [41] [42] [43]	
	<i>Capsicum annum</i>	Red pepper	Fruit	Ethanolic	Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) ester Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) ester Capsaicin, Dihydrocapsaicin, hytolkauran16-ol, Eugenol, 1, 2-benedicarbixylic acid	Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) este Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) ester Cap-Saicin, dihydrocapsaicin, tridecanoic acid, phytol, Kauran-16-ol, eugenol and 1, 2-benzenedicarboxylic Acid, mono (2-ethylhexyl) ester Capsaicin, Dihydrocapsaicin, hytolkauran16-ol, Eugenol, 1, 2-benedicarbixylic acid	300 mg/kg	Type 2 diabetes rats.	Amelioration of polyphagia, polydipsia And hyperglycemia, improving glucose tolerance Ability, insulin sensitivity, pancreatic β -cell histology And functions, and improving dyslipidemia Improved hyperglycemia, polydipsia, polyphagia, glucose tolerance ability, insulin sensitivity, and dyslipidemia	-	
	<i>Solanum torvum Swartz</i>	Tit baegun	Fruit	Methanolic	Methyl caffeate		10 mg/kg, 20 mg/kg, 40 mg/kg	Streptozotocin-induced diabetic rats	Decreased glucose level, increased insulin level, Increased expression level of GLUT4	-	
	<i>Datura stramonium L</i>	Shada dhutura	Leaves	Aqueous crude extract	Flavonoids, Glycosides, Alkaloids, Tannins, Saponins, Phenols		-	<i>In vitro</i>	Showed α -amylase inhibitory activity	-	
	<i>Withania somnifera (L.) Dunal</i>	Aswagandha	Root, Leaf	Ethanolic	Flavonoids	100 mg/kg, 200 mg/kg	Alloxen induced diabetic rat	Increased insulin secretion, Decreased oxidative stress.	Showed high antioxidant activity		
Annonaceae	<i>Annona squamosa</i>	Ata	Leaf	Aqueous	Flavonoid	55 mg/kg	Streptozotocin-induced diabetic rats	Reduced the concentration of glucose	A. squamosa leaf extracts showed antioxidant activity due to presence of flavonoids in the leaf, which in turn act as strong superoxide radicals and singlet oxygen quenchers.	[44] [45] [46]	

Continued

	<i>Ananus Cosmosus</i>	Anaras	Leaves		Flavonoids, Alkaloids, Saponins, Terpenoids, Annonaceous, Acetogenins, Megastigmanes, Cyclopeptides, and Essential oils	–	<i>In vitro</i>	Enhanced insulin sensitivity, Regulated the activities of glycogen metabolizing enzymes	Showed antioxidant activity	
Acanthaceae	<i>Andrographis paniculata</i>	Kalomegh	Leaf	Ethanol	Diterpenoid, Flavonoids, and Polyphenols	0.1, 0.2, and 0.4 g/body weight	Normal and Streptozotocin-induced diabetic rats.	Showed antidiabetic effect by increasing glucose metabolism. It also gave hypotriglyceremic effect	Showed antioxidant activity	[47] [48]
Lauraceae	<i>Cinnamon zeylanicum</i>	Daruchini	Bark	Chloroformic	Cinnamaldehyde (71.50%), Linalool (7.00%), Beta-caryophyllene (6.40%), Eucalyptol (5.40%), and Eugenol (4.60%)	20 mg/kg	Streptozotocin-induced diabetic rats	Insulinotropic effect due to glucose uptake increased through the glucose transporter translocation	–	[49]
Zingiberaceae	<i>Curcuma longa</i>	Holud	Bark	Ethanol	Curcumin, Demethoxycurcumin, Bisdemethoxycurcumin, and Ar-turmerone	1500 mg/kg/d	Type 2 diabetic KK-A(y) mice	EtOH extract stimulated human adipocyte differentiation in a dose-dependent manner, and reduced the blood glucose level	In the DPPH assay, <i>Curcuma longa</i> showed excellent scavenging against the radical	[50]
Caesalpiniaaceae	<i>Tamarindus indica</i>	Tetul	Seed	Aqueous	–	80 mg/100g	Streptozotocin-induced diabetic rats	Showed some important antidiabetic activity with attenuation of hyperglycemia	–	[51]
Piperaceae	<i>Piper betle</i>	Betel	Leaf	Aqueous, Ethanol	–	100, 200, 300 and 1500 mg/kg	Normoglycemic and Streptozotocin-induced diabetic rats	Showed strong antidiabetic activity	Showed high antioxidant activity	[28] [52] [53] [54] [55]
	<i>Piper cubeba L.f</i>	Kabab chini	Fruit	Methanol	Phenols, Flavonoids	–	<i>In vitro</i>	Showed α -amylase inhibitory activity, Increased glucose uptake	Showed high antioxidant activity	
	<i>Piper longum L.</i>	Pipul, Pipla	Roots	Aqueous	Piperin, Pippalartin, Piperleguminin, Sterols, and Glycosides, Piperlongumine	200 mg/kg for 30 days	Streptozotocin-induced diabetic rats	Decreased glucose level, Increased insulin level	Showed high antioxidant activity	
Amaranthaceae	<i>Amaranthus esculentus</i>	Dherosh	Grain	–	–	500 g/kg	Streptozotocin-induced diabetic rats	Reduced the glucose level	Showed antioxidant activity	[56] [57] [58] [59] [60] [61]
	<i>Achyranthes aspera L.</i>	Udvid nagra	Leaves	Ethanol	Alkaloids, Phenol, Flavonoids, Saponins, Carbohydrates, Steroids, and Terpenoids	1000 mg/kg	Streptozotocin-induced diabetic rats	Decreased blood glucose	Showed high antioxidant activity	
	<i>Amaranthus spinosus L.</i>	Katanotey	Leaf	Ethanol	Alkaloids, Saponin, Carbohydrate, Tannin, Protein, Glycoside, Flavonoid, and Phenol	450 mg/kg	Diabetic rat	Inhibited alpha amylase activity	Showed antioxidant activity	
Scrophulariaceae	<i>Scoparia dulcis</i>	Chingura	Whole plant	Aqueous	–	200 mg/kg	Streptozotocin-induced diabetic rats	Reduced blood glucose, increased plasma insulin level	Decreased free radical formation in tissues (liver and kidney)	[62]
Gramineae	<i>Hordeum vulgare</i>	Barley	Fruit	Ethanol	Phenol	200 mg/kg	Streptozotocin-induced diabetic rats	Decreased blood glucose level	–	[63]
Apiaceae	<i>Cuminum nigrum</i>	Cumin	Seeds	–	Flavonoids, Alkaloids	0.5 to 1.5 g/kg	Alloxan-diabetic rabbits	Exhibited considerable hypoglycemic activity	–	[4] [56] [64]
	<i>Centella asiatica (L.)</i>	Thankuni	Leaf	Ethanol	Alkaloids, Glycosides, Steroids, Flavonoids, Tannins, and Reducing sugars	500 mg/kg	Alloxan induced diabetic rat	Decreased glucose level	Showed antioxidant activity	
Gentianaceae	<i>Swertia chirayita</i>	Tite or Pothi Chirayita or Tikta	Leaf	Aqueous	Mangiferin	200 mg/kg	Streptozotocin-induced diabetic rats	Increased the plasma insulin, Reduced blood sugar	Showed antioxidant activity	[65]
Asteraceae	<i>Eugenia jambolana</i>	Jam	Fruit pulp	Ethanol	Flavonoids, Saponins, Glycosides, and Triterpenoids	50, 100, 200 mg/kg	Alloxan-induced diabetic rat	Increased glucose tolerance activity	–	[66] [67] [68] [69] [70] [71]
	<i>Eclipta alba L.</i>	Bhringoraj or Bhringraj	Leaves	Ethanol	Carbohydrates, Terpanoids, Lactones, Glycosides, Flavonoids, Esters, Steroids, Tannins	200 mg/kg for 30 days	Alloxan induced diabetic mice	Decreased glucose level, increased insulin level	–	
	<i>Enhydra fluctuans Lour.</i>	Helench, harkuch	Aerial Herbs	Ethanol	Flavonoids, Tannins, Saponins, Phenolic, Terpenoids, Steroids, and Coumarin compounds	500 mg/kg	Alloxan induced type –2 diabetic rat	Decreased blood glucose, Showed hypoglycemic effect	Showed antioxidant activity	

Continued

	<i>Blumea lacera</i>	Bonomula, shealmoti.	Aerial part, Leaf	Methanolic	Carbohydrates, Saponins, Tannins, Phenols, Polyphenols, Flavonoids, Reducing Sugar, Steroids, Alkaloids, Glycosides	400 mg/kg	Streptozotocin induced diabetic rat	Reduced blood glucose level, showed antihyperglycemic effect, Showed α -amylase inhibitory activity	Showed antioxidant activity	
Meliaceae	<i>Azadirachta indica</i>	Neem	Leaf	-	Flavonoids, Tannins, Sterols, Saponins, Anthraquinones, and Alkaloids	25, 48.4, 93.5, 180.9 and 350 mg/kg	Alloxan-induced white male albino mice	The extract lowered blood glucose at all dosages in a dose independent manner	-	[72] [73]
	<i>Swietenia mahagoni</i> L. Jacq.	mahagoni	Bark	Methanolic	-	25 and 50 mg/kg	Streptozotocin-induced diabetic rats	Showed strong hypoglycemic activity	Showed high antioxidant activity	
Anacardiaceae	<i>Mangifera indica</i>	Mango	Leaf	Aqueous	-	1 g/kg	Normoglycaemic, glucose-induced hyperglycaemic mice, Streptozotocin-induced diabetic mice	Reduced blood glucose level	-	[74] [75]
	<i>Spondias pinnata</i>	Amra	Fruits	Methanolic	Flavonoids, Tannins, Saponins, and Terpenoids	-	Alloxen induced diabetic rat	Decreased glucose level, Decreased triglycerides and total cholesterol, Decreased LDL, and VLDL, Increased HDL	-	
Malvaceae	<i>Abelmoschus moschatus</i>	Muskok dana		Aqueous	Myricetin	100 mg/kg	Streptozotocin-induced diabetic rats	Significantly attenuated the increase of plasma glucose	-	[76]
Legumes	<i>Glycine max</i>	Soyabeans	Seed	-	3-O-methyl-D-chiro-inositol (D-pinitol)	1.2 g	Type 2 diabetes mellitus (seven men, eight women; 60.3 +/- 3.1 years old)	Controlled postprandial capillary blood glucose the most effectively	-	[77]
Moraceae	<i>Ficus bengalensis</i>	Bot	Arial root	Aqueous	-	300 mg/kg	Sub and mild diabetic rats	Decreased blood glucose level	-	[36] [78] [79] [80] [81]
	<i>Ficus benghalensis</i> L.	Bat	Leaves	Ethanolic	Phenolics, Terpenoids/ Terpenes	200 mg/kg, 400 mg/kg body weight	Alloxen induced diabetic rat	Decreased glucose level, Decreased triglycerides, and cholesterol,	Showed antioxidant activity	
	<i>Ficus racemosa</i> L.	Joiggi dumur	Stem, Bark	Aqueous, Ethanolic	Tannin, Wax, Saponingluanol acetate, B-sitosterol, Leucocyanidin-3-O-b-d-glucopyranoside, Leucopelargonidin-3	Aqueous extract - 500 mg/kg & ethanol extract - 400 mg/kg)	Streptozotocin-induced diabetic rats	Decreased blood glucose	Showed antioxidant activity	
Moringaceae	<i>Moringa oleifera</i>	Shojna	Pods	Methanolic	-	150 or 300 mg/kg	Streptozotocin-induced diabetic rats	Reduced serum glucose levels, increased serum insulin levels	Increased antioxidant levels in pancreatic tissue	[82]
Caricaceae	<i>Carica papaya</i>	Papaya	Leaf	Ethanolic	Flavonoids, Tannins, Saponins, Triterpenoids, Alkaloids	250, 500, 1000 mg/kg	Alloxan induced diabetic rats	Decreased blood glucose level	-	[83]
Lamiaceae	<i>Mentha piperita</i>	Pudina	Leaf	Ethanolic	-	300 mg/kg	Streptozotocin-induced diabetic rats	Decreased the level of blood glucose and creatinine as well as increased level of insulin, glycogen, and body weight	-	[84] [85]
	<i>Ocimum sanctum</i> L.	Tulsi	Leaves	Ethanolic	Saponins, Triterpenes, and Flavonoids, Tannins, Alkaloids,	200 mg/kg i.p	Alloxen induced diabetic rat	Reduced blood glucose level	Showed antioxidant activity	
Musaceae	<i>Musa paradisiaca</i>	Kala	Leaves, Fruit peels	Ethanolic	-	500 mg/kg	Streptozotocin-induced diabetic rats	Reduced blood glucose level	-	[86] [87]
Euphorbiaceae	<i>Emblica officinalis</i> or <i>Phyllanthus emblica</i> L.	Amla,	Seeds	Aqueous	Flavonoids, Gallic acids	300 mg/kg	Streptozotocin-induced diabetic rats	Reduced blood glucose level and improved glucose tolerance	Showed antioxidant activity	[88]

Continued

Araceae	<i>Amorphophallus campanulatus</i> (Roxb.) Blume	Ol	Tuber	Methanolic	Betulinic acid, Tricontane, Lupeol, Stigmasterol, Sitosterol	400 mg/kg	Oral glucose tolerance tests (OGTT)	Inhibited amylase activity, Decreased glucose level, Reduced glycated hemoglobin, and nitric oxide, Increased insulin level	Showed antioxidant activity	[28] [45] [89] [90]
	<i>Alocasia macrorrhizos</i> (L.) G. Don Or <i>Alocasia indica</i>	Mankachu	Rizome	Methanolic	Flavonoids, Cynogenetic glycosides, Ascorbic acid, Gallic acid, Malic acid, Oxalic acid, Alocasin, Amino acids, Succinic acid, and Beta-lectins	500 mg/kg	Alloxen induced diabetic rat	Stimulated insulin secretion from beta cell	Showed antioxidant activity	
Combretaceae	<i>Terminalia bellerica</i> (Gaertn.) Roxb.	Bohera	Fruit	Aqueous	Termilignan, Thannilignan, 7-hydroxy-30, 40 -(methyleneedioxy) flavone, Anolignan B, Gallic acid, Ellagic acid, b-sitosterol, Arjungenin, Belleric acid, Bellericoside, and Cannogenol 3-O-b-D-galactopyranosyl-(1 → 4)-O-a-L-rhamnopyranoside	p.o.-up to 11 days; 120 mg/kg	Alloxen induced diabetic rat	Decreased glucose level, Stimulated both the secretion and action of insulin, inhibited protein glycation	Showed antioxidant activity	[1] [7] [10] [36]
	<i>Terminalia chebula</i>	Hortoki	Fruit, Seed	Ethanol, Chloroform, Aqueous	Shikimic, Gallic, Triacontanoic, Palmitic acid, β -sitosterol, Daucoesterol	p.o.-1 month; 200 mg/kg	Streptozotocin-induced diabetic rats	Decreased glucose level		
Cucurbitaceae	<i>Coccinia grandis</i> (L.)	Telakucha,	Leaves, Fruits	Aqueous	β-Sitosterol, Cephalandrol, Cephalandrine A & B,	200 mg/kg for 14 days	Alloxen induced diabetic rat	Reduced glucose level	Showed antioxidant activity	[1] [7] [18] [91] [92] [93] [94] [95] [96] [97] [98]
	<i>Momordica charantia</i> L.	Karolla, Uchhe, Usta	Whole plant	Methanolic, Aqueous, Chloroformic	Momordicin, Galactose-binding lectin non -bitter, Diosgenin, Cholesterol, Lanosterol, β -sitosterol, Cucurbitacin glycoside	p.o., 27 -30d; 10 -20 mg/kg	Streptozotocin-induced diabetic mice	Decreased glucose level, glycosylated hemoglobin, and oxidative stress, increased glycogen, Decreased lipid peroxidation	Showed antioxidant activity	
	<i>Momordica dioica</i>	Kakrol	Fruit	Methanolic	Lectins, β-sitosterol, Saponin glycosides, Triterpenes of ursolic acid, Hederagenin, Oleanolic acid, Stearic acid, Gypsogenin, and Momodicaursenol	300 mg/kg	Streptozotocin-induced diabetic rats	Reduced serum glucose, Increased serum insulin and urea levels	Showed antioxidant activity	
	<i>Luffa cylindrica</i> L.	Dhundul.	Fruit	Methanolic	Alkaloids, Glycosides, Flavonoids, Tannins, Terpenoids, Sterols, Saponins, Fats, and Sugars	400 mg/kg	Alloxan induced hyperglycemic rats	Showed hypoglycemic and antihyperglycemic effects	Showed antioxidant activity	
	<i>Luffa acutangula</i>	Zinga, Sponge ground	Fruit	Methanolic	Carbohydrates, Saponins, Phenol, Amino acid, Flavonoids	400 mg/kg	Streptozotocin induced diabetic rat	Showed alpha-glucosidase inhibitory activity	Showed antioxidant activity	
Flacourtiaceae	<i>Flacourtia indica</i> (Burm.f.) Merr	Katabohori/ Boichi	Leaves	Ethanol	-	-	Streptozotocin-induced diabetic rats	Altered fasting blood glucose, serum cholesterol and triglycerides	-	[99] [100]
Rutaceae	<i>Aegle marmelos</i>	Bael	Leaf, Seed, Fruit	Ethanol, Aqueous	Aegeline 2, Coumarin, Flavonoid, Alkaloid	I. p. 14d; p. o. 14d; 1.0 g/kg	Streptozotocin-induced diabetic rats	Decreased glucose level, Decreased glycosylated hemoglobin, Increased C peptide, increased glucose tolerance, increased glycogen,	Showed antioxidant activity	[7] [101]
Sterculiaceae	<i>Heretiera fomes</i>	Sundri	Leaves	Methanolic	Saponins, reducing sugars, Alkaloids, Glycosides, Flavonoids, Tannins, Steroids, and Gums	-	<i>In vitro</i>	Showed α amylase inhibitory activity, Increased glucose adsorption	Showed antioxidant activity	[102] [103]
Asparagaceae	<i>Asparagus racemosus</i> L.	Sotomuli	Whole plant	Ethanol	Alkaloids, Flavonoids, Tannins, Saponins, Phenols, Terpenes, Polysaccharides, and Steroids	200 and 400 mg/kg for 21 days	Streptozotocin-induced diabetic rats	Significantly decreased the blood glucose level, significantly decreased thiobarbituric acid reactive substances and significantly increased in reduced glutathione, superoxide dismutase and catalase	Showed antioxidant activity	[36] [104]

Continued

Poaceae	<i>Cynodon dactylon</i> (L.)	Durva, Dub, Dubla, Durba, Doorva, Neel Doorva	Whole plant	Aqueous	Flavonoids, Sterols	500 mg/kg	Streptozotocin-induced diabetic rats	Decreased glucose level, Decreased TC, Decreased TG, Decreased LDL, Increased HDL	-	[105] [106]
Lythraceae	<i>Lagerstroemia speciosa</i> (L.) Pers	Jarul	Leaves	90% ethanolic	Polyphenol, Corosolic acid	-	Streptozotocin-induced diabetic rats	Increased insulin secretion, decreased glucose level, improved hyperlipidemia, and recovered the beta cell of langerhans islet	-	[107] [108]
Missociaceae	<i>Mimosa pudica</i> L.	Lojjaboti	Leaves	Ethanolic	Alkaloids, Tannins and Flavonoids	600 mg/kg	Alloxan induced diabetic rat	Reduced blood glucose level with respect to corresponding control	-	[109] [110]
Fabaceae	<i>Trigonella foenum-graecum</i> L	Methi	Leaves	Ethanolic	Vitamin C, E and β carotene, Phenolics	250 and 500 mg/kg for 28 days	Streptozotocin-induced diabetic rats	Decreased glucose level,	Showed high antioxidant activity	[36] [111] [112] [113] [114] [115]
	<i>Sesbania sesban</i>	jainti, jayant	Leaves	Aqueous	Triterpenoids, Carbohydrates, Vitamins, Amino acids, Proteins, Tannins, Saponins Glycosides	500 mg/kg	Streptozotocin-induced diabetic rats	Reduced serum glucose level	Showed antioxidant activity	
	<i>Erythrina variegata</i>	Tiger's Claw, coral tree	Leaf	Methanolic	Alkaloids, Flavonoids, Triterpenoids, and Lectin	900 mg/kg	Streptozotocin induced diabetic rat	Showed plasma glucose lowering activity, Stimulated pancreatic β cells	Showed antioxidant activity	
Menispermaceae	<i>Tinospora cordifolia</i>	Gulancha lota	Stem	Methanolic	-	500 mg/kg for 6 weeks	Alloxan induced diabetic rat	Reduced blood glucose, decreased glycosylated, decreased hemoglobin, decreased cholesterol, Decreased free radicals' levels	Showed high antioxidant activity	[38] [116]
Anacardiaceae	<i>Lannea coromandelica</i> (Houtt.) Merr.	Jiga	Bark	Ethanolic	Flavonoids, Alkaloids, Glycosides, Carbohydrate and Phenolics	500 mg/kg	Alloxan induced type-2 diabetic rat	Reduced blood glucose level significantly, Showed antihyperglycemic effect	Showed antioxidant activity	[117] [118]
Polypodiaceae	<i>Drynaria quercifolia</i> (L.) J. Smith	Pankhiraj, ferns	Stem, Rhizome	Methanolic	Epifriedelinol, Beta-amyrin, Beta-sitosterol, Beta-sitosterol 3-beta-D-glucopyranoside, 3, 4 dihydroxyl benzoic acid, Acetyl lupeol, Aglycone naringenin, and Flavones glycoside	100 μ /ml	Alpha-amylase inhibition assay and glucose uptake assay	Showed antihyperglycemic effect Limited intestinal carbohydrate digestion, Maintained plasma glucose concentration	Showed antioxidant activity	[28] [119]
Verbenaceae	<i>Clerodendrum viscosum</i> Vent	Ghetu, Bhat.	Leaves	Ethanolic	Saponin, Flavonoids, Alkaloids, A new glycoside, Clerodendroside, Lupeol, Benzoic acid derivatives, and Beta-itosterol	500 mg/kg	Alloxan induce diabetic rat.	Reduced blood glucose level	Showed antioxidant activity	[120] [121]
	<i>Premna integrifolia</i>	Agnimantha	Bark	Methanolic	Alkaloids, Glycosides, Terpenoids, Carotenoids, and Flavonoids	300 mg/kg	Alloxan induced diabetic rat	Decreased blood glucose level	Showed antioxidant activity	
Rubiaceae	<i>Xeromphis uliginosa</i> Retz		Root	Methanolic	Flavonoids, Tannins	500 mg/kg	Alloxan induced diabetic rat	Showed glucose lowering effect, and anti-hyperglycemic effect, Protected beta cell from destroy, and regeneration	Showed antioxidant activity	[122]
Asclepiadaceae	<i>Gymnema sylvestre</i>	Medha singi, Gorshar	Whole plant	Methanolic	Alkaloids, Polypeptide, Flavonoids, Tannins, Phenols, Cinnamic acid, Folic acid, Ascorbic acid, Butyric acid, Tartaric acid	30 mg/day	Streptozotocin Induced Diabetic Rats	Lowered blood glucose level (sugar destroyer), Showed antihyperglycemic effect	Showed antioxidant activity (reducing the oxidative stress in cells)	[28] [123] [124]
Scrophulariaceae,	<i>Scoparia dulcis</i>	Bon dhonia, Sweet Broom	Aerial Part	Ethanolic	Alkaloids, Carbohydrates, Glycoside, Flavonoid, & Tannins	200 mg/kg	Alloxan induced diabetic rat	Changed serum glucose level, Showed antihyperglycemic effect	Showed antioxidant activity	[14] [125]

Continued

Brassicaceae	<i>Brassica juncea</i>	Mustard, Sarsu	Seed	Aqueous	Tannin, Terpenoid, Saponin, Flavonoid, Alkaloid, Phenols, and Cardiac glycoside	100 mg/kg	Streptozotocin induced diabetic rats	Showed antidiabetic activity, and hypoglycemic activity, Depleted serum insulin levels	Showed antioxidant activity	[126] [127]
Berberidaceae	<i>Berberis aristata</i>	Daruhaldi	Stem	Methanolic	Alkaloids, Glycosides, Carbohydrates, Saponins	500 mg/kg	Streptozotocin induced diabetic rat	Reduced blood glucose level	Showed antioxidant activity	[128] [129]
Magnoliaceae	<i>Michelia champaca</i>	Sornochampa, champaka	Bark	Ethanollic	Phenolics, Flavonoids, Flavonols, and Proanthocyanidins, Alkaloids, Saponins, Tannins, Sterols	–	<i>In vitro</i>	Showed α -amylase inhibitory activity	Showed antioxidant activity	[130]
Actnidaceae	<i>Saurauia roxburghii</i>	Bhola kadam	Leaves	Ethanollic	Alkaloids, Carbohydrates, Cardiac glycosides, Flavonoids, Saponins, Terpenes, Steroids	2 g/kg	Alloxan induced diabetic rat	Lowered blood glucose level	Showed antioxidant activity	[121]
Leguminosae	<i>Acacia catechu</i>	Khoyer/ Khair	Bark	Ethanollic	Flavonoids, Alkaloids, Glycosides, Flavones, Tannins, Terpenes, Sterols, Saponins, Fats, and Sugars	400 mg/kg	Alloxan induce diabetic rat	Stimulated insulin secretion from beta cell	Showed antioxidant activity	[90] [131] [132]
Palmaceae	<i>Areca catechu L.</i>	Supari, betel nut, areca nut	Leaf	Methanolic	Flavonoids, Triterpenoid Compounds, Alkaloids, Steroids, Saponins, Tannins, Glycosides and Carbohydrates	200 mg/kg	Streptozotocin-induced diabetic rats	Reduced and normalized of elevated blood sugar levels	Showed antioxidant activity	[133] [134]

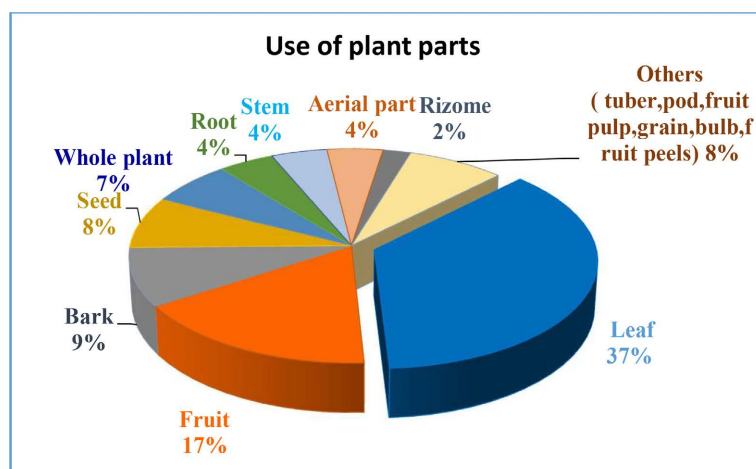


Figure 1. Percentage of plant parts used for the treatment of diabetes and oxidative stress.

4. Discussion

Diabetes mellitus is expanding shockingly. In Bangladesh, diabetes mellitus is becoming a common disorder. Uncontrolled diabetes mellitus leads to many chronic complications such as blindness, heart disease, renal failure, etc. The oral hypoglycemic drugs available on the market have a number of drawbacks such as gastrointestinal discomfort, weight gain, and hepatic dysfunction [7]. Regularly used glucose-lowering drugs may affect the etiology of COVID 19, which may affect the people with diabetes mellitus and initiating the risk of development of severe disease. That's what motivates researchers to look for novel anti-diabetic drugs [92]. So, scientists are trying to treat diabetes mellitus with plant-derived compounds. Moreover, medicinal plants are used to treat diabetes in developing nations, especially to lessen the economic burden of synthetic drugs on people [57]. Plants are known to possess a wide variety of pharmaco-

logical effects and therapeutic activities.

The generation of free radicals is one of the most common causes of the development of diabetes. Oxidative stress in diabetes is accompanied by a reduction in the antioxidant status, which can increase the harmful effect of free radicals [135]. Oxidative stress increases complications of diabetes, which include stroke, neuropathy, retinopathy, and nephropathy. It plays a part in the growth of complications of diabetes, especially type-2 diabetes. It leads to endothelial dysfunction in the blood vessels of diabetic patients [136].

Our study revealed that 81 plant species of Bangladesh belonging to 51 families showed significant anti-diabetic and antioxidant activity. Most of the medicinal plants showed anti-diabetic activity by decreasing blood glucose levels. The use of fruits and leaves were the most common plant parts against the diseases (Figure 1). Most common active chemical constituents which show antidiabetic and antioxidant effects are flavonoid, saponin, terpenoid, alkaloid, tannins, phenol, steroid, and glycoside. Alloxan induced diabetic rats and Streptozotocin (STZ)-induced diabetic rats were the most frequently utilized diabetic models.

From this review the most significant plants were noted based on their substantial effect against diabetes and oxidative stress namely *Psidium guajava* (L.), Aloe vera, *Catharanthus roseus*, *Allium sativum*, *Annona squamosa*, *Cinnamom zeylanicum*, *Amaranthus esculentus*, *Eugenia jambolana*, *Azadirachta indica*, *Moringa oleifera*, *Spondias pinnata*, *Coccinia grandis* (L.), *Momordica charantia* L, *Heretiera fomes*, *Trigonella foenum-graecum* (Table 1) Few of them are narrated below:

Psidium guajava (L.) (Local Name: Peyara) is one of the most common plants in Bangladesh. The hypoglycemic effect of the plants' extract was examined in normal and diabetic rats using a streptozotocin (STZ)-induced diabetes mellitus model. Doses of 50 - 800 mg/kg were given and the effects were dose-dependent. Tannins, polyphenolic compounds, flavonoids, pentacyclic triterpenoids, guajaverin, quercetin, and other chemical compounds present in the plant are responsible for the hypoglycemic effect. It also showed good antioxidant activity, but fruit extracts exhibited weaker antioxidant effects than leaf extracts because many phytochemical compounds such as flavonoids, phenolic acids and carotenoid were responsible for antioxidant activity. So, fruit part and leaf part showed different antioxidant activity [19] [20] [21].

Aloe vera is another popular plant in Bangladesh. (Local name: Ghritkumari, Family: Liliaceae). Alloxan 200 and 300 mg/kg of plant extract doses were given to alloxan-induced diabetic rats. Pseudoprotinosaponin AIII and protinosaponin AIII showed anti-diabetic activity. Significant role of these constituents is to uptake glucose and insulin release by glycogenolysis or gluconeogenesis pathway. It showed high antioxidant activity [21].

Catharanthus roseus, which is also known as noyon tara, can be found anywhere in Bangladesh. It produced a maximum reduction in blood glucose of 19.6% (8 h), 31.4% (18 h), and 36.5% (20 h). Observed with *C. roseus* at doses of

0.5, 0.75, and 1.0 ml/kg body weight respectively. So, the effects were dose dependent. It showed good antioxidant activity (81.70%). *C. roseus* showed rapid normalization of blood glucose levels rather than to control because some β -cells were still surviving to act upon by *C. roseus* to exert its insulin releasing effect. It can be said that, the mode of action of the active ingredients of *C. roseus* is probably arbitrated by an enhanced secretion of insulin, like sulphonylureas. *C. roseus* has indicated a prolonged duration of antidiabetic action after treatment in the period 18 - 24 h. Moreover, the juice has a more prolonged effect (at 1.0 ml/kg) than the glibenclamid, an antidiabetic drug [29] [30].

The anti-diabetic effect of garlic (Bulb Part) ethanolic extract (*Allium sativum* Lantioxidants, garlic.) was investigated in streptozotocin-induced diabetic rats. The result showed that garlic alcoholic extract significantly decreased serum glucose. *Allium sativum* (L.) is also rich in antioxidants, which help destroy free radicals [33] [34] [35].

The leaves of *Annona squamosa* (ata) displayed an anti-diabetic effect. Flavonoids found in the leaf part are responsible for the anti-diabetic effect. Oral administration of *A. squamosa* aqueous extract to diabetic rats for 30 days significantly reduced the levels of blood glucose, lipids, and lipid peroxidation. The study also revealed that *A. squamosa* can partially reduce the imbalance between the generation of reactive oxygen species (ROS) and scavenging enzyme activity. So, it can be supplemental as an antioxidant therapy [44].

Terminalia chebula (Combretaceae) has been widely used as a medicinal plant for the treatment of diabetes. The chloroform extract of the seed part showed an anti-diabetic effect by reducing the blood glucose level. The active chemical constituents are Shikimic, gallic, triacontanoic, palmitic acid, β -sitosterol, daucosterol. *T. chebula* produced a maximum reduction of blood glucose with doses of 200 mg/kg. Effects were dose-dependent reduction of blood glucose. The study also revealed that *T. chebula* more effectively inhibited the incidence of diabetic nephropathy [1] [7].

Cinnamomum zeylanicum is widely used in alternative systems of medicine for the treatment of diabetes. Cinnamaldehyde is the active ingredient which works against diabetes. Oral administration of CND (20 mg/kg bw) to diabetic rats for 2 months showed significant improvement (p 0.001). The insulinotropic effect of CND increases the glucose uptake through glucose transporter (GLUT4) translocation in peripheral tissues. Additionally, it has been demonstrated to have anti-oxidative, anti-inflammatory, and tumor cell growth inhibitory properties by inhibiting the formation of nitric oxide by LPS-stimulated macrophages and it signifies a wide margin of safety [49].

The oral anti-hyperglycemic effect of the ethanolic extracts of the fruit-pulp of *Eugenia jambolana* (EJ) was investigated in alloxan-induced diabetic rats. Flavonoids, saponins, glycosides, and triterpenoids are the active ingredients responsible for increasing the glucose tolerance level [68].

The hypoglycemic effect of aqueous leaf extracts of *A. indica* was evaluated in

alloxan-induced white male albino mice. Flavonoids, tannins, sterols, saponins, anthraquinones and alkaloids were responsible for antidiabetic effect. Intraperitoneal administration of *A. indica* decreases the blood glucose level dose dependently at the five doses of 5 mg/kgbw, 48.4 mg/kgbw, 93.5 mg/kgbw, 180.9 mg/kgbw, and 350 mg/kgbw [34].

The presence of bioflavonoids in *Moringa oleifera* is responsible for the stimulation of glucose uptake in peripheral tissues and the regulation of the activity and or expression of the rate-limiting enzymes involved in carbohydrate metabolism and flavonoids work as an insulin secretagogue or insulin mimetic. It also increased antioxidant levels in pancreatic tissue and decreased oxidative stress. So, it has significant antidiabetic and antioxidant properties [83].

The hypoglycemic effect of aqueous leaf extracts of *A. indica* was evaluated in alloxan-induced white male albino mice. Flavonoids, tannins, terpenoids, sterols, saponins, anthraquinones and alkaloids were responsible for the antidiabetic effect. Among them, terpenoids are heart-friendly since they assist in lowering blood sugar levels and diastolic blood pressure. Anthraquinones are also used to treat peripheral neuropathy. Intraperitoneal administration of *A. indica* decreases the blood glucose level dose dependently at the five doses of 5 mg/kgbw, 48.4 mg/kgbw, 93.5 mg/kgbw, 180.9 mg/kgbw, and 350 mg/kgbw [34].

Administration of *Carica papaya* methanolic leaf extract of 250, 500, and 1000 mg/kg B.W was examined on alloxan-induced diabetic rats. Continuous induction of *Carica papaya* leaf extracts decreased blood glucose levels, especially at a dose of 1000 mg/kg body weight [84].

Momordica charantia L (Karolla) has an effective dose of 10 - 20 mg/kg for 27 - 30 days and contains bitter substances such as charantin, vicine, glycosides, and karavilosides along with polypeptide-p, plant insulin. These compounds are hypoglycemic in action and lower blood sugar levels by increasing glucose uptake and glycogen synthesis in the liver, muscles, and fat cells. They also enhance insulin release from pancreatic beta cells and restore or encourage the proliferation of insulin-secreting cells. Another bioactive substance found in bitter melon, called lectin, has insulin-like activity and aids in appetite suppression [7] [137].

From above discussion, we can see that most of the plants contain alkaloids, flavonoids, tannins, saponins, and phenolic compounds. Some possibilities concern how they affect the creation and release of pancreatic β cells, the improvement of insulin sensitivity, or the insulin-like activity of plant extracts. According to, flavonoids increase lipogenesis and glucose transport in adipocytes, resulting in a reduction in blood sugar. The alkaloids encourage pancreatic islet regeneration, which in turn restores insulin secretion. It has also been demonstrated that tannins and saponins have hypoglycemic properties [72]. The elimination or reduction of diabetic complications may be the result of all of these activities.

Among 81 medicinal plants, *Psidium guajava* (L.), *Aloe vera*, *Catharanthus roseus*, *Allium sativum*, *Annona squamosa*, *Cinnamomum zeylanicum*, *Amaranthus*

esculentus, *Azadirachta indica*, *Moringa oleifera*, *Spondias pinnata*, *Coccinia grandis* (L.), *Momordica charantia* L, *Trigonella foenum-graecum* responded more swiftly and relatively at a lower dose than others in mice or rats, making them more successful in treating lower blood glucose levels. Although the reduction of blood glucose has been observed only in test animals like rabbit, mice or rat, the effects can be same as human body due to their anatomical, physiological and genetic similarity. When blood glucose levels drop, there is a significant likelihood that the number of complications related to diabetes will also decrease in humans. So, these most significant plants also help reduce diabetic complications.

5. Conclusions

Diabetes is a metabolic illness that has become a global health concern. Many studies manifest oxidative stress playing a vital role during diabetes, including impairment of insulin action and advancement of the complication incidence. As a result, there is a significant need for proper and adequate therapy for this disorder, which requires the development of treatments with minimal side effects and lower costs than the currently available synthetic medications. Because of several limitations related to the use of established synthetic anti-diabetic drugs, the hunt for innovative anti-diabetic therapies from natural sources should be a matter of concern to us.

From this review, it can be concluded that various research institutions or organizations should give more attention to medicinal plants available in Bangladesh and take necessary steps to extract active phytochemicals from them and it will certainly assist the treatment and management process of diabetes and oxidative stress. And we hope that it will be the groundbreaking step in the health sector.

Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

References

- [1] Islam, M.M., Jarna, R.N., Jain, P., Alam, M.A., Reza, H.M., Hossain, M. and Rahman, M.M. (2019) Potential Anti-Diabetic Medicinal Plants in Bangladesh: A Comprehensive Review. *World Journal of Pharmaceutical Research*, **8**, 140-150.
- [2] Kaul, K., Tarr, J.M., Ahmad, S.I., Kohner, E.M. and Chibber, R. (2013) Introduction to Diabetes Mellitus. In: Ahmad, S.I., Ed., *Diabetes: An Old Disease, a New Insight*, Advances in Experimental Medicine and Biology, Vol. 771, Springer, Berlin, 1-11. https://doi.org/10.1007/978-1-4614-5441-0_1
- [3] Reza, M.S., Shuvo, M.S.R., Hassan, M.M., Basher, M.A., Islam, M.A., Naznin, N.E. and Daula, A.S.U. (2020) Antidiabetic and Hepatoprotective Potential of Whole Plant Extract and Isolated Compounds of *Aeginetia indica*. *Biomedicine & Pharmacotherapy*, **132**, Article ID: 110942. <https://doi.org/10.1016/j.biopha.2020.110942>
- [4] Rahman, M.M., Sayeed, M.S.B., Haque, M.A., Hassan, M.M. and Islam, S.A. (2012)

- Phytochemical Screening, Antioxidant, Anti-Alzheimer and Anti-Diabetic Activities of *Centella asiatica*. *Journal of Natural Product and Plant Resources*, **2**, 504-511.
- [5] American Diabetes Association (2005) Diagnosis and Classification of Diabetes Mellitus. *Diabetes Care*, **28**, S37. https://doi.org/10.2337/diacare.28.suppl_1.S37
- [6] Mazumdar, S., Akter, R. and Talukder, D. (2015) Antidiabetic and Antidiarrhoeal Effects on Ethanolic Extract of *Psidium guajava* (L.) Bat. Leaves in Wister Rats. *Asian Pacific Journal of Tropical Biomedicine*, **5**, 10-14. [https://doi.org/10.1016/S2221-1691\(15\)30163-5](https://doi.org/10.1016/S2221-1691(15)30163-5)
- [7] Mamun-or-Rashid, A.N.M., Hossain, M.S., Hassan, N., Dash, B.K., Sapon, M.A. and Sen, M.K. (2014) A Review on Medicinal Plants with Antidiabetic Activity. *Journal of Pharmacognosy and Phytochemistry*, **3**, 149-159.
- [8] Jakhmola, V. and Tangri, P. (2012) Diabetes Mellitus a Silent Killer: Role of DPP4 Inhibitors in Treatment. *Journal of Pharmaceutical Science and Bioscientific Research*, **2**, 49-53.
- [9] Kasabri, V., Flatt, P.R. and Abdel-Wahab, Y.H. (2010) *Terminalia bellirica* Stimulates the Secretion and Action of Insulin and Inhibits Starch Digestion and Protein Glycation *in Vitro*. *British Journal of Nutrition*, **103**, 212-217. <https://doi.org/10.1017/S0007114509991577>
- [10] Alam, U., Asghar, O., Azmi, S. and Malik, R.A. (2014) General Aspects of Diabetes Mellitus. In: *Handbook of Clinical Neurology*, Vol. 126, Elsevier, Amsterdam, 211-222. <https://doi.org/10.1016/B978-0-444-53480-4.00015-1>
- [11] Rahman, A.M. (2015) Ethno-Botanical Survey of Anti-Diabetic Medicinal Plants Used by the Santal Tribe of Joypurhat District, Bangladesh. *International Journal of Research*, **2**, 19-26.
- [12] Choudhury, M.E., Mostofa, M. and Awal, M.A. (2006) Effects of Four Indigenous Medicinal Plants on Some Serum Parameters in Streptozotocin Induced Diabetic Rats. *Journal of the Bangladesh Agricultural University*, **4**, 259-264.
- [13] Prottoy, N.I., Sarkar, B., Ullah, A., Hossain, S., Boby, A.S. and Araf, Y. (2019) Molecular Docking and Pharmacological Property Analysis of Antidiabetic Agents from Medicinal Plants of Bangladesh against Type II Diabetes: A Computational Approach. *PharmaTutor*, **7**, 6-15.
- [14] Zulfiker, A.H.M., Ripa, F.A., Rahman, M.M., Ullah, M.O., Hamid, K., Khan, M.M.R. and Rana, M.S. (2010) Antidiabetic and Antioxidant Effect of *Scoparia dulcis* in Alloxan Induced Albino Mice. *International Journal of PharmTech Research*, **2**, 2527-2534.
- [15] Bhushan, M.S., Rao, C.H.V., Ojha, S.K., Vijayakumar, M. and Verma, A. (2010) An Analytical Review of Plants for Antidiabetic Activity with Their Phytoconstituent & Mechanism of Action. *International Journal of Pharmaceutical Sciences and Research*, **1**, 29-46.
- [16] Rahman, A.H.M.M. and Kumar, A.K. (2015) Investigation of Medicinal Plants at Katakhal Pouroshova of Rajshahi District, Bangladesh and Their Conservation Management. *Applied Ecology and Environmental Sciences*, **3**, 184-192.
- [17] Rahmatullah, M., Das, A.K., Mollik, M.A.H., Jahan, R., Khan, M., Rahman, T. and Chowdhury, M.H. (2009) An Ethnomedicinal Survey of Dhamrai Sub-District in Dhaka District, Bangladesh. *American Eurasian Journal of Sustainable Agriculture*, **3**, 881-888.
- [18] Bushnak, R., El Hajj, M. and Jaber, A. (2021) A Review on the Antidiabetic Potential of Medicinal Plants. *Journal of Medical and Healthcare*, **4**, 172-189. <https://doi.org/10.47752/sjmh.44.172.189>

- [19] Ojewole, J.A.O. (2005) Hypoglycemic and Hypotensive Effects of *Psidium guajava* Linn. (Myrtaceae) Leaf Aqueous Extract. *Methods and Findings in Experimental and Clinical Pharmacology*, **27**, 689-696. <https://doi.org/10.1358/mf.2005.27.10.948917>
- [20] Rai, P.K., Mehta, S. and Watal, G. (2010) Hypolipidaemic & Hepatoprotective Effects of *Psidium guajava* Raw Fruit Peel in Experimental Diabetes. *Indian Journal of Medical Research*, **131**, 820-824.
- [21] Chen, H.Y. and Yen, G.C. (2007) Antioxidant Activity and Free Radical-Scavenging Capacity of Extracts from Guava (*Psidium guajava* L.) Leaves. *Food Chemistry*, **101**, 686-694. <https://doi.org/10.1016/j.foodchem.2006.02.047>
- [22] Singh, L.W. (2011) Traditional Medicinal Plants of Manipur as Antidiabetics. *Journal of Medicinal Plants Research*, **5**, 677-687.
- [23] Ayodhya, S., Kusum, S. and Anjali, S. (2010) Hypoglycaemic Activity of Different Extracts of Various Herbal Plants. *International Journal of Research in Ayurveda and Pharmacy (IJRAP)*, **1**, 212-224.
- [24] Heś, M., Dziedzic, K., Thanh-Blicharz, J.L., Kmieciak, D. and Górecka, D. (2016) Antioxidant Activity of True Aloe (*Aloe vera*) Extract in Model Systems. *Nauka Przyroda Technologie*, **10**, Article No. 53. <https://doi.org/10.17306/J.NPT.2016.4.53>
- [25] Kumari, K., Mathew, B.C. and Augusti, K.T. (1995) Antidiabetic and Hypolipidemic Effects of S-methyl Cysteine Sulfoxide Isolated from *Allium cepa* Linn. *Indian Journal of Biochemistry & Biophysics*, **32**, 49-54.
- [26] Hannan, J.M.A., Ali, L., Khaleque, J., Akhter, M., Flatt, P.R. and Abdel-Wahab, Y.H.A. (2012) Antihyperglycaemic Activity of *Asparagus racemosus* Roots Is Partly Mediated by Inhibition of Carbohydrate Digestion and Absorption, and Enhancement of Cellular Insulin Action. *British Journal of Nutrition*, **107**, 1316-1323. <https://doi.org/10.1017/S0007114511004284>
- [27] Karuna, D.S., Dey, P., Das, S., Kundu, A. and Bhakta, T. (2017) *In Vitro* Antioxidant Activities of Root Extract of *Asparagus racemosus* Linn. *Journal of Traditional and Complementary Medicine*, **8**, 60-65. <https://doi.org/10.1016/j.jtcme.2017.02.004>
- [28] Rahman, M., Uddin, M., Reza, A.S.M., Tareq, A.M., Emran, T.B. and Simal-Gandara, J. (2021) Ethnomedicinal Value of Antidiabetic Plants in Bangladesh: A Comprehensive Review. *Plants*, **10**, Article No. 729. <https://doi.org/10.3390/plants10040729>
- [29] Nammi, S., Boini, M.K., Lodagala, S.D. and Behara, R.B.S. (2003) The Juice of Fresh Leaves of *Catharanthus roseus* Linn. Reduces Blood Glucose in Normal and Alloxan Diabetic Rabbits. *BMC Complementary and Alternative Medicine*, **3**, Article No. 4. <https://doi.org/10.1186/1472-6882-3-4>
- [30] Patharajan, S. and BalaAbirami, S. (2014) Antioxidant Activity and Phytochemical Analysis of Fractionated Leaf Extracts of *Catharanthus roseus*. *International Journal of Pharmaceutics*, **1**, 138-143.
- [31] Dhruvi, M., Bhavika, P. and Meonis, P. (2016) Studies on Phytochemical Constituents and Antioxidant Activity of *Alstonia scholaris*. *International Journal of Life Science*, **4**, 529-538
- [32] Chatim, Devil's Tree, *Alstonia scholaris*—Flora of Bangladesh. <http://www.floraofbangladesh.com/2016/12/chatim-devils-tree-alstonia-scholaris.html>
- [33] Eidi, A., Eidi, M. and Esmaili, E. (2006) Antidiabetic Effect of Garlic (*Allium sativum* L.) in Normal and Streptozotocin-Induced Diabetic Rats. *Phytomedicine*, **13**, 624-629. <https://doi.org/10.1016/j.phymed.2005.09.010>

- [34] Bnouham, M., Ziyat, A., Mekhfi, H., Tahri, A. and Legssyer, A. (2006) Medicinal Plants with Potential Antidiabetic Activity—A Review of Ten Years of Herbal Medicine Research (1990-2000). *International Journal of Diabetes and Metabolism*, **14**, 1-25. <https://doi.org/10.1159/000497588>
- [35] Ayodele, E.T., Hudson, H.R., Ojo, I.A. and Pianka, M. (2000) Organosulfur Compounds as Potential Fungicides: The Preparation and Properties of Some Substituted Benzyl 2-Hydroxyethyl Oligosulfides. *Phosphorus, Sulfur, and Silicon and the Related Elements*, **159**, 123-142. <https://doi.org/10.1080/10426500008043656>
- [36] Rahman, M.M., Mishuk, A., Halder, S. and Rouf, A.S.S. (2013) A Survey on Traditional Medicinal Plants Used for the Treatment of Diabetes in Urban Areas of Dhaka and Khulna, Bangladesh. *Global Journal of Medical Research: (B) Pharma, Drug Discovery, Toxicology & Medicine*, **13**, 21-26.
- [37] Gandhi, G.R., Ignacimuthu, S., Paulraj, M.G. and Sasikumar, P. (2011) Antihyperglycemic Activity and Antidiabetic Effect of Methyl Caffeate Isolated from *Solanum torvum* Swartz. Fruit in Streptozotocin Induced Diabetic Rats. *European Journal of Pharmacology*, **670**, 623-631. <https://doi.org/10.1016/j.ejphar.2011.09.159>
- [38] Biswas, K.R., Ishika, T., Rahman, M., Swarna, A., Khan, T., Monalisa, M.N. and Rahmatullah, M. (2011) Antidiabetic Plants and Formulations Used by Folk Medicinal Practitioners of Two Villages in Narail and Chuadanga Districts, Bangladesh. *American Eurasian Journal of Sustainable Agriculture*, **5**, 158-167.
- [39] Udayakumar, R., Kasthuriangan, S., Mariashibu, T.S., Rajesh, M., Anbazhagan, V.R., Kim, S.C. and Choi, C.W. (2009) Hypoglycaemic and Hypolipidaemic Effects of *Withania somnifera* Root and Leaf Extracts on Alloxan-Induced Diabetic Rats. *International Journal of Molecular Sciences*, **10**, 2367-2382. <https://doi.org/10.3390/ijms10052367>
- [40] Shobha, G., Soumya, C., Shashidhara, K.S. and Moses, V. (2014) Phytochemical Profile, Antibacterial and Antidiabetic Effects of Crude Aqueous Leaf Extract of *Datura stramonium*. *Pharmacophore*, **5**, 273-278.
- [41] Rahmatullah, M., Hasan, A., Parvin, W., Moniruzzaman, M., Khatun, A., Khatun, Z. and Jahan, R. (2012) Medicinal Plants and Formulations Used by the Soren Clan of the Santal Tribe in Rajshahi District, Bangladesh for Treatment of Various Ailments. *African Journal of Traditional, Complementary and Alternative Medicines*, **9**, 350-359. <https://doi.org/10.4314/ajtcam.v9i3.8>
- [42] Islam, M.S. and Choi, H. (2008) Dietary Red Chilli (*Capsicum frutescens* L.) Is Insulinotropic Rather than Hypoglycemic in Type 2 Diabetes Model of Rats. *Phytotherapy Research*, **22**, 1025-1029. <https://doi.org/10.1002/ptr.2417>
- [43] Mohammed, A., Koorbanally, N. and Md, S.I. (2017) Anti-Diabetic Effect of *Capsicum annum* L. Fruit Acetone Fraction in a Type 2 Diabetes Model of Rats. *Acta Poloniae Pharmaceutica*, **74**, 1767-1779.
- [44] Kaleem, M., Asif, M., Ahmed, Q.U. and Bano, B. (2006) Antidiabetic and Antioxidant Activity of *Annona squamosa* Extract in Streptozotocin-Induced Diabetic Rats. *Singapore Medical Journal*, **47**, 670-675. <https://doi.org/10.31254/jahm.2018.4108>
- [45] Hasan, T. and Sultana, M. (2018) Antidiabetic Potency of Bangladeshi Medicinal Plants. *Journal of Ayurvedic and Herbal Medicine*, **4**, 35-42. <https://doi.org/10.1016/j.heliyon.2020.e05413>
- [46] Badmus, J.A., Oyemomi, S.A., Adedosu, O.T., Yekeen, T.A., Azeez, M.A., Adebayo, E.A. and Marnewick, J.L. (2020) Photo-Assisted Bio-Fabrication of Silver Nanoparticles Using *Annona muricata* Leaf Extract: Exploring the Antioxidant, Anti-Diabetic, Antimicrobial, and Cytotoxic Activities. *Heliyon*, **6**, e05413.

- [47] Zhang, X.F. and Tan, B.H. (2000) Anti-Diabetic Property of Ethanolic Extract of *Andrographis paniculata* in Streptozotocin-Diabetic Rats. *Acta Pharmacologica Sinica*, **21**, 1157-1164.
- [48] Dandu, A.M. and Inamdar, N.M. (2009) Evaluation of Beneficial Effects of Antioxidant Properties of Aqueous Leaf Extract of *Andrographis paniculata* in STZ-Induced Diabetes. *Pakistan Journal of Pharmaceutical Sciences*, **22**, 49-52.
- [49] Anand, P., Murali, K.Y., Tandon, V., Murthy, P.S. and Chandra, R. (2010) Insulinotropic Effect of Cinnamaldehyde on Transcriptional Regulation of Pyruvate Kinase, Phosphoenolpyruvate Carboxykinase, and GLUT4 Translocation in Experimental Diabetic Rats. *Chemico-Biological Interactions*, **186**, 72-81.
<https://doi.org/10.1016/j.cbi.2010.03.044>
- [50] Kuroda, M., Mimaki, Y., Nishiyama, T., Mae, T., Kishida, H., Tsukagawa, M. and Kitahara, M. (2005) Hypoglycemic Effects of Turmeric (*Curcuma longa* L. Rhizomes) on Genetically Diabetic KK-Ay Mice. *Biological and Pharmaceutical Bulletin*, **28**, 937-939. <https://doi.org/10.1248/bpb.28.937>
- [51] Maiti, R., Das, U.K. and Ghosh, D. (2005) Attenuation of Hyperglycemia and Hyperlipidemia in Streptozotocin-Induced Diabetic Rats by Aqueous Extract of Seed of *Tamarindus indica*. *Biological and Pharmaceutical Bulletin*, **28**, 1172-1176.
<https://doi.org/10.1248/bpb.28.1172>
- [52] Muchandi, A.A., Jadhav, A.S., Patil, S.B., Patil, S.A. and Jadhav, N.B. (2018) Antioxidant and *in Vitro* Antidiabetic Activity of Methanol Extract of *Piper cubeba* L. *International Research Journal of Pharmacy and Medical Sciences*, **1**, 1-4.
- [53] Srivastava, P. (2014) Therapeutic Potential of *Piper longum* L. for Disease Management—A Review. *International Journal of Pharma Sciences*, **4**, 692-696.
- [54] Nabi, S.A., Ali, M.S., Natava, R., Tilak, T.K., Rao, C.A. and Rao, C.A. (2012) Antidiabetic and Antioxidant Activities of *Piper longum* Root Aqueous Extract in STZ Induced Diabetic Rats. *Journal of Pharmacy and Chemistry*, **6**, 30-35.
- [55] Arambewela, L.S.R., Arawwawala, L.D.A.M. and Ratnasooriya, W.D. (2005) Antidiabetic Activities of Aqueous and Ethanolic Extracts of *Piper betle* Leaves in Rats. *Journal of Ethnopharmacology*, **102**, 239-245.
<https://doi.org/10.1016/j.jep.2005.06.016>
- [56] Rahmatullah, M., Mollik, M.H., Ahmed, M.N., Bhuiyan, M.A., Hossain, M.M., Azam, M.K. and Jahan, R. (2010) A Survey of Medicinal Plants Used by Folk Medicinal Practitioners in Two Villages of Tangail District, Bangladesh. *American Eurasian Journal of Sustainable Agriculture*, **4**, 357-362.
- [57] Edwin, S., Jarald, E.E., Deb, L., Jain, A., Kingler, H., Dutt, K.R. and Raj, A.A. (2008) Wound Healing and Antioxidant Activity of *Achyranthes aspera*. *Pharmaceutical Biology*, **46**, 824-828. <https://doi.org/10.1080/13880200802366645>
- [58] Kumar, A., Gnananath, K., Gande, S., Goud, E., Rajesh, P. and Nagarjuna, S. (2011) Anti-Diabetic Activity of Ethanolic Extract of *Achyranthes aspera* Leaves in Streptozotocin Induced Diabetic Rats. *Journal of Pharmacy Research*, **4**, 3124-3125.
- [59] Kim, H.K., Kim, M.J., Cho, H.Y., Kim, E.K. and Shin, D.H. (2006) Antioxidative and Anti-Diabetic Effects of Amaranth (*Amaranthus esculantus*) in Streptozotocin-Induced Diabetic Rats. *Cell Biochemistry and Function: Cellular Biochemistry and Its Modulation by Active Agents or Disease*, **24**, 195-199.
<https://doi.org/10.1002/cbf.1210>
- [60] Khanal, D.P., Raut, B. and Dangol, K.S. (2015) Phytochemical Screening, Pharmacognostic Evaluation and Biological Activity of *Amaranthus spinosus* L. *Journal of Manmohan Memorial Institute of Health Sciences*, **1**, 29-34.

<https://doi.org/10.3126/jmmihs.v1i4.11999>

- [61] Bavarva, J.H. and Narasimhacharya, A.V. (2013) Systematic Study to Evaluate Anti-Diabetic Potential of *Amaranthus spinosus* on Type-1 and Type-2 Diabetes. *Cellular and Molecular Biology*, **59**, 1818-1825.
- [62] Pari, L. and Latha, M. (2005) Effect on Lipid Peroxidation in Streptozotocin Diabetes. *General Physiology and Biophysics*, **24**, 13-26.
- [63] Minaiyan, M., Ghannadi, A., Movahedian, A. and Hakim-Elahi, I. (2014) Effect of *Hordeum vulgare* L. (Barley) on Blood Glucose Levels of Normal and STZ-Induced Diabetic Rats. *Research in Pharmaceutical Sciences*, **9**, 173-178.
- [64] Ahmad, M., Akhtar, M.S., Malik, T. and Gilani, A.H. (2000) Hypoglycaemic Action of the Flavonoid Fraction of *Cuminum nigrum* Seeds. *Phytotherapy Research: An International Journal Devoted to Pharmacological and Toxicological Evaluation of Natural Product Derivatives*, **14**, 103-106.
[https://doi.org/10.1002/\(SICI\)1099-1573\(200003\)14:2<103::AID-PTR578>3.0.CO;2-P](https://doi.org/10.1002/(SICI)1099-1573(200003)14:2<103::AID-PTR578>3.0.CO;2-P)
- [65] Dey, P., Singh, J., Suluvoy, J.K., Dilip, K.J. and Nayak, J. (2020) Utilization of *Swerdia chirayita* Plant Extracts for Management of Diabetes and Associated Disorders: Present Status, Future Prospects and Limitations. *Natural Products and Bioprospecting*, **10**, 431-443. <https://doi.org/10.1007/s13659-020-00277-7>
- [66] Jahan, R., Al-Nahain, A., Majumder, S. and Rahmatullah, M. (2014) Ethnopharmacological Significance of *Eclipta alba* (L.) Hassk. (Asteraceae). *International Scholarly Research Notices*, **2014**, Article ID: 385969.
<https://doi.org/10.1155/2014/385969>
- [67] Vijay, P. (2015) Anti-Diabetic Effects of *Eclipta alba* on Alloxan-Induced Diabetic Mice. *International Journal of Pharmaceutical Sciences and Research*, **6**, 308-314.
- [68] Sharma, S.B., Nasir, A., Prabhu, K.M. and Murthy, P.S. (2006) Antihyperglycemic Effect of the Fruit-Pulp of *Eugenia jambolana* in Experimental Diabetes Mellitus. *Journal of Ethnopharmacology*, **104**, 367-373.
<https://doi.org/10.1016/j.jep.2005.10.033>
- [69] Delfita, R., Tjong, D.H., Dahelmi, D. and Suhatri, S. (2021) Hypoglycemic Effects of *Enhydra fluctuans* Aerial Extract on Alloxan-Induced Diabetic Rats. *Journal of Physics: Conference Series*, **1940**, Article ID: 012058.
<https://doi.org/10.1088/1742-6596/1940/1/012058>
- [70] Ali, R., Billah, M., Hassan, M. and Dewan, S.M.R. (2013) *Enhydra fluctuans* Lour: A Review. *Research Journal of Pharmacy and Technology*, **6**, 927-929.
- [71] Rath, D., Panigrahy, S.R., Panigrahi, S.K., Kar, D.M. and Maharana, L. (2017) Anti-diabetic Effect of Extracts of *Blumea lacera* dc. in Streptozotocin Induced Hyperglycemic Rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, **9**, 218-223. <https://doi.org/10.22159/ijpps.2017v9i10.19851>
- [72] Arika, W.M., Nyamai, D.W., Agyirifo, D.S., Ngugi, M.P. and Njagi, E.N.M. (2016) *In Vivo* Antidiabetic Effect of Aqueous Leaf Extract of *Azadirachta indica*, A. *juss* in Alloxan Induced Diabetic Mice. *Journal of Diabetic Complications and Medicine*, **1**, Article ID: 1000106.
- [73] Panda, S.P., Haldar, P.K., Bera, S., Adhikary, S. and Kandar, C.C. (2010) Antidiabetic and Antioxidant Activity of *Swietenia mahagoni* in Streptozotocin-Induced Diabetic Rats. *Pharmaceutical Biology*, **48**, 974-979.
<https://doi.org/10.3109/13880200903390051>
- [74] Sutradhar, A., Sarkar, A.P., Saleh, M.A., Mondal, M., Wahed, T.B., Ahmed, S. and Kundu, S.K. (2018) Investigation of Antidiabetic and Antilipidemic Effect of Fruit

- Extract of *Spondias pinnata* (Amra) in Alloxan Induced Hyperglycemic Rats. *Journal of Pharmacognosy and Phytochemistry*, **7**, 2785-2789.
- [75] Aderibigbe, A.O., Emudianughe, T.S. and Lawal, B.A.S. (2001) Evaluation of the Antidiabetic Action of *Mangifera indica* in Mice. *Phytotherapy Research*, **15**, 456-458. <https://doi.org/10.1002/ptr.859>
- [76] Liu, I.M., Liou, S.S., Lan, T.W., Hsu, F.L. and Cheng, J.T. (2005) Myricetin as the Active Principle of *Abelmoschus moschatus* to Lower Plasma Glucose in Streptozotocin-Induced Diabetic Rats. *Planta Medica*, **71**, 617-621. <https://doi.org/10.1055/s-2005-871266>
- [77] Kang, M.J., Kim, J.I., Yoon, S.Y., Kim, J.C. and Cha, I.J. (2006) Pinitol from Soybeans Reduces Postprandial Blood Glucose in Patients with Type 2 Diabetes Mellitus. *Journal of Medicinal Food*, **9**, 182-186. <https://doi.org/10.1089/jmf.2006.9.182>
- [78] Bhavsar, S.K. (2017) Evaluation of Phytochemical Profile and Antidiabetic Activity of *Ficus racemosa* (Linn.) Stem Bark in Rats. *Indian Drugs*, **54**, 49-54. <https://doi.org/10.53879/id.54.01.10279>
- [79] Murugesu, S., Selamat, J. and Perumal, V. (2021) Phytochemistry, Pharmacological Properties, and Recent Applications of *Ficus benghalensis* and *Ficus religiosa*. *Plants*, **10**, Article No. 2749. <https://doi.org/10.3390/plants10122749>
- [80] Uddin, M.Z. and Hassan, M.A. (2016) Plant Diversity of Dhaka University Campus, Bangladesh. *Journal of the Asiatic Society of Bangladesh, Science*, **42**, 49-68. <https://doi.org/10.3329/jasbs.v42i1.31750>
- [81] Singh, R.K., Mehta, S., Jaiswal, D., Rai, P.K. and Watal, G. (2009) Antidiabetic Effect of *Ficus bengalensis* Aerial Roots in Experimental Animals. *Journal of Ethnopharmacology*, **123**, 110-114. <https://doi.org/10.1016/j.jep.2009.02.017>
- [82] Gupta, R., Mathur, M., Bajaj, V.K., Katariya, P., Yadav, S., Kamal, R. and Gupta, R.S. (2012) Evaluation of Antidiabetic and Antioxidant Activity of *Moringa oleifera* in Experimental Diabetes. *Journal of Diabetes*, **4**, 164-171. <https://doi.org/10.1111/j.1753-0407.2011.00173.x>
- [83] Chandrasegaran, G., Elanchezhian, C., Suhasini, S. and Babby, A. (2014) Antihyperglycemic Activity of *Mentha piperita* Ethanol Leaves Extract on Streptozotocin Induced Diabetic Rats. *International Journal for Pharmaceutical Research Scholars (IJPRS)*, **3**, 113-117.
- [84] Khan, M.R.I., Islam, M.A., Hossain, M.S., Asadujjaman, M., Wahed, M.I.I., Rahman, B.M. and Ahmed, M. (2010) Antidiabetic Effects of the Different Fractions of Ethanolic Extracts of *Ocimum sanctum* in Normal and Alloxan Induced Diabetic Rats. *Journal of Scientific Research*, **2**, 158-168. <https://doi.org/10.3329/jsr.v2i1.2769>
- [85] Solikhah, T.I., Setiawan, B. and Ismukada, D.R. (2020) Antidiabetic Activity of Papaya Leaf Extract (*Carica papaya* L.) Isolated with Maceration Method in AlloxanInduces Diabetic Mice. *Systematic Reviews in Pharmacy*, **11**, 774-778. <https://www.sysrevpharm.org/articles/antidiabetic-activity-of-papaya-leaf-extract-carica-papaya-l-isolated-with-maceration-method-in-alloxaninduces-diabetic-.pdf>
- [86] Lakshmi, V., Agarwal, S.K., Ansari, J.A., Mahdi, A.A. and Srivastava, A.K. (2014) Antidiabetic Potential of *Musa paradisiaca* in Streptozotocin-Induced Diabetic Rats. *The Journal of Phytopharmacology*, **3**, 77-81. <https://doi.org/10.31254/phyto.2014.3201>
- [87] Uddin, M.Z., Hassan, M.A., Rahman, M. and Arefin, K. (2012) Ethno-Medico-Botanical Study in Lawachara National Park, Bangladesh. *Bangladesh Journal of Botany*, **41**, 97-104. <https://doi.org/10.3329/bjb.v41i1.11087>
- [88] Mehta, S., Singh, R.K., Jaiswal, D., Rai, P.K. and Watal, G. (2009) Anti-Diabetic Ac-

- tivity of *Emblica officinalis* in Animal Models. *Pharmaceutical Biology*, **47**, 1050-1055. <https://doi.org/10.3109/13880200902991532>
- [89] Rahaman, M.M., Hasan, M.M., Badal, I.H., Swarna, A., Rahman, S. and Rahmatulla, M. (2014) A Preliminary Antihyperglycemic and Antinociceptive Activity Evaluation of *Amorphophallus campanulatus* Corms. *International Journal of Pharmacy and Pharmaceutical Sciences*, **6**, 613-616.
- [90] Rahman, M., Hossain, M.A., Siddique, S.A., Biplab, K.P. and Uddin, M.H. (2012) Antihyperglycemic, Antioxidant and Cytotoxic Activities of *Alocasia macrorrhizos* (Linn.) Rhizomes Extract. *Turkish Journal of Biology*, **36**, 574-579. <https://doi.org/10.3906/biy-1112-11>
- [91] Nagare, S., Deokar, G.S., Nagare, R. and Phad, N. (2015) Review on *Coccinia grandis* (L.) voigt (Ivy Gourd). *World Journal of Pharmaceutical Research*, **4910**, 728-743.
- [92] Islam, M.S., Parvin, S., Uddin, M.N. and Mazid, M.A. (2014) Antidiabetic and Antioxidant Activities of Decoctions of *Coccinia grandis* Linn. and *Centella asiatica* (L.) on Alloxan-Induced Diabetic Rats. *Bangladesh Pharmaceutical Journal*, **17**, 86-91. <https://doi.org/10.3329/bpj.v17i1.22322>
- [93] Gupta, R., Katariya, P., Mathur, M., Bajaj, V.K., Yadav, S., Kamal, R. and Gupta, R.S. (2011) Antidiabetic and Renoprotective Activity of *Momordica dioica* in Diabetic Rats. *Diabetologia Croatica*, **40**, 81-88.
- [94] Jha, D.K., Koneri, R. and Samaddar, S. (2017) Potential Bio-Resources of *Momordica dioica* Roxb: A Review. *International Journal of Pharmaceutical Sciences Review and Research*, **45**, 203-209.
- [95] Bulbul, I.J., Zulfiker, A.H.M., Hamid, K., Khatun, M.H. and Begum, Y. (2011) Comparative Study of *in Vitro* Antioxidant, Antibacterial and Cytotoxic Activity of Two Bangladeshi Medicinal Plants—*Luffa cylindrica* L. and *Luffa acutangula*. *Pharmacognosy Journal*, **3**, 59-66. <https://doi.org/10.5530/pj.2011.23.9>
- [96] Hazra, M., KunduSen, S., Bhattacharya, S., Halder, P.K., Gupta, M. and Mazumder, U.K. (2011) Evaluation of Hypoglycemic and Antihyperglycemic Effects of *Luffa cylindrica* Fruit Extract in Rats. *Journal of Advanced Pharmacy Education and Research*, **2**, 138-146. <https://doi.org/10.5402/2011/869273>
- [97] Pimple, B.P., Kadam, P.V. and Patil, M.J. (2011) Antidiabetic and Antihyperlipidemic Activity of *Luffa acutangula* Fruit Extracts in Streptozotocin Induced NIDDM Rats. *Asian Journal of Pharmaceutical and Clinical Research*, **4**, 156-163.
- [98] Yasmin, A., Bharathi, R.V. and Radha, R. (2019) Review Article on *Luffa acutangula* (L) Roxb. *Research Journal of Pharmacy and Technology*, **12**, 2553-2558. <https://doi.org/10.5958/0974-360X.2019.00429.3>
- [99] Islam, A.R., Das, S.K., Alam, M.F. and Rahman, A.H.M.M. (2019) Documentation of Wild Edible Minor Fruits Used by the Local People of Barishal, Bangladesh with Emphasis on Traditional Medicinal Values. *Journal of Bio-Science*, **27**, 69-81. <https://doi.org/10.3329/jbs.v27i0.44672>
- [100] Amritha, M.S. and Santhosh, C. (2021) Phytochemical and Pharmacological Profiling of Aghori (*Flacourtia indica* (Burm. f.) Merr)—An Exploration of the Evidence of a Potent Folklore Medicine. *Journal of Research in Indian Medicine*, **16**, 1-8.
- [101] Chaubey, A. and Dubey, A.K. (2018) Phytochemical Profiling and Antioxidant Activity of Aqueous Extract of *Aegle marmelos* Fruit Shell. *MOL2NET 2018, International Conference on Multidisciplinary Sciences*, Paris, 15 January 2018-20 January 2019, 1-4. <https://doi.org/10.3390/mol2net-04-05432>
- [102] Sarkar, B.K., Barman, S.K., Akhter, S., Akter, R., Das, J., Sarkar, A.P. and Kundu,

- S.K. (2019) Evaluation of *in Vitro* Anti Diabetic Activity of Two Mangrove Plant Extracts: *Heritiera fomes* and *Sonneratia apetala*. *Journal of Pharmacognosy and Phytochemistry*, **8**, 2376-2380.
- [103] Mahmud, I., Islam, M.K., Saha, S., Barman, A.K., Rahman, M.M., Anisuzzman, M. and Rahmatullah, M. (2014) Pharmacological and Ethnomedicinal Overview of *Heritiera fomes*: Future Prospects. *International Scholarly Research Notices*, **2014**, Article ID: 938543. <https://doi.org/10.1155/2014/938543>
- [104] Vadivelan, R., Dipanjan, M., Umasankar, P., Dhanabal, S.P., Satishkumar, M.N., Antony, S. and Elango, K. (2011) Hypoglycemic, Antioxidant and Hypolipidemic Activity of *Asparagus racemosus* on Streptozotocin-Induced Diabetic in Rats. *Advances in Applied Science Research*, **2**, 179-185.
- [105] Singh, S.K., Kesari, A.N., Gupta, R.K., Jaiswal, D. and Watal, G. (2007) Assessment of Antidiabetic Potential of *Cynodon dactylon* Extract in Streptozotocin Diabetic Rats. *Journal of Ethnopharmacology*, **114**, 174-179. <https://doi.org/10.1016/j.jep.2007.07.039>
- [106] Nitu, S.K., Islam, S.M.S. and Tarique, M.H. (2019) Morphological Characteristics of Different Accessions of *Cynodon dactylon* (L.) Pers. and Physico-Chemical Properties of Soil of Their Growing Region in Bangladesh. *International Journal of Biosciences*, **15**, 350-369.
- [107] Morshed, A., Hossain, M.H., Shakil, S., Nahar, K., Rahman, S., Ferdousi, D. and Rahmatullah, M. (2010) Evaluation of Antinociceptive Activity of Two Bangladeshi Medicinal Plants, *Kalanchoe pinnata* (Lam.) Pers. and *Lagerstroemia speciosa* (L.) Pers. *Advances in Natural and Applied Sciences*, **4**, 193-197.
- [108] Roh, S.G., Kim, K.H. and Choi, W.C. (2009) Antidiabetic Effects of Leaves Extracts of *Psidium guajava* L. and *Lagerstroemia speciosa* L. in STZ-Induced Rats. *Journal of Life Science*, **19**, 40-45. <https://doi.org/10.5352/JLS.2009.19.1.040>
- [109] Sutar, U.N. and Behera, B.C. (2009) Antidiabetic Activity of the Leaves of *Mimosa pudica* Linn. in Albino Rats. *Journal of Herbal Medicine and Toxicology*, **3**, 123-126.
- [110] Khatun, A., Jannat, K., Ahamed, T., Jahan, R. and Rahmatullah, M. (2018) Some Esoteric Home Remedies Practiced in Narayanganj District, Bangladesh. *Journal of Medicinal Plants*, **6**, 166-168.
- [111] Premanath, R., Lakshmidivi, N., Jayashree, K. and Suresh, R.N. (2012) Evaluation of Anti-Diabetic Effect of *Trigonella foenum graecum* Linn. Leaf Extract in Streptozotocin Induced Diabetic Rats. *International Journal of Diabetes in Developing Countries*, **32**, 138-144. <https://doi.org/10.1007/s13410-012-0081-3>
- [112] Pandhare, R.B., Sangameswaran, B., Mohite, P.B. and Khanage, S.G. (2011) Anti-diabetic Activity of Aqueous Leaves Extract of *Sesbania sesban* (L) Merr. in Streptozotocin Induced Diabetic Rats. *Avicenna Journal of Medical Biotechnology*, **3**, 37-43.
- [113] Gomase, P.V. (2012) *Sesbania sesban* Linn: A Review on Its Ethnobotany, Phytochemical and Pharmacological Profile. *Asian Journal of Biomedical and Pharmaceutical Sciences*, **2**, 11-14.
- [114] Kumar, A., Lingadurai, S., Shrivastava, T.P., Bhattacharya, S. and Haldar, P.K. (2011) Hypoglycemic Activity of *Erythrina variegata* Leaf in Streptozotocin-Induced Diabetic Rats. *Pharmaceutical Biology*, **49**, 577-582. <https://doi.org/10.3109/13880209.2010.529615>
- [115] Kumar, A., Lingadurai, S., Jain, A. and Barman, N.R. (2010) *Erythrina variegata* Linn: A Review on Morphology, Phytochemistry, and Pharmacological Aspects.

- Pharmacognosy Reviews*, **4**, 147-152. <https://doi.org/10.4103/0973-7847.70908>
- [116] Sivakumar, V. and Rajan, M.D. (2011) Hypoglycemic and Antioxidant Activity of *Tinospora cordifolia* in Experimental Diabetes. *International Journal of Pharmaceutical Sciences and Research*, **2**, 608-613.
- [117] Islam, R., Khan, M.A.S., Islam, M.S., Benozir, S. and Alam, M.J. (2018) Anti-Diabetic Properties of *Lannea coromandelica* L. Bark Extract on Alloxan Induced Type-2 Diabetic Rats. *European Journal of Pharmaceutical and Medical Research*, **5**, 31-38.
- [118] Amelia, P., Ivada, P.A.K., Fitriana, N., Komala, I., Bahri, S. and Hanafi, M. (2021) Antioxidant and Antimicrobial Activity of Secondary Metabolite Produced by Endophytic Fungi Isolated from *Lannea coromandelica* (Houtt.) Merr. *International Journal of Pharmaceutical Sciences and Research*, **12**, 1588-1592.
- [119] Prasanna, G., Devi, R. and Ishwarya, G. (2019) *In Vitro* Evaluation of Antidiabetic and Cytotoxicity Potentials of the Rhizome Extract of *Drynaria quercifolia* (L.) J. Smith. *Asian Journal of Pharmaceutical and Clinical Research*, **12**, 2455-3891. <https://doi.org/10.22159/ajpcr.2019.v12i11.35078>
- [120] Hossain, M.S., Islam, J., Sarkar, R. and Hossen, S.M. (2014) Antidiarrheal, Antidiabetic, Antioxidant and Antimicrobial Activity of Methanolic Extracts of Leaves of *Clerodendrum viscosum* (vent.). *International Journal of Pharmaceutics*, **7**, 449-453.
- [121] Islam, F., Rahman, M.S., Akter, R., Bakar, M.A., Abdullah, A.M. and Uddin Ahmed, N. (2012) Antidiabetic and Antioxidant Effect with Phytochemical Screening of Ethanol Extract of *Saurauia roxburghii*. *Pharma Science Monitor*, **3**, 2601-2612.
- [122] Mohammad, F.K., Zahirul, I.K., Md, R.U., Mohammad, S.R. and Mohammad, A.R. (2015) *In Vivo* Hypoglycemic and Alloxan Induced Antidiabetic Activity of *Xeromphis uliginosa* Retz. *African Journal of Pharmacy and Pharmacology*, **9**, 363-366. <https://doi.org/10.5897/AJPP2015.4293>
- [123] Laha, S. and Paul, S. (2019) *Gymnema sylvestre* (Gurmar): A Potent Herb with Anti-Diabetic and Antioxidant Potential. *Pharmacognosy Journal*, **11**, 201-206. <https://doi.org/10.5530/pj.2019.11.33>
- [124] Kumar, P., Rani, S., Arunjoyothi, B., Chakrapani, P. and Rojarani, A. (2017) Evaluation of Antidiabetic Activity of *Gymnema sylvestre* and *Andrographis paniculata* in Streptozotocin Induced Diabetic Rats. *International Journal of Pharmacognosy and Phytochemical Research*, **9**, 22-25. <https://doi.org/10.25258/ijpapr.v9i1.8034>
- [125] Goat Weed (বন ধন্যৈ ফুল)—*Scoparia dulcis*. <https://icflora.blogspot.com/2015/04/goat-weed-scoparia-dulcis.html>
- [126] Shafi, S. and Tabassum, N. (2013) Survey on Anti-Diabetic Plants in Kashmir. *Journal of Advanced Pharmacy Education & Research*, **3**, 306-318.
- [127] Ogidi, I.O., Omu, O. and Ezeagba, P.A. (2019) Ethno Pharmacologically Active Components of *Brassica juncea* (Brown Mustard) Seeds. *International Journal of Pharmaceutical Research and Development*, **1**, 9-13.
- [128] Upwar, N., Patel, R., Waseem, N. and Mahobia, N.K. (2011) Hypoglycemic Effect of Methanolic Extract of *Berberis aristata* DC Stem on Normal and Streptozotocin Induced Diabetic Rats. *International Journal of Pharmacy and Pharmaceutical Sciences*, **3**, 222-224.
- [129] Bhatt, L.R., Wagle, B., Adhikari, M., Bhusal, S., Giri, A. and Bhattarai, S. (2018) Antioxidant Activity, Total Phenolic and Flavonoid Content of *Berberis aristata* DC. and *Berberis thomsoniana* C.K. Schneid. from Sagarmatha National Park, Nepal.

- Pharmacognosy Journal*, **10**, s167-s171. <https://doi.org/10.5530/pj.2018.6s.29>
- [130] Yesmin, R., Das, P.K., Belal, H., Aktar, S., Ayesha, M.A., Rakib, M.A. and Khanam, J.A. (2019) *In Vitro* Antioxidant and Antidiabetic Assessment of Extracts from the Bark of *Michelia champaca*, a Medicinal Plant in Bangladesh. *World Journal of Pharmaceutical Research*, **8**, 1505-1526.
- [131] Jarald, E., Joshi, S.B. and Jain, D.C. (2009) Biochemical Study on the Hypoglycaemic Effects of Extract and Fraction of *Acacia catechu* willd in Alloxan-Induced Diabetic Rats. *International Journal of Diabetes and Metabolism*, **17**, 63-69. <https://doi.org/10.1159/000497675>
- [132] Stohs, S.J. and Bagchi, D. (2015) Antioxidant, Anti-Inflammatory, and Chemoprotective Properties of *Acacia catechu* Heartwood Extracts. *Phytotherapy Research*, **29**, 818-824. <https://doi.org/10.1002/ptr.5335>
- [133] Mondal, S., Bhattacharya, S. and Biswas, M. (2012) Antidiabetic Activity of *Areca catechu* Leaf Extracts against Streptozotocin Induced Diabetic Rats. *Journal of Advanced Pharmacy Education & Research*, **2**, 10-17.
- [134] Wetwitayaklung, P., Phaechamud, T., Limmatvapirat, C. and Keokitichai, S. (2013) The Study of Antioxidant Capacity in Various Parts of *Areca catechu* L. *Naresuan University Journal: Science and Technology (NUJST)*, **14**, 1-14.
- [135] Luo, Y., Peng, B., Wei, W., Tian, X. and Wu, Z. (2019) Antioxidant and Anti-Diabetic Activities of Polysaccharides from Guava Leaves. *Molecules*, **24**, Article No. 1343. <https://doi.org/10.3390/molecules24071343>
- [136] Asmat, U., Abad, K. and Ismail, K. (2016) Diabetes Mellitus and Oxidative Stress—A Concise Review. *Saudi Pharmaceutical Journal*, **24**, 547-553. <https://doi.org/10.1016/j.jsps.2015.03.013>
- [137] Gupta, M., Sharma, S., Gautam, A.K. and Bhadauria, R. (2011) *Momordica charantia* Linn. (Karela): Nature's Silent Healer. *International Journal of Pharmaceutical Sciences Review and Research*, **11**, 32-37.