

Algae from Fresh and Brackish Waters, Côte d'Ivoire

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Abstract

The aim of this work is to study the diversity of phytoplanktonic from some fresh and brackish waters. The methodology employed consisted of sampling the organisms using the Van Dorn bottle at the surface (0.5 m) of fresh and sea waters from Container Terminal from San-Pedro Port Area (CTSP) during June and July 2021 and Ebrié Lagoon in June 2023. A total of 324 taxa were collected with 274 taxa for CTSP and 82 taxa for Ebrié Lagoon. The identified phyla were, in order of importance, the Bacillariophyta (151 taxa), Dinophyta (80 taxa), Euglenophyta (34 taxa), Chlorophyta (31 taxa), Chrysophyta (12 taxa), Cyanobacteria (11 taxa), Cryptophyta and Rhodophyta with 2 taxa respectively and Ochrophyta (1 taxon). In the CTSP, the species *Prorocentrum lima* and *Prorocentrum micans* have contributed to the highest number of the taxa during the first mission, but with low contribution during the second mission. At this mission, the contribution of the Dinophyta *Proto-peridinium quinque-corne* and the Bacillariophyta *Melosira* spp., *Coscinodiscus nodulifer*, *Thalassionema frauenfeldii* and *Thalassionema nitzschoides* was revealed. The Ebrié Lagoon was characterized by the high contribution of the species *Cyclotella meneghiniana* during the sampling mission. As a recommendation, we propose the monitoring in time and space of toxin-producing species and the evaluation of their toxins that could constitute a risk for the fishery, recreational and commercial activities, and the ecosystems studied providing important ecosystem and economic services.

Keywords

Phytoplankton, Diversity, Abidjan, San-Pédro

1. Introduction

Studies of the autecology of different phytoplankton species can help in developing a greater understanding of growth niches, prediction of when blooms may

occur and potential control strategies for bloom minimization. The understanding of phytoplankton growth responses to temperature is important due to their regulating role in determining the predominance of certain species [1]. Significant proliferations of certain species occur regularly, especially in spring, sometimes forming “colored waters”, the color of the pigments of phytoplankton cells. These efflorescences or “blooms” are natural phenomena, but they are sometimes amplified by significant enrichment of the environment, mainly due to nutrient inputs from rivers and runoff. The species in question are in most cases harmless, but these phenomena sometimes have harmful consequences: indeed, the sudden decrease in available oxygen, created by the decomposition of dead phytoplankton near the bottom, can lead to anoxia, of the environment and therefore to marine animal mortalities [2].

This study, which the aim is to study the diversity of phytoplanktonic from some fresh and brackish waters, is a contribution to the work carried out on the diversity of phytoplanktonic from several Ivorian aquatic ecosystems in which the proliferation of phytoplankton was registered under extreme pollution conditions [3] [4] [5] [6] [7].

2. Material and Methods

2.1. Study Area and Sampling Stations

Both geolocalization and description of stations from the Container Terminal from San-Pedro Port Area (CTSP) were presented in [8]. Concerning Ebrié Lagoon, the geographic coordinates of study stations are presented in **Table 1**. Sampling Stations P1, P2, P3 and P4 are located in the lagoon between the EOLIS fruit terminal and the National Customs School. Sampling Stations P5, P6, P8, P9 and P10 are located in the lagoon between the ADDOHA lagoon city, the Municipal High School of Attécoubé, Abobo-Doumé Beach and the Abobo-Doumé Lagoon station. Sampling Station P7 is located between Stations P4 and P9.

Table 1. Geographic coordinates of Ebrié Lagoon sites.

Sites	Latitude Nord (ddmns)	Longitude Ouest (ddmns)
P1	5° 19'10.81"	4° 1'48.35"
P2	5° 18'59.30"	4° 1'33.88"
P3	5° 18'55.28"	4° 1'28.39"
P4	5° 18'46.97"	4° 1'17.65"
P5	5° 19'5.09"	4° 1'49.94"
P6	5° 18'56.21"	4° 1'39.34"
P7	5° 18'45.95"	4° 1'28.56"
P8	5° 18'51.02"	4° 1'42.66"
P9	5° 18'42.42"	4° 1'39.83"
P10	5° 18'33.86"	4° 1'33.52"

2.2. Sampling and Study of Phytoplankton Community

The samples used for the study of phytoplankton were obtained 1) from the remaining volume of sample taken using the Van Dorn bottle for the analysis of nutrient salts in Stations S1, S2, S3, S4, S5, S6, S7, S8, S9 and S10 between 8 a.m. and 3 p.m. for the Container Terminal from San-Pedro Port Area (CTSP) and 2) using a plastic bottle of 1-liter capacity at 0.5 m from the lagoon water body in 10 sampling Stations P1, P2, P3, P4, P5, P6, P7, P8, P9 and P10 between 8 a.m. and 3 p.m. The TCSP samples were filtered through a 20 μm diameter plankton net, stored in 100 mL pillboxes and then fixed in 5% formalin for subsequent analysis in the laboratory. In total, 27 samples including 17 samples for the TCSP and 10 samples for the Ebrié Lagoon were collected from the surface water sampling. Observations and counting of organisms were carried out using an Optika-type inverted microscope. The observed taxa were photographed using a Sony Cyber-shot DSC-W800 type camera. The methodology of [9] was used for the preparation of diatoms. The identification of the taxa observed was carried out using general and specific identification books and keys.

Concerning the CTSP samples, the counting of all the organisms was made using the sedimentation cups of volume 3.4 mL, 5 mL, 10 mL, and 25 mL from the inverted microscope according to the method of [10] modified (standard NF EN 15204) by [11]. Replica counts of organisms were carried out on all samples except for samples from Stations S1, S2, S3, S5, S6 and S7 of the second mission. For this mission, 3.4 mL sedimentation cups were used for the samples from Stations S1 and S6, those of 10 mL were used for the samples from Stations S2, S5 and S7. The samples from the Station S3 were counted using the 5 mL volume sedimentation dish. Counting replicas of samples from the first mission were made with 3.4 mL and 5 mL sedimentation cups for samples from Stations S2, S3, S7, S9 and S10. The 5 mL and 10 mL sedimentation cups were used for the samples from Stations S1 and S4. The samples from Stations S5, S6 and S8 were counted using 5 mL and 25 mL sedimentation cups. The sample from the Station S4 of the second mission was counted with 3.4 mL and 25 mL cups. The samples from the Ebrié Lagoon, sub-samples were made in Petri dishes for the observation and counting of the taxa contained in 1 liter of water sample.

The results are expressed in number of individuals per milliliter for the CTSP samples and per liter for the samples from the Ebrié Lagoon.

3. Results and Discussion

3.1. Results

A total of 324 taxa were collected from fresh and brackish waters studied (**Table 2**). The Container Terminal from San-Pedro Port Area (CTSP) was characterized by 274 taxa while Ebrié Lagoon was represented by 82 taxa belonging to Cyanobacteria (11 taxa), Euglenophyta (34 taxa), Chlorophyta (31 taxa), Dinophyta (80 taxa), Cryptophyta (2 taxa), Chrysophyta (12 taxa), Rhodophyta (2 taxa), Ochrophyta (1 taxon) and Bacillariophyta (151 taxa). The genus contributing to

Table 2. Phytoplanktonic list and individuals' number of taxa from Container Terminal from San-Pedro Port Area (CTSP) and Ebrié Lagoon.

Organisms	Study stations																										
	CTSP										Ebrié Lagoon																
	E		C		BL-Z		M		Em		E		C		BL-Z		P1-P8		P9-P10								
	S1	S2	S3	S4	S7	S5	S6	S8	S9	S10	S1	S2	S3	S4	S7	S5	S6	P1	P2	P3	P4	P5	P6	P7	P8	P9	P10
Cyanobacteria																											
<i>Aphanocapsa incerta</i>						10	2			2				1													
<i>Chroococcus minutus</i>									2																		
<i>Coelomonon pusillum</i>																									1		
<i>Limnothrix</i> sp.																			1		1	1					
<i>Merismopedia tenuissima</i>				2																							
<i>Microcystis wesenbergii</i>										1																	
<i>Planktothrix agardhii</i>																						1					
<i>Oscillatoria</i> spp.														2									1				
<i>Pseudanabaena catenata</i>						1	3	1				4	12	4	1	3	3										
<i>Pseudanabaena</i> spp.					1										1	1	1										
<i>Plectonema</i> sp.																2											
Euglenophyta																											
<i>Cyclidiopsis acus</i>				3	1																						
<i>Euglena acus</i>								5	1																		
<i>Euglena erhenbergii</i>						1						2		1	4					1					1		
<i>Euglena oxyuris</i>				1			2							2	1	5											
<i>Euglena polymorpha</i>		1	1	1								1	1	1	1	9											
<i>Euglena</i> spp.															1	2											
<i>Lepocinclis ovum</i>						1		2							1	1											
<i>Lepocinclis</i> sp.									1																		
<i>Phacus curvicauda</i>					1		1																				
<i>Phacus glaber</i>							1								1												
<i>Phacus lefevrei</i>			1	1	1				2				2														
<i>Phacus platalea</i>																									1		
<i>Phacus ranula</i>							2																				
<i>Phacus suecicus</i>							2																			1	
<i>Phacus</i> spp.				2									3														
<i>Ploeotia</i> sp.									1	1				2		1											
<i>Strombomonas acuminata</i>			1		1			1								1											
<i>Strombomonas fluviatilis</i>					1		2	1		1																	
<i>Strombomonas girardiana</i>	1	1					2							1													
<i>Strombomonas gibberosa</i>						2																					
<i>Strombomonas ovalis</i>										1																	

Continued

<i>Strombomonas</i> spp.	2			1	2			1		1	1							
<i>Trachelomonas armata</i> var. <i>steinii</i>													2					
<i>Trachelomas curta</i>	1																	
<i>Trachelomonas duplex</i>	1																	
<i>Trachelomonas hispida</i>								2					1					
<i>Trachelomonas oblonga</i>	1			2	3	2	3	2	1				4	1				
<i>Trachelomonas planctonica</i>	1	3			8	4	2	2					1	1				
<i>Trachelomonas radiosa</i>	1				2			1		2								
<i>Trachelomonas scabra</i>				1														
<i>Trachelomonas similis</i>					1													
<i>Trachelomonas volvocina</i>	1	3	1	2	1	4	12	4	1			4	3	2	8	1		
<i>Trachelomonas volvocinopsis</i>	1	4		2		5	2	1				2	2					
<i>Trachelomonas</i> spp.	1		2		1	2	1			1	1	2		1	1	2		
Chlorophyta																		
<i>Actinastrum hantzschii</i>		2				2							11					
<i>Ankistrodesmus spiralis</i>							1		2									
<i>Characium angustum</i>	1							1										
<i>Coelastrum sphaericum</i>															1			
<i>Cosmarium curcubita</i>												1						
<i>Debarya</i> sp.	1					1	2	2					1					
<i>Dictyosphaerium pulchellum</i>				3		1	2											
<i>Eremosphaera viridis</i>															1			
<i>Entromorpha (Ulva) compressa</i>	1												1					
<i>Golenkiena radiata</i>				3		1												
<i>Korshikoviella</i> sp.	1	1																
<i>Monoraphidium arcuatum</i>																1		
<i>Oocystis solitaria</i>	1			2				1						3		1		
<i>Pediastrum simplex</i>									1	2								
<i>Scenedesmus acunae</i>																1	1	2
<i>Scenedesmus acutiformis</i>																1		
<i>Scenedesmus bicaudatus</i>				1			2			2						1		
<i>Scenedesmus disciformis</i>																	2	
<i>Scenedesmus obtusus</i> f. <i>alternans</i>														1				
<i>Scenedesmus opoliensis</i>							2											
<i>Scenedesmus quadricauda</i>																		2
<i>Scenedesmus quadrispina</i>																		1
<i>Spirotaenia kirchneri</i> var. <i>erythropunctata</i>												1						

Continued

<i>Teilingia granulata</i>												1
<i>Tetraedron muticum</i>				2	2							
<i>Tetraedron pentaedricum</i>				1								
<i>Tetraselmis cordiformis</i>										2	2	
<i>Tetraselmis</i> sp.												1
<i>Ulothrix fimbriata</i>												1
<i>Ulva flexuosa</i>	4		2	1	6	3	1			1	2	
<i>Ulva</i> spp.	1	1				5				2	1	
Dinophyta												
<i>Adnatosphaeridium multispinosum</i>												1
<i>Anisonema acinus</i>										1		
<i>Amphisolenia</i> sp.			1									
<i>Amphidinium</i> sp.											1	
<i>Centrodinium</i> spp.		1		1	1	1	1					
<i>Ceratium</i> cf. <i>balchii</i>		1										
<i>Ceratium contortum</i> var. <i>karstenii</i>		1	1								1	
<i>Ceratium dens</i>				1								
<i>Ceratium furca</i>					1							
<i>Ceratium fusus</i> var. <i>fuscus</i>				2								
<i>Ceratium fusus</i> var. <i>seta</i>			1									
<i>Ceratium lunula</i>		3		3	2		1					
<i>Ceratium massiliense</i>		1										
<i>Ceratium minutum</i>										1		
<i>Ceratium pentagonum</i>		2	4	2	1		1			1		
<i>Ceratium trichoceros</i>		3	4		1		1					
<i>Ceratium tripos</i> var. <i>atlanticum</i>		1		1	1							
<i>Ceratium</i> spp.		7	2		2					4	2	1
<i>Cochlodinium strangulatum</i>										1		
<i>Cochlodinium</i> cf. <i>strangulatum</i>												1
<i>Cochlodinium</i> spp.	1									1	1	2
<i>Cordosphaeridium</i> cf. <i>minimum</i>												1
<i>Diplocystis antarctica</i>		1		1					1	12	4	1
<i>Dinophysys caudata</i>		1		2			1					
<i>Ebria</i> spp.	2	1	3	1	3	1						5
<i>Glenodinium lenticula</i>	1				1	16	1	1		2	4	
<i>Gonyaulax</i> spp.					1					2	1	1
<i>Gymnodinium pseudonoctiluca</i>												1
<i>Gymnodinium</i> spp.				4		2	3	1	2	1		

Continued

<i>Gyrodinium instriatum</i>									1	1						
<i>Gyrodinium lachryma</i>									1			1				
<i>Gyrodinium</i> spp.			4	1	1							3	1			
<i>Heteraulacus acuminatus</i>						1										
<i>Heterocapsa triquetra</i>									2							
<i>Heterodinium leiorhynchus</i>	1	1				3						1				
<i>Heterodinium</i> spp.	2	5	4	3		3			1		2	2	5	3	6	16
<i>Histioneis</i> sp.															1	
<i>Kenleyia</i> sp.															1	1
<i>Kofooidinium</i> spp.	4	3	2	7	2	2	2	1	1	2	3	5	6	2	10	11
<i>Ornithocercus</i> spp.	1	1			1	7									3	
<i>Ostreopsis</i> sp.	1															1
<i>Oxytoxum</i> spp.						1	2					4		3		
<i>Peridiniopsis cunningtoni</i>																1
<i>Peridinium cinctum</i>	1		1	8	2		14	14	3	1						
<i>Peridinium umbulatum</i>	1	1			2		1					1				
<i>Peridinium</i> spp.		1	2		1		1	5	1			1	3	1	4	4
<i>Phalacroma</i> sp.	1														1	
<i>Polykrikos hartmannii</i>															1	1
<i>Polykrikos</i> sp.																3
<i>Pronoctiluca pelagica</i>												1				4
<i>Prorocentrum aporum</i>	1															
<i>Prorocentrum arcuatum</i>				5	5	7		1			2					
<i>Prorocentrum</i> cf. <i>obtusidens</i>			1													
<i>Prorocentrum concavum</i>	2															
<i>Prorocentrum compressum</i>	1															
<i>Prorocentrum gracile</i>		2	1	6		17	3		1						5	
<i>Prorocentrum lima</i>						36	4	3								
<i>Prorocentrum micans</i>		1	6	3	22	23	1	2	11	2		2	2	1	13	6
<i>Prorocentrum oblongum</i>				3												
<i>Prorocentrum robustum</i>			1													
<i>Prorocentrum schillerii</i>					3											
<i>Prorocentrum triestinum</i>																1
<i>Prorocentrum</i> spp.	9	6		2	15	10		2	4		3		1	5	3	
<i>Protoberidinium depressum</i>				1		5		4				2				
<i>Protoberidinium minutum</i>	1				1						2					
<i>Protoberidinium oviforme</i>																1
<i>Protoberidinium pellucidum</i>	1		1												1	
<i>Protoberidinium pyrophorum</i>	2	1		1	2		2				2	1		1	3	5
<i>Protoberidinium quinquecorne</i>			5		3						4	18	7	11	3	25

Continued

<i>Protopteridinium</i> spp.						1						1	1	5
<i>Pselodinium pimum</i>														1
<i>Ptychodiscus inflatus</i>						1								
<i>Ptychodiscus noctiluca</i>						1		1						
<i>Ptychodiscus</i> spp.							1	1	2					
<i>Pyrophacus horologium</i>	1			3										2
<i>Pyrophacus</i> spp.	17	6	4	9	1	1	1	5	11	3				1
<i>Spatulodinium</i> sp.			1						1					
<i>Torodinium teredo</i>														1
<i>Warnowia</i> sp.												1		1 2
<i>Warnowia virescens</i>	2	1	3	1	16	1				4	2	3	8	
Cryptophyta														
<i>Cryptomonas ovata</i>														3
<i>Cryptomonas</i> sp.			1		3							1		4
Chrysophyta														
<i>Chrysococcus rufescens</i>			4	3		1		1	5	2	1		14	1
<i>Chrysococcus rufescens</i> f. <i>tripora</i>			5	5	1		1		1		5	1	3	1
<i>Chrysococcus triporus</i>			1		1			1	3	1		1	7	1
<i>Chrysococcus</i> spp.					1	1	1					1		
<i>Dinobryon cylindricum</i> var. <i>divergens</i>					1									
<i>Dinobryon sociale</i>					1			1						
<i>Dinobryon</i> sp.	1				1	1								
<i>Kephyrion spirale</i>	1				2	1					1	1		1
<i>Kephyrion</i> spp.	4		2		1	1			1					2
<i>Ochromonas</i> spp.						2								
<i>Mallomonas</i> spp.	2				2		2				2	2		1
<i>Pseudokephyrion</i> sp.					3	1								1
Rhodophyta														
<i>Audouinella</i> sp.					1									
<i>Polysiphonia</i> sp.												1		
Ochrophyta														
<i>Botrydium granulatum</i>									1					
Bacillariophyta														
<i>Actinopterychus senarius</i>												1		
<i>Amphiprora alata</i>													1	2
<i>Amphiprora alata</i> f. <i>minor</i>													1	
<i>Amphora coffeaeformis</i>												1		
<i>Amphora proboscidea</i>													1	

Continued

<i>Amphora</i> sp.									1											1															
<i>Asterionella formosa</i>																						1													
<i>Asterionella glacialis</i>																						1													
<i>Asterionella</i> sp.																						1													
<i>Asterolampra</i> cf. <i>vanheurcki</i>																						1													
<i>Asterolampra marylandica</i>		1																			9	1	2	3					1						
<i>Asterolampra</i> sp.																							1												
<i>Asteromphalus arachne</i>																						1													
<i>Asteromphalus heptactis</i>																								1											
<i>Aulacoseira ambigua</i>																						1		2											
<i>Aulacoseira distans</i>																														1					
<i>Aulacoseira granulata</i>		3	1	3	1	9	16	4	1													2	1			3	2	4	5	8					
<i>Aulacoseira granulata</i> var. <i>angustissima</i>																																4			
<i>Aulacoseira granulata</i> var. <i>angustissima</i> f. <i>spiralis</i>																																1	1	1	
<i>Aulacoseira islandica</i>																											1	1							
<i>Auliscus sculptus</i>																							1	1											
<i>Auliscus</i> sp.																							1												
<i>Auricula complexa</i>																							2												
<i>Auricula flabelliformis</i>																									1										
<i>Bacillaria paradoxa</i>																																		1	
<i>Bacillaria paxillifer</i>																								3											
<i>Bacillaria</i> spp.				1																			1												
<i>Bacteriastrum hyalinum</i>																											7								
<i>Bellerochea malleus</i>																								1											
<i>Bellerochea</i> spp.																							1	1	1										
<i>Biddulphia alternans</i>																									1			2	2				3	7	3
<i>Biddulphia aurita</i>																							1	1											
<i>Biddulphia aurita</i> var. <i>obtusa</i>																							1												
<i>Biddulphia mobiliensis</i>																										1									3
<i>Biddulphia regina</i>																											1							4	2
<i>Campylodiscus echemeis</i>																							1												
<i>Campylodiscus</i> sp.																							1												
<i>Catenula exigua</i> sp. nov.																											2								
<i>Cerataulina pelagica</i>																								1											
<i>Chaetoceros brevis</i>																							1			1									
<i>Chaetoceros decipiens</i>																											1								
<i>Chaetoceros didymus</i>																							1												
<i>Chaetoceros diversus</i>																											1								

Continued

<i>Chaetoceros peruvianus</i>								1											1					
<i>Chaetoceros pseudocurvisetus</i>									1	1			2	1	2									
<i>Chaetoceros</i> spp.								2		1	4													
<i>Climacodium frauenfeldianum</i>										1	6	1												
<i>Climacosphenia elongata</i>		1								1				1	1									
<i>Climacosphenia moniligera</i>										1														
<i>Cocconeis</i> spp.	1	1	5				2						2		2									
<i>Coscinodiscus centralis</i>																1			1 1 3					
<i>Coscinodiscus concinnus</i>														2		1			1					
<i>Coscinodiscus divisus</i>																			1					
<i>Coscinodiscus granii</i>															1									
<i>Coscinodiscus jonesianus</i> var. <i>commutatus</i>										1					1	2			2					
<i>Coscinodiscus nodulifer</i>		2					1	1	1	2	8		34	3										
<i>Coscinodiscus</i> sp.			1	1	1				1				1											
<i>Cyclotella meneghiniana</i>																2	1	21	5	16	35	2	4	1
<i>Cyclotella</i> sp.		2	3																	1				
<i>Cymbella</i> spp.							1								2									
<i>Detonula pumila</i>										1	2													
<i>Diatoma mesodon</i>																				2				
<i>Diatoma</i> sp.																				1				
<i>Dictyocha fibula</i>											1		2	1										
<i>Distephanus speculum</i>								1	1															
<i>Donkinia recta</i>														1	1	3		1	4	3	5	3		
<i>Encyonema</i> sp.																				1				
<i>Epithemia</i> spp.	1	1	3		2	2	3	1																
<i>Eunotia minor</i>	2						1	5					1	2	2									
<i>Eunotia serra</i>	1																							
<i>Eunotia</i> sp.		1					1	2												1				
<i>Eunotogramma</i> sp.										1														
<i>Fragilaria striatula</i>																				1				
<i>Fragilaria ulna</i>														1	1									
<i>Fragilaria</i> spp.										1				2										
<i>Gomphonema</i> sp.																				1				
<i>Grammatophora</i> cf. <i>hamulifera</i>																				1				
<i>Grammatophora oceanica</i>									1					1										
<i>Grammatophora oceanica</i> var. <i>macilenta</i>																				1				
<i>Grammatophora marina</i>															2					1				
<i>Grammatophora serpentina</i>															1	1								

Continued

<i>Grammatophora</i> spp.						1														
<i>Guinardia flaccida</i>	3	1	1	2		1	1		1	1	10					2		2		
<i>Gyrosigma</i> spp.	2			1			1	4	3	3	4	2	5	1		1				
<i>Helicotheca tamesis</i>									2					1						
<i>Hemiaulus membranaceus</i>										1						1				
<i>Isthmia</i> sp.									1		2									
<i>Lampriscus</i> sp.	2			1	1	4	9													
<i>Lauderia annulata</i>				1						1										
<i>Leptocylindrus danicus</i>	1					13	1	3									3	2	3	3
<i>Lithodesmium undulatum</i>										1	1									
<i>Mastogloia</i> sp.																1			1	
<i>Mediopyxis helysia</i>									1											
<i>Melosira nummulus</i>														1						
<i>Melosira nummuloides</i>										1										
<i>Melosira varians</i>																10	1	1	11	
<i>Melosira</i> spp.	2				1			3	5	4	22	1	20							
<i>Meridion circulare</i>																			1	
<i>Navicula distans</i>																			2	
<i>Navicula pennata</i>																			1	
<i>Navicula pupula</i>																			1	
<i>Navicula</i> sp.																			2	
<i>Neidium</i> sp.																			1	
<i>Nitzschia closterium</i>																			1	
<i>Nitzschia delicatissima</i>					1	1														
<i>Nitzschia polaris</i>										1										
<i>Odontella sinensis</i>																			1	
<i>Odontidium mesodon</i>																1			1	
<i>Pinnularia mesolpepta</i>																			1	
<i>Pinnularia</i> spp.								1	2							2		1		
<i>Plagiogramma staurophorum</i>																			1	
<i>Plagiogramma</i> sp.										1										
<i>Plagiotropis lepidoptera</i>																			1	
<i>Planktoniella sol</i>										1	1									
<i>Pleurosigma delicatulum</i>								1											1	
<i>Pleurosigma elongata</i>																			1	
<i>Pleurosigma marinum</i>																			1	
<i>Pleurosigma simonsenii</i>									1											
<i>Pleurosigma strigosum</i>																			1	
<i>Pleurosigma</i> sp.																			1	

ridinium (7 taxa) for Dinophyta, Aulacoseira (6 taxa), Biddulphia (5 taxa), Chaetoceros (7 taxa), Coscinodiscus (7 taxa), Grammatophora (6 taxa), Pleurosigma (6 taxa) and Thalassiosira (7 taxa).

The Stations S1 (100 taxa), S2 (130 taxa) and S3 (106 and 153 taxa) of site E and all stations of sites C and BL-Z except Station S7 of the site C from TCSP of the second mission were characterized by a high number of individuals taxa. The Station S8 of the site M was represented by 118 taxa at the first mission. Concerning the Ebrié Lagoon, the high number of individuals was obtained at the Station P7 with 116 taxa during sampling mission.

During the first mission of June 2021, the species contributed at the highest number in the CTSP were the Dinophyta *Prorocentrum lima* (S5: 36 individuals) and *Prorocentrum micans* (S5: 23 individuals; S7: 22 individuals). The number of the other communities contributed between 1 and 17 individuals. The second mission carried out in July 2021 was characterized by the contribution of the Dinophyta *P. quinquecorne* (S6: 25 individuals) and the Bacillariophyta *Melosira* spp. (S4: 22 individuals; S5: 20 individuals), *Coscinodiscus nodulifer* (S5: 34 individuals), *Thalassionema frauenfeldii* (S3: 21 individuals; S4: 30 individuals), *T. nitzschioides* (S2: 21 individuals). At this mission, the *P. lima* and *P. micans* contribution were lowest in the different stations. In the lagoon Ebrié, the species *Cyclotella meneghiniana* has contributed at the highest number of individuals in the Stations P4 (21 individuals) and P7 (35 individuals) during mission of June 2023.

3.2. Discussion

The study zone was characterized by 324 taxa with 274 taxa for TCSP and 82 Taxa for Ebrié Lagoon. The high number of taxa in the TCSP compared to the Ebrié Lagoon would be due on the one hand to the number of sampling campaigns and on the other hand to their sensibility to environmental conditions as varied as temperature, light, pH, current, grazing by herbivores, oxidizable organic matter, nitrate and phosphate concentrations, dissolved salts, silica content. The low number of taxa in Ebrié Lagoon would be due to its position and many activities carried out in its catchment area. Our results agree with those of [12] who indicated in their work that because of its position, the Ebrié Lagoon in Abidjan records high levels of pollution, particularly in the Banco watershed by the discharges of the countless scrubbers or “laundry washers”, liquid and solid effluents from the municipalities of Adjamé, Plateau, Attécoubé and Yopougon leading to an increase in nitrate and phosphate concentrations (organic pollution) which leads to the multiplication of microalgae. These microalgae disappear when the salinity of the water exceeds 15 mg/L.

The contribution of species *P. lima* and *P. micans* in the total number of TCSP taxa in Stations S5 and S7 could be harmful to the human and aquatic (plants and animals) populations due to the toxins they are likely to produce. According to [13] and [14], toxic or otherwise harmful phytoplankton blooms may be in-

creasing in frequency worldwide. Accumulation of phytoplankton toxins in shellfish with subsequent poisoning of humans [15] and fish kills [16] [17] are widely known. *P. lima* is a benthic and epiphytic species that can be phytoplanktonic [18] [19]. According to [20], this species produces a pale-colored resting cyst as part of its life cycle. In contrast, *P. micans* is one of the most common and diversified species in the genus *Prorocentrum*. It is a planktonic species commonly found in neritic and estuarine waters, but it is also found in oceanic environments. It is cosmopolitan in cold temperatures to tropical waters. This species is also known to tolerate very high salinity: population has been reported from hypersaline salt lagoons (>90‰) in the Caribbean islands [19]. Cells are active swimmers [19] [21]. This species forms extensive red tides in many parts of the world [22] [23]. The contribution of *Cyclotella meneghiniana* in the Ebrié Lagoon during the sampling period can be explained by its appearance in warm, nutrient-rich environments as well as low-productivity environments, however, when the salinity of the water does not exceed 15 mg/L.

4. Conclusions

Phytoplankton diversity was relatively high with 324 taxa collected in fresh and brackish waters studied. The CTSP and Ebrié Lagoon studied were characterized by 274 taxa and 82 taxa respectively. The species *Prorocentrum lima* and *Prorocentrum micans* observed in the CTSP and *Cyclotella meneghiniana* collected in the Ebrié Lagoon were contributed highly in terms of individual number.

The ecosystems studied provide important ecosystem and economic services, monitoring in time and space of toxin-producing species as well as the evaluation of their toxins that could constitute a risk for the fishery, recreational and commercial activities.

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Conflicts of Interest

The author declares no conflicts of interest regarding the publication of this paper.

References

- [1] Harris, G.P. (1986) *Phytoplankton Ecology*. Chapman and Hall Ltd., London, 384 p. <https://doi.org/10.1007/978-94-009-4081-9>
- [2] Belin, C. and Soudant, B. (2018) *Trente années d’observation des micro-algues et des toxines d’algues sur le littoral*. Librairie Quae: Des Livres au Coeur des Sciences. <https://www.quae.com/produit/1545/9782759229413/trente-annees-d-observation-des-micro-algues-et-des-toxines-d-algues-sur-le-littoral>

- [3] Ouattara, A. (2000) Premières données systématiques et écologiques du phytoplancton du lac d'Ayamé (Côte d'Ivoire). Thèse de Doctorat, Katholieke Universiteit Leuven, Belgique, 207 p.
- [4] Niamien-Ebrottié, E.J., Konan, K.F., Gnagne, T., Ouattara, A. and Ouattara, M. (2008) Etude diagnostique de l'état de pollution du système fluvio-lagunaire Aby-Bia-Tanoé (Sud-Est, Côte d'Ivoire). *Sud Sciences et Technologies*, No. 20, 1-9.
- [5] Adon, M.P., Ouattara, A. and Gourène, G. (2014) Variability of Phytoplankton Community in a Shallow Tropical Reservoir (Adzopé, Côte d'Ivoire). *Asian Academic Research Journal of Multidisciplinary*, 1, 2319-2801.
- [6] Konan, K.F., Bony, K.Y., Adon, M.P. and Potgieter, J. (2019) Appendix 17: Hydrobiological Study of the Bandama Basin in Yaoure Gold Project's Area of Influence Yaoure Gold Project, Côte d'Ivoire.
<https://perseusmining.com/wp-content/uploads/2019/11/Yaoure-ESIA-Appendix-17-Freshwater-Aquatic-Baseline.pdf>
- [7] Adon, M.P., Kotchy, Y. and Kone, T. (2021) Microorganisms as Bioindicators of Saprobiology. LAP Lambert Academic Publishing, London.
- [8] Adon, M.P. (2022) Macroalgae Diversity from San-Pedro Port Area. *European Journal of Applied Sciences*, 10, 609-613. <https://doi.org/10.14738/aivp.106.13745>
- [9] Des Clers, S., Philippe, M., Barbe, J., Mouthon, J., Faessel, B., Coste, M., Wasson, J.G. and Lafont, M. (1982) Etude des méthodes biologiques d'appréciation quantitative de la qualité des eaux. Division Qualité des Eaux Pêche et Pisciculture, Lyon.
- [10] Utermöhl, H. (1958) Zur Vervollkommung der quantitativen Phytoplankton methodik. *Mitteilungen Internationale Vereinigung für Theoretische und Angewandte Limnologie*, 9, 1-38.
- [11] Laplace-Treyture, C., Barbe, J. and Dutartre, A. (2007) Protocole standardisé d'échantillonnage, de conservation et d'observation du phytoplancton en plan d'eau. Département Milieux Aquatiques, Cemagref, 1-19.
- [12] Adingra, A.A. and Kouassi, A.M. (2011) Pollution en lagune Ebrié et ses impacts sur l'environnement et les populations riveraines.
<https://aquadocs.org/handle/1834/5812?show=full>
- [13] Smayda, T.J. (1989) Primary Production and the Global Epidemic of Phytoplankton Blooms in the Sea: A Linkage? In: Cosper, E.M., et al., Eds., *Novel Phytoplankton Blooms, Causes and Impacts of Recurrent Brown Tides and Other Unusual Blooms*, Springer, Berlin, 449-483. <https://doi.org/10.1029/CE035p0449>
- [14] Hallegraeff, G.M. (1993) A Review of Harmful Algal Blooms and Their Apparent Global Increase. *Phycologia*, 32, 79-99.
<https://doi.org/10.2216/i0031-8884-32-2-79.1>
- [15] Shumway, S.E. (1990) A Review of the Effects of Algal Blooms on Shellfish and Aquaculture. *Journal of the World Aquaculture Society*, 21, 65-105.
<https://doi.org/10.1111/j.1749-7345.1990.tb00529.x>
- [16] Steidinger, K.A. (1983) A Re-Evaluation of Toxic Dinoflagellate Biology and Ecology. *Progress in Phycological Research*, 2, 147-188.
- [17] Burkholder, J.M. and Glasgow, J.R.H.B. (1995) Interactions of a Toxic Estuarine Dinoflagellate with Microbial Predators and Prey. *Archiv für Protistenkunde*, 145, 177-188. [https://doi.org/10.1016/S0003-9365\(11\)80314-3](https://doi.org/10.1016/S0003-9365(11)80314-3)
- [18] Fukuyo, Y. (1981) Taxonomical Study on Benthic Dinoflagellates Collected in Coral Reef. *Bulletin of the Japanese Society for the Science of Fish*, 47, 967-978.
<https://doi.org/10.2331/suisan.47.967>

- [19] Steidinger, K.A. and Tangen, K. (1996) Dinoflagellates. In: Tomas, C.R., Ed., *Identifying Marine Diatoms and Dinoflagellates*, Academic Press, New York, 387-598.
<https://doi.org/10.1016/B978-012693015-3/50006-1>
- [20] Faust, M.A. (1993) Sexuality in a Toxic Dinoflagellate, *Prorocentrum lima*. In: Smayda, T.J. and Shimizu, Eds., *Toxic Phytoplankton Blooms in the Sea*, Elsevier, Amsterdam, 121-126.
- [21] Dodge, J.D. (1982) Marine Dinoflagellates of the British Isles. Her Majesty's Stationery Office, London.
- [22] Fukuyo, Y., Takano, H., Chihara, M. and Matsuoka, K. (1990) Red Tide Organisms in Japan. An Illustrated Taxonomic Guide. Uchida Rokakuho Co., Ltd., Tokyo.
- [23] Fukuyo, Y., Yoshida, K., Ogata, T., Ishimaru, T., Kodama, M., Pholpunthin, P., Wiessang, S., Phanichyakarn, V. and Piyakarnchana (1989) Suspected Causative Dinoflagellates of Thailand. In: Okachi, T., Anderson, D.M. and Nemoto, T., Eds., *Red Tides: Biology, Environmental Science, and Toxicology*, Elsevier, New York, 403-406.