

Soil Amendments Enhanced Summer Squash Yield, Fruit Composition, Quality, and Soil Enzymes Activity

George F. Antonious^{1*}, Mohammad H. Dawood², Eric T. Turley¹, Thomas G. Trivette¹

¹Division of Environmental Studies, College of Agriculture, Community, and the Sciences, Kentucky State University, Frankfort, KY, USA

²Department of Soil Science and Water Resources, College of Agriculture, University of Kufa, El-Najaf, Iraq
Email: *george.antonious@kysu.edu

How to cite this paper: Antonious, G.F., Dawood, M.H., Turley, E.T. and Trivette, T.G. (2022) Soil Amendments Enhanced Summer Squash Yield, Fruit Composition, Quality, and Soil Enzymes Activity. *Agricultural Sciences*, 13, 684-701.
<https://doi.org/10.4236/as.2022.136045>

Received: February 24, 2022

Accepted: June 10, 2022

Published: June 13, 2022

Copyright © 2022 by author(s) and Scientific Research Publishing Inc.
This work is licensed under the Creative Commons Attribution International License (CC BY 4.0).

<http://creativecommons.org/licenses/by/4.0/>



Open Access

Abstract

Summer squash, *Cucurbita pepo* was field grown under fourteen soil treatments: sewage sludge (SS); horse manure (HM); chicken manure (CM); vermicompost; inorganic fertilizer (Inorg); commercial organic fertilizer (Org); and no-mulch (NM) control treatment. Soil treatments were also mixed with biochar to make a total of 14 treatments to assess the impact on 1) squash fruit yield and quality, 2) fruit vitamin C, total phenols, and soluble sugars content, and 3) soil microbial activity expressed as urease and invertase secretions. Results revealed that SS treatments increased squash yield and fruit number by 114% and 116%, respectively compared to NM control treatment. Fruits of plants grown in Inorg mixed with biochar (InorgBio) increased fruits' vitamin C, total phenols, and soluble sugars by 73%, 52%, and 7%, respectively compared to Inorg with no-biochar treatment. However, biochar was not consistent in increasing soil urease and invertase activities. The use of animal manure is an affordable way to reduce dependence on mineral fertilizers. Results revealed that the addition of biochar to Org fertilizer increased squash fruit weight and numbers of fruits compared to Org not treated with biochar. No single amendment increased all fruit composition and soil urease and invertase activities.

Keywords

Sewage Sludge, Horse Manure, Chicken Manure, Vitamin C, Urease, Invertase

1. Introduction

Currently, the world generates 1.3 billion tons per year of municipal sewage

sludge (SS), a by-product of sewage treatment plants, also known as biosolids. By 2025, the world could generate 2.2 billion tons of biosolids per year [1]. The use of SS and other animal manures as organic amendments in agricultural production systems would reduce the need for synthetic fertilizers and improve soil nutrient status at affordable or no cost to limited-resource farmers. SS compost promotes soil health and microbiological activity [2]. Manures intensify soil organic matter, improve soil physical structure, enhance soil fungal and bacterial activity, reduce eutrophication (excess N and P in natural water resources), provide low-cost adsorbents that bind with agricultural contaminants and prevent natural water contamination by pesticide residues and inorganic fertilizers, reducing the impact of agricultural chemicals on natural surface and groundwater quality [3].

Microorganisms in animal manures break down complex forms of organic nutrients and facilitate the slow release of N, P, and K from soil organic matter for plant uptake. In addition, soil enzymes, such as urease, invertase, dehydrogenase, cellulase, amylase, and phosphatase secreted by microorganisms (bacteria, fungi, protozoa, and algae) in animal manure are primary means of mineralization and are sensitive indicators of soil health [4]. The literature review on the use of organic amendments in agriculture has been clearly demonstrated [5]. Animal manures contain humus substances, macro- and micro-nutrients important for plant growth. The use of SS as a soil conditioner to enhance soil physical, chemical, and microbial conditions might also enhance soil bioremediation [6]. Agricultural use of SS has been successful in the production of vegetables [7] [8]. SS improves soil physical properties, nutrient and water holding capacity, total pore space, aggregate stability, soil erosion, and decreases soil density, and increases soil organic matter, such as humic acid and fulvic acid [9] [10] [11] that improve soil aeration and moisture retention. Chicken manure (CM) also enhances soil biological activity and fertility, nutrient status and growth of several groups of microorganisms, such as bacteria, fungi, and actinomycetes [12]. CM contains several essential plant nutrients (N, P, K, S, Ca, Mg, B, Cu, Fe, Mn, Mo, and Zn), and has been documented as an excellent fertilizer [13]. Because of the rapid growth in the poultry industry, CM has become available in increasing quantities. Compared to other livestock species, poultry has relatively high dietary requirements for the sulfuric amino acids (methionine and cysteine). Commercial diets for laying hens, broilers and turkeys are usually supplemented with synthetic methionine. Consequently, the sulfur content tends to be higher in poultry manure than in manures from other farm animals. Investigators reported that the concentration of dipropyl disulfide and dipropyl trisulfide that are useful in preventing cardiovascular diseases were greatest in onion plants grown in soil amended with CM [14].

The use of horse manure (HM) as organic fertilizer revealed that a ton of HM contains 11N:2P:8K [15]. HM nutrient value is relatively small and depends largely on the type of bedding material, the food source and type, age and condi-

tion of the animal. Vermicompost (Vermi) is a product of the interaction of earthworms (*Eisenia foetida*) with microorganisms and other fauna within a decomposer, designed for earthworm incubation. The NPK essential plant nutrients and C/N ratio of Vermicompost revealed its agronomic value as an organic soil conditioner. Researchers found that Vermicompost used in crop production as organic amendment promotes plant nutrient availability. According to Ramnarain *et al.* [16], Vermicompost can be of significant value to farmers as a replacement for inorganic fertilizers that secure better prices for organic produce.

Studies on biochar have indicated that biochar (product of waste incineration) could increase plant nutrient availability, soil cation exchange capacity (CEC), soil organic matter, and soil microbial activities [17] [18] and crop yields [19]. There is a lack of information on the impact of organic amendments on plants' nutritional and antioxidant properties. Studies had focused on crop yield and soil physical and chemical properties after the addition of animal manures and other organic amendments with very little information on the nutritional and antioxidant contents of edible plants. Several vegetable species and varieties of the same species have not been completely analyzed for vitamin C and phenolic contents, which have a number of human health benefits. Plant phenols may interfere with stages of the cancer process resulting in a reduction of cancer risk. Phenols prevent oxidative damage to biomolecules, such as DNA, lipids and proteins that play a role in chronic diseases such as cancer and cardiovascular diseases [20]. The role of phenols as antioxidants with properties like vitamins C, E, and β -carotene has prompted several studies on these phytochemicals.

Shi [21] reported the agricultural and ecological impact of soil enzymes secreted by microorganisms in animal manures on nutrient recycling. Urease (urea amidohydrolase, EC 3.5.1.5) hydrolyzes urea fertilizers into NH_3 and CO_2 , which are associated with rise in soil pH [22], resulting in a rapid N loss to the atmosphere due to NH_3 volatilization [23]. Soil enzymes, such as urease and invertase are important for breaking down complex forms of organic matter and release of C and N sources for the growth and multiplication of soil microorganisms. Urease activity in soil has received great attention, due to its vital role in the regulation of N supply to plants after urea fertilization. Soil urease originates mainly from plants [24] and microorganisms [25]. Invertase (β -D-fructofuranosidase) is the enzyme that splits sucrose into glucose and fructose and is available in microorganisms, animals, and plants [26]. Invertase hydrolysis occurs in both acidic and alkaline conditions [27].

Summer squash (*Cucurbita pepo*, variety Raven) is among the most widely grown cucurbits worldwide, it is a seasonal crop that contains beneficial minerals, carotenoids, vitamin C, and phenolic compounds [28] that has antioxidant/antiradical, anticarcinogenic, anti-inflammatory, antiviral, and antimicrobial activities [29]. Accordingly, the present objectives were established to investigate the impact of soil mixed with animal manures (sewage sludge, chicken manure, horse manure, vermicompost), organic and inorganic com-

mercial fertilizers, with and without biochar on: 1) summer squash fruit yield and quality; 2) fruit vitamin C, total phenols, and soluble sugar content; 3) soil urease and invertase activity after the incorporation of soil amendments to native soil; and 4) to answer Kentucky farmers' questions if animal manure could reduce dependence on the costly inorganic mineral fertilizers, increase squash yield, and improve soil quality, and fruit nutritional composition.

2. Materials and Methods

2.1. Field Study

A field experiment at the University of Kentucky Horticulture Research Farm (Lexington, KY, USA) was established in a randomized complete block design (RCBD). Each plot was 1.2 m × 3 m (3.6 m²) and the entire study area contained 42 plots (3 replicate × 14 treatments). Seven soil treatments were investigated: 1) sewage sludge (SS), 2) horse manure (HM), 3) chicken manure (CM), 4) vermicompost (Vermi; worm casting), 5) organic (Org) commercial fertilizer (Nature Safe 10:2:8), 6) inorganic (Inorg) commercial fertilizer (Southern State 20:20:20), and 7) control (no-mulch NM untreated soil). The soil in each of the seven treatments also was mixed with 10% (w/w) biochar obtained from Wakefield Agricultural Carbon (Columbia, MO) to make total of 14 treatments. Properties of biochar used in this investigation are: surface area 366 m²·g⁻¹ dry, bulk density 480.6 kg·m⁻³, total organic carbon 88%, N 0.27%, P 2.06 mg·kg⁻¹, K 280 mg·kg⁻¹, Ca 1881 mg·kg⁻¹, Cu 2.45 mg·kg⁻¹, Mg 558 mg·kg⁻¹, Zn 2.09 mg·kg⁻¹, 54% moisture, preparation temperature 200 °C, total inorganic carbon 0.34%, particle size (<0.5 mm), and pH 7.4. The control (no-mulch NM untreated soil) is a Bluegrass-Maury Silty Loam that contains 2.2% organic matter and pH of 6.2 located at the blue grass region (Fayette County, KY) that has 56%, 38%, and 6% silt, clay, and sand, respectively.

Each soil amendment was applied at the rate of 5% N on dry weight basis to eliminate variations among soil treatments due to N content. SS from the Metropolitan Sewer District, Louisville, KY applied at 0.83 kg plot⁻¹. CM (1.1% N) from the Department of Animal and Food Sciences, University of Kentucky, Lexington, Kentucky applied at 3.78 kg plot⁻¹, HM (0.7% N) from the Kentucky horse park, Lexington, Kentucky and applied at 5.94 kg plot⁻¹. Vermicompost (1.5% N, worm castings) purchased from Worm Power (Montpelier, Vermont, USA) and applied at 2.78 kg plot⁻¹. Organic (Nature Safe 10N:2P:8K) and inorganic (20N:20P:20K) commercial fertilizers (10% and 20% N, respectively), were obtained from the Southern States Cooperative Stores (Lexington, KY, USA) and used at 0.45 and 0.23 kg plot⁻¹, respectively. Each soil amendment was mixed with native soil to a depth of 15 cm (area of increased soil microbial activity). Seedlings of Summer squash (*Cucurbita pepo*, cultivar Raven) of 25 d old were planted in raised black plastic mulch (Figure 1) of freshly tilled soil of 42 plots (14 treatments × 3 replicates each) and watered using a drip irrigation system. All agricultural operations were implemented regularly as needed. The plants



Figure 1. Laying black plastic mulch (A) to maintain soil moisture and control weeds, summer squash (zucchini), *Cucurbita pepo* plants grown in animal manures amended soil showing no symptoms of plants necrosis (B), glossy fruit on squash plant grown in soil amended with sewage sludge compost (C), and yield of dark-colored squash (*Cucurbita pepo*) variety Raven (D).

were sprayed with the insecticide esfenvalerate (Asana XL) and fungicide chlorothalonil (Bravo) three times during the growing season at the recommended rates of application and the plants were grown according to Kentucky Vegetable Production Guide [30], but no other fertilizers were applied.

2.2. Squash Yield and Fruit Quality Characteristics

At harvest (50 d after planting) and before bolting (flowering) when fruit skin had a glossy appearance, fruits were collected, counted, weighed, and graded into U.S. No.1, U.S. No. 2, and unclassified according to the USDA Standards for Grades of Summer Squash (2016) [31]. U.S. No. 1 consists of squash fruits with stems or portions of stems attached, fairly young, tender and well formed, firm, free from decay and breakdown, free from damage caused by discoloration, cuts, bruises, and scars (wounds), freezing, dirt, disease, insects, mechanical or other means. U.S. No. 2 consists of fruits which are not old and tough, but firm, free from decay and breakdown, free from damage caused by freezing, discoloration, cuts, bruises, scars, dirt, disease, insects, mechanical or other means. Accordingly, squash pickers should use plastic containers and wear soft gloves to avoid fruit bruises, scratches, and fingernail punctures. Ungraded soft-shell squash

fruits are considered “unclassified”.

2.3. Quantification of Total Phenols, Vitamin C, and Soluble Sugars

Fruits were cleaned with tap water, cut into small cubes, and a representative 20 g samples from each plot were blended with 150 mL of ethanol to extract phenols. The homogenates were filtered through Whatman No. 1 filter paper and 1 mL aliquots of filtrate were used for determination of total phenols colorimetrically using the Folin-Ciocalteu method [32] and a standard calibration curve of 10 - 80 $\mu\text{g}\cdot\text{mL}^{-1}$ of chlorogenic acid (Fisher Scientific Company, Pittsburg, PA, USA). Ascorbic acid (vitamin C) was extracted by blending representative 20 g of fruits with 100 mL of 0.4% (w/v) oxalic acid solution [33] and determined colorimetrically using the potassium ferricyanide method [34] and a standard curve of ascorbic acid in the range of 90 - 300 $\mu\text{g}\cdot\text{mL}^{-1}$. Soluble sugars in 20 g fruits were extracted with 80% ethanol and quantified using a calibration curve in the range of 100 - 800 $\mu\text{g}\cdot\text{mL}^{-1}$ of glucose [35]. Concentrations of ascorbic acid, total phenols, and soluble sugars in squash fruit extracts were calculated using linear regression equations ($y = a + b \times$) established for each parameter, where y is the absorbance of the color formed, x is the concentration (ppm), a is the intercept of the regression line, and b is the slope of the line.

2.4. Collection and Preparation of Soil Samples

Soil samples ($n = 3$) were collected from the rhizosphere (a zone where soil and plant root make contact) of growing squash plants to a depth of 15 cm (a zone of increased microbial and enzyme activity). Soil samples collected using a core sampler (Clements Associates, Newton, IA) equipped with plastic liner tubes of 2.5 cm i.d. were air-dried at room temperature, passed through a 2 mm sieve, and kept in plastic bags at 4 °C up to 24 h before use.

2.5. Soil Enzymes Analysis

For determination of soil urease activity, five-g of soil collected from each treatment and 10 mL of 0.1 M phosphate buffer (pH 6.7) in 50 mL volumetric flasks were kept in an incubator at 37 °C for 24 h, and the procedure was completed as described by Tabatabai and Bremner [36]. The method was developed by measuring the concentrations of NH_4^+ ions released in the soil solutions by the selective electrode method [37]. A series of standard solutions of NH_4Cl covering the concentrations of 0.1 - 100 $\mu\text{g NH}_4\text{-N mL}^{-1}$ of water was used for calibration. Urease activity was expressed as $\mu\text{g NH}_4\text{-N released g}^{-1}$ dry soil during the incubation time.

Invertase activity in soil was measured by the method described by Balasubramanian *et al.* [38]. A standard calibration curve was obtained with each group of samples using analytical grade glucose in the range of 10 - 50 $\mu\text{g}\cdot\text{mL}^{-1}$ glucose (Sigma Chemical Company, St. Louis, MO, USA).

2.6. Statistical Analysis

Data containing squash yield, fruit numbers and quality, fruit composition (phenols, vitamin C, and soluble sugars), soil urease, and invertase activity were statistically analyzed using analysis of variance (ANOVA) and the means were compared using Duncan's multiple range test [39], which provided significant differences among means.

3. Results and Discussion

Average weight of squash fruits of plants grown in soil amended with municipal SS, Inorg, Vermi, CM, HM were significantly ($P \leq 0.05$) greater than weight of fruits obtained from plants grown in Org and NM control treatment. Biochar added to SS (SSBio), Vermi (VermiBio), Inorg (InorgBio), CM (CMBio), HM (HMBio) did not impact squash fruit weight. Whereas, Biochar added to Org (OrgBio) and NM (NMBio) significantly increased squash yield (**Figure 2(A)**). Average number of squash fruits collected from plants grown in soil amended with SS, Inorg, Vermi, and CM were greater compared to fruits of plants grown in NM control plots (5.5 fruits plant^{-1}) (**Figure 2(B)**).

Regarding fruit quality, **Table 1** revealed that SS increased the number of U.S. No. 1 fruits compared to Org and NM control treatments. Biochar added to SS (SSBio) treatments did not significantly increase number of U.S. No.1 and U.S. No.2 fruits compared to SS treatment not amended with biochar, whereas biochar added to each of the soil amendments did not affect number of unclassified fruits. Plants grown in soil amended with Vermi had greater number of U.S. No. 2 fruits compared to plants grown in HM, Org, and NM treatments. Other than that, no significant differences ($P \geq 0.05$) were found in U.S. No. 2 fruit grades among soil amendments tested. Overall, the number of unclassified fruits plant^{-1} was not significantly different among soil treatments. Accordingly, other than some impact on U.S. No. 1 fruits, soil amendments impacted squash yield, but did not impact fruit grades.

Regarding fruit composition, it is recognized that that diet constituents may contain cancer-causing substances as well as many cancer-preventive agents [40]. Antioxidants, such as vitamin C and phenols in plants tend to give their electrons to free radicals to neutralize them, preventing the cells from potential damage, which in turn cure numerous human diseases. Squash fruits of plants grown in Inorg fertilizer mixed with biochar (InorgBio) as well as organic fertilizer (Org) exhibited a higher concentration of vitamin C (ascorbic acid) by 11% and 12% compared to plants grown in NM treatments (**Figure 3(A)**). InorgBio increased vitamin C in squash fruits by 73% compared to squash fruits of plants grown in soil amended with Inorg commercial fertilizer (Inorg) not mixed with biochar, indicating the role of biochar in elevating the concentration of vitamin C. On the contrary, biochar added to SS (SSBio) reduced vitamin C concentration by 22% compared to SS with no biochar addition. In addition, plants grown in soil amended with Inorg fertilizer mixed with biochar (InorgBio) increased

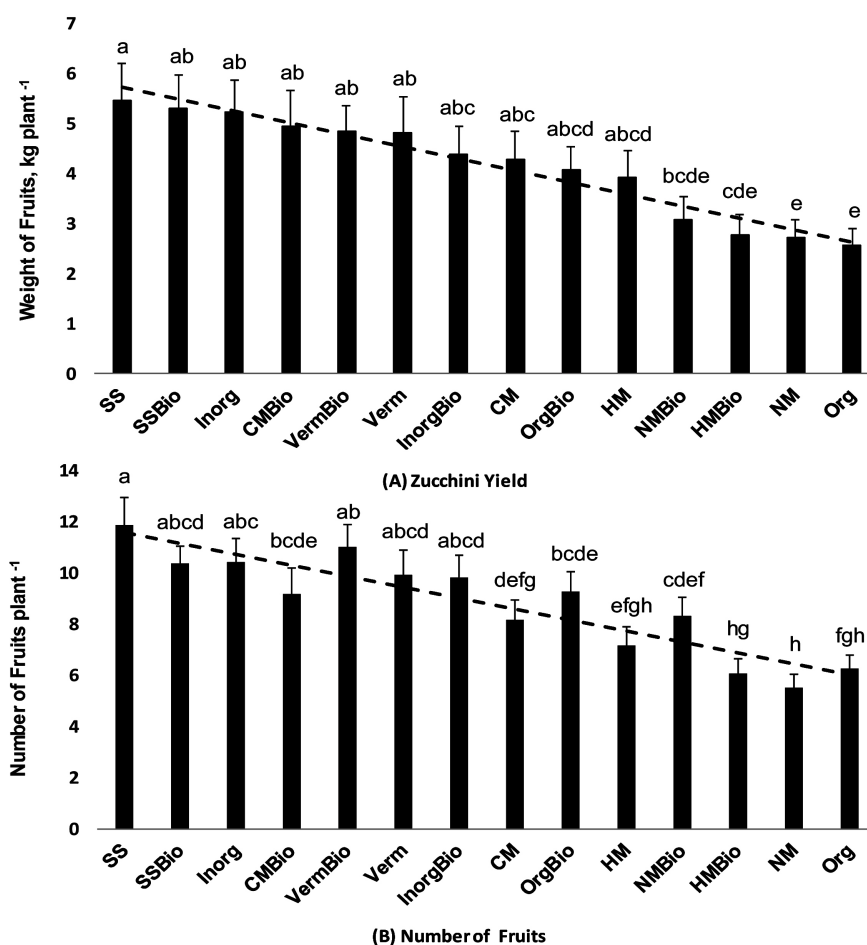


Figure 2. Yield of summer squash (zucchini) fruits expressed as weight of fruits (A) and number of fruits (B) of plants grown under fourteen soil management practices: sewage sludge (SS), SS mixed with biochar (SSBio), inorganic commercial fertilizer (Inorg), chicken manure mixed with biochar (CMBio), vermicompost mixed with biochar (VermBio), vermicompost (Vermi), inorganic commercial fertilizer mixed with biochar (InorgBio), chicken manure (CM), organic commercial fertilizer mixed with biochar (OrgBio), horse manure (HM), no-mulch native soil mixed with biochar (NMBio), horse manure mixed with biochar (HMBio), no-mulch soil (NM), and organic commercial fertilizer (Org). Statistical comparisons were carried out among fourteen soil management practices. Bars accompanied by the same letter(s) are not significantly different ($P > 0.05$) using Duncan's multiple range tests.

total phenols in squash fruits from 118 to 179 $\mu\text{g}\cdot\text{g}^{-1}$ fresh fruits indicating 52% increase due to biochar addition to Inorg fertilizer (**Figure 3(B)**). **Figure 4** also revealed that plants grown in CMBio, CM, and InorgBio significantly increased total phenols compare to all other treatments tested, whereas plants grown in soil amended with SS did not increase total phenols content in fruits of plants grown in NM treatment. **Figure 4** revealed that plants grown in SS amended with biochar (SSBio) increased soluble sugars in squash fruits compared to all other 13 treatments tested, whereas addition of biochar to Vermi, CM, and Inorg (VermBio, CMBio, and InorgBio, respectively) did not increase soluble

Table 1. Summer squash fruit grading based on USDA marketable fruit quality characteristics.

Soil Amendment	Number of U.S. No. 1 Plant ⁻¹	Number of U.S. No. 2 Plant ⁻¹	Number of Unclassified Plant ⁻¹
SS	4.48 ± 0.66 a	6.81 ± 1.27 ab	0.57 ± 0.21 a
CM	3.52 ± 0.58 ab	4.43 ± 0.36 ab	0.19 ± 0.11 a
HM	3.33 ± 0.52 abc	3.38 ± 0.51 b	0.43 ± 0.16 a
Vermi	3.62 ± 0.49 ab	5.76 ± 0.73 a	0.53 ± 0.22 a
Organic	2.09 ± 0.36 d	3.86 ± 0.39 b	0.29 ± 0.12 a
Inorganic	4.05 ± 0.53 a	6.0 ± 0.86 ab	0.38 ± 0.17 a
No-Mulch	2.10 ± 0.38 d	3.1 ± 0.29 b	0.3 ± 0.16 a
SSBio	4.19 ± 0.51 a	5.76 ± 0.55 ab	0.38 ± 0.12 a
CMBio	3.43 ± 0.53 ab	5.28 ± 0.25 ab	0.43 ± 0.11 a
HMBio	2.19 ± 0.28 cd	3.62 ± 0.36 ab	0.24 ± 0.11 a
VermiBio	4.19 ± 0.67 a	6.21 ± 0.59 ab	0.62 ± 0.22 a
OrgBio	3.43 ± 0.23 ab	5.57 ± 0.50 ab	0.24 ± 0.09 a
InorgBio	3.57 ± 0.49 ab	5.75 ± 0.28 ab	0.27 ± 0.17 a
No-Mulch Bio	2.62 ± 0.44 abc	5.04 ± 0.33 ab	0.62 ± 0.22 a

SS = sewage sludge, CM = chicken manure, HM = horse manure, Vermi = vermicompost, Organic = commercial organic fertilizer, Inorganic = inorganic commercial fertilizer, No-Mulch = no-mulch native soil, SSBio = ss mixed with biochar, CMBio = chicken manure mixed with biochar, HMBio = horse manure mixed with biochar, VermiBio = vermicompost mixed with biochar, OrgBio = organic commercial fertilizer mixed with biochar, InorgBio = inorganic commercial fertilizer mixed with biochar, and No-Mulch Bio = no-mulch native soil mixed with biochar. Each value in the table is an average of 21 replicates ± standard deviation. Values in each column accompanied by the same letter(s) are not significantly different ($P > 0.05$) using Duncan's multiple range test (SAS Institute 2016).

sugars concentrations after biochar addition.

Vitamin C and total phenols have antioxidant properties and are thus important quality attributes in edible plants. Concentrations of these two phytochemicals in squash fruits varied significantly among soil treatments. One can investigate whether the higher content of vitamin C, phenols, and soluble sugars in some treatments is due to higher synthesis of these water-soluble compounds by squash plants, or due to increased absorption from soil by the plant roots. Alternatively, these elevated concentrations in the fruits of some treatments might be due to increased soil organic matter and microbial activity after addition of certain soil amendments. Based on the results in **Figure 3(A)**, plants grown in NM native soil (control plants) contained low concentrations of vitamin C compared to InorgBio and organic commercial fertilizer (Org) treatments. Total phenols concentrations in fruits of plants grown in NM native soil were

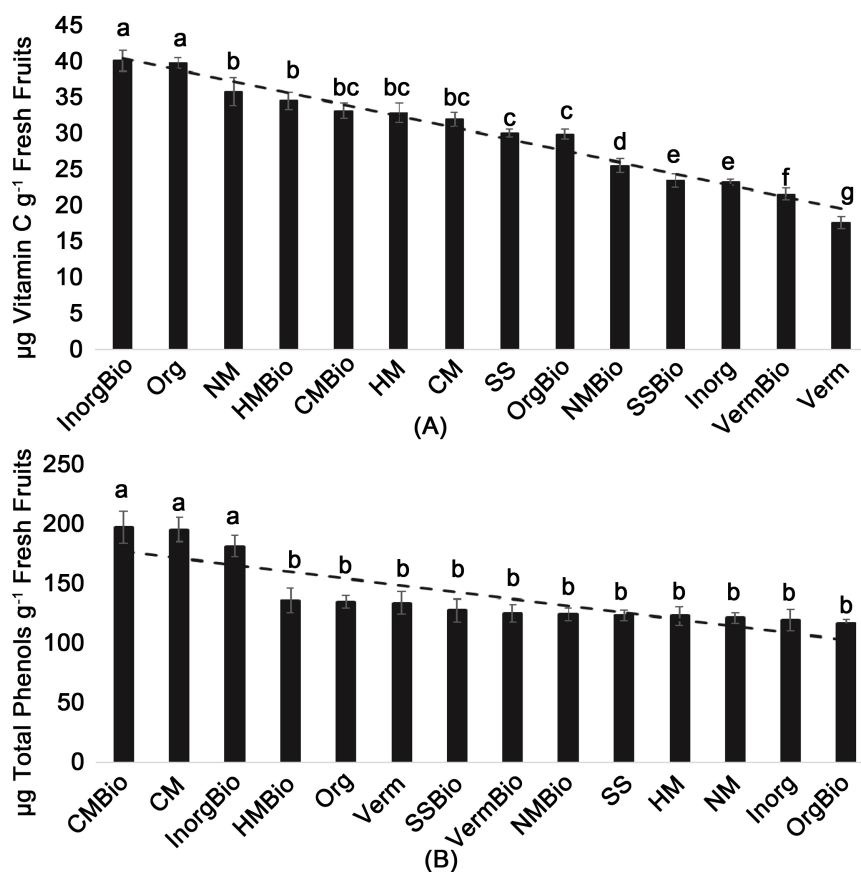


Figure 3. Average concentrations of ascorbic acid (A) and total phenols (B) \pm std. error in fresh summer squash (zucchini) fruits of plants grown under fourteen soil management practices: inorganic commercial fertilizer mixed with biochar (InorgBio), organic commercial fertilizer (Org), no-mulch native soil (NM), horse manure mixed with biochar (HMBio), organic commercial fertilizer (Org), vermicompost (Verm), sewage sludge mixed with biochar (SSBio), vermicompost mixed with biochar ((VermBio), no-mulch native soil mixed with biochar (NMBio), sewage sludge (SS), horse manure (HM), no-mulch native soil (NM), inorganic commercial fertilizer (Inorg), and organic commercial fertilizer mixed with biochar (OrgBio). Statistical comparisons were carried out among soil management practices for each parameter. Bars accompanied by the same letter are not significantly different ($P > 0.05$) using Duncan's multiple range tests.

also lower than fruits of plants grown in CM, CMBio, and InorgBio indicating the role of these treatments in promoting the concentrations of phenols (Figure 3(B)). Concentration of soluble sugars were also greater in SS mixed with biochar (SSBio) compared to fruits of plants grown in other treatments including the NM control (Figure 4). Animal manures (SS, CM, HM, and Vermi) contain several enzyme substrates, such as urea, sucrose, and orthophosphates and enzymes that secrete soil urease, invertase, and phosphatase, respectively allowing the breakdown of complex forms of organic materials in soil and release nutrients for plant uptake.

Accordingly, the pronounced increase in soil urease and invertase activity (Figure 5) could be attributed to increased microbial activity and the enzymes

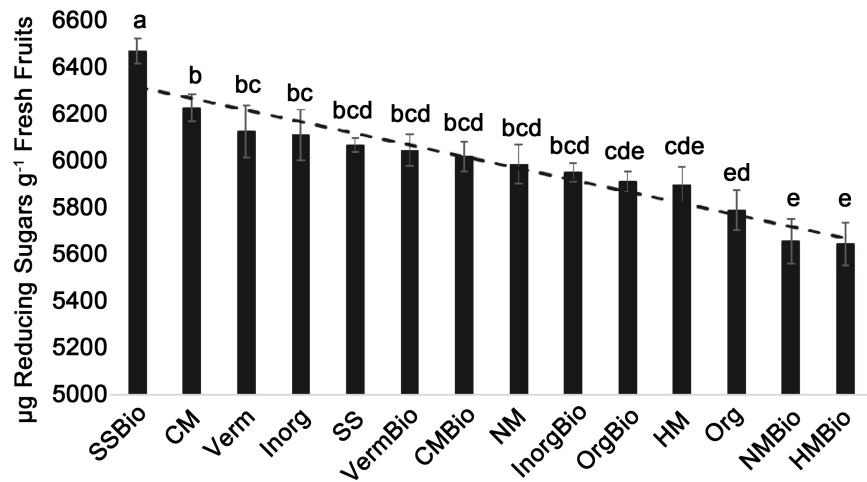


Figure 4. Average concentrations of soluble sugars \pm std. error in fresh summer squash (zucchini) fruits of plants grown under fourteen soil management practices: sewage sludged mixed with biochar (SSBio), chicken manure (CM), vermicompost (Verm), inorganic commercial fertilizer (Inorg), sewage sludge (SS), vermicompost mixed with biochar ((VermiBio), chicken manure mixed with biochar (CMBio), no-mulch native soil (NM), inorganic commercial fertilizer mixed with biochar (InorgBio), organic commercial fertilizer mixed with biochar (OrgBio), horse manure (HM), organic commercial fertilizer (Org), no-mulch native soil mixed with biochar (NMBio), and horse manure mixed with biochar (HMBio). Statistical comparisons were carried out among soil management practices. Bars accompanied by the same letter(s) are not significantly different ($P > 0.05$) using Duncan's multiple range tests.

they produce. Recent studies carried out by Antonious *et al.* [5] revealed that animal manures increased the activities of soil urease and invertase. Soil enzymes secreted by soil microorganisms also promote soil processes such as synthesis of humus substances and breakdown of organic matter present in animal manure and consequently release soil nutrients through mineralization. CM amended soil increased soil urease activity by 119% compared to NM native soil (Figure 5(A)). This increase in soil urease revealed the transformation of N in CM mixed soil from urea to ammonium ions (NH_4^+). On the contrary, CMBio, VermiBio, InorgBio, and OrgBio did not increase urease activity, indicating that biochar has no role in increasing soil urease activity in these treatments. Figure 5(B) also indicated that biochar added to SS (SSBio), Verm (VermiBio), Organic fertilizer (OrgBio), and Inorganic fertilizer (InorgBio) reduced soil invertase activity.

In fact, some animal manures, such as SS, CM, and HM are associated with inorganic and organic toxic compounds, such as trace metals, hormones, antibiotics and pesticides that when incorporated into soil, can cause a pollution problem and consequently toxic effects to soil microorganisms that control nutrients availability to growing plants. The increased demand for animal protein is leading to a great use of antibiotics in agriculture to raise food-producing animals in intensive production systems. More antibiotics are currently used in poultry, horses, swine, and cattle raising to promote growth and prevent diseases

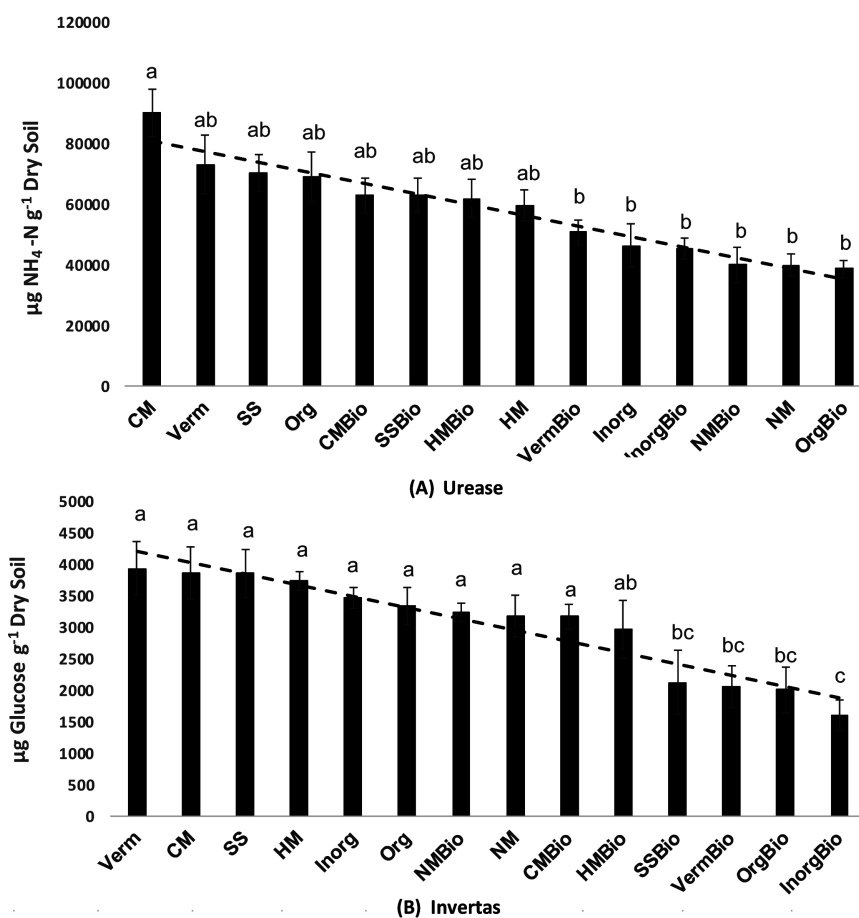


Figure 5. Urease activity \pm std. error expressed as $\mu\text{g NH}_4\text{-N}$ released g^{-1} dry soil (A) and invertase activity \pm std. error expressed as μg glucose released g^{-1} dry soil (B) in soil mixed with chicken manure (CM), vermicompost (Verm), sewage sludge (SS), organic commercial fertilizer (Org), chicken manure mixed with biochar (CMBio), sewage sludge mixed with biochar (SSBio), horse manure mixed with biochar (HMBio), horse manure (HM), vermi-compost mixed with biochar (VermBio), inorganic commercial fertilizer (Inorg), inorganic commercial fertilizer mixed with biochar (InorgBio), no-mulch native soil mixed with biochar (NMBio), no-mulch soil (NM), and organic commercial fertilizer mixed with biochar (OrgBio). Statistical comparisons using analysis of variance (ANOVA) procedure were used to test the differences among means of soil management practices. Bars accompanied by the same letter(s) are not significantly different ($P > 0.05$) using Duncan's multiple range tests.

(CDDEP) [41]. According to the Lexington Herald-Leader on November 14, 2012, Kentucky is the highest five states in the USA in overuse of antibiotics (Kentucky Health News [42]. Yang *et al.* [43] studied the influence of CM fertilization on antibiotic-resistant bacteria in soil and the endophytic bacteria of Pakchoi (a type of Chinese cabbage, *Brassica rapa*). They detected these bacterial populations in CM, CM amended soil, and in harvested vegetables grown in manure-amended soil and concluded that they presented a potential threat to human health.

In addition, a considerable area of agricultural land is contaminated with

cadmium (Cd) and lead (Pb) due to land application of fertilizers, animal manures, and atmospheric deposition [44]. Cd has a retention time of 150 years in soil [45]. Pb is also estimated to have a long soil retention time [46]. These metals have negative impact of soil hydrolyzing enzymes that break down complex forms of organic matter and release of plant nutrients. Accordingly, our results are not surprising since other investigators reported that the increased concentrations of Cd and Pb have negative impacts on soil microbes, and the effect of Cd on soil urease activity is more than that on invertase, while Pb has more effect on invertase activity than Cd [47]. Investigators reported variability of biochar effect on soil enzymes activity that might be due soil type and chemical composition of soil, trace metals in biochar, or variations in biochar properties of absorbing and retaining water molecules that impact microbial activity and enzymes secretions. Park *et al.* [48] and Kumar *et al.* [49] reported positive effect of biochar on soil enzymes, whereas Lehmann *et al.* [50] reported negative effects, while Wu *et al.* and Lammirato *et al.* [51] [52] reported a non-biochar effect on soil enzymes. Accordingly, continuous monitoring of soil enzymes secreted by microorganisms in relation to the antioxidant contents of plants grown in animal manure is recommended when using animal manures for growing edible plants. These results revealed that each soil amendment used in this investigation has a unique impact on increasing each of the fruit quality attributes and no single amendment increased squash yield and all nutrients in squash fruits and/or soil urease and invertase activities. Further work will be continued in our future studies to investigate the potential impact of using mixtures of animal manures incorporation with biochar on elevating the nutritional composition of squash fruits as well as soil enzymatic activities.

4. Conclusion

The present investigation provided new information on the nutritional value of summer squash fruits, along with some beneficial effects in relation to composting. Municipal SS and CM generation is expected to be available in increasing quantities due to increased municipal SS composting facilities and the rapid growth in the poultry industry. Using animal waste as a low-cost organic fertilizer has a positive effect on the growth and crop yield. SS treatments were superior in increasing squash yield and fruit number by 114% and 116%, respectively compared to NM control treatments. The presence of organic matter in recycled manure often improves soil's physical and chemical properties and promotes soil biological activities. Composts improved summer squash yield. Fruits of plants grown in inorganic fertilizer mixed with biochar (InorgBio) as well as organic fertilizer with no biochar (Org) exhibited a higher increase of vitamin C (ascorbic acid) by 11% and 12%, respectively compared to plants grown in NM control treatments. Plants grown in soil amended with Inorg fertilizer mixed with biochar (InorgBio) increased total phenols in squash fruits from 118 to 179 $\mu\text{g}\cdot\text{g}^{-1}$ fresh fruits indicating a 52% increase due to biochar addition to Inorg fer-

tilizer. Soluble sugars in plants grown in soils amended with SS mixed with biochar (SSBio) were significantly increased compared to other soil treatments. CM amended soil increased soil urease activity by 119% compared to NM native soil, whereas the addition of biochar to SS, Vermi, Org, and Inorg reduced soil invertase activity. Organic amendments from animal manure can be used as an alternative to inorganic fertilizers. The increase of organic matter in the soil after the addition of animal manure has a great impact on the biological and biochemical properties of soil. However, soil amendments such as SS could be contaminated with inorganic and organic compounds such as heavy metals, hormones, antibiotics, and pesticides that when incorporated into soil might constitute a pollution problem and therefore impact the activity of soil microorganisms and their enzymatic secretions. On the other hand, increasing costs of commercial fertilizers and the release of large amounts of SS, CM, and HM worldwide have made cropland application of this waste an attractive disposal option. Results of this investigation revealed a significant increase in total vitamin C, and total phenols in squash fruits of plants grown with Inorg fertilizer mixed with biochar (Inorg-Bio).

Acknowledgements

The authors thank Steven Diver and his farm crew for preparation of the soil amendments and maintaining the field experimental plots.

Author Contributions

George F. Antonious designed the field study, implemented the laboratory analysis, and wrote the manuscript. Mohammad H. Dawood conducted the statistical analysis and field work, Eric T. Turley and Thomas G. Trivette organized the field work and collected the plant and soil samples.

Funding

This investigation was funded by a Grant No. KYX-10-18-P65, Accession No. 1017900 from the United States Department of Agriculture, National Institute of Food and Agriculture (USDA/NIFA) to Kentucky State University.

Conflicts of Interest

The authors declare that there is no conflict of interest.

References

- [1] Moya, D., Aldás, C., López, G. and Kaparaju, P. (2017) Municipal Solid Waste as a Valuable Renewable Energy Resource: A Worldwide Opportunity of Energy Recovery by Using Waste-to-Energy Technologies. *Energy Procedia*, **134**, 286-295. <https://doi.org/10.1016/j.egypro.2017.09.618>
- [2] Antonious, G.F. (2009) Enzyme Activities and Heavy Metals Concentration in Soil Amended with Sewage Sludge. *Journal of Environmental Science and Health, Part*

- A, **44**, 1019-1024. <https://doi.org/10.1080/10934520902996971>
- [3] Antonious, G.F. (2015) Decontamination of Pesticide Residues for Sustainable Agriculture. *JSM Environmental Science & Ecology*, **3**, 1-7.
- [4] Angelovicova, L., Lodenius, M., Tulisalo, E. and Fazekasova, D. (2014) Effect of Heavy Metals on Soil Enzyme Activity at Different Field Conditions in Middle Spis Mining Area (Slovakia). *Bulletin of Environmental Contamination and Toxicology*, **93**, 670-675. <https://doi.org/10.1007/s00128-014-1397-0>
- [5] Antonious, G., Turley, E. and Dawood, M. (2020) Monitoring Soil Enzymes Activity before and after Animal Manure Application. *Agriculture*, **10**, 166. <https://doi.org/10.3390/agriculture10050166>
- [6] Antonious, G.F. and Snyder, J.C. (2007) Accumulation of Heavy Metals in Plants and Potential Phytoremediation of Lead by Potato, *Solanum tuberosum* L. *Journal of Environmental Science and Health*, **42**, 811-816. <https://doi.org/10.1080/10934520701304757>
- [7] Antonious, G.F., Turley, E.T., Hill, R.R. and Snyder, J.C. (2013) Effect of Municipal Refuse and Chicken Manure Applications on Kale and Collard Green Yields and Quality. University of Kentucky, College of Agriculture, Food and Environment, Lexington, Fruit and Vegetable Research Report, PR-673, 37-39.
- [8] Antonious, G.F. (2014) Impact of Soil Management Practices on Yield, Fruit Quality, and Antioxidant Contents of Pepper at Four Stages of Fruit Development. *Journal of Environmental Science and Health, Part B*, **49**, 769-774. <https://doi.org/10.1080/03601234.2014.929908>
- [9] Antonious, G.F. (2016) Chapter 7. Soil Amendments for Agricultural Production. In: Larramendy, M.L. and Soloneski, S., Eds., *Organic Fertilizers: From Basic Concepts to Applied Outcomes*, Intech, Rijeka, 157-187.
- [10] Antonious, G.F. (2016) Recycling Organic Waste for Enhancing Soil Urease and Invertase Activity. *International Journal of Waste Resources*, **6**, 219-225. <https://doi.org/10.4172/2252-5211.1000219>
- [11] Plaza, C., Polo, A., Brunetti, G., Garcia-Gil, J. and D'Orazio, V. (2003) Soil Fulvic Acid Properties as a Means to Assess the Use of Pig Amendment. *Soil and Tillage Research*, **74**, 179-190. <https://doi.org/10.1016/j.still.2003.07.002>
- [12] Wanner, U., Fuhr, F. and Burauel, P. (2005) Influence of the Amendment of Corn Straw on the Degradation Behaviour of the Fungicide Dithianon in Soil. *Environmental Pollution*, **133**, 63-70. <https://doi.org/10.1016/j.envpol.2004.04.013>
- [13] Subramanian, B. and Gupta, G. (2006) Adsorption of Trace Elements from Poultry Litter by Montmorillonite Clay. *Journal of Hazardous Materials B*, **128**, 80-83. <https://doi.org/10.1016/j.jhazmat.2005.05.036>
- [14] Antonious, G.F., Perkins, E. and Cantor, A.H. (2009) Chicken Manure Increased Concentration of Organic Sulfur Compounds in Field Grown Onions. *Journal of Environmental Science and Health Part B*, **44**, 481-487. <https://doi.org/10.1080/03601230902935303>
- [15] Westendorf, M. and Krogmann, U. (2014) Horses and Manure. New Jersey Agricultural Experiment, New Brunswick, Cooperative Extension Fact Sheet FS036.
- [16] Ramnarain, Y.I., Abdel Ansari, A. and Ori, L. (2019) Vermicomposting of Different Organic Materials Using the Epigeic Earthworm, *Eisenia foetida*. *International Journal of Recycling of Organic Waste in Agriculture*, **8**, 23-36. <https://doi.org/10.1007/s40093-018-0225-7>
- [17] Antonious, G.F. (2018) Biochar and Animal Manure Impact on Soil, Crop Yield and

- Quality. In: *Agricultural Waste and Residue*, IntechOpen Science Books, Rijeka, 45-67. <https://doi.org/10.5772/intechopen.77008>
- [18] Wu, H., Lai, C., Zeng, G., Liang, J., Chen, J., Xu, J. and Lu, L. (2017) The Interactions of Composting and Biochar and Their Implications for Soil Amendment and Pollution Remediation: A Review. *Critical Reviews in Biotechnology*, **37**, 754-764. <https://doi.org/10.1080/07388551.2016.1232696>
- [19] Ferreira, C., Verheijen, F., Puga, J., Keizer, J. and Ferreira, A. (2017) Biochar in Vineyards: Impact on Soil Quality and Crop Yield Four Years after the Application. *Proceedings of the 19th EGU General Assembly, EGU2017*, Vienna, 23-28 April 2017, 1600.
- [20] Hollman, P.C.H. (2001) Evidence for Health Benefits of Plant Phenols: Local or Systemic Effects? *Journal of the Science of Food and Agriculture*, **81**, 842-852. <https://doi.org/10.1002/jsfa.900>
- [21] Shi, W. (2011) Chapter 3. Agricultural and Ecological Significance of Soil Enzymes: Soil Carbon Sequestration and Nutrient Cycling. In: Shukla, G. and Varma, A., Eds., *Soil Enzymology*, Springer, Berlin/Heidelberg, 43-60. https://doi.org/10.1007/978-3-642-14225-3_3
- [22] Andrews, R.K., Blakeley, R.L. and Zerner, B. (1989) Urease: A Ni(II) Metalloenzyme. In: Lancaster, J.R., Ed., *The Bioinorganic Chemistry of Nickel*, VCH, New York, 141-166.
- [23] Simpson, J.R., Freney, J.R., Wetselaar, R., Muirhead, W.A., Leuning, R. and Denmead, O.T. (1984) Transformations and Losses of Urea Nitrogen after Application to Flooded Rice. *Australian Journal of Agricultural Research*, **35**, 189-200. <https://doi.org/10.1071/AR9840189>
- [24] Polacco, J.C. (1977) Is Nickel a Universal Component of Plant Ureasases? *Plant Science Letters*, **10**, 249-255. [https://doi.org/10.1016/0304-4211\(77\)90136-5](https://doi.org/10.1016/0304-4211(77)90136-5)
- [25] Mobley, H.L.T. and Hausinger, R.P. (1989) Microbial Urease: Significance, Regulation and Molecular Characterization. *Microbiological Reviews*, **53**, 85-108. <https://doi.org/10.1128/mr.53.1.85-108.1989>
- [26] Lef, L.K. and Nannipieri, P. (1995) *Methods in Soil Microbiology and Enzyme Activities*, Academic Press/Harcourt Brace and Company, London, 225-230.
- [27] Splading, B.P. (1979) Effect of Divalent Metal Cations Respiration and Extractable Enzymes Activities of Douglass-Fir Needle Litter. *Journal of Environmental Quality*, **8**, 105-109. <https://doi.org/10.2134/jeq1979.00472425000800010023x>
- [28] Martínez-Valdivieso, D., Gómez, P., Font, R. and Del Río-Celestino, M. (2015) Mineral Composition and Potential Nutritional Contribution of 34 Genotypes from Different Summer Squash Morphotypes. *European Food Research and Technology*, **240**, 71-81. <https://doi.org/10.1007/s00217-014-2308-7>
- [29] Oloyede, F., Agbaje, G.O., Obuotor, E.M. and Obisesan, I.O. (2012) Nutritional and Antioxidant Profiles of Pumpkin (*Cucurbita pepo* Linn.) Immature and Mature Fruits as Influenced by NPK Fertilizer. *Food Chemistry*, **135**, 460-463. <https://doi.org/10.1016/j.foodchem.2012.04.124>
- [30] (2018) *Vegetable Production Guide for Commercial Growers*. University of Kentucky, Department of Horticulture, College of Agriculture, Food and Environment, University of Kentucky, Lexington, ID-36.
- [31] United States Department of Agriculture (2016, September 6) United States Standards for Grades of Summer Squash. Specialty Crops Inspection Division Specialty Crops Program, USDA, Washington DC.

- <https://www.ams.usda.gov/sites/default/files/media/SummerSquashStandard.pdf>
- [32] McGrath, R.M., Kaluza, W.Z., Daiber, K.H., Van der Riet, W.R. and Glennie, C.W. (1982) Polyphenols of Sorghum Grain, Their Change during Malting and Their Inhibitory Nature. *Journal of Agricultural and Food Chemistry*, **30**, 450-456. <https://doi.org/10.1021/jf00111a010>
- [33] Antonious, G.F. and Kasperbauer, M.J. (2002) Color of Light Reflected to Leaves Modifies Nutrient Content of Carrot Roots. *Crop Science*, **42**, 1211-1216. <https://doi.org/10.2135/cropsci2002.1211>
- [34] Hashmi, M.H. (1973) Spectrophotometric Determination with Potassium Ferricyanide. In: *Assay of Vitamins in Pharmaceutical Preparations*, John Wiley and Sons, Hoboken, 311-312.
- [35] VanEtten, C.H., McGrew, C.E. and Daxenbichler, M.E. (1974) Glucosinolate Determination in Cruciferous Seeds and Meals by Measurement of Enzymatically Related Glucose. *Journal of Agricultural and Food Chemistry*, **22**, 483-487. <https://doi.org/10.1021/jf60193a013>
- [36] Tabatabai, M.A. and Bremner, J.M. (1972) Assay of Urease Activity in Soils. *Soil Biology and Biochemistry*, **4**, 479-487. [https://doi.org/10.1016/0038-0717\(72\)90064-8](https://doi.org/10.1016/0038-0717(72)90064-8)
- [37] American Public Health Association (APHA) (1995) Standard Methods for the Determination of Water and Wastewater. 19th Edition.
- [38] Balasubramanian, D., Bagyaraj, D.J. and Rangaswami, G. (1970) Studies on the Influence of Foliar Application of Chemicals on the Microflora and Certain Enzyme Activities in the Rhizosphere of *Eleusine coracana* Gaertn. *Plant and Soil*, **32**, 198-206. <https://doi.org/10.1007/BF01372858>
- [39] SAS Institute Inc. (2016) SAS/STAT Guide, Version 6.4. SAS Inc., Campus Drive.
- [40] Kuno, T., Tsukamoto, T., Hara, A. and Tanaka, T. (2012) Cancer Chemoprevention through the Induction of Apoptosis by Natural Compounds. *Biophysical Chemistry*, **3**, 156-173. <https://doi.org/10.4236/jbpc.2012.32018>
- [41] CDDEP (2015) The State of the World's Antibiotics. Center for Disease Dynamics, Economics & Policy, Washington DC. https://cddep.org/publications/state_worlds_antibiotics_2015/
- [42] Kentucky Health News (2012, November 14) The Land Report Kentucky Business News Source. <http://www.Kentucky.com/living/health-and-medicine/article44389116.html>
- [43] Yang, Q., Zhang, H., Guo, Y. and Tian, T. (2016) Influence of Chicken Manure Fertilization on Antibiotic-Resistant Bacteria in Soil and the Endophytic Bacteria of Pakchoi. *International Journal of Environmental Research and Public Health*, **13**, 662. <https://doi.org/10.3390/ijerph13070662>
- [44] Wuana, R.A. and Okieimen, F.E. (2011) Heavy Metals in Contaminated Soils: A Review of Sources, Chemistry, Risks and Best Available Strategies for Remediation. *International Scholarly Research Notices*, **2011**, Article ID: 402647. <https://doi.org/10.5402/2011/402647>
- [45] Yang, X., Feng, Y., He, Z. and Stoffella, P.J. (2005) Molecular Mechanism of Heavy Metals Hyperaccumulation and Phytoremediation. *Journal of Trace Elements in Medicine and Biology*, **18**, 339-353. <https://doi.org/10.1016/j.jtemb.2005.02.007>
- [46] Szczygłowska, M., Piekarska, A., Konieczka, P. and Namiesnił, J. (2011) Use of Brassica Plants in the Phytoremediation and Biofumigation Processes. *International Journal of Molecular Sciences*, **12**, 7760-7771. <https://doi.org/10.3390/ijms12117760>
- [47] Liu, S., Yang, Z., Wang, X., Gao, R. and Liu, X. (2007) Effects of Cd and Pb Pollu-

- tion on Soil Enzymatic Activities and Soil Microbiota. *Frontiers of Agriculture in China*, **1**, 85-89. <https://doi.org/10.1007/s11703-007-0016-9>
- [48] Park, J., Choppala, G., Bolan, N., Chung, J. and Chuasavathi, T. (2011) Biochar Reduces the Bioavailability and Phytotoxicity of Heavy Metals. *Plant Soil*, **348**, 439-451. <https://doi.org/10.1007/s11104-011-0948-y>
- [49] Kumar, S., Mastro, R., Ram, L., Sarkar, P., George, J. and Selvi, V. (2013) Biochar Preparation from *Parthenium hysterophorus* and Its Potential Use in Soil Application. *Ecological Engineering*, **55**, 67-72. <https://doi.org/10.1016/j.ecoleng.2013.02.011>
- [50] Lehmann, J., Rillig, M.C., Thies, J., Masiello, C.A., Hockaday, W.C. and Crowley, D. (2011) Biochar Effects on Soil Biota—A Review. *Soil Biology and Biochemistry*, **43**, 1812-1836. <https://doi.org/10.1016/j.soilbio.2011.04.022>
- [51] Wu, F., Jia, Z., Wang, S., Chang, S. and Startsev, A. (2013) Contrasting Effects of Wheat Straw and Its Biochar on Greenhouse Gas Emissions and Enzyme Activities in a Chernozemic Soil. *Biology and Fertility of Soils*, **49**, 555-565. <https://doi.org/10.1007/s00374-012-0745-7>
- [52] Lammirato, C., Miltner, A. and Kaestner, M. (2011) Effects of Wood Char and Activated Carbon on the Hydrolysis of Cellobiose by b-Glucosidase from *Aspergillus niger*. *Soil Biology and Biochemistry*, **43**, 1936-1942. <https://doi.org/10.1016/j.soilbio.2011.05.021>