

Stability of Soil and Biosolid Nanocolloid and Macrocolloid Particles in the Absence and Presence of Arsenic, Selenium, Copper and Lead

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Abstract

Due to their enhanced stability and contaminant transport potential, environmental nanoparticles derived from soil and biosolid materials may pose a considerable risk to groundwater quality. Very little information exists on the stability and transportability of environmental or natural nanocolloids in the presence of As, Se, Pb and Cu contaminants, all of which are considered to represent substantial threats to human and animal populations through groundwater contamination. This study involved stability settling experiments of nanocolloids (NCs) (<100 nm) and macrocolloids (MCs) (100 - 2000 nm) fractionated from Bt horizons of three Kentucky soils and one biosolid waste material in water suspensions of 0, 2, and 10 mg·L⁻¹ of As, Se, Pb and Cu. The results indicated greater stability in the mineral than the biosolid colloid fractions, and enhanced stability of NCs over corresponding MCs in the presence or absence of contaminants at low contaminant loads. At high contaminant loads nearly all colloids were unstable except for the bio-nanocolloids which still sustained considerable stability. At low contaminant loads, the MC fraction stability sequence was smectitic > mixed > kaolinitic > biosolid. Among the nano-fractions, the smectitic and kaolinitic colloids demonstrated lower stability than the MCs, but higher than those of the mixed and biosolid fractions. Physicochemical characterizations indicated that extensive organic carbon surface coatings and higher Al/Fe:Si ratios may have induced higher stability in the NC fractions, but their overall stability may also have been hindered in some cases by nano-aggregation phenomena.

Keywords

Nanocolloids, Macrocolloids, Colloid Stability, Contaminant Transport, Nanoparticles

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1. Introduction

Water dispersed colloids (WDC) are known to disperse from soil aggregates and remain mobile in subsurface environments [1] [2]. They have average diameters between 1 and 1000 nm, which includes nanoparticles, with one dimension equal to or less than 100 nm [3]-[5]. These naturally derived NCs are prevalent in the environment and play various roles in environmental processes such as the cycling of nutrients, remediation procedures, contaminant transport and soil genesis [6].

Nanoparticle stability can be used as an indicator of potential transport into groundwater supplies. The tendency for particles to aggregate or remain stable in solution may be affected by the complex mineralogical and physico-chemical attributes of the particle, including particle size, surface chemistry, and the aqueous environment surrounding transport [5] [6]. Even though the small size of nanoparticles may promote long-range stability and transport due to the proportionality of their displacement to the inverse square root of their radius, their high surface energy may also result in multiple interparticle collisions and aggregation, especially at increased ionic strengths [6] [7]. Chemically, colloid stability can be influenced by solution ionic strength and composition, particle surface coatings and functional groups, pH, and zeta potential [6] [8]. Stability can also be a function of the mineralogical composition of the particles. Studies have shown that kaolinitic particles tend to flocculate, while smectitic particles tend to remain dispersed or stable over time [1] [9]. An important consideration in natural systems includes their complex composition, which consists of mixed colloidal phases. This encompasses the interactions with other NCs and solutes in the system, particularly since nanoparticle mobility is controlled mainly by Brownian motion and not by gravitational settling like their larger counterparts [10].

Nanoparticles and NCs can carry contaminants, illustrating their potential role in environmental pollution, water quality, and remediation processes [11] [12]. Four contaminants that are of particular concern include arsenic (As), selenium (Se), copper (Cu), and lead (Pb), all of which are considered toxic to humans, wildlife and plants at relatively high concentrations. All four of these contaminants are accumulated in the environment from anthropogenic sources/industrial runoff, including car batteries, where arsenic is used to strengthen copper and lead alloys [13], or from naturally occurring sources like seleniferous and arsenopyrite-derived soils [14] [15]. They may be readily transported into groundwater by naturally occurring soil and biosolid colloids. While studies have characterized, modeled, and predicted the movement of water dispersible colloids [1] [11] [16] [17], there is a lack of information comparing the stability of macro-sized (0.1 - 2.0 μ m) to nano-sized colloids (<0.1 μ m) of differing mineralogy from subsurface horizons, and their potential to transport contaminants such as Se, As, Cu, and Pb. Additionally, there is little to no information available on the role of nano-sized biosolid-derived colloids, despite studies showing that larger sized bio-colloids enhance contaminant transport through soil [18]-[20].

The objectives of this study were to evaluate and compare the stability of water suspended NCs and MCs derived from Bt horizons of 3 Kentucky soils with kaolinitic, montmorillonitic, and mixed mineralogies, as well as from an aerobically digested biosolid with three levels of Cu, Pb, As, and Se contaminant loads.

2. Methods and Materials

Mineral colloids were fractionated from the Bt horizons of three Kentucky soils with differing mineralogy: Caleast-variant (fine, smectitic, mesic mollic Hapludalf), Tilsit (fine-silty, mixed, mesic Typic Fragiudult), and Trimble (fine-loamy, siliceous, mesic Typic Paleudult). The recovered colloids are referred to as smectitic, mixed, and kaolinitic NCs or MCs, respectively. An aerobically digested municipal sewage sludge, obtained from Jessamine County, Kentucky, was fractionated to obtain the biosolid NCs and MCs. Centrifugation was used to fractionate the WDCs into two size classes (NCs < 100 nm and MCs 100 - 2000 nm) using a Centra GP8R Model 120 centrifuge (ThermoIEC) in deionized water (resistivity of 1 $\mu\Omega$ cm at 25°C). The clay fraction was separated from the bulk soil using centrifugation at 107 RCF for 3.5 minutes, and then at 4387 RCF for 46 minutes to separate the NCs from the MCs [6]. Centrifugation times were determined using Stokes law, and separation of the NC from MC fraction was performed on a centrifuge with a rotor radius of 170 mm, a speed of 4387 RCF, a density difference from water of 1650 kg·m⁻³, and viscosity of 0.0008904 Pas. All collected sample suspensions were then diluted to 50 mg·L⁻¹ concentrations for additional analysis.

2.1. Stability Experiments

Settling kinetics experiments were used to determine the stability of the NCs and MCs over time. Duplicate 400

mL suspensions of 50 mg colloid L^{-1} in de-ionized water (D-H₂O) were used to generate stability graphs based on sampled concentrations at times 0, 30 minutes, 1, 2, 4, 8, 24 and 48 hours. Additional stability experiments were performed using the same procedure with the addition of 2 and 10 mg L^{-1} mixed contaminant concentrations of Pb, Cu, As, and Se [prepared as aqueous solutions from: PbCl₂ (98% purity, Aldrich Chemicals, Milwaukee, WI), CuCl₂ (>99% purity, Sigma Chemical Company, St. Louis, MO), arsenic acid Na₂HAsO₄·7H₂O (98% purity, Sigma Chemical Company, St. Louis, MO), and sodium selenate decahydrate Na₂SeO₄·10H₂O (99.9% purity, Sigma Chemical Company, St. Louis, MO)]. The suspended colloid concentrations were determined using a colorimetric procedure on a Molecular Devices Versa Max Microplate Reader at 450 nm [2] [20].

2.2. Particle Size, Morphology, and Surface Area Analysis

Primary particle size of NC particles was determined using a high resolution transmission electron microscope (JEOL 2010F) attached to an Oxford EDS detector (TEM-EDS; JEOL 2010F, Tokyo, Japan) [21] [22]. Primary particle size of MC particles was determined using a S-4300 scanning electron microscope, equipped with a Princeton Gamma-Tech EDS Microanalysis system (SEM-EDS; Hitachi S-4300, Tokyo, Japan) [23]. The average diameters were calculated using ImageJ software (ImageJ 1.46r, Wayne Rasband, National Institutes of Health, USA). A Malvern Instruments ZetasizerNano ZS (Malvern, United Kingdom) was used to obtain intensity weighted mean particle hydrodynamic diameters (z-average diameter, d_h) on suspensions of 50 mg collo-id·L⁻¹ using dynamic light scattering (173° backscatter analysis method). This procedure also allowed estimates of NC and MC aggregation potentials in the absence and presence of 2 mg·L⁻¹ additions of As, Cu, Pb and Se contaminants. Surface area analysis was performed on both the NCs and MCs using the Ethylene Glycol Monoethyl Ether (EGME) method.

2.3. Mineralogical Characterization

A combination of X-ray diffraction (XRD) and Thermogravimetric analysis (TG) was employed for mineralogical characterization. For XRD analysis, K, K-heat treated, Mg, and Mg-glycol saturated samples were collected on glass slides and analyzed on a Phillips PW 1840 diffractometer and PW 1729 X-ray generator (Mahwah, NJ) fitted with a cobalt X-ray tube and run at 40 kV and 30 mA using a Bragg-Bretano design goniometer at a scanning rate of $0.05^{\circ}2\theta$ per minute from 2° to 40° with a scattering slit of 0.1° . Mg-saturated colloids were also used for TG analysis on a Thermal Analyst 2000 (TA Instruments) equipped with a 951 Thermogravimetric Analyzer (DuPont Instruments) with a heating rate of 20° C/min under N₂ atmosphere [24]. The TG Analysis was used to verify the presence of kaolinite, as well as goethite and gibbsite, and to compliment quantification interpretations derived from the XRD patterns [24].

2.4. Chemical Characterization

A Denver Instruments Model 250 pH*ISE* electrical conductivity meter (Arvada, CO) was used to measure pH and electrical conductivity (EC) of the NC and MC suspensions. Ionic strength (IS) was derived by multiplying electrical conductivities by 0.0127 [25]. A Varian Spectr AA 50B atomic absorption spectrometer was used to determine the base cation concentrations (Ca^{2+} , Mg^{2+} , K^+ , and Na^+), which were summed and used to report the cation exchange capacity (CEC) from triplicate samples of the NCs and MCs using an adapted version of the ammonium acetate method. Concentrations of Ca^{2+} , Mg^{2+} , and Na^+ were used to calculate sodium adsorption ratios (SAR). A Flash EA 1112 Series Soil Analyzer (Thermo Electron Corporation) with a Mettler Toledo MX5 microbalance was used to determine organic carbon (OC). Zeta potential measurements in absence of contaminants were used to estimate surface and point of zero charge (PZC), as well as suspension stability at adjusted pH values of 4, 6, 8 and 10 using 0.01 N HCl and NaOH. The Smoluchowski approximations determined zeta potentials from electrophoretic mobilities on suspensions in 0.001 M NaCl background electrolyte as measured on a Malvern Zetasizer Nano ZS (Malvern, United Kingdom). Additionally, zeta potentials were measured in the presence of 2 and 10 mg·L⁻¹ additions of Cu, Pb, As and Se.

2.5. Statistical Analysis

The accepted error levels for all duplicate and triplicate measurements were $\leq 15\%$. Significance of differences between means was tested using Analysis of Variance (ANOVA) (SAS PROC GLM) and Fisher's protected

least significant difference test (LSD) in SAS 9.3 (SAS Institute Inc., Cary, NC, USA). The statistical significance level used was $\alpha = 0.05$.

3. Results and Discussion

3.1. Colloid Stability

3.1.1. Effect of Particle Size

Primary particle size analysis by scanning electron microscopy (SEM) indicated size ranges between 288 and 549 nm for the MCs, while transmission electron microscopy (TEM) portrayed size ranges between 7 and 50 nm for the NCs (**Table 1**). Stability settling characteristics of the colloid suspensions as affected by size and contaminant load are shown in **Figure 1** and **Table 2**. A particle is considered stabile if it tends to remain dispersed in solution [2]. Based on size alone, the settling experiments showed NCs to be significantly more stable in suspension ($\alpha = 0.05$) after 48 hours than the MCs in the presence and absence of contaminants (**Figure 1**). After 48 hours, in the absence of contaminants mineral MCs had an average of 80% colloids in suspension while NCs had an average of 96% (**Table 2**). In comparison, the bio-MCs and bio-NCs had 36% and 58%, respectively (**Table 2**). This highlights two things: first, the enhanced mobility and contaminant transport potential of the NCs over their corresponding MCs, and second, the greater stability of the mineral NCs had an average of 35% more colloids in suspension than corresponding MCs (65% versus 30%, respectively) (**Table 2**). Despite greater stability in the mineral colloids at 2 mg/L contaminant additions, the bio-NCs had as much as 18% colloids in suspension in the presence of 10 mg/L contaminant, compared to only 1% on average for the mineral colloids (**Table 2**) [26] [27].

The average intensity weighted (Z-average) hydrodynamic diameters (d_h) can be used to indicate stability as well (Figure 2). Since d_h is related to the size of aggregates formed by the colloids, the smaller values would

Properties		Size Class	Surface Area $(m^2 \cdot g^{-1}) \pm SD^{\ddagger}$	$\frac{\text{SEM/TEM}^{\$} \text{ Mean Smallest}}{\text{Particle Size} \pm \text{SD}^{\ddagger} (nm)}$	SEM Images (Macro)	TEM Images (Nano)	
	Smectitic	Macro	708 ± 137	328 ± 144	K	1 al	
		Nano	879 ± 76	37 ± 13	- WD15-1mm 20.70kV \$255 2um	50 nm	
	Mixed	Macro	420 ± 105	549 ± 394			
		Nano	466 ± 10	7 ± 5	PD15_3Bar 40_00XV (2051) 1000	<u>50</u> nm	
Colloids	Kaolinitic	Macro	333 ± 37	288 ± 184		and the second	
		Nano	389 ± 44	41 ± 19	2 jun movement of and the	<u>5 mi</u>	
	Biosolid	Macro	1674 ± 70	363 ± 338		s.F	
		Nano	1303 ± 63	50 ± 19	Amite cam at over where	20 nm	

Table 1. Selected physical and morphological characteristics of the macrocolloid and nanocolloid fractions.

[‡]SD = Standard Deviation from duplicate or triplicate measurements. [§]SEM = Scanning Electron Microscopy, TEM= Transmission Electron Microscopy data represent the average smallest dimension of 300 representative particles from three to eight images of each size fraction as measured until the average and standard deviation values had less than 10% variation.





Figure 1. Stability of the macro- and nano-colloids in mixtures of equal parts (0, 2, and 10 mg·L⁻¹) of each contaminant (As, Se, Cu and Pb) expressed as % of colloids remaining in suspension after 48 hours (error bars represent standard error of means; upper case letters represent significant differences ($\alpha = 0.05$) between mineralogy at the same level of contaminant concentration; lower case letters represent trends within the same mineralogy across the 0, 2, and 10 mg·L⁻¹ contaminant concentration gradients).

Table 2. Average % of mineral and biosolid colloids still in suspension at 48 nours.									
Composition:	Mine	eral	Biosolid						
Size:	Macro	Nano	Macro	Nano					
Without contaminants (Control)	80%	96%	36%	58%					
With 2 mg/L contaminants	45%	65%	8%	30%					
With 10 mg/L contaminants	1%	1%	1%	18%					



Figure 2. Dynamic Light Scattering (DLS) intensity weighted hydrodynamic diameters (d_h) of macrocolloids (a) and nanocolloids (b) in the absence and presence of 2 mg/L contaminants after a 48 hour interaction (error bars represent standard error of the means). indicate smaller aggregate size, thus more dispersive colloids. In the absence of contaminants the d_h values complemented the stability trends, indicating greater stability in the mineral NCs over that of the bio-NCs (**Figure 1** and **Figure 2**). For the MCs, the anomaly was the kaolinitic MC fraction, which showed the smallest d_h (**Figure 2**), yet in the stability settling experiments (**Figure 1**) portrayed lower stability than the smectitic and mixed MC fraction. Other than the MC kaolinitic fraction, the smectitic colloids showed consistently small d_h values and high stability in the settling studies compared to other colloid compositions (**Figure 1** and **Figure 2**). Since d_h also reflects on the size of aggregates formed by the colloids, the smaller values indicate smaller aggregate size thus more dispersive colloids. The d_h values in the absence of contaminants suggested that the MCs were nearly four-times larger than the NCs, with an average d_h of 842 nm for the MCs and 224 nm for the NCs (**Figure 2**). The d_h also showed compositional trends, with mean comparisons showing the bio-colloids to be significantly larger than the mineral colloids ($\alpha = 0.05$). While the larger d_h values (as measured by DLS) compared to the crystallite diameters (as measured by TEM/SEM) may indicate some aggregation in both size fractions, the larger DLS size range may also be partially explained by the disproportionate influence of the larger particles on the overall average diameter compared to the smaller particles (**Figure 1**), or by potential surface coatings on the particles [8] [16].

The d_h values of the kaolinitic MCs, the smectitic and mixed NCs were unaffected by the addition of contaminants, but increased significantly for the smectitic MCs, and the kaolinitic, and biosolid NCs. Surprisingly, the biosolid MCs experienced a considerable reduction in size in the presence of contaminants (Figure 2). The higher d_h values in the presence of contaminants were probably caused by enhanced aggregation due to increased ionic strength from the addition of polyvalent ions (Figure 2) [8] [16]. MCs showed greater aggregation potentials in the presence of contaminants than NCs through larger shifts in dh values, especially in the biosolid and mixed MCs (Figure 2). The largest d_h shifts with contaminant additions occurred in the kaolinitic and biosolid NCs and the mixed and biosolid MCs (Figure 2). Greater contaminant loading potential of cation contaminants onto larger surface areas could cause aggregation of other negatively charged colloids, thus creating pseudo-aggregates that increase the intensity weighted averages measured (Figure 2) [8] [16]. In spite of a size increase of about 230 nm, the smectitic MC fraction showed a slight reduction in stability comparable to that of the NC fraction. In contrast, the stability of the kaolinitic, mixed, and biosolid MC fraction decreased significantly regardless of the d_h size changes (Figure 2) following the mixed > kaolinitic > biosolid trend. The addition of the 2 mg·L⁻¹ contaminant load caused a consistent stability reduction across all NCs following the sequence smectitic = kaolinitic > mixed > biosolid. However, the stability of the NCs was generally higher than that of the MCs under the 2 mg \cdot L⁻¹ contaminant load, with the exception of the smectitic MCs which showed similar stability to the smectitic NCs.

3.1.2. Effects of Mineralogy

Mineralogy appears to have played a partial role in the stability processes of the MCs and NCs (Table 3). In the absence of contaminants, the smectitic MCs had greater stability than the kaolinitic and biosolid MCs, but equal stability to the mixed MCs (Figure 1). In the absence of contaminants, the smectitic, mixed and kaolinitic NCs were more stable than the biosolid NCs (Figure 1). After additions of 2 mg L^{-1} contaminants the mineral colloids showed more surface repulsion than the biosolid colloids. The smectitic and kaolinitic NCs were more stable than the mixed and biosolid colloids, while within the MC fraction, the smectitic and mixed MCs were more stable than the kaolinitic and biosolid colloids (Figure 1). With the exception of the bio-NCs, the addition of 10 $mg \cdot L^{-1}$ contaminants overwhelmed the available surface area of the colloids and induced significant flocculation (Figure 1). The greater stability (Figure 1) of the smectitic colloids over that of other mineralogies is likely due to higher surface charge potential, with the greater charge densities in smaller sized colloids resulting in greater repulsion (Table 1, Figure 1) [1] [28]. Lower stability in colloids containing kaolinite are likely due to the tendency of kaolinite to remain flocculated at pH less than 7.5 [1] [9]. Additionally, smaller sized fractions that are high in kaolinite, Al and Fe hydroxides may have enhanced surface charge density contributing to destabilization potentials [2] [8]. This is evidenced by the attachment of smaller nanoparticles to several MC surfaces (Table 1). Additionally, the presence of hydroxy-interlayered vermiculite may have released Al and promoted flocculation processes in the mixed and kaolinitic colloid fractions (Figure 1 and Figure 2; Table 3) [2]. In the stability kinetics experiments, mixed MCs were more stable than kaolinitic MCs after the addition of $2 \text{ mg} \cdot \text{L}^{-1}$, with a reversed stability sequence for the NCs (Figure 1). The mixed NCs were less stable than the kaolinitic NCs in the presence of 2 mg L^{-1} contaminants, probably due to their higher Fe-hydroxide content (Figure 1,

rable 5. Mineralogical composition of nano- and macro-conoid fractions.						
g	Macro	$^{*}K_{29}, Ge_{7}, Gi_{0}, Q_{6}, M_{10}, Sm_{48}, MVI_{0}, HIV_{0}$				
Smectitic	Nano	*K ₂₉ , Ge ₇ , Gi ₀ , Q ₆ , M ₁₀ , Sm ₄₈ , MVI ₀ , HIV ₀ *K ₃₀ , Ge ₉ , Gi ₀ , Q ₄ , M ₆ , Sm ₅₁ , MVI ₀ , HIV ₀ *K ₄₂ , Ge ₅ , Gi ₀ , Q ₅ , M ₃₁ , Sm ₀ , MVI ₇ , HIV ₁₀ *K ₄₆ , Ge ₇ , Gi ₀ , Q ₃ , M ₃₀ , Sm ₀ , MVI ₇ , HIV ₇ *K ₅₂ , Ge ₁₂ , Gi ₅ , Q ₄ , M ₃ , Sm ₀ , MVI ₀ , HIV ₂₄ *K ₅₅ , Ge ₁₅ , Gi ₆ , Q ₂ , M ₃ , Sm ₀ , MVI ₀ , HIV ₁₉ *ND				
Minad	Macro	$^{*}K_{42}, Ge_{5}, Gi_{0}, Q_{5}, M_{31}, Sm_{0}, MVI_{7}, HIV_{10}$				
Mixed	Nano	[*] K ₄₆ , Ge ₇ , Gi ₀ , Q ₃ , M ₃₀ , Sm ₀ , MVI ₇ , HIV ₇				
Vaclinitia	Macro	[*] K ₅₂ , Ge ₁₂ , Gi ₅ , Q ₄ , M ₃ , Sm ₀ , MVI ₀ , HIV ₂₄				
Kaominuc	Nano	[*] K ₅₅ , Ge ₁₅ , Gi ₆ , Q ₂ , M ₃ , Sm ₀ , MVI ₀ , HIV ₁₉				
Dissolid	Macro	*ND				
BIOSOIIU	Nano	*ND				

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 aı	710	· •	111110	Jaiogical	composition o	i nano- ano	

K = Kaolinite, Ge = Goethite, Gi = Gibbsite, Q = Quartz, M = Mica, Sm = Smectite, MVI = Mica-Vermiculite Interstratified, HIV = Hydroxy-interlayered Vermiculite, ND = Not Detected.

Table 3). One confounding observation is that there are higher quantities of flocculating agents (Fe and Al hydroxides, goethite, and gibbsite) in the NCs, yet they remain more stable than their corresponding MCs (**Table 3**, **Figure 1**). The answer to this dilemma may lie in the EDS data which indicated greater (Al + Fe):Si ratios in the NCs than in their corresponding MCs. Increased (Al + Fe):Si ratios have shown to enhance humic acid sorption capacities and stabilize colloid suspensions [16] [29].

3.1.3. Effects of Particle Morphology

The smectitic MCs showed typical montmorillonitic honeycomb shapes with edge-face interactions (**Table 1**). Montmorillonite can be more dispersive than other minerals because weak Van der Waals' forces between oriented tactoids allow penetration between layers by water and exchangeable cations, which may inhibit stable aggregate formation [9]. The smectitic NCs indicated less particle to particle attachment than the MCs (**Table 1**). Morphologies displayed by the mixed mineralogy MCs showed tightly aggregated honeycomb-like plates, while the mixed NCs had various shapes encompassing tubes, rods, plates and hexagons within a tightly aggregated mineral matrix (**Table 1**). The greater morphological variation in the shapes of the mixed NCs in TEM images may indicate some surface deformation (**Table 1**). Similar HIV images were shown with associated iron minerals [21]. Kaolinitic MC SEM images had the typical hexagonal shapes displayed by kaolinitic minerals, while TEM images of the NCs showed much smaller hexagonal shapes with a noted increase in iron minerals embedded within and binding the hexagonal aggregates (**Table 1**) [22]. Kaolinitic minerals tend to flocculate below pH 7, which is representative of the colloid suspension pH range [9] (**Table 4**).

The morphological compositions of the biosolid MCs, as shown in their SEM images, displayed various aggregations of organic materials with some quartz grains present, while the NC TEM images lacked distinct particle shapes or displays of individual particle morphology, indicating less integrity of sample and potential dissolution (**Table 1**). Overall, within the mineral colloids, SEM images displayed more tightly held aggregates within the MCs (**Table 1**), indicating greater flocculation potential (**Figure 1**) as opposed to the more loosely held aggregates displayed by their corresponding NCs in TEM images (**Table 1**).

Additionally, TEM images suggested potential particle morphology alterations from surface disorder, which may have modified NC stability behavior (**Table 1**). TEM images of the smectitic and mixed NC showed similar disorder and potential structural abnormalities (**Table 1**), with particles adhering to each other to display extended and oblong morphologies. The cause of such abnormalities has been suggested to be impurities or oriented aggregation resulting from forced structural incorporation of previously sorbed species that could alter surface sites and influence stability and reactivity characteristics [5] [30]-[32].

3.1.4. Surface Area Effects

With the exception of the biosolids, NCs had greater surface area than the MCs despite the SEM indications of nanoparticle adhesion to MC surfaces (**Table 1**) [5] [6] [30]. Surface area showed trends by composition, with the largest coming from the biosolid materials, followed by the smectitic, mixed, and kaolinitic colloids, respectively (**Table 1**). In addition to size effects, increased surface area values measured in the NCs could be the result of organic surface coatings or the presence of iron hydroxides maximizing sorption of the EGME [33] [34].

The larger surface areas in the NCs also indicate greater potential for contaminant sorption and increased repulsion of similarly charged particles, thus enhancing stability potentials.

3.1.5. Effect of Chemical Characteristics

Unadjusted pH values ranged from 4.9 to 5.4 for both the NC and MC fractions (**Table 4**). Such a narrow pH range evinces that pH values may not have promoted differences in stability between the differing colloid compositions or sizes, but may have contributed to a greater overall flocculation tendency (**Table 4**, **Figure 1**) [2]. However, in spite of this narrow pH range, there were significant differences in stabilization potentials (**Figure 1**) across most of the studied colloid fractions. The same colloids in higher pH environments would likely be even more dispersive, as evidenced by the zeta potential titrations above pH 6 (**Figure 3**).

The electrical conductivity (EC) and ionic strength (IS) of the colloids were relatively low with the NCs showing higher overall values than their corresponding MCs (**Table 4**). Additionally, the bio-colloids had greater EC and ionic strength values than most mineral colloids except for the mixed NCs (**Table 4**). Higher ionic strength values for the NCs could decrease stability [35] and may have contributed to lower fractional recoveries as compared to MCs (**Table 1** and **Table 4**). However, at the natural range these higher EC and ionic strength values of the NC suspensions did not appear to significantly deter the overall stability of the NCs as shown in **Figure 1**. Increased ionic strength effects from contaminant additions resulted in lower NC and MC stability, likely due to flocculation in the presence of contaminants, with greater flocculation at higher contaminant concentrations (**Figure 1**) [6] [16]. Increased ionic strength has been shown to reduce the effectiveness of Coulomb repulsion by shielding the charge of two approaching particles [6]. This effect on Coulomb repulsion may explain the stability findings after contaminant additions of 10 mg·L⁻¹, where significantly lower colloid suspension concentrations indicated flocculation in all but the bio-NC fractions (**Figure 1**). Coulomb repulsions appeared to overcome ionic strength effects at 2 mg·L⁻¹ contaminant concentrations.

The higher SAR values of the NCs over that of the MCs are more consistent with their stability trends than the IS values, indicating greater dispersion potentials in the NCs over their corresponding MCs (**Table 4**). The higher CEC values of the NCs over the MCs are also complimentary to their larger and likely more reactive surface areas (**Table 4**). As expected, the smectitic colloids had the greatest CEC within the mineral colloids, followed by the mixed mineralogy and kaolinitic colloids (**Table 4**). The higher OC content of the bio-colloids



Figure 3. Zeta potential of nanocolloid and macrocolloid fractions at pH 4, 6, 8, and 10.

Р	roperties	Size Class	Electrical Conductivity (mmhos·cm ⁻¹)	Ionic Strength [§] $(mol \cdot L^{-1})$	Natural pH	$\begin{array}{c} \text{CEC}^{\#} \\ (\text{cmol}_{c} \cdot \text{kg}^{-1}) \\ \pm \text{SD}^{\ddagger} \end{array}$	Ca^{2+} (cmol _c ·kg ⁻¹)	Mg^{2+} (cmol _c ·kg ⁻¹)	K^+ (cmol _c ·kg ⁻¹)	Na^+ (cmol _c ·kg ⁻¹)	$SAR^{\dagger\dagger}$	$OC (mg \cdot kg^{-1})^{\ddagger \ddagger}$
Colloids	Smectitic	Macro	3.93×10^{3}	4.99×10^{-5}	4.92	35.05 ± 12.84	27.60	6.27	0.57	0.61	0.15	658
		Nano	$6.07\times10^{\text{-3}}$	$7.71 imes 10^{-5}$	5.12	42.19 ± 15.12	32.64	8.00	0.78	0.77	0.17	897
	Mixed	Macro	$2.91\times 10^{\text{-3}}$	$3.70 imes 10^{-5}$	5.07	8.89 ± 1.62	3.80	3.40	1.15	0.54	0.28	645
		Nano	3.09×10^{-2}	$3.92 imes 10^{-4}$	4.92	10.51 ± 1.67	4.00	3.47	2.80	0.24	0.12	774
	Kaolinitic	Macro	2.87×10^{3}	$3.64 imes 10^{-5}$	4.91	6.94 ± 1.85	4.40	1.60	0.45	0.49	0.28	430
		Nano	3.80×10^{-3}	4.83×10^{-5}	5.38	13.12 ± 2.84	7.12	3.73	1.23	1.04	0.45	647
	Biosolid	Macro	1.56×10^{-2}	$1.97 imes 10^{-4}$	5.39	37.61 ± 14.85	31.60	3.60	1.64	0.77	0.18	1.3K
		Nano	$4.69 imes 10^{-2}$	5.96×10^{-4}	5.25	70.99 ± 22.98	51.68	12.13	3.98	3.20	0.57	16K

Table 4. Selected chemical characteristics of the macro- and nano-colloid fractions.

 † SD = Standard Deviation from duplicate or triplicate measurements. $^{\$}$ Ionic Strength (IS) = Estimated using Griffin and Jurinak's equation where IS (mol·L⁻¹) = 0.0127 × Electrical Conductivity (millimhos·cm⁻¹) (1973). $^{\#}$ CEC = Cation Exchange Capacity by sum of cations. †† SAR = Sodium Adsorption Ratio. ‡ OC = Organic Carbon (Total Carbon-Dissolved Organic Carbon).

may have contributed to their higher CEC over that of the mineral colloids (**Table 4**). There were higher exchangeable Ca^{2+} and Mg^{2+} concentrations in the NCs than the corresponding MC fractions (**Table 4**). Divalent cations can promote flocculation and may explain why NCs were found attached to MC surfaces likely due to bridging effects (**Table 1** and **Table 4**) [16].

NCs also contained higher OC content than their corresponding MCs which may have enhanced their stability over corresponding MCs (**Figure 1**, **Table 4**) [2] [8] [26]. While some studies indicated enhanced dispersibility and stability with increased OC content due to both charge and stearic stabilization [4] [16] [26] [27] [36], other findings suggested weak correlations [1]. It is likely, that the presence of differing OC functional groups caused a different behavior. Referring back to the d_h findings (**Figure 2**), the greatest aggregation potentials with contaminant additions occurred in the biosolid colloids that had a combination of greater OC content and surface area (**Table 1** and **Table 4**). Correlations between amount of C measured per unit surface area (SA) showed that the greatest aggregation potentials occurred in the colloids with the highest C:SA ratios (kaolinitic and biosolid NCs, respectively) (**Figure 2**, **Table 1** and **Table 4**). The difference in OC functional group as well as C:SA ratio may also explain differences in stability findings (**Table 1** and **Table 4**; **Figure 1**). It is also possible that OC bridges together micro-nano-aggregates that are more stable in suspension than individual particles. Despite higher OC and potential formations of micro-nano-aggregates, NCs were generally still more stable in the absence and presence of contaminants than were MCs (**Figure 1**).

3.1.6. Zeta Potential Effects

NC zeta potentials were more negative than that of their corresponding MCs, with both sizes showing increasingly negative zeta potentials with increased pH (Figure 3). Mineral NCs exhibited more negative zeta potentials than their corresponding MC counterparts and the bio-colloids, especially above pH 6 (Figure 3). Based on the assumption that zeta potentials approximate the charge residing in the diffuse layer of the electrical double layer, the negative zeta potential values suggest that all colloids bear a net negative surface charge. The increased negative charge with increasing pH is ascribed to the deprotonation of phyllosilicate edge sites, and indicates repulsion energies of the particles, suggesting increased stability potential of NCs over corresponding MCs (Figure 3), especially in negatively charged subsurface environments [37].

In the mineral colloids, there was never a point where the zeta potential reached zero (**Figure 3**), suggesting isoelectric points (the pH of zero mobility) of <4 for all colloids [38]. This was especially surprising in fractions where mineral compositions were dominated by kaolinite, which has an isoelectric point of 4.25 [38]. Apparently, the presence of other minerals and organic carbon surface coatings have considerably altered the isoelectric behavior of the mineral colloids [39].

The biosolid colloids had differing trends from the mineral colloids, where the bio-NCs became less negative with increasing pH as opposed to bio-MCs that showed an increased negative charge with increasing pH

(Figure 3). This may suggest a lower stability potential in the bio-NCs than in the bio-MCs, which is the opposite of what was seen in stability kinetics experiments (Figure 1). The less negative zeta potentials exhibited by the bio-colloids than the mineral colloids may be due to the greater ionic strength of the bio-colloids, especially those of the bio-NCs (Table 3), whose higher ionic strength likely overwhelmed potential negative surface charges generated by carboxyl groups above pH's 2.5 and 6 [38].

In the single point zeta potential measurements, additions of 2 and 10 mg L^{-1} mixed contaminants resulted in NC zeta potentials which were still more negative than those of their corresponding MCs, with both sizes showing trends based on composition (Figure 4). This display of greater stability in the NCs over the MCs complements the stability kinetics studies, suggesting that their stabilization potentials overcame their supposed high surface energy, even in the presence of up to 2 mg \cdot L⁻¹ contaminants [6]. It is noted here that in order to mimic natural conditions, the pH of the zeta potential measurements varied, so the change in zeta potential is likely a reference to colloidal stability potential (Figure 3 and Figure 4). Overall, the mineral colloids indicated greater stability over bio-colloids through higher negative zeta potentials in the presence of 0, 2 and even 10 mg $\cdot L^{-1}$ contaminant concentrations, although evidence of flocculation was more prevalent after 10 mg \cdot L⁻¹ contaminant additions (Figure 1 and Figure 4). While some aggregation was implied with increasing ionic strengths of contaminants by slight shifts towards less negative zeta potentials (Figure 4), corresponding to increased d_h values in the presence of contaminants (Figure 2), the NCs indicated significantly greater stability potentials over their corresponding MCs (Figure 1). Colloids with the most negative zeta potential usually have the largest resistance to flocculation [16], which was true for the smectitic, mixed, kaolinitic and biosolid NCs in the absence and presence of 2 and 10 mg L^{-1} contaminants (Figure 3). The MCs did not show the same stability trends based on composition, and surprisingly, the smectitic MCs were the least stable upon contaminant additions as compared to the other mineral (and even the biosolid) MCs (Figure 1 and Figure 4). The lack of stability indicated by lower negative zeta potentials in the smectitic MCs upon addition of contaminants (despite their higher initial zeta potential) may be a function of their smaller C:SA ratio (Table 1 and Table 4). The greater shifts in their zeta potentials (v27 mV without contaminants shifting to -18, then -11 mV with 2 and 10 mg·L⁻¹ contaminant additions, respectively) were most likely materialized by greater cation contaminant attraction to their high initial negative charge and to larger surface area availability (Table 1, Figure 4). Despite this large shift in the smectitic MC zeta potential, the stability kinetics studies indicated that smectitic MCs were more stable than any of the other mineral and biosolid MCs (Figure 1). The disparity of the zeta potential data to that of the stability kinetics data suggests that while the zeta potentials are consistent in predicting colloid stability of minerals versus biosolids, they maybe inconsistent in predicting stability trends based on mineralogy.

The implications may be that colloid stability may not be accurately predicted from zeta potential and d_h measurements in the presence of contaminants. Also, it emphasizes placing more weight on stability settling experiments than chemical data alone for transport predictions (although chemical characterization should still be considered important for explaining portions of the overall model). Finally, this showcases how irrelevant the average surface charge of colloids may become in contaminant transport behavior because of spatial heterogeneity and the importance of considering additional multiple relevant factors to achieve reliable assessments of environmental risks [17].



Figure 4. Zeta potential of macrocolloids and nanocolloids in mixtures with equal parts (0, 2, and 10 mg·L⁻¹) of each contaminant (As, Se, Cu and Pb) (error bars represent standard error of triplicate measurements on duplicate samples).

4. Conclusion

This study filled a previous void on the stability behavior of natural soil and biosolid water dispersible NCs and their differences from their corresponding larger MC fractions. The findings demonstrated that soil and biosolid NCs are more stable in the absence and presence of up to $2 \text{ mg} \cdot L^{-1}$ As, Se, Cu and Pb contaminants than corresponding MCs. Only the bio-NCs showed considerable stability at higher contaminant loads. The importance of mineralogy in stability was best shown by the MCs, where the smectitic and mixed colloids were more stable than the kaolinitic and biosolid compositions. In the NCs, the mineral colloids were more stable than the bio-colloids. Kaolinitic NCs and MCs showed surprisingly high stability potentials, even in the presence of goethite and gibbsite minerals, likely due to stearic stabilization effects of surface OC functional groups. Overall, NCs, regardless of mineralogy, where shown to have as much as 20% more colloids in suspension after 48 hours in the absence and presence of As, Se, Cu and Pb contaminants. While size played a very important role in the colloid stability behavior, NCs were also found to enhance aggregation of MCs via attachment to their surfaces. Further study is needed to better understand the causes of this nano-mediated aggregation behavior, but the presence of OC surface coatings and increased Al/Fe:Si ratios of NCs may have contributed to MC coagulation. Overall, the findings of this investigation demonstrated that soil NCs through a combination of physical, chemical, mineralogical and morphological properties that contribute to enhanced stability in natural environments, possess a higher potential to transport contaminants to greater distances than their corresponding larger size MC fractions. This potential was even greater with the bio-nano-colloids at higher contaminant loads. However, coagulation phenomena through nano-macro-colloid interactions may complicate their behavior in natural environments and result in misleading predictions. Therefore, multiple physicochemical and mineralogical parameters need to be considered in contaminant transport models in order to accurately assess environmental pollution risks and develop efficient remediation strategies.

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