

Crystal Structure, Synthesis and Biological Activity of Ether and Ester *Trans***-Ferulic Acid Derivatives**

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Abstract

The structures of methoxymethyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate, **2**; (E)-3-(3-methoxy-4-(methoxymethoxy)phenyl)acrylic acid, **3**; methyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate, **4**; benzyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate, **6**; and (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylic acid, **7**; were established by spectroscopic and X-ray diffraction studies. Structure **2** is a new compound. Compounds with free phenolic hydroxyls *v.gr.* methyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate **1**, **2** and benzyl(E)-3-(4-hydroxy-3-methoxyphenyl)acrylate **5**, showed scavenging freeradical and antioxidant activity while moderate scavenging free-radical was observed in compound **3**. Moderate inhibition of lipid peroxidation was observed for **7**. Compound **5** exerted significant inhibition of cell growth in PC-3, K562 tumor cell lines and **4** exhibited the largest cytotoxic effect upon the K562 cell line.

Keywords

Trans-Ferulic Acid, Crystal Structures, Antioxidant Activity, Antitumor, Scavenging Free-Radicals

1. Introduction

Ferulic acid (FA) is a phenolic derivative of cinnamic acid, which exhibits a wide spectrum of biomedical activities [1]. Many staple foods such as grain bran,

whole-grain foods, citrus fruits, banana, coffee, orange juice, eggplant, bamboo shoots, beetroot, cabbage, spinach and broccoli are among the richest sources of FA [2]. This compound is present in seeds and leaves, both in its free form and in its covalent-conjugated form, in plant cell-wall polysaccharides, glycoproteins, polyamides, lignin and hydroxy fatty acids. FA confers rigidity to the cell-wall and it is a precursor to other important organic compounds such as diferulic acid and curcumin [3]. Several reports have described the biological effects of FA, for example, as an antioxidant [4], anti-inflammatory and anticarcinogenic [5].

The well-known antioxidant activity of FA has been related to its phenolic nucleus and extensive chain conjugation that forms a resonance-stabilized phenoxy radical ion [4]. Since the hydroxyl and phenoxy groups of FA donate electrons to quench free radicals [6], FA can be used as a renewable, aromatic scaffold for conversion into novel, and useful products in the food, pharmaceutical and cosmetic industries. The ability of FA to inhibit peroxidation of fatty acids finds uses such as preservative for oranges or to inhibit the autoxidation of linseed oil. It is also employed in the industry of cosmetic skin lotions [4]. FA is used during oxidative-reductive depolymerization (ORD) of polysaccharides to protect both guar gum and cassava starch from thermal degradation [7].

The synthesis of new derivatives of FA in general has been aimed to improve its chemical, physicochemical and biological properties, in particular in its esterified form at the acidic moiety, which increases both radical scavenging activity and partition coefficient [8]. However, it is known that the blockade of the phenolic groups of FA can limit its antioxidant potential.

In the present report, the synthesis, spectroscopic characterization, X-ray analysis and biological activity of ether and ester derivatives of FA is described and it is depicted in **Scheme 1**. We also highlight the importance of the free hydroxyl and phenolic group in relation to the antioxidant and cytotoxic activity of FA and how the protection of these functional groups reduces their biological activity.

Compounds **1**, **3**, **4**, **5**, **6** and **7** have been previously reported while **2** is a new compound [9] [10] [11] [12] [13].

2. Experimental

2.1. Physical Measurements

All chemicals were reagent grade and were used as purchased. Solvents were purified by standard methods [14].

Melting points were determined on an Electrothermal IA9100 digital melting point apparatus and are uncorrected. IR absorption spectra were recorded in the 4000 - 400 cm⁻¹ range as KBr pellets on a Perkin Elmer 283-B spectrophotometer. ¹H and ¹³C NMR spectra were recorded in CDCl₃ on a Bruker 500 MHz spectrometer using TMS as an internal reference. NMR spectra were processed with MestReNOVA 12.0.0 software.



Scheme 1. Synthetic pathway for FA derivatives.

The mass spectrometry spectra were recorded in equipment MStation JMS-700 or Jeol, SX 102A using electron ionization impact or fast atom bombardment (FAB) mode.

X-ray data were collected on a Bruker APEX2 CCD area detector with a graphite monochromator and Mo *Ka* radiation ($\lambda = 0.71073$ Å) at room temperature with ω scan mode. All collected data were reduced using SAINTV8.27B [15]. These data are shown in **Table 1**.

2.2. Synthesis of Compounds

Methyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate (1). 5 meq of FA were dissolved in 20 mL of anhydrous methanol and 0.2 ml of H₂SO₄ were added and the mixture was refluxed for 6 hours. The reaction was quenched by the addition of NaHCO₃ until it reached pH 6 and the mixture was extracted with ethyl acetate (3 × 20 mL). The combined extracts were dried over anhydrous Na₂SO₄, then filtered and the solvent removed *in vacuo*, 82% yield, yellowish liquid. ¹H NMR (500 MHz, chloroform-d); δ 7.62 (d, *J* = 15.9 Hz, 1H), 7.07 (ddd, *J* = 8.2, 2.0, 0.5 Hz, 1H), 7.02 (d, *J* = 2.0 Hz, 1H), 6.92 (d, *J* = 8.2 Hz, 1H), 6.29 (d, *J* = 15.9 Hz, 1H), 5.96 (s, 1H), 3.92 (s, 3H), 3.80 (s, 3H) spectrum is shown in Figure 1. ¹³C NMR (125 MHz, chloroform-d) δ 167.70, 147.98, 146.76, 144.93, 126.92, 122.99, 115.12, 114.72, 109.38, 55.90, 51.56, IR; 3379.82, 2942.22, 1705.06, 1510.97, 1429.46, 1236.41 cm⁻¹, MS [208⁺].

Methoxymethyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate (2). To a solution of 1.0 meq of FA (0.1940 g) in 30 ml of CH_2Cl_2 , 1.1 meq of triethylamine

and 1.1 meq of chloromethyl methyl ether (Cl-MOM) were added. The reaction mixture was stirred at room temperature until the disappearance of the starting material was observed by TLC (Hexane:EtOAc: 70:30). The reaction was quenched by evaporation of the solvents of reaction and the mixture was extracted with EtOAc (3×20 mL), yield 68%, mp 80°C. ¹H NMR (500 MHz, chloroform-d); δ 7.68 (d, J = 15.9 Hz, 1H), 7.08 (m, 1H), 7.04 (d, J = 2.0 Hz, 1H), 6.92 (d, J = 8.2 Hz, 1H), 6.31 (d, J = 15.9 Hz, 1H), 6.00 (s, 1H), 5.37 (s, 2H), 3.93 (s, 3H), 3.52 (s, 3H) spectrum is shown in **Figure 2**. ¹³C NMR (125 MHz, chloroform-d) δ 166.59, 148.18, 146.80, 145.78, 126.76, 123.19, 114.86, 114.76, 109.44, 90.35, 57.55, 55.90. IR; 3266.48, 2944.22, 1716.75, 1627.83, 1508.79 cm⁻¹. MS [238⁺].

(E)-3-(3-methoxy-4-(methoxymethoxy)phenyl)acrylic acid (**3**). To a solution of the 1.0 meq of FA (0.1940 g) in 10 ml of acetone 1.0 meq of K₂CO₃ anhydrous and 1.1 meq of chloromethyl methyl ether (Cl-MOM) were added. The reaction mixture was stirred at room temperature until the disappearance of the starting material was observed by TLC (Hexane:EtOAc: 70:30). The reaction was quenched by filtration of the potassium salts and the solvent was removed in vacuum, yield 92%, mp 134.8°C. ¹H NMR (500 MHz, chloroform-d); δ 7.74 (d, *J* = 15.99 Hz, 1H), 7.17 (d, *J* = 8.28 Hz, 1H), 7.10 (m, 2H), 6.34 (d, *J* = 15.99 Hz, 1H), 5.28 (s, 2H), 3.93 (s, 3H), 3.52 (s, 3H) spectrum is shown in Figure 3. ¹³C NMR (125 MHz, chloroform-d) δ 172.56, 149.89, 148.97, 146.86, 128.43, 122.74, 115.82, 115.54, 110.57, 95.19, 56.37, 55.97, IR; 3438.01, 1688.78, 1622.87, 1506.04 cm⁻¹, MS [239⁺].

Table 1. Data collection and handling for compounds 2, 3, 4, 6, and 7.

Compound	2	3	4	6	7	
Empirical formula	$C_{12}H_{14}O_5$	$C_{12}H_{14}O_5$	$C_{18}H_{18}O_4$	$C_{24}H_{22}O_4$	$C_{17}H_{16}O_4$	
Formula weight	238.23	238.23	298.32	374.41	284.30	
Crystal external appearance	Colorless prism	Colorless prism	Colorless prism	Colorless prism	Colorless prism	
Diffractometer	Bruker APEX2 CCD	Bruker APEX2 CCD	Bruker APEX2 CCD	Bruker APEX2 CCD	Bruker APEX2 CCD	
Scanmode	Omega scans	Omega scans	Omega scans	Omega scans	Omega scans	
Theta range for data collection	2.89 to 25.40 deg	2.73 to 25.37 deg	2.37 to 25.29 deg	2.30 to 25.41 deg	2.48 to 25.32 deg	
Wavelength	Mo K <i>a</i> (0.71073 Å)	Mo K <i>a</i> (0.71073 Å)	Mo K <i>a</i> (0.71073 Å)	Mo K <i>a</i> (0.71073 Å)	Mo K <i>a</i> (0.71073 Å)	
Program system	SHELXTL	SHELXTL	SHELXTL	SHELXTL	SHELXTL	
Structure determination	Structure determination Direct methods (SHELXS-2014/7)		Direct methods (SHELXS-2014/7)	Direct methods (SHELXS-2014/7)	Direct methods (SHELXS-2014/7)	
Refinement method	Full-matrix Refinement method least-squares on F ² (SHELXL-2014/7)		Full-matrix least-squares on F ² (SHELXL-2014/7)	Full-matrix least-squares on F ² (SHELXL-2014/7)	Full-matrix least-squares on F ² (SHELXL-2014/7)	











Figure 3. ¹H NMR spectrum of compound **3**.

Methyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate (**4**). 1 meq of compound **1** was dissolved in 30 mL of anhydrous acetone and 1 meq of K₂CO₃ and 1.1 meq of benzyl bromide were stirred at room temperature. The reaction was followed by TLC (Hexane:EtOAc: 70:30). The solvent was removed *in vacuo*, yield 78%, mp 97.1°C. ¹H NMR (500 MHz, chloroform-d); δ 7.61 (d, *J* = 15.9 Hz, 1H), 7.42 (ddt, *J* = 7.6, 1.4, 0.7 Hz, 2H), 7.36 (tt, *J* = 6.6, 0.8 Hz, 2H), 7.30 (m, 1H), 7.06 (d, *J* = 2.0 Hz, 1H), 7.02 (ddd, *J* = 8.2, 2.1, 0.5 Hz, 1H), 6.86 (d, *J* = 8.3 Hz, 1H), 6.30 (d, *J* = 15.9 Hz, 1H), 5.18 (s, 2H), 3.91 (s, 3H), 3.79 (s, 3H) spectrum is shown in **Figure 4**. ¹³C NMR (125 MHz, chloroform-d) δ 167.58, 150.24, 149.74, 145.12, 144.69, 143.62, 136.55, 128.58, 127.96, 127.71, 127.16, 122.30, 115.59, 113.44, 110.27, 70.82, 55.95, 51.54, IR; 2942.82, 1700.03, 1627.76, 1592.64, 1508.58, 1255.42 cm⁻¹, MS [300⁺].

Benzyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate (**5**). To a solution of 1.0 meq of FA (0.1940 g) in 30 ml of CH₂Cl₂ 1.1 meq of triethylamine 1.1 meq of benzyl bromide were added. The reaction mixture was stirred at room temperature and followed by TLC (Hexane:EtOAc: 70:30). After removal of solvent reaction was partitioned with EtOAc (3×20 mL) and H₂O, the combined extracts were dried over anhydrous Na₂SO₄, filtered, and the solvent was removed in vacuum, yield 73%, mp 57.3°C. ¹H NMR (500 MHz, chloroform-d) δ 7.65 (d, *J* = 15.9 Hz, 1H), 7.38 (m, 5H), 7.06(ddd, *J* = 8.1, 2.0, 0.5 Hz, 1H), 7.02 (d, *J* = 2 Hz, 1H) 6.91 (d, *J* = 8.2 Hz, 1H), 6.34 (d, *J* = 15.9 Hz, 1H), 5.90 (s, 1H), 5.24 (s, 2H), 3.90 (s, 3H) spectrum is shown in Figure 5. ¹³C NMR (125 MHz, chloroform-d) δ 167.05, 148.03, 146.75, 145.23, 136.17, 128.56, 128.24, 128.19, 126.93, 123.13,



115.22, 114.71, 109.33, 66.22, 55.91, IR; 3404.3, 2941.23, 1703.49, 1632.44, 1593.13, 1514.53 cm⁻¹, MS [286⁺].

Figure 4. ¹H NMR spectrum of compound 4.





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Benzyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate (**6**). 1 meq of compound **5** was solved in 30 mL of acetone anhydrous and was added 1 meq of K₂CO₃ and finally was added 1.1 meq of benzyl bromide. The reaction mixture was stirred at room temperature until the disappearance of the starting material was observed by TLC (Hexane:EtOAc: 70:30). The reaction was quenched by filtration of potassium salts and the solvent was removed in vacuum, yield 84%, mp 86.3°C. ¹H NMR (500 MHz, chloroform-d); δ 7.65 (d, *J* = 15.9 Hz, 1H), 7.45 - 7.27 (m, 10H), 7.06 (d, *J* = 2.0 Hz, 1H), 7.02 (ddd, *J* = 8.3, 2.1, 0.5 Hz, 1H), 6.86 (d, *J* = 8.3 Hz, 1H), 6.35 (d, *J* = 15.9 Hz, 1H), 5.24 (s, 2H), 5.18 (s, 2H), 3.90 (s, 3H) spectrum is shown in **Figure 6**. ¹³C NMR (125 MHz, chloroform-d) δ 166.97, 150.32, 149.76, 145.62, 145.03, 136.56, 136.16, 128.61, 128.57, 128.24, 128.19, 128.00, 127.70, 127.18, 122.44, 115.67, 113.43, 110.22, 70.85, 66.23, 55.96, IR; 2937.39, 1697.95, 1628.78, 1505.19, 1255.53, 1157.91, 1135.79 cm⁻¹, MS [molecular ion not observed].

(E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylic acid (7). To 1 meq of compound **4** was solved in 20 mL of ethanol and 10 mL of one solution 1M of NaOH were added the mixture refluxed for 40 minutes. The reaction was terminated by addition of HCl 1N until pH 6 and the mixture partitioned with ethyl acetate (3 × 20 mL) and H₂O. The combined extracts were dried over anhydrous Na₂SO₄, filtered, and the solvent was removed in vacuum; yield 74%. mp 192.9°C. ¹H NMR (500 MHz, chloroform-d); δ 7.58 (d, *J* = 15.9 Hz, 1H), 7.43 (m, 2H), 7.37 (m, 2H), 7.31 (m, 1H), 7.08 (d, *J* = 2.0 Hz, 1H), 7.03 (m, 1H), 6.89 (d, *J* = 8.3 Hz, 1H), 6.29 (d, *J* = 15.9 Hz, 1H), 5.17 (s, 2H), 3.91 (s, 3H) spectrum is shown in **Figure 7**. ¹³C NMR (125 MHz, chloroform-d) δ 168.86, 150.03, 149.70, 144.38, 136.58, 128.56, 127.96, 127.92, 127.24, 122.23, 116.74, 113.50, 110.30, 70.78, 55.95, IR; 2968.68, 1669.68, 1592.66, 1423.76, 1265.73 cm⁻¹, MS [285⁺].

2.3. Scavenging Activity of DPPH Free Radicals

The free radical scavenging activity was measured using a modified method from Mellors and Tappel [16]. The test was carried out on 96-well microplates. A 50 μ L aliquot of the solution of the test compound was mixed with 150 μ L of an ethanol solution of DPPH (final concentration 100 μ M). This mixture was incubated at 37°C for 30 min, and the absorbance was then measured at 515 nm using a BioTek microplate reader SYNERGY HT. The inhibition percent for each compound was determined by comparison with a 100 μ M DPPH ethanol blank solution.

2.4. Inhibition of Lipid Peroxidation on Rat Brain

Inhibition of lipid peroxidation of FA derivatives was assayed following previous described method [17] [18].

2.5. Cytotoxicity Assay

All synthetized compounds were screened in vitro at doses of 50 µg/mL against



Figure 6. ¹H NMR spectrum of compound 6.



Figure 7. ¹H NMR spectrum of compound **7**.

monkey kidney cells and cancer cell lines: U251 (central nervous system glia cancer), PC-3 (prostate adenocarcinoma), K562 (human chronic myelogenous

leukemia), HCT-15 (colon adenocarcinoma), MCF-7 (human mammary adenocarcinoma) and SKLU-1 (human lung adenocarcinoma). Cell lines were supplied by U.S. National Cancer Institute (NCI). The human tumor cytotoxicity was determined using the protein-binding dye sulforhodamine B (SRB) in microculture assay to measure cell growth, as described in the protocols established by the NCI [19].

3. Results and Discussion

3.1. Refinement Details

All reflections were defined based on F^2 . The weighted R-factor wR and goodness of fit S are based on F^2 , conventional R-factors R are based on F, with F set to zero for negative F^2 . The threshold expression of $F^2 > \sigma(F^2)$ is used only for calculating R-factors (gt) etc., and is not relevant to the choice of reflections for refinement. R-factors based on F^2 are statistically about twice as large as those based on F, and R-factors based on all data will be even larger.

There is an intramolecular hydrogen bond and one intermolecular hydrogen bond between the hydroxyl group at C7 and O5 with a symmetry-related molecule forming sheets along the *a*-axis. In structure 3, the bond lengths and angles are similar to those found in 3-(4-Hydroxy-3-methoxyphenyl)-2-propenoic acid (ferulic acid). Molecular packing of molecules in the crystal shows an intermolecular O-H...O hydrogen-bonded carboxylic acid dimers that together with several C-H...O interactions form a sheet-like structure. In structure 4 is a new benzoyloxy and acrylate derivative of FA. The molecules in the crystal are linked by one C-H...O intermolecular interaction involving H atom of the phenyl ring C4-C9 and carbonyl oxygen of the acrylate group. This interaction links crystal and packed molecules at normal van der Waals distances. In structure 6, there are two independent molecules in the crystal. Structure 6 contains three substituents mainly at C4 a methoxyphenylacrylate moiety; at C6 a methoxy group and C7 a benzoyloxy moiety. The dihedral angle between phenyl rings C4-C9 and C28-C33 is only 4.76 (12)°. There are seven C-H...O interactions that link two independent molecules in the crystal and packed structures at normal van der Waals distances. Structure 7 contains three substituents mainly at C4 an acrylic acid group; at C6 a methoxy group and C7 a benzoyloxy moiety. There is one intermolecular hydrogen bond O-H...O with a symmetry related molecule (+2 - x, +2 - y, +2 - z) forming sheets along the *b*-axis and two C-H...O interactions linked the molecules in the crystal and packed at normal van der Waals distances.

3.2. Geometric Details

All estimation of standard deviation (esd's) (except the esd in the dihedral angle between two least square planes) are estimated using the full covariance matrix. The cell esds are taken into account individually in the estimation of esds in distances, angles and torsion angles; correlations between esds in cell parameters are only used when they are defined by crystal symmetry. An approximate (isotropic) treatment of cell esds was used for estimating esds involving least square planes.

3.3. Structural X-Ray Data

All structures were solved by direct methods [20], and then refined by full-matrix least-squares technique; the position and anisotropic parameters of all non-hydrogen atoms were obtained. **Table 1** shows the data collection and handling for compounds 2, 3, 4, 6, and 7. **Table 2** shows the crystal and refinement parameters for compounds 2, 3, 4, 6 and 7. **Figure 8** contains the molecular structures of 2, 3, 4, 6 and 7. This figure was drawn with 50% displacement ellipsoids using ORTEP-3 for Windows [21]. The geometry of the molecule was calculated using PARST [22] [23] software. In all structures, the bond distances and bond angles are in good agreement with the corresponding values and are within normal ranges.



Figure 8. The molecular structures of 2, 3, 4, 6 and 7.

Compound	2	3	4	6	7		
Crystal system	Monoclinic	Triclinic	Monoclinic	Monoclinic	Monoclinic		
Unitcell dimensions	a = 12.2068 (11) Å a = 90.00 deg b = 9.0967 (8) Å $\beta = 113.431 (5) \text{ deg}$ c = 11.9634 (10) Å $\gamma = 90.00 \text{ deg}$	a = 6.8359 (6) Å a = 93.459 (3) deg b = 7.8261 (6) Å $\beta = 101.485 (3) \text{ deg}$ c = 11.9252 (10) Å $\gamma = 106.323 \text{ deg}$	a = 9.5021 (3) Å a = 90.00 deg b = 20.2456 (6) Å $\beta = 93.783 (2) \text{ deg}$ c = 8.1203 (2) Å $\gamma = 90.00 \text{ deg}$	a = 5.6645 (3) Å a = 90.00 deg b = 34.863 (2) Å $\beta = 96.742 (2) \text{ deg}$ c = 20.7049 (14) Å $\gamma = 90.00) \text{ deg}$	a = 5.3864 (2) Å a = 90.00 deg b = 8.5106 (3) Å $\beta = 94.1040 (10) \text{ deg}$ c = 31.6002 (10) Å $\gamma = 90.00 \text{ deg}$		
Volume	1218.89 (19)Å ³	595.40 (9) Å ³	1558.75 (8) Å ³	595.40 (9) Å ³	1444.89 (9) Å ³		
Z/Calculated density	4/1.298 mg/m ³	2/1.329 mg/m ³	4/1.271 mg/m ³	8/1.225 mg/m ³	4/1.307 mg/m ³		
Absorption Coefficient	0.101 mm^{-1}	0.104 mm^{-1}	0.089 mm^{-1}	0.083 mm^{-1}	0.093 mm^{-1}		
Space group/F (000)	<i>P</i> 2 ₁ / <i>c</i> / 504	P-1/252	<i>P</i> 2 ₁ / <i>c</i> /632	Cc/1584	<i>P</i> 2 ₁ / <i>c</i> /600		
Limiting indices	$-14 \le h \le 14$ $-10 \le k \le 10$ $-14 \le l \le 14$	$-8 \le h \le 8$ $-9 \le k \le 9$ $-14 \le l \le 14$	$-9 \le h \le 11$ $-24 \le k \le 20$ $-9 \le l \le 9$	$-6 \le h \le 6$ $-42 \le k \le 41$ $-24 \le l \le 24$	$-6 \le h \le 6$ $-10 \le k \le 10$ $-37 \le l \le 37$		
Reflection scollected unique/R (int)	9089/2224/0.0367	9223/2181/0.0622	8788/2835/0.0528	32,474/6927/0.1218	16172 /2654/0.0702		
Completeness to $\theta = 25.10$	99.9%	99.9%	99.9%	99.8%	99.9%		
Max/min Transmission	0.961/0.990	0.958/0.984	0.965/0.994	0.976/0.995	0.970/0.983		
Data/restraints/parameters	2224/0/160	2181/1/159	2835/0/201	6927/2/507	2654/1/194		
Final R indices	R1 = 0.0403	R1 = 0.0441	R1 = 0.0414	R1 = 0.0484	R1 = 0.0458		
$I > 2\sigma(I)$	wR2 = 0.1182	wR2 = 0.1228	wR2 = 0.0880	wR2 = 0.1137	wR2 = 0.1111		
R indices (all data)	R1 = 0.0553 wR2 = 0.1260	R1 = 0.0517 wR2 = 0.1297	R1 = 0.0717 wR2 = 0.1045	R1 = 0.0773 wR2 = 0.1286	R1 = 0.0790 wR2 = 0.128		
Goodness-of-fit on F ²	1.059	1.070	0.998	1.030	1.019		
Largest diff. peak/hole	0.211/-0.185 e.Å ⁻³	0.178/-0.162 e.Å ⁻³	0.208/-0.241 e.Å ⁻³	0.150/-0.138 e.Å ⁻³	0.189/-0.178 e.Å ⁻³		
CCDC deposition number	1,825,526	1,825,528	1,825,527	1,825,529	1,825,523		

Compound **2** crystallizes in the monoclinic space group P_1/c with unit cell parameters a = 12.2068 (11), b = 9.0967 (8), c = 11.963421 (10) Å, $\beta = 113.431$ (5), Z = 4. Compound **3** crystallizes in the triclinic space group *P*-1 with unit cell parameters a = 6.8359 (6), b = 7.8261 (6), c = 11.9252 (10) Å, a = 93.459 (3), $\beta = 101.485$ (3), $\gamma = 106.323$ (2)°, Z = 2. Compound **4** crystallizes in the monoclinic space group P_1/c with unit cell parameters a = 9.5021 (3), b = 20.2456 (6), c = 8.1203 (2) Å, $\beta = 93.783$ (2), Z = 4. Compound **6** crystallizes in the monoclinic space group *C c* with unit cell parameters a = 5.6645 (3), b = 34.863 (2), c = 20.7049 (14) Å, $\beta = 96.742$ (2), Z = 8. Compound **7** crystallizes in the monoclinic space group P_1/c with unit cell parameters a = 5.3864 (2), b = 8.5106 (3), c = 31.6002 (10) Å, $\beta = 94.1040$ (10), Z = 4.

The molecular structure of **2** is not planar, with a fully extended methoxymethyl group of the acrylate residue as shown in **Figure 8**. The dihedral angle between the side chain at C4 and the phenyl ring is $32.7(2)^\circ$. The hydroxyl group at C7 has an intramolecular hydrogen with O3 and one intermolecular hydrogen bond between the hydroxyl group at C7 and O5 with a symmetry related molecule (-x - 1, +y, +z) forming sheets along the a-axis. There are no stacking interactions between phenyl rings. The crystal packing diagram is shown in **Figure 9**.



Figure 9. The molecular packing of structures **2**, **3**, **4**, **6** and **7**.

Figure 8 shows the molecular structure of **3**. Bond lengths and angles are similar to those found in 3-(4-Hydroxy-3-methoxyphenyl)-2-propenoic acid (FA) [24]. The dihedral angles between the side chain at C4, the acrylate and the phenyl ring are 48.31 (5) and 9.32 (5)°, respectively. The molecular packing of **3** in the crystal shows an intermolecular O-H...O hydrogen-bonded carboxylic

acid dimers that together with several C-H...O interactions form a sheet-like structure (Figure 9).

The molecular structure of **4** corresponds to a new benzoyloxy and acrylate derivative of ferulic acid. There are no significant molecular distortions in terms of bond lengths and angles. The dihedral angles between benzene ring C4 to C9 and the side chains at C4 and C7 are 21.25 (5) and 8.84 (5)°, respectively. However, the benzene ring C4 to C9 makes and angle of 86.00 (5)° with the benzene ring C11 to C16. The molecule **4** in the crystal are linked by one C-H...O intermolecular interaction involving an H atom of the phenyl ring C4-C9 and carbonyl oxygen of the acrylate group. This interaction linked the molecules in the crystal and packed at normal van der Waals distances (**Figure 9**).

Figure 8 shows the molecular structure of compound **6**. There are two independent molecules 4 in the crystal. Molecule **6** contains three substituents mainly at C4 a methoxyphenyl acrylate moiety; at C6 a methoxy group and C7 a benzoyloxy moiety. The dihedral angles between phenyl rings C11-C16 and C19-C24 with the central phenyl ring C4-C9 are 70.50 (12) and 69.61 (14), respectively. The dihedral angles between phenyl rings C35-C40 and C43-C48 with the central phenyl ring C28-C33 are 77.00 (14) and 73.92 (16), respectively. The dihedral angle between the phenyl rings C4-C9 and C28-C33 is only 4.76 (12)°. There are seven C-H...O interactions linked the two independent molecules in the crystal packed at normal van der Waals distances (**Figure 9**).

The molecular structure of **7** is shown in **Figure 8**. The molecule contains three substituents mainly at C4 an acrylic acid group; at C6, a methoxy group and at C7 a benzoyloxy moiety. The dihedral angles between phenyl ring C12-C17; the linker C7-O4-C11-C12 and acrylic acid group at C4 with the central phenyl ring C4-C9 are 74.62 (6), 12.17 (9) and 7.37 (5)°, respectively. There is one intermolecular hydrogen bond O-H...O with a symmetry related molecule (+2 - x, +2 - y, +2 - z) forming sheets along the *b*-axis and two C-H...O interactions linked the molecules in the crystal and packed at normal van der Waals distances (**Figure 9**).

For geometrical details and notations of all of these hydrogen bonds and intermolecular interactions are given in Table 3.

3.4. Biological Assay

Interestingly, compounds 1, 2 and 5, with free hydroxyl group, showed to have DPPH free radicals scavenging activity *in vitro* in moderate to high level (see **Table 4**). These results are consistent with a previous report, where it is postulated that antioxidant property of FA and its derivatives is associated with the number of free hydroxyl groups on the benzene ring and *ortho* substitution with the electron donor methoxy group, due to stabilization of the phenoxy radical [25]. FA derivatives, especially those with etherified phenolic groups, did not show effect upon the free radical essay.

Compound	D-HA	D-H	HA	DA	D-HA	Symmetrycode
	O4-H4O3	0.82 (2)	2.25 (3)	2.661 (2)	111 (2)	<i>x</i> , <i>y</i> , <i>z</i>
2	O4-H4O5	0.82 (2)	1.97 (2)	2.731 (2)	156 (2)	<i>x</i> , <i>y</i> , <i>z</i>
	C8-H8O2	0.930 (2)	2.920 (1)	3.431 (2)	116.1 (1)	2 - x, -y, 2 - z
	C8-H8O5	0.930 (1)	2.553 (1)	3.081 (2)	116.4 (1)	<i>x</i> , <i>y</i> , <i>z</i>
	O1-H1O2	0.86 (2)	1.77 (2)	2.635 (2)	179 (2)	+3 - x, $+1 - y$, $+1 - z$
3	C8-H8O2	0.930(1)	2.612 (1)	3.294 (2)	130.7 (1)	-1 + x, -1 + y, z
	С9-Н9О2	0.930 (2)	2.889 (1)	3.434 (2)	118.7 (1)	-1 + x, -1 + y, z
	C12-H12O3	0.960 (3)	2.720 (1)	3.348 (3)	123.61 (1)	+1 - x, $+1 - y$, $+2 - z$
4	С9-Н9О2	0.950 (2)	2.394 (1)	3.324 (2)	165.8 (1)	-x, +1 - y, +1 - z
	С3-Н3О6	0.930 (4)	2.516 (3)	3.411 (5)	161.7 (3)	<i>x</i> , <i>y</i> , <i>z</i>
	C20-H20O2	0.930 (5)	2.686 (3)	3.294 (5)	123.7 (3)	<i>x</i> , <i>y</i> , <i>z</i>
	C27-H27O2	0.931 (4)	2.526 (3)	3.425 (5)	162.5 (3)	<i>x</i> , <i>y</i> , <i>z</i>
6	C48-H48O6	0.930 (4)	2.612 (3)	3.216 (5)	123.1 (3)	<i>x</i> , <i>y</i> , <i>z</i>
	C3-H3O1	0.930 (4)	2.958 (3)	3.410 (5)	111.5 (3)	+1 + x, y, z
	C17-H17O3	0.959 (6)	2.685 (3)	3.370 (7)	128.8 (4)	+1 - x, $+1 - y$, $+2 - z$
	C18-H18AO2	0.970 (5)	2.560 (3)	3.489 (5)	160.6 (3)	-1 + x, y, z
	O1-H1O2	1.01 (3)	1.62 (3)	2.6213 (2)	175 (2)	+2 - x, $+2 - y$, $+2 - z$
7	C8-H8O2	0.930 (2)	2.509 (2)	3.433 (3)	172.3 (1)	-1 + x, $-1 + y$, z
	С9-Н9О1	0.930 (2)	2.701 (2)	3.437 (3)	136.7 (1)	+1 + x, $+1 - y$, $+2 - z$

Table 3. Geometry of the hydrogen bonds and hydrogen bonding interactions (Å, °) for compounds 2,3,4, 6 and 7.

	Scavenging activity DPPH		Inhibition of lipid peroxidation TBARS			Cytotoxic activity Percentage of inhibition of cell lines at 50 μM						
Compound	10 µM	100 µM	10 µM	100 µM	U251	PC-3	K562	HCT-15	MCF-7	SKLU-1	COS-7	
FA	19.90 ± 0.26	$76.81 \pm 0.26^{*}$	-4.07 ± 3.82	4.06 ± 1.36	NC	NC	NC	NC	NC	NC	ND	
1	11.76 ± 0.31	$60.39 \pm 0.13^{*}$	10.51 ± 4.03	34.52 ± 3.05*	NC	4.8	26.6	NC	NC	18.6	1.2	
2	9.95 ± 0.38	56.76 ± 0.29*	12.24 ± 2.37	30.19 ± 2.13*	NC	NC	7.7	NC	NC	NC	NC	
3	5.39 ± 0.32	$30.10\pm0.39^{\ast}$	2.02 ± 2.20	1.45 ± 3.14	NC	8	20.3	NC	6.5	12	7.0	
4	0.49 ± 0.38	0.39 ± 0.05	4.31 ± 1.45	7.48 ± 2.80	NC	24.8	72.5	16.3	NC	20.9	17.7	
5	12.84 ± 0.24	$59.95 \pm 0.25^{*}$	$40.61 \pm 1.60^{*}$	96.68 ± 0.65*	25.5	59.4	48.0	26.9	25.4	20.2	52.6	
6	1.03 ± 0.31	0.83 ± 0.13	6.08 ± 0.98	9.16 ± 2.71	NC	15.3	32.4	15.5	14.6	27.4	31.3	
7	1.27 ± 0.19	3.19 ± 0.47	7.46 ± 12.71	13.8 ± 25.14	5.8	16.6	17	5.7	7.8	18.1	22.2	

Data represent media and standard error. * $P \le 0.05$ compared to FeSO₄ control. * $P \le 0.05$ compared to DPPH control. FA: ferulic acid. Cell lines: U251: central nervous system glia cancer, PC-3: prostate adenocarcinoma, K562: human chronic myelogenous leukemia, HCT-15: colon adenocarcinoma, MCF-7: human mammary adenocarcinoma, SKLU: human lung adenocarcinoma, COS7: non-cancerous monkey kidney cell line. NC: non-cytotoxic, ND: non determinated.

Moreover, compounds 1, 2 and 5 also inhibited lipoperoxidation induced by ferric sulphate in the rat brain, mainly due to preservation of phenolic group and a similar mechanism of action of antioxidant effect described above. Remarkably, compound 7 showed moderate inhibition of lipid peroxidation, despite the fact of it has the phenol group blocked probably due to higher lipophilicity, which aids to penetrate the cell membrane where lipid peroxidation takes place

[26].

The FA derivatives showed low cytotoxic activity in the preliminary screening in different cancer cell lines (all compounds were tested at 50 μ M). As seen in **Table 4**, only compound **5** inhibited cancer cell lines PC-3 and K562 by 59% and 48% respectively, while to healthy cell line COS-7 it did so by 53%. Compounds **4** and **6** also inhibited the growth of K562 cell line by 72% and 32% respectively. Since there were not relevant cytotoxic agents, it is remarkable that addition of benzyl groups to the FA improved cytotoxic activity *in vitro*, possibly by the increment of lipophilicity of compounds and a more efficient molecular action within the cell, as it has been reported previously [27] [28]. Our findings do not coincide with Jayaprakasam *et al.* (2006), who reported anti-inflammatory, anticancer and antioxidant activity of FA esters at low concentrations [29]. New refined analyses are required in order to elucidate molecular effects of FA and its derivatives.

4. Conclusion

The synthesis with moderate to good yields and unambiguous assignment of chemical structures of methoxymethyl (E)-3-(4-hydroxy-3-methoxyphenyl)acrylate ($C_{12}H_{14}O_5$) **2**; (E)-3-(3-methoxy-4-(methoxy methoxy)phenyl)acrylac acid ($C_{12}H_{14}O_5$) **3**; methyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate($C_{18}H_{18}O_4$) **4**; benzyl (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate($C_{24}H_{22}O_5$) **6** and (E)-3-(4-(benzyloxy)-3-methoxyphenyl)acrylate($C_{17}H_{16}O_4$) **7** was achieved using ¹H/¹³C NMR and X-ray techniques of compounds **2**, **3 4**, **6** and **7** were successfully grown single crystals. Compounds containing free phenolic hydroxyls e.g. **1**, **2** and **5** exhibited scavenging free radical and antioxidant activity. Compound **5** induced significant inhibition of cell growth in PC-3, K562 and COS-7 cell lines by 59.4%, 52.6% and 48% respectively while **4** exhibited the largest cytotoxic effect upon K562 cell line *i.e.* 72.5%.

Supplementary Materials

CCDC-1825526, CCDC-1825528, CCDC-1825527, CCDC-1825529 and CCDC-1852523 contain the supplementary crystallographic data for compound **2**, **3**, **4**, **6** and **7**, respectively. These data can be obtained free of charge via https://www.ccdc.cam.ac.uk/structures/ by e-mailing data_request@ccdc.cam.ac.uk, or by contacting The Cambridge Crystallographic Data Centre, 12 Union Road, Cambridge CB2 1EZ, UK, fax; +44(0)1223-336033.

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Conflicts of Interest

The authors declare no conflicts of interest regarding the publication of this paper.

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