

Self-Referencing Method for Relative Color Intensity Analysis Using Mobile-Phone

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How to cite this paper: Souza, W.S., de Oliveira, M.A.S., de Oliveira, G.M.F., de Santana, D.P. and de Araujo, R.E. (2018) Self-Referencing Method for Relative Color Intensity Analysis Using Mobile-Phone. *Optics and Photonics Journal*, 8, 264-275. <https://doi.org/10.4236/opj.2018.87022>

Received: May 25, 2018

Accepted: July 24, 2018

Published: July 27, 2018

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Abstract

Mobile-phones have been widely explored on colorimetric evaluations. However, their use with different built-in image systems and acquisition configuration, in an environment with non-controlled illumination, limits the establishment of an accurate color analysis. To overcome this restriction, the determination of the absolute color of an object can be overlooked and a relative color value determined. In this work, we establish a new approach for spectroscopic evaluation based on cell-phone imaging, with no previous calibration, determining relative color values. The standalone relative color intensity method is evaluated under the use of four distinct mobile-phones and different illumination conditions. The capability to distinguish different color shades exploring the proposed self-referenced relative color intensity technique is appraised. Moreover, the potential use of the method is demonstrated by evaluating the chemical-adsorption process of Cysteamine molecules on gold nanoparticle surfaces. The proposed self-referenced technique can improve and expand the use of mobile-phones in spectroscopic applications.

Keywords

Spectrometry, Colorimetry, Mobile-Phone

1. Introduction

Colorimetric test have been widely used on several application such as industrial control [1], immunoassays [2], food industry [3], water quality evaluation [4]. Color evaluations are mainly performed by exploring optical absorption or reflection spectroscopy techniques, which may require expensive devices, controlled illumination and the use of color calibrators. However, in a real world

scenario there are several applications demanding color's evaluation in an imperfect illumination environment, with no available calibrated reference color object. On that context, mobile-phones could be explored as an important tool for colorimetric analysis [5] [6]. Several mobile-phones applications (Apps) are available to perform a roughly color evaluation of objects, as Catch Color Free (developed by CeSnow), Color Grab (by Loomatix), LifeDropper (by Bitjutsu Software). However, the color analysis of an object using distinct cell-phones and Apps leads to disparate results [6]. The popularization of mobile-phones and the improvement of their components allowed the implementation of different colorimetric analysis techniques in several areas such as forensic science [7], biotechnology [8] [9], and life sciences [10]. Colorimeters base on mobile-phone can be classified as: 1) applications with hardware components and 2) standalone devices [11]. Colorimetric systems classified in this first category require external tailor-made apparatus for each device, as LED light sources, are powered by a battery to maintain constant illumination. These types of applications have as main advantage the control of ambient light, which may enable continuous monitoring of color changes of a particular sample. As an example, mobile-phone based colorimetric readers for enzyme-linked immunosorbent assays (ELISA) have recently been developed, requiring the use of mechanical platform and illumination set-up [12] [13]. Moreover, the feasibility of a colorimetric biosensor based on a smartphone spectrometer with an integrated grating was demonstrated [14]. The need of external apparatus may limit the use of the mobile system (first category). On the other hand, standalone devices have the advantage that they do not require external media such as housings, light sources or batteries. Moreover, they usually explore correction algorithm to minimize the variable light condition. Mobile colorimeters with a standalone application designed to process color information are an excellent low-cost alternative to expensive commercial color readers. Several applications of standalone colorimetric systems are described in the literature, such as: To quantify the concentrations of pH, glucose, and protein [15]; to evaluate the presence of albumin in urine [16]; to monitor chlorine concentration and pH in water [17] [18]; and to estimate the level of hemoglobin present in blood [19]. Usually, to establish a precise color analysis with standalone mobile-phone devices, calibration procedures are required. To overcome this limitation, the determination of the absolute color value of an object can be overlooked and a relative color value determined [8] [17] [18] [20]. However, in addition to the restrictions regarding the type of ambient lighting, different built-in image systems and acquisition configuration of cell-phones may limit the color analysis accuracy. In this work we establish a new approach for relative color intensity evaluation based on mobile-phone imaging. An algorithm to determine relative color values of an image is proposed. The standalone spectroscopic method has been evaluated under the use of four distinct cell-phones and different illumination conditions. The capability to distinguish different color shades exploring the self-referenced relative color intensity technique is also appraised. Moreover, the potential use of the

method is demonstrated evaluating chemical-adsorption process of Cysteamine molecules on gold nanoparticle surfaces.

2. Materials and Methods

2.1. Relative Color

Images, obtained by cell-phones, usually provide color information on RGB space. Even under same illumination, different image capture devices can lead to distinct color output information (RGB values) [6]. Therefore, an accurate color analysis requires a calibration procedure, that is performed exploring a reference object with know RGB values [21]. Moreover, even the evaluation of color differences of two regions in a single image can be device dependent. In many cases, the relation between the input and output signal strength of a given device is characterized by a non-linear equation. The output color information of an image capture devices maybe described as

$$C' = C^\gamma, \quad (1)$$

where C is the real object color, C' is the colorimage output and γ is known as the correction gamma factor [22] [23]. Therefore, if two or more cell phones have approximately the same value of γ , the color perception at the output of these devices will be similar. However, if γ values are different, the output values of the measured color will be distinct. In a first approximation, γ also carries the information of the illumination source. Moreover, as color is described on a RGB space, Equation (1) can be express as

$$R' = R^\gamma, G' = G^\gamma, B' = B^\gamma \quad (2)$$

where (R, G, B) and (R', G', B') are respectively the real object color and the cell-phone color output. On the color evaluation of two different regions in a single image, we define the relative color parameters (r, g, b) as:

$$r = \frac{\ln(R'_i)}{\ln(R'_r)} = \frac{\ln(R_i)}{\ln(R_r)} \quad (3)$$

$$g = \frac{\ln(G'_i)}{\ln(G'_r)} = \frac{\ln(G_i)}{\ln(G_r)} \quad (4)$$

$$b = \frac{\ln(B'_i)}{\ln(B'_r)} = \frac{\ln(B_i)}{\ln(B_r)} \quad (5)$$

where the index i and r denotes the two different regions of an image, the region of interest and the reference region, respectively. Notices that the relative colors are independent of γ , and therefore they are only dependent of the real colors of the object.

2.2. Spectroscopic Measurements

Two homemade color patterns, shown in **Figure 1**, were used in this study. The pattern in **Figure 1(a)** comprised four areas with four different colors (red, green, blue and white). The pattern in **Figure 1(b)** presents 15 distinct areas

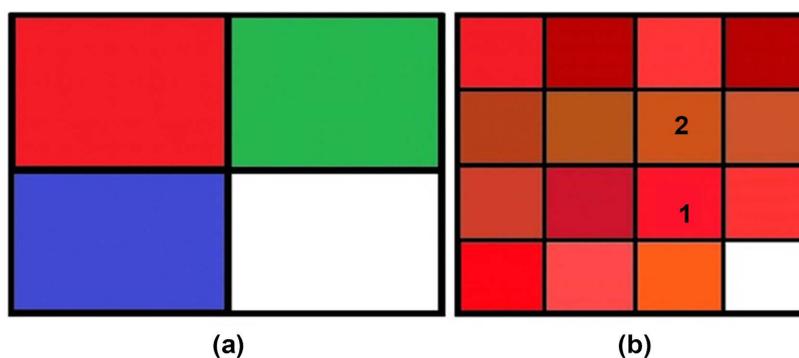


Figure 1. Patterns (a) with different colors and (b) with different shades of red.

with different shades of red color. All color patterns areas were determined using a *CR-400* colorimeter from *Konica Minolta*. However, the *CR-400* colorimeter provides color information in the XYZ-CIE space and is required to convert these values to the RGB color space. For this, a code in Matlab was used considering the CIE standard illuminant D65. Images of the patterns were obtained using 4 different cell-phones (*CP1: LG Optimus L3; CP2: iPhone 4s; CP3: Samsung Galaxy B; CP4: Microsoft Lumia 640*). The *CP1* device has Android operating system, 3.1 MP camera and 2048×1533 pixels image resolution. *CP2* has iOS 9 operating system, 8 MP camera and 3264×2448 pixels image resolution. The *CP3* phone features Android operating system, 8MP camera and 3264×2448 pixels image resolution. Finally, *CP4* runs Windows Phone 8.1 platform, 8 MP camera and 1280×721 pixels image resolution. Despite these technical differences, the color images recorded by these devices are True Color type, meaning that each pixel of the RGB image has 24 bits color information. Therefore each of the three-color components (red, green, blue) has a value ranging from 0 to 255 (8 bits).

Pictures were taken without flash and at approximately 30 cm away from the target. Automatic focus was also explored on the image capture procedure. An Android application (App), developed on the Android Studio platform, was developed to evaluate the relative color intensity (Equation (3), (4) and (5)) of two different regions of the same picture. Images captured using the App were saved in PNG format. The developed smartphone application features an intuitive interface, ability to zoom the images and allows the evaluation of images taken by other devices (stored in the mobile-phone).

The color evaluation procedure was also explored on the analysis of molecule adsorption on metallic nanoparticles. A 0.01 g of Cysteamine, from Sigma-Aldrich, was added to 1.3 ml of a gold colloidal solution, with 50 nm nanospheres, from Sigma-Aldrich. Photographs of the colloid were taken before and after the addition of Cysteamine. Absorption spectra were also obtained by using a spectrometer (*USB2000 Ocean Optics*).

3. Results and Discursion

By imaging the color pattern on **Figure 1(a)** with all four different mo-

mobile-phones, significant difference on the output color image were identified. The images were taken under the same illumination condition (under fluorescent light, with 200 Lux) and the color evaluations were performed on an $80 \times 80\text{ pixels}$ area. In addition, all color areas on **Figure 1(a)** were also examined using the colorimeter. **Table 1** shows the RGB color vales for three-color areas of this color pattern. The mobile-phones RGB mean values and the standard deviations were obtained by analyzing 16 images (4 pictures from 4 cell-phones). Huge differences were observed on the RGB colors values obtained by the colorimeter and by the mobile-phones. Those discrepancies are expected since the colorimeter explores an internal light source with optical characteristics distinct from the illumination used on the cell-phone analysis. As shown in **Table 1**, variations on RGB value, obtained from the mobile-phones images, have reached over 28%. Furthermore, one can notice that the evaluation of the R color component of the red color area, and the G component of the green area, and the B color component of the blue area have presented the smaller standard deviations (11.38%, 12.41% and 5.70% respectively).

It is well known in the literature that color evaluation by distinct mobile-phones leads to significant difference of color values [7]. The obtained color values of an acquired picture are influenced by the ambient illumination, the camera color filters, and the white balance algorithms explored [24]. To reduce this limitation, the determination of the absolute color value of an object can be overlooked and a relative color value determined. Ahuja and coauthors have established a method to evaluate relative color, based on RGB components differences in the RGB color space [25]. On the analysis of colors with mobile-phones, Yestisen *et al.* have explored color distances on the 2D (x, y) CIE 1931 chromaticity space, [15]. Sumriddetchkajorn *et al.* have analyzed colors by defining a specific color ratio [17]. Moreover, the described measures presented in references [7]-[12] [17] [19] [20] were performed using a single mobile-phone.

Here we evaluate and compared the performance of several methods, exploring 4 different mobile-phones. Based on the references [15] [17] [25] and on Equation (3) the values of $|R_i - R_r|$, R_i/R_r , $|x_i - x_r|$ and r were determined. The x chromaticity values of the relative color distance $|x_i - x_r|$ was define as $x = R/(R + G + B)$. The index i and r designates the region of interest and the reference region, respectively. For that, the red area of **Figure 1(a)** was set as the region of interest and the white area was established as the reference region. The methods were also appraised using the G and B color channels. **Table 2** shows the averaged relative color intensity values for each color channel and the standard deviations obtained using all four cell-phones. One can observed that the relative color values (r, g, b) , obtained using the proposed method, present the lowest standard deviation (about 2%, 3% and 3.5%, respectively), indicating that proposed procedure is weakly dependent on the mobile-phone used. The results presented in **Table 2** indicate that the self-referenced method for color evaluation based on Equations (3), (4) and (5) reduces the discrepancies of the mobile-phones output color values.

Table 1. RGB values of color regions (red, green and blue) using colorimeter and mobile-phones.

Colorimeter	Region Red			Region Green			Region Blue		
	R	G	B	R	G	B	R	G	B
Mean	27.38	19.93	14.47	19.47	27.43	17.92	16.38	14.96	14.51
S. Deviation (%)	0.11	0.16	0.32	0.35	0.23	0.59	0.22	0.23	0.15
Smartphone	R	G	B	R	G	B	R	G	B
Mean	147.72	69.15	74.17	73.52	125.08	77.43	45.57	64.21	139.44
S. Deviation (%)	11.38	16.40	12.30	19.34	12.41	14.83	28.34	13.35	5.70

Table 2. The relative color values of the **Figure 1(a)** red region using different mobile-phones. White area was used as the reference region.

Relative color	Mobile-phone (mean value)	Standard Deviation (%)
$ R_i - R_r $	48.90	37.9
R_i/R_r	0.75	9.72
$ x_i - x_r $	0.18	10.06
r	1.06	1.93
$ G_i - G_r $	127.32	16.53
G_i/G_r	0.35	14.86
$ y_i - y_r $	0.09	11.11
g	1.25	3.2
$ B_i - B_r $	134.35	23.08
B_i/B_r	0.36	15.83
$ z_i - z_r $	0.091	13.19
b	1.25	3.46

To evaluate the performance of proposed method to distinguish different color shades, the color pattern of **Figure 1(b)** was imaged and analyzed, using Equation (3). **Figure 2** shows the values of the relative color intensity r of different areas of **Figure 1(b)**. In **Figure 2**, the $|R_i - R_r|$ values of abscissa axis were obtained by measuring the R-color component of the red shades areas with the colorimeter. **Figure 2** indicates that the proposed self-referenced colorimetric method can distinguish two different color shades areas with $|R_i - R_r| > 5$, establishing a relative red color threshold, r_p of ~ 1.02 . Similar analyses were performed to identify the limitation of the proposed method in the evaluation of regions with green and blue tonalities. The measured relative green and blue color thresholds were $g_i \sim 1.00$ and $b_i \sim 1.03$.

The relative color values of **Table 2** and **Figure 2** were obtained taking photographs under the same illumination condition. However the real color of an object is greatly dependent of the illumination light source. Relative color

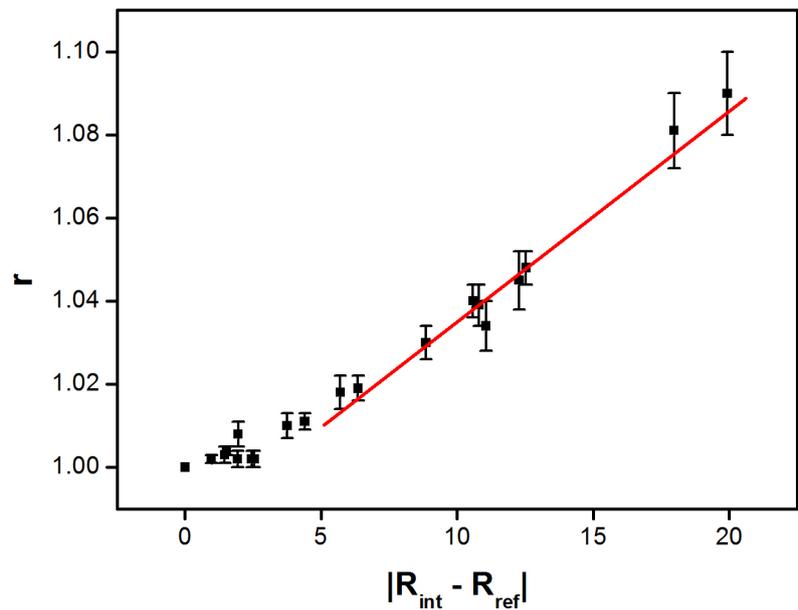


Figure 2. Relative r color intensity values of the **Figure 1(b)** red shades areas as function of the color differences ($|R_i - R_r|$), measured with the colorimeter. The solid line is a guide to the eye.

intensity analysis can also reduce the illumination dependence of the spectroscopic evaluation. **Figure 3** shows the normalized average relative color intensity values of the red shades areas (using regions 1 and 2 from **Figure 1(b)**) under different illuminations. Here a fluorescent lamp (175 Lux), an incandescent lamp (33 Lux) and a white LED (5 Lux) were used as light source. Sunlight (355 Lux) was also explored on the illumination of the color pattern. The relative color average value obtained under sun illumination was used to normalize all the measured values. The smallest deviation (0.53%) of the measured average relative color values was observed using the proposed method (Equation (3)), under different illumination conditions. The proposed self-referenced relative color intensity method reduces the mobile-phone spectroscopic evaluation dependence on the illumination source.

To demonstrate the potential use of the method, the chemical-adsorption of a molecule on nanoparticle surface was evaluated, using a mobile-phone (*CP3*). Optical properties (scattering and absorption) of metallic nanoparticle maybe changed by the adsorption of molecules on the metal surface [26]. In particular, 50 nm gold sphere colloid presents a faint pink color, with an extinction spectrum characterized by a peak (Plasmon peak) at 523.94 nm, and FWHM of 69.13 nm. The addition of Cysteamine to colloid volume and its adsorption on nanostructures induces a red shift of the Plasmon peak to 544.91 nm. **Figure 4** presents the extinction spectra of gold colloid with and without Cysteamine. The interaction of Cysteamine's thiol group with the gold surface is well described in the literature [27] [28]. Thiol-gold interaction establishes the basis to the development of robust self-assembled monolayers for several applications, as optical biosensors

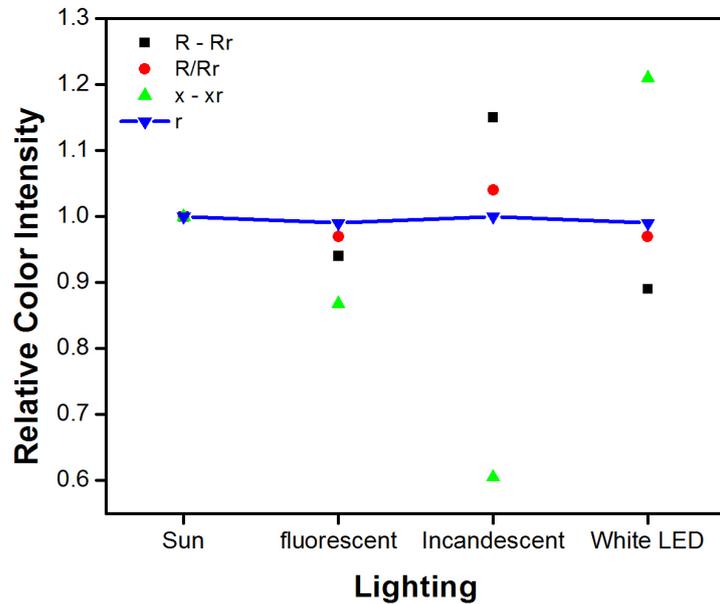


Figure 3. Dependence of the relative color parameters as a function of the light source.

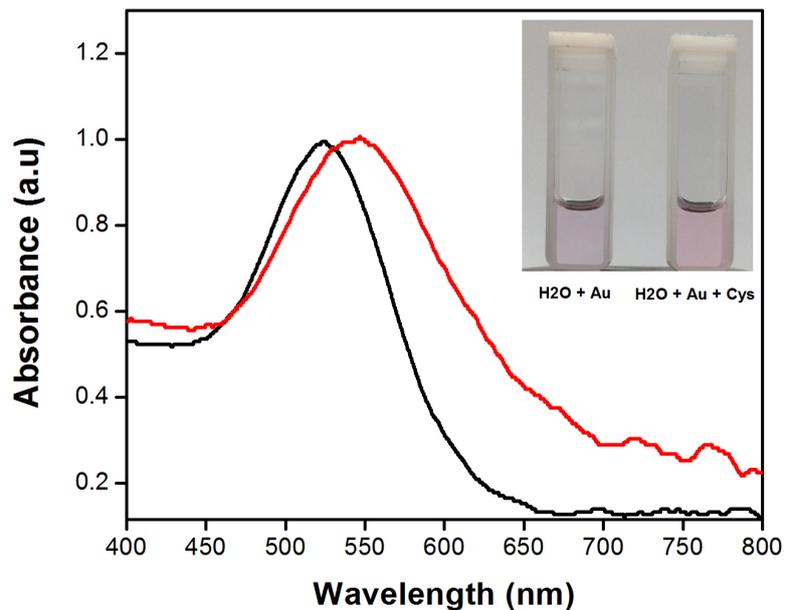


Figure 4. Extinction spectra of the 50 nm Au sphere colloid with (red) and without (black) Cysteamine. Inset: Mobile-phone image of the colloid with and without Cysteamine (Cys).

[29] [30] [31]. The inset of **Figure 4** shows a picture of the two solutions (colloid with and without Cysteamine) in 1 cm optical length glass cuvettes. The color difference of the samples is hardly noticed.

The colloid's relative color value was determined using the developed mobile-phone application, installed on the *CP3* mobile phone. For that, the colloid without Cysteamine image was chosen to be the reference region and an area of

the gold-Cysteamine solution image was selected as the interested region. The App interface with the processing result for a given image is shown in **Figure 5**. After taking a photo with the mobile-phone application, the image is shown on the display of the device with two regions markers (black and red). The black marker was positioned over the region of the image used as a color reference, while the red marker was placed on the region of interest. The color analysis was performed by selecting the render button (process) of the App.

The time behavior of the Cysteamine adsorption on the gold surface was evaluated with the self-referenced relative color assay. **Figure 6** shows the time evolution of the sample's red component of the relative color value with the addition of Cysteamine in the gold colloid and the peak shift in the spectrum due to the presence of the Cysteamine. Although the method cannot precisely identify color changes smaller than r_p , we were able to verify that the adsorption process takes approximately 40 min to be complete established. A good correlation ($r^2 = 0.9824$) between the red component of the relative color and the corresponding extinction peak wavelength were identified, as shown in the inset of **Figure 6**. The measured r value change ($r = 1.013$), associated to the colloid color modification, was close to the relative color threshold value, r_p , determined on **Figure 2**, indicating the capability of the method to distinguish slight colors changes. Moreover we were able, with a mobile-phone, to identify a 20.99 nm spectrum shift of a 69.13 nm FWHM spectrum band.

4. Conclusion

The color analysis of an object using distinct mobile-phones can lead to disparate results. To overcome this obstacle a mobile-phone based standalone spectroscopic method, without calibration procedures, to determine relative color values of an image was proposed and evaluated. On the analysis of a color pattern, no more than 3.5% of deviation was observed on the relative color values obtained by four different mobile-phones, indicating that the proposed color

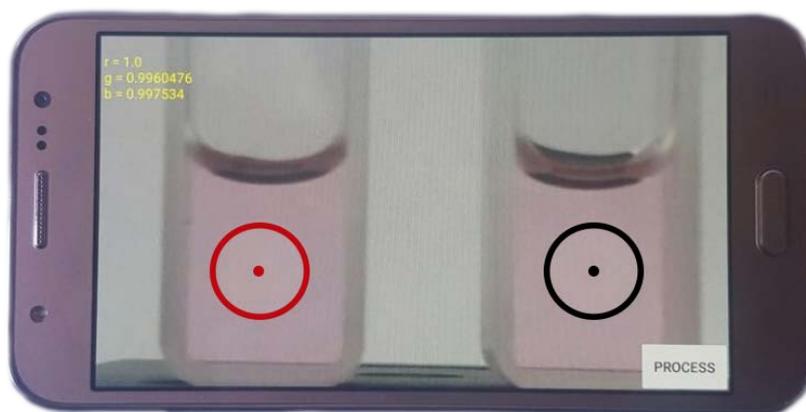


Figure 5. Picture of the mobile-phone running the self-referencing colorimetric App. The black and red-circled areas indicate the region of interest and the reference region.

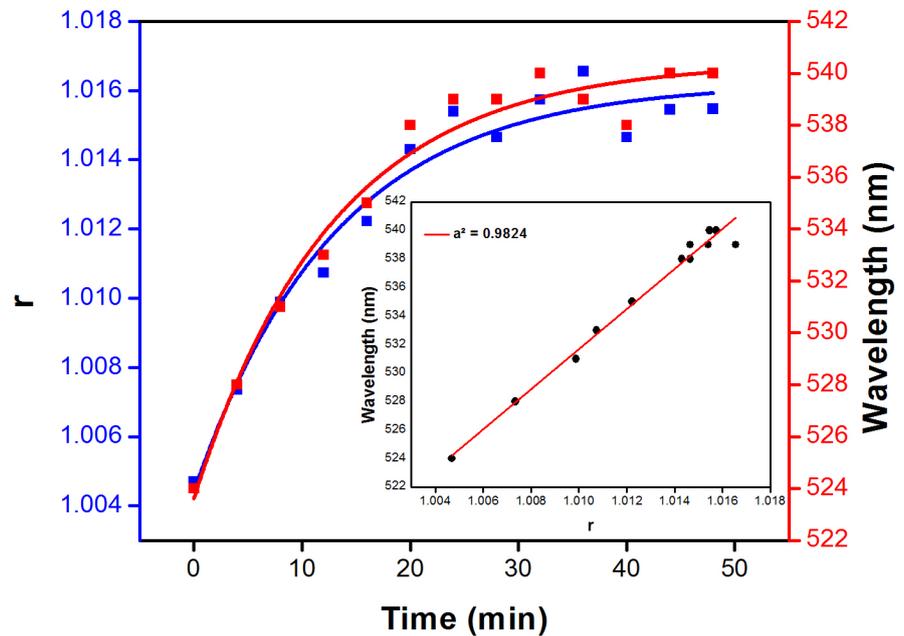


Figure 6. Time evolution of the goldcolloid extinction spectrum peak (red) and red component of the relative color intensity value (blue), with the addition of Cysteamine. The inset shows a linear correlation ($a^2 = 0.9824$) of the r and the colloid extinction peak.

evaluation procedure is weakly dependent of the image device used. The proposed self-referenced spectroscopic method can distinguish 2% ($|R_i - R_r| > 5$) difference on the R-color component of red shades areas. We also demonstrated that relative color analysis could also reduce the illumination dependence on the spectroscopic evaluation. Under different illumination conditions we observed a 0.53% deviation of the measured relative color values of a color pattern. Moreover, the method allowed identifying a 20.99 nm spectrum shift of a 69.13 nm FWHM spectrum band of colloidal gold, due to the Cysteamine adsorption on the metal surface. The proposed self-referenced technique leads to relative color intensity values that are weakly dependent of the illumination condition and mobile-phone used. Therefore the color analysis method can improve and expand the use of cell-phones in standalone spectroscopic applications. Moreover, mobile-phones spectroscopic platforms can contribute to the democratization of diagnostics technologies globally, by offering high sensitivity cost-effective sensors [32].

Acknowledgements

The authors acknowledge the financial support from the Brazilian agency CAPES/PNPD, FACEPE, CNPq and the National Institute of Photonics.

References

- [1] Cleve, E., Bach, E. and Schollmeyer, E. (2000) Using Chemometric Methods and NIR Spectrophotometry in the Textile Industry. *Analytica Chimica Acta*, **420**, 163-167. [https://doi.org/10.1016/S0003-2670\(00\)00888-6](https://doi.org/10.1016/S0003-2670(00)00888-6)

- [2] Yin, Y., Cao, Y., Xu, Y. and Li, G. (2010) Colorimetric Immunoassay for Detection of Tumor Markers. *International Journal Molecular Sciences*, **11**, 5077-5094. <https://doi.org/10.3390/ijms11125077>
- [3] Xing, J., Ngadi, M., Gunenc, A., Prasher, S. and Garipey, C. (2007) Use of Visible Spectroscopy for Quality Classification of Intact Pork Meat. *Journal of Food Engineering*, **82**, 135-141. <https://doi.org/10.1016/j.jfoodeng.2007.01.020>
- [4] Zhang, C. and Suslick, K.S. (2005) A Colorimetric Sensor Array for Organics in Water. *Journal of the American Chemical Society*, **127**, 11548-11549. <https://doi.org/10.1021/ja052606z>
- [5] Kim, H., Awofeso, O., Choi, S., Juang, Y. and Bal, E. (2017) Colorimetric Analysis of Saliva-Alcohol Test Strips by Smartphone-Based Instruments Using Machine-Learning Algorithms. *Applied Optics*, **56**, 84-92. <https://doi.org/10.1364/AO.56.000084>
- [6] Ilja, Z. (2013) Colour Management Using Mobile Phone Camera. Bachelor's Thesis. Helsinkimetropolia University of Applied Sciences, Finland.
- [7] Shin, J., Choi, S., Yang, J., Song, J., Chui, J. and Juang, H. (2017) Smart Forensic Phone: Colorimetric Analysis of a Bloodstain for Age Estimation Using a Smartphone. *Sensors and Actuators B*, **243**, 221-225. <https://doi.org/10.1016/j.snb.2016.11.142>
- [8] Xu, W., Lu, S., Chen, Y., Zhao, T., Jiang, Y., Wang, Y. and Chen X. (2015) Simultaneous Color Sensing of O₂ and pH Using a Smartphone. *Sensors and Actuators B*, **220**, 326-330. <https://doi.org/10.1016/j.snb.2015.05.088>
- [9] Zachary, J.S., Kaiqin, C., Alyssa, R.E., Mehdi, R., Amy, G., Marco, M., Denis, M.D., Stephen, L., Dennis, M. and Sebastian, W.H. (2011) Cell-Phone-Based Platform for Biomedical Device Development and Education Applications. *PLoS ONE*, **6**, 1-11.
- [10] Sewoong, K., Dongrae, C., Jihun, K., Manjae, K., Sangyeon, Y., Jae, E.J., Minkyu, J., Dong, H.L., Boreom, L., Daniel, L.F. and Jae, Y.H. (2016) Smartphone-Based Multispectral Imaging: System Development and Potential for Mobile Skin Diagnosis. *Biomedical Optics Express*, **7**, 5294-53071. <https://doi.org/10.1364/BOE.7.005294>
- [11] Karthik, R.K., Surya, C. and Michael, G. (2017) mHealth Dipstick Analyzer for Monitoring of Pregnancy Complications. *IEEE Sensors Journal*, **17**, 7311-7316. <https://doi.org/10.1109/JSEN.2017.2752722>
- [12] Sandeep, K.V., Thomas, V.O., Marion, S., Roland, Z., Felix, V.S. and John, L. (2015) A Smartphone-Based Colorimetric Reader for Bioanalytical Applications Using the Screen-Based Bottom Illumination Provided by Gadgets. *Biosensors and Bioelectronics*, **67**, 248-255. <https://doi.org/10.1016/j.bios.2014.08.027>
- [13] Brandon, B., Bingen, C., Derek, T., Haydar, O., Steve, F., Qingshan, W., et al. (2015) Cellphone-Based Hand-Held Microplate Reader for Point-of-Care Testing of Enzyme-Linked Immunosorbent Assays. *ASC Nano*, **9**, 7857-7866. <https://doi.org/10.1021/acsnano.5b03203>
- [14] Wang, Y., Liu, X., Chen, P., Tran, N.T., Zhang, J., Chia, W.S., et al. (2016) Smartphone Spectrometer for Colorimetric Biosensing. *Analyst*, **141**, 3233-3238. <https://doi.org/10.1039/C5AN02508G>
- [15] Yetisen, A.K., Martinez-Hurtado, J.L., Garcia-Melendrez, A., Vasconcelos, F.C. and Lowe, C.R. (2014) A Smartphone Algorithm with Inter-Phone Repeatability for the Analysis of Colorimetric Tests. *Sensors and Actuators B*, **196**, 156-160. <https://doi.org/10.1016/j.snb.2014.01.077>
- [16] Mathaweasurn, A., Maneerat, N. and Choengchan, N. (2017) A Mobile Phone-Based Analyzer for Quantitative Determination of Urinary Albumin Using

Self-Calibration Approach. *Sensors and Actuators B*, **24**, 476-483.

<https://doi.org/10.1016/j.snb.2016.11.057>

- [17] Sumriddetchkajorn, S., Chaitavon, K. and Intaravanne, Y. (2013) Mobile Device-Based Self-Referencing Colorimeter for Monitoring Chlorine Concentration in Water. *Sensors and Actuators B*, **182**, 592-597.
<https://doi.org/10.1016/j.snb.2013.03.080>
- [18] Samuel, S. (2014) Colorimetric Water Quality Sensing with Mobile Smart Phones. PhD Thesis, University of British Columbia, Vancouver.
- [19] Erika, A.T., Scott, E.G., William, A.S., Robert, G.M., Alexander, J.W., Alexa, F.S., *et al.* (2014) Disposable Platform Provides Visual and Color-Based Point-of-Care Anemia Self-Testing. *Journal of Clinical Investigation*, **124**, 4387-4394.
<https://doi.org/10.1172/JCI76666>
- [20] Yuttana, I., Sarun, S. and Jiti, N. (2012) Cell Phone-Based Two-Dimensional Spectral Analysis for Banana Ripeness Estimation. *Sensors and Actuators B*, **168**, 390-394. <https://doi.org/10.1016/j.snb.2012.04.042>
- [21] Reza, S. and Milivoje, A. (2010) An Efficient Spectral-Based Calibration Method for RGB White-Balancing Gains under Various Illumination Conditions for Cell-Phone Cameras.
- [22] Stephen, W. and Catarina, R. (2004) Computational Colour Science Using Matlab. John Wiley & Sons Ltd., Hoboken.
- [23] Charles, A.P. (1993) Gamma and Its Disguises: The Nonlinear Mappings of Intensity in Perception, CRTs, Film, and Video. *SMPTE Journal*, **102**, 1099-1108.
- [24] Felix, K., Peter, B., Sangam, C. and Tran, Q.K. (2013) Optimized Flash Light-Emitting Diode Spectra for Mobile Phone Cameras. *Applied Optics*, **36**, 8779-8788.
- [25] Punkaj, A., Maria, A.P., Brian, D.H. and Miklós, G. (2014) Minimizing Color Interference from Biological Samples in Optode-Based Measurements. *Sensors and Actuators B*, **204**, 319-325. <https://doi.org/10.1016/j.snb.2014.07.113>
- [26] Willets, K.A. and Van Duyne, R.P. (2007) Localized Surface Plasmon Resonance Spectroscopy and Sensing. *Annual Review of Physical Chemistry*, **58**, 267-297.
<https://doi.org/10.1146/annurev.physchem.58.032806.104607>
- [27] Yurui, X., Xun, L., Hongbin, L. and Wenke, Z. (2014) Quantifying Thiol-Gold Interactions towards the Efficient Strength Control. *Nature Communications*, **5**, Article No. 4348. <https://doi.org/10.1038/ncomms5348>
- [28] Michel, G., Martin, B., Matthias, R. and Hermann, E. (1999) How Strong Is a Covalent Bond? *Science*, **283**, 1727-1730.
- [29] Anker, J.N., Hall, W.P., Lyandres, O., Shah, N.C., Zhao, J. and Van Duyne, R.P. (2008) Biosensing with Plasmonicnanosensors. *Nature Materials*, **7**, 442-453.
<https://doi.org/10.1038/nmat2162>
- [30] Jie, C., Tong, S. and Kenneth, G. (2014) Gold Nanorod-Based Localized Surface Plasmonresonance Biosensors: A Review. *Sensors and Actuator B*, **195**, 332-351.
<https://doi.org/10.1016/j.snb.2014.01.056>
- [31] Alexandre, C., Paula, G., Ana, C., Arthur, M., Rosa, D., Renato, E.A., *et al.* (2013) Dengue Immunoassay with an LSPR Fiber Optic Sensor. *Optics Express*, **21**, 27023-27031. <https://doi.org/10.1364/OE.21.027023>
- [32] Aydogan, O. (2014) Mobile Phones Democratize and Cultivate Next-Generation Imaging, Diagnostics and Measurement Tools. *Lab on a Chip*, **14**, 3187-3194.
<https://doi.org/10.1039/C4LC00010B>