

Induction of Apoptosis in the Human Prostate Cancer Cell Line DU-145 by a Novel Micronutrient Formulation

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Abstract

Prostate cancer, the most frequently diagnosed cancer in men, primarily affects males aged 55 and older and is more common in African Americans than Caucasians. Once the cancer has metastasized, current treatments are generally ineffective. We have identified a novel anti-neoplastic agent, a specifically designed nutrient mixture (NM), containing ascorbic acid, lysine, proline and green tea extract that demonstrates a broad spectrum of anti-tumor activity against a number of human cancer cell lines. In a previous study NM significantly inhibited prostate tumor in nude mice. In this study, we tested whether the formulation exerts its anti-tumor effects through induction of apoptosis on prostate cancer cell line DU-145. The effect of the nutrient mixture (NM) on cell growth inhibition in DU-145 cells was examined by 3-(4,5-dimethylthiazole-2-yl)-2,5-diphenyltetrazolium bromide (MTT) assay. Morphological changes and caspase activation associated with apoptosis induction was checked by H&E staining and Live Green Caspase assay, respectively. The NM was found to be slightly toxic to DU-145 cells at 100 µg/ml, but significantly toxic at 500 µg/ml and 1000 µg/ml. Percentage of cells undergoing apoptosis also increased from 6% at 100 µg/ml to 49% at 500 µg/ml and 83% at 1000 µg/ml, with greater number of cells showing morphological changes such as condensed nuclei and an acidophilic cytoplasm at higher concentrations. For the purpose of comparison, NM was also tested on a normal human dermal fibroblast (NHDF) cell line which exhibited far less apoptosis induction as compared to DU-145 cells. The percentage of cells undergoing apoptosis in case of NHDF cells was 7% at 100 µg/ml, 25.6% at 500 µg/ml and 76.5% at 1000 µg/ml. Our results demonstrate that the NM is effective in inhibiting cancer cell viability and inducing apoptosis in prostate cancer DU-145 cells and can thus be used as an effective treatment for prostate cancer.

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Keywords

Prostate Cancer DU-145, NHDF, Apoptosis, Cytotoxicity, Caspase, MTT

1. Introduction

Prostate cancer, the leading non-cutaneous cancer and the second leading cause of death in men in the U.S., primarily affects males of age 55 and above and is more common in African American males than white males [1]. It accounted for approximately 15% of all cancer diagnosis and 6.6% of all cancer deaths in men in 2012. According to the estimates of world cancer prevalence, prostate cancer is likely to contribute 25.6% of all the cancers in men in the next 5 years [2].

Standard treatment of prostate cancer consists of surgery (prostatectomy), hormonal therapy and radiotherapy. While prostatectomy is successful for treating early stage cancer, its use is limited to localized tumors and can cause nerve damage resulting in impotence, incontinence or rectal injury [3]-[5]. Hormonal therapy acts by blocking the action of cancer supporting male hormones and is only effective till the time the cancer can spread independent of male hormones [6] [7]. Similarly, radiotherapy is also effective for early stage prostate cancer and becomes progressively ineffective in later stages. Thus, while prostate carcinomas are initially responsive to surgery and hormonal therapies when localized, they eventually become more aggressive and unresponsive to standard treatments.

Due to the limitations of the current treatment modalities in tackling prostate cancer, there is an urgent need for safe and effective therapeutic approaches. Epidemiological studies have suggested that consumption of a plant based diet has been associated with the prevention of cancer. Diet and nutritional supplementation have often been analyzed by several studies for beneficial effect on prostate cancer attenuation [8] [9]. Several studies have also implicated the phytochemicals and micro nutrients such as epigallocatechin gallate (EGCG—present in green tea), ascorbic acid, N-acetyl cysteine and selenium individually have anti-cancer properties. However, a study combining the micronutrients in the appropriate quantities for synergistic and enhanced action is lacking.

We have developed strategies to inhibit cancer development and its spread by using naturally occurring nutrients such as ascorbic acid, green tea extract, lysine and proline in a unique combination. This unique nutrient mixture (NM) demonstrates a broad spectrum of anti-tumor activity against a number of human cancer cell lines. In a previous study we found that NM significantly inhibited prostate tumors in nude mice. In this study we investigated in a DU-145 prostate cancer cell line whether the inhibition of prostate tumors by NM was due to induction of apoptosis.

The anti-proliferative and apoptotic effect of NM was determined *in vitro* by checking for the loss of cell viability, cell morphology changes and caspase activation in human prostate DU-145 cells. Apoptosis induction was also analyzed in normal cells represented by human dermal fibroblasts upon treatment with the NM to check for distinct effects on cancer and normal cells.

2. Materials and Methods

2.1. Composition of the Nutrient Mixture

The stock solution of the nutrient mixture (total weight 4.4 g) used for testing was composed of the following in the quantities indicated: Vitamin C (as ascorbic acid and as Mg, Ca and Palmitate ascorbate) 700 mg, L-lysine 1000 mg; L-proline 750 mg; L-arginine 500 mg, N-acetyl cysteine 200 mg; standardized green tea extract 1000 mg (derived from green tea leaves, was obtained from US Pharma Lab; the certificate of analysis indicated the following characteristics: total polyphenol: 80%, Catechins: 60%; Epigallocatechin gallate (EGCG): 35% and Caffeine: 1%), selenium 30 µg, copper 2 mg, manganese 1 mg.

2.2. Cells and Cell Culture

Human prostate cancer cell line DU-145 and normal human dermal fibroblasts (NHDF) were obtained from ATCC (American Type Culture Collection, Rockville, MD, USA) and maintained in Dulbecco's Modified Eagle's Medium (DMEM) and supplemented with 10% FBS, 100 U/ml penicillin and 100 µg/ml streptomycin.

The media and sera were obtained from ATCC, and antibiotics from Gibco BRL (Long Island, NY, USA). The cells were grown at 37°C in 5% CO₂.

2.3. MTT Cytotoxicity Assay

Cell viability was evaluated by MTT assay, a colorimetric assay based on the ability of viable cells to reduce a soluble yellow tetrazolium salt (3-[4,5-dimethylthiazol-2-yl]-2,5-diphenyltetrazolium bromide) to a blue formazan crystal by mitochondrial succinate dehydrogenase activity of viable cells. This test is a good index of mitochondrial activity and thus of cell viability. Briefly, cell suspensions were plated in 24-well tissue culture plates (Nunc, Denmark) at a concentration of 3×10^4 cells/well. After incubating the plates for 24 hours at 37°C in a humidified incubator, the cells were treated with the nutrient mixture at concentrations of 100, 500 and 1000 µg/ml for 24 hours or left untreated in the control group. 500 µl of MTT assay reagent (Sigma No. M-2128-0.5 mg/ml in media) was added to each well followed by 2-hour incubation at 37°C. Following incubation, the solution was carefully aspirated from the wells, the formazan product was dissolved in 1 ml DMSO and the absorbance (OD) was measured on a microplate reader at a wavelength of 570 nm in BioSpec 1601, Shimadzu spectrometer. The OD₅₇₀ of the DMSO solution in each well was considered to be proportional to the number of cells. The OD₅₇₀ of the control (treatment without supplement) was considered to be 100%.

2.4. Live Green Caspase Assay

The cells were grown to confluence and either left in media alone or challenged with the NM dissolved in media at 100, 500 and 1000 µg/ml and incubated for 24 hours. The cell culture was washed with PBS and treated with the caspase reagent as specified in the manufacturer's protocol (Molecular Probes Image-IT Live Green Poly Caspases Detection Kit 135104, Invitrogen). The cells were photographed under the fluorescence microscope and counted. Green colored cells represent viable cells, while orange and red colors represent early and late apoptotic cells respectively.

2.5. H & E Staining of Cells

The cells were cultured in 24-well plates and were either kept untreated or treated with NM in test concentrations of 100, 500 and 1000 µg/ml. After 24 hour incubation, the cells were washed with PBS, fixed with cold methanol and then stained with haematoxylin and eosin for 5 min each. The stained cells were then observed and imaged by microscopy.

2.6. Statistical Analysis

The results were expressed as mean ± standard deviation (SD) for the groups. Data was analyzed by the independent t-test.

3. Results

3.1. Nutrient Mixture Decreased the Viability of Prostate Cancer Cells

To determine whether the nutrient formulation is cytotoxic to prostate cancer cells DU-145, we used the MTT assay to check for cell viability at various NM dosages. The cells in the control group were left untreated and those in the test group were treated with 100, 500 and 1000 µg/ml of nutrient mixture for 24 hr. Our results demonstrated that the nutrient formulation can induce cell death in the prostate cancer cells in a dose dependent manner after 24 hours of incubation. As can be seen in **Figure 1**, the cell viability showed a significant dose dependent decrease with 84% ± 6% viable cells at 100 µg/ml NM, 63% ± 12% at 500 µg/ml NM and 53% ± 4% at 1000 µg/ml NM, respectively, as compared to the untreated control group.

3.2. Nutrient Mixture Induced Apoptosis through Induction of Caspases

Quantitative analysis of DU-145 cells in early and late apoptotic stages was done by using the Live Green Caspase assay. Analysis with Live Green Caspase detection kit revealed a dose dependent increase in apoptosis of DU-145 cells with slight apoptosis found at 100 µg/ml, moderate at 500 µg/ml and significant at 1000 µg/ml

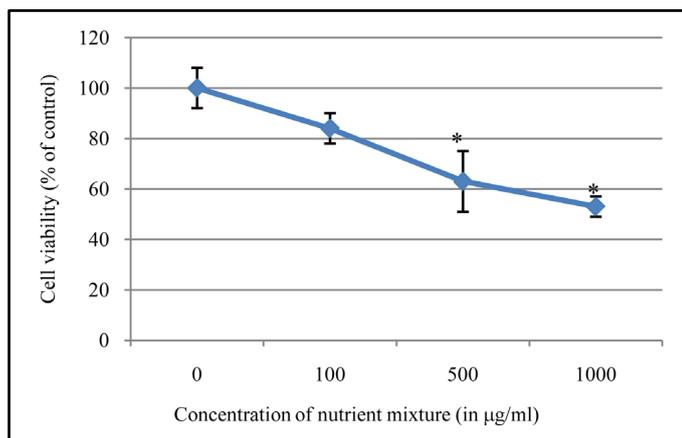


Figure 1. Effect of NM on cell viability: MTT assay using prostate cancer DU-145 cells treated with (a) 0 µg/ml, (b) 100 µg/ml, (c) 500 µg/ml, (d) 1000 µg/ml concentrations of NM respectively ($p < 0.01$).

NM. Quantitative analysis of the data revealed the percentage of apoptotic cells to be 6% at 100 µg/ml (all in early phase apoptosis), 49% at 500 µg/ml (10% in early phase; 39% in late phase) and 83% for 1000 µg/ml NM (8% in early phase; 75% in late phase). It was observed that there was a corresponding decrease in percentage of live cells and an increase in percentage of cells in late apoptotic stages with increasing NM concentrations. A quantitative analysis of apoptosis induction in normal human dermal fibroblasts showed equivalency to DU-145 cells at 100 µg/ml. However, at higher concentrations of NM the induction of apoptosis was significantly lower in NHDF cells as compared to DU-145 cells. The percentage of apoptotic NHDF cells was 7% at 100 µg/ml (2% in early phase, 5% in late phase), 25.6% at 500 µg/ml (4.6% in early phase, 21% in late phase) and 76.5% at 1000 µg/ml (23.5% in early phase, 53% in late phase). Photomicrographs of apoptotic DU-145 cells and NHDF are shown in **Figures 2(a)-(d)** and **Figures 3(a)-(d)**, respectively. The quantitative analysis of live, early and late DU-145 prostate cancer cells and NHDF cells are shown in **Figure 2(e)** and **Figure 3(e)**, respectively.

3.3. Nutrient Mixture Induced Morphological Changes Associated with Apoptosis

The DU-145 cells were stained by using H & E to detect apoptotic cells which show the characteristic morphological changes such as the shrinkage of the cytoplasm and darkly stained nuclei with intensely acidophilic cytoplasm. H & E staining of DU-145 cells treated with the nutrient formulation showed that with increasing concentrations of NM the cells showed extensive cell shrinkage, rounding and nuclear condensation (**Figure 4**). The changes were dose dependent, increasing in intensity with increasing NM concentrations.

4. Discussion

The stimulation of suppressed apoptotic pathways in cancer cells and the induction of apoptosis is a predominant mechanistic approach to target cancer. It has previously been reported that NM inhibited prostate tumor growth in xenografts [10]. In this study we analyzed whether the tumor inhibitory effect may be due to the cytotoxic and apoptotic effect of NM treatment on DU-145 prostate cancer cells.

The cytotoxic effect of NM on prostate cancer cell line DU-145 was evident from the significant dose dependent increase in cytotoxicity that was observed with increasing NM concentrations. The cell viability showed a substantial decrease from 84% \pm 6% at 100 µg/ml to 63% \pm 12% at 500 µg/ml and 53% \pm 4% at 1000 µg/ml. Thus, the NM is capable of inhibiting cell viability in a dose dependent fashion.

Activation of caspase enzymes is a distinctive feature of the early stages of apoptosis and these enzymes participate in a series of reactions that are triggered in response to pro-apoptotic signals and result in the cleavage of protein substrates and in the subsequent disassembly of the cell. Upon treatment with 100, 500 and 1000 µg/ml NM, activated caspases were detected. A dose dependency was seen in the activation of caspases and induction of apoptosis, with cells progressing from 6% in early stage apoptosis at 100 µg/ml to 49% at 500 µg/ml and to 83% at 1000 µg/ml NM concentration. A dose dependent induction of apoptosis was also observed in case of

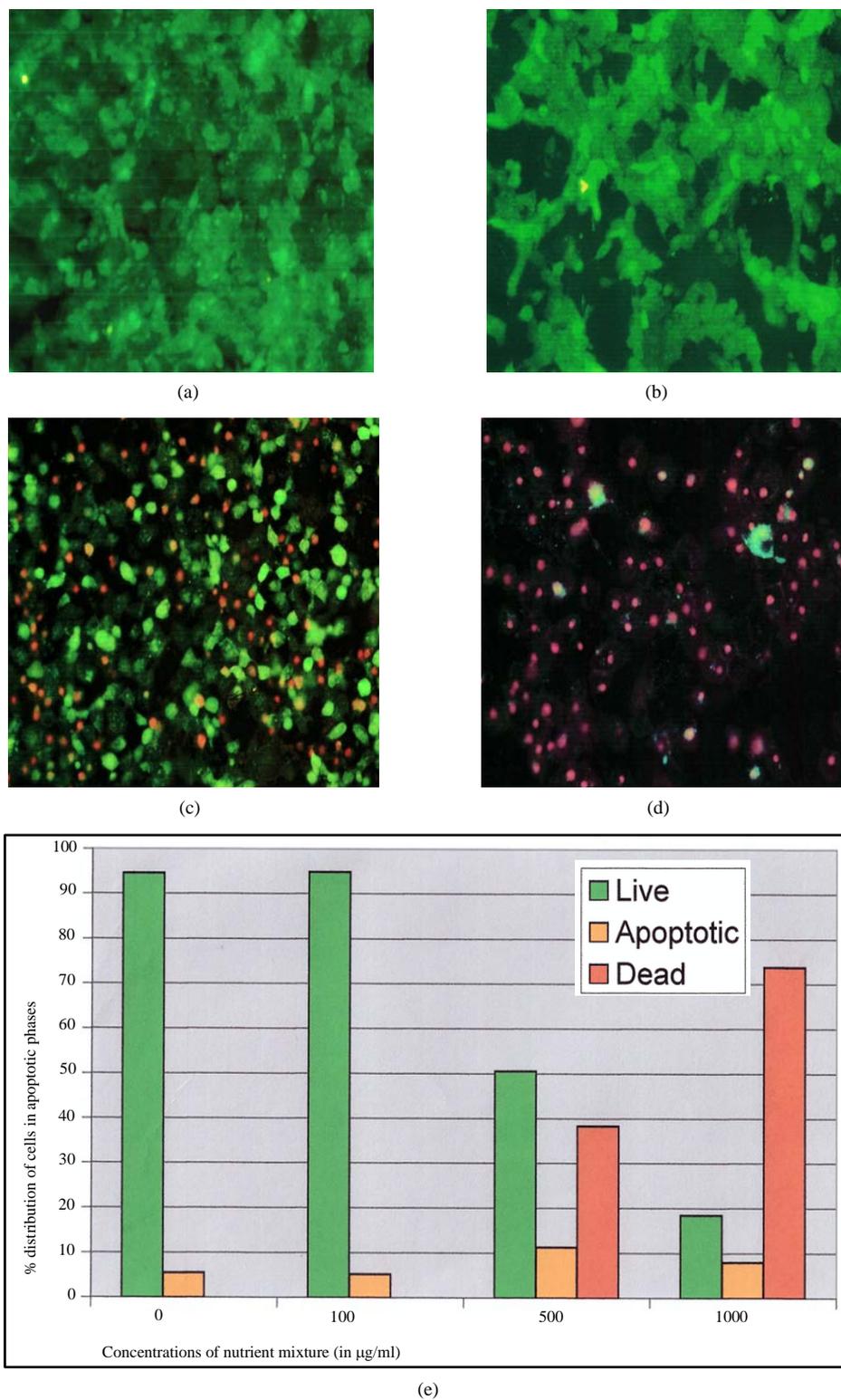


Figure 2. Effect of NM on induction of apoptosis in prostate cancer DU-145 cells. Photomicrographs of DU-145 cells ($\times 20$ magnification) treated with (a) 0 $\mu\text{g/ml}$, (b) 100 $\mu\text{g/ml}$, (c) 500 $\mu\text{g/ml}$, (d) 1000 $\mu\text{g/ml}$ concentrations of NM respectively. (e) Analytical representation of the differential distribution of cells in early or late phases of apoptosis upon treatment with 0, 100, 500 and 1000 $\mu\text{g/ml}$ NM concentrations.

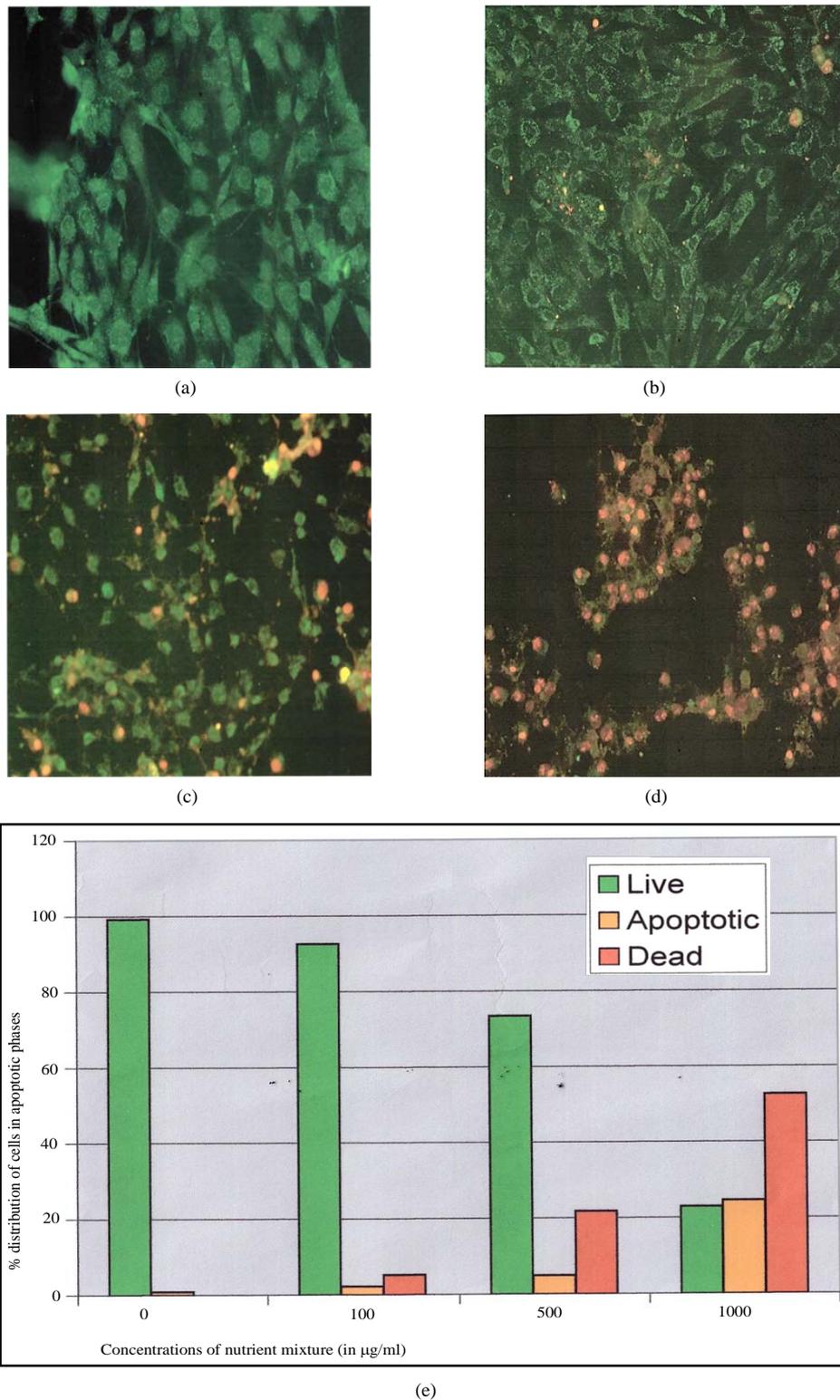


Figure 3. Effect of NM on induction of apoptosis in normal NHDF cells. Photomicrographs of NHDF cells ($\times 20$ magnification) treated with (a) 0 $\mu\text{g/ml}$, (b) 100 $\mu\text{g/ml}$, (c) 500 $\mu\text{g/ml}$, (d) 1000 $\mu\text{g/ml}$ concentrations of NM respectively. (e) Analytical representation of the differential distribution of cells in early or late phases of apoptosis upon treatment with 0, 100, 500 and 1000 $\mu\text{g/ml}$ NM concentrations.

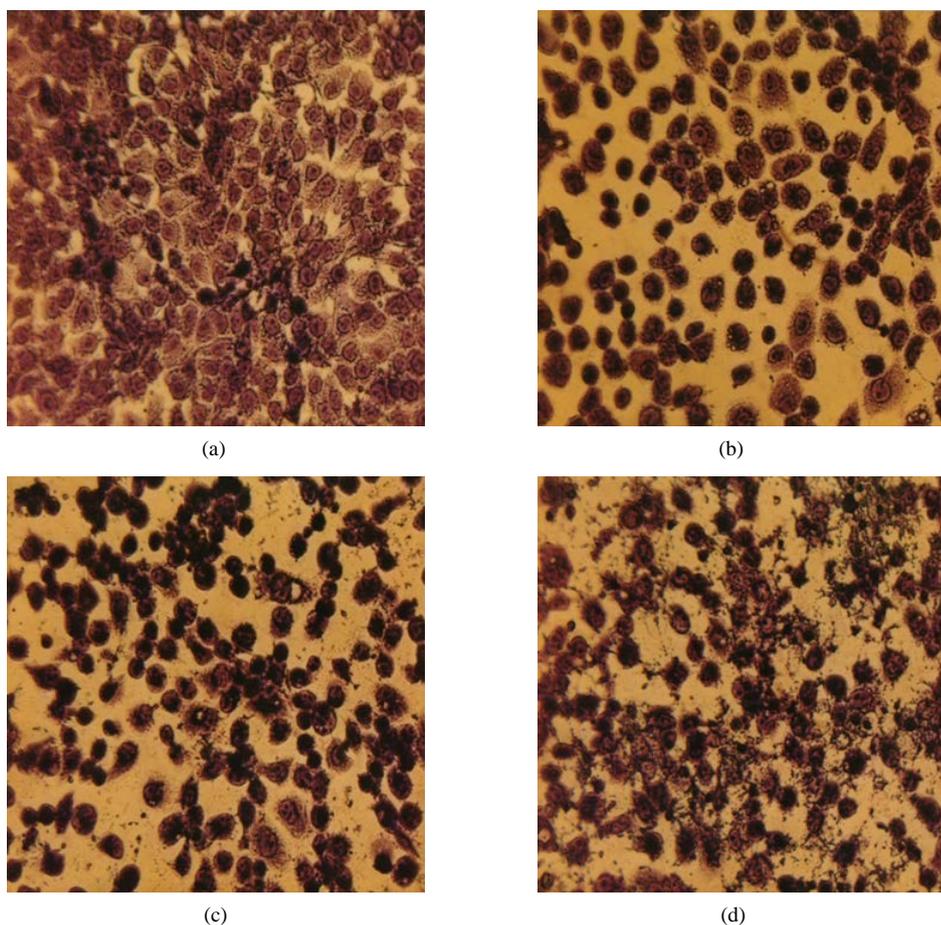


Figure 4. Effect of NM on morphological changes: H & E staining of prostate cancer DU-145 cells treated with (a) 0 µg/ml, (b) 100 µg/ml, (c) 500 µg/ml, (d) 1000 µg/ml concentrations of NM respectively.

NHDF cells treated with increasing concentrations of NM. While the percentage of apoptotic cells was 7% at 100 µg/ml, it increased to 25.6% at 500 µg/ml and to 76.5% at 1000 µg/ml NM.

Morphological changes characteristic of apoptotic cells such as cell shrinkage, nuclear compaction, loss of membrane symmetry and condensation of the cytoplasm were also analyzed. Upon treatment of DU-145 prostate cancer cells with 100, 500 and 1000 µg/ml of NM, it was found that the progressive rounding of cells and nuclear condensation showed a dose dependent increase.

From the results, it can thus be seen that at 100µg/ml NM a very small percentage of cells showed cytotoxicity (16%), apoptosis induction (6%) and morphological changes. On the other hand, at 500 µg/ml NM, significant cytotoxicity (37%) and apoptosis induction (49%) was observed. At 1000 µg/ml NM, apoptosis induction (83%) and cytotoxicity (47%) were even higher. The NHDF cells in turn showed a lower induction of apoptosis as compared to the prostate cancer DU-145 cells, especially at higher treatment doses. A comparison of apoptosis induction at 500 µg/ml and 1000 µg/ml NM revealed a 23.4% and 6.5% lesser percentage of apoptotic NHDF cells as compared to DU-145 cells. It can thus be inferred that the effective concentration of NM would ideally be 500 µg/ml which can induce apoptosis through caspase activation in a significant number of cancer cells while not significantly effect of normal cells.

The nutrient formulation used in the study was specifically developed to combine the individual anti-tumorigenic and pro-apoptotic properties of the component micronutrients. The inhibitory effects of the individual nutrients comprising the novel nutrient formulation have been reported in both experimental and clinical studies. Ascorbic acid is increasingly recognized as an agent with broad biological function. Among its well-known functions are its antioxidant and free radical scavenging functions and detoxification of exogenous compounds

[11]. Previous studies have described the mechanisms of action of ascorbic acid in cancer prevention [12]-[15], including a role in collagen synthesis and basement membrane integrity and hyaluronidase inhibition [16] [17], which may be important in inhibiting tumor spread and micrometastases. It has also been linked to the induction of both necrosis and apoptosis in prostate cancer [18].

Green tea has been shown to have chemopreventive and protective effects on prostate cancer incidence in a case controlled study in China [19] and a proof of principle study in Italy [20]. The green tea catechins such as (-)-epigallocatechin-3-gallate (EGCG), (-)-epigallocatechin (EGC), (-)-epicatechin-3-gallate (ECG), and (-)-epicatechin (EC) have been proven to be chemopreventive agents *in vitro* and in many *in vivo* animal models of induced carcinogenesis [21] [22]. EGCG on its own is also a potent anti-cancer agent and has been reported to have a growth inhibitory effect against certain human cancer cell lines [23]-[25].

Supplementation with micronutrients such as selenium, studied alone in intervention trials [26]-[29] has shown chemopreventive effects in prostate cancer. Even in combination with other nutrients, selenium has been shown to have chemopreventive effects in prostate and colorectal cancer [30].

The nutrient mixture was formulated based on a number of studies conducted in our lab to test various combinations of micronutrients. The selection of a particular formulation was dependent on demonstration of significant synergy in inhibiting cancer progression and metastasis. It has been observed in previous studies that a combination of nutrients such as ascorbic acid, EGCG, lysine and proline show a synergistic anticancer effect which is much more effective than any of the individual nutrients alone [31].

The NM has also proven to be effective in inhibiting a variety of other cancer types which has been demonstrated by both *in vitro* and *in vivo* studies. As seen in **Table 1**, tumor growth in a variety of tumor types was inhibited by treatment with NM in diet. While a reduction in tumor weight of 42% was observed in hepatocellular carcinoma SK-Hep-1 xenografts [32], it went down by 59% in cervical HeLa [33] and 63% in colon HCT-116 xenografts [34]. Similar results were obtained with *in vitro* cancer models in which 1000 µg/ml NM resulted in growth inhibition of various cancer cell lines as seen in **Table 2**. The breast cancer cell line MDA-MB-231 [35] showed a growth inhibition of 50%, while melanoma A2058 [36] and liposarcoma SW-872 [37] showed inhibition of 64% and 61%, respectively. A relatively lower growth inhibition was seen in the osteosarcoma cell line MNNG which showed an inhibition of only 10% [38].

In positive correlation with the results of this study, the prostate cancer cell line LNCaP showed a growth inhibition of 80% [39] while prostate cancer PC-3 xenografts showed a reduction in tumor weight by 47% [10]. NM has also been substantiated by other prostate cancer studies that demonstrate its effectivity in inhibiting MMP expression, invasion and migration of LNCaP and DU-145 cells [39]. Thus, treatment with NM can serve

Table 1. Inhibition of tumor weight by NM in various cancer cell xenografts.

Cancer cell type	Reduction of tumor weight by NM diet over control diet
Prostate PC-3	47% (p < 0.0001)
Hepatocellular carcinoma	42% (p = 0.09)
Colon HCT-116	63% (p = 0.0002)
Cervical HeLa	59% (p = 0.001)
Lung A549	44% (p = 0.001)

Table 2. Inhibition of cancer cell proliferation upon treatment with NM.

Cancer cell type	% Inhibition of cell growth at 1000 µg/ml NM
Prostate LNCaP	80%
Osteosarcoma U ₂ OS	60%
Liposarcoma SW-872	61%
Breast MDA-MB-231	50%
Osteosarcoma MNNG	10%

as a multipronged approach to target prostate cancer.

The results of the present study showed that supplementation with NM was effective in inhibiting proliferation of prostate cancer cells by inducing apoptosis using a caspase dependent mechanism. The severity of apoptosis showed dose dependence, increasing with increasing concentrations of NM. Interestingly, a comparison of apoptosis induction in normal human dermal fibroblasts and prostate cancer DU-145 cells suggested a decreased apoptotic effect of the NM on normal cells, thereby supporting the hypothesis of preferential inhibition of prostate cancer cells.

Though a validation of the effectivity of the NM for different cancers and a selection of therapeutic dose can only be obtained upon testing the response of each cancer type and dose individually, in the absence of effective therapies for prostate cancer treatment, the novel nutrient formulation offers a good alternative. In contrast to the toxic side effects of chemotherapy, the NM has also proven to be a safe therapeutic agent. In a previous study addressing safety issues, we found that gavaging adult female ODS rats (weighing 250 - 300 g) with the NM (at 30, 90, or 150 mg per day for 7 days) had no adverse effects on either of the vital organs (heart, liver, kidney) or on the associated functional serum enzymes. This indicates that the mixture is safe to use even at these high doses, which far exceed the normal equivalent dosage [40]. In light of the above fact and based on the results in this study, the NM shows promise and offers a good alternative to chemotherapy in the treatment of prostate cancer.

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Conflicts of Interest

The authors have declared that they have no competing interest.

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