

# P53 protein expression and cell viability in irradiated peripheral blood mononuclear cells as bioindicators of radiosensitivity

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## ABSTRACT

Cellular radiosensitivity is directly correlated with the mechanism of DNA repair, in which p53 protein plays a major role. In this context, this study correlated cell death with p53 expression in lymphocytes irradiated *in vitro* with different doses of gamma-radiation. For this, peripheral blood samples were collected from 10 healthy subjects. Each sample was divided in aliquots and, separately, irradiated with doses of 0.5; 2 and 4 Gy. After this, peripheral blood mononuclear cells (PBMCs) were isolated and cultivated during 72 hours in 5% CO<sub>2</sub> at 37°C without mitogen stimulation. The expression of p53 protein was evaluated by flow cytometry. In parallel, cell viability was determined by trypan blue staining. Statistical analysis was performed using analysis of variance (ANOVA), differences were considered as statistically significant when  $p < 0.05$ . The results showed an increase of p53 expression with the absorbed dose, which was proportional to cell death, suggesting that p53 can be used as bioindicator of individual radiosensitivity.

**Keywords:** P53 Protein; Radiosensitivity; Lymphocytes; Flow Cytometry; Cell Viability

## 1. INTRODUCTION

About 50% of all cancer patients in the World undergo radiotherapy at some point of their treatment, being 60% of them treated with curative intent [1]. The invasive characteristic of cancer implies that target volume to be irradiated is generally outlined beyond

the tumor boundaries, which leads to side effects of radiotherapy that are caused by damage of normal tissues. Therefore, in treatments of radioresistant tumors, connective tissue tolerance to irradiation has an important role in the protocol definition as well as for the effectiveness of radiotherapy.

It has been shown that even when submitted to similar protocols, patients differ widely in terms of normal tissue reactions after irradiation [2,3], approximately 80% of these differences cannot be explained by physical factors [4]. Post-irradiation side effects occur as a function of individual radiosensitivity, which is correlated with genetically determined intrinsic differences in the cellular and molecular response to the radio induced damage [4-6]. This feature was firstly emphasized from patients with rare genetic syndromes, related to mutations in genes involved in the detection and repair of DNA damage [7].

Depending on multiple factors, including source and extent of stress, DNA damage leads either to cell cycle arrest in which the cell is given the opportunity to repair damaged DNA or to the complete disposal of the cell by a process so-called apoptosis. Although the mechanisms responsible for this decision are still far from being elucidated, they are not mutually exclusive, being orchestrated by components that are critical for two processes: DNA repair and apoptosis [8-10].

After DNA damage, cellular decision is mediated by the expression of the p53 protein, which can modulate these events in a transcription-dependent and independent manner. Normally, p53 protein can be detected in the cell cytoplasm at small concentration and with a short average lifetime. However, when cells suffer the action of ionizing radiation (IR), an increase of p53 protein expression is observed as well as alterations in

its conformational structure, increasing the protein mean life [11-14].

Recently, Cavalcanti *et al.* observed the rise of p53 protein expression with the absorbed dose in human lymphocytes. The authors suggested the evaluation of this protein, by Flow Cytometry (FC), as biomarker of individual exposure to IR [15]. In the last years, FC has been applied in fast evaluations of antigens located on the cell surface, cytoplasm or nucleus, in a large number of cells in short time. The use of FC to investigate cellular changes after exposure to ionizing radiation may constitute a fast and reliable method for individual monitoring in cases of accidental or suspected exposures to IR [14-16].

In this context, this work was designed to evaluate the expression of p53 protein as possible bioindicator of radiosensitivity based on *in vitro* irradiation of peripheral blood mononuclear cells with different doses of  $^{60}\text{Co}$  gamma source.

## 2. MATERIALS AND METHODS

### 2.1. Sampling and *in Vitro* Irradiation

This work was priory approved by the Pernambuco Hemocenter Ethics Committee (Hemope Foundation, N<sup>o</sup> 046/06). All participants signed an informed consent.

The study group consisted of 5 females (mean age, 29 years; range, 19 - 42) and 5 males (mean age, 30 years; range, 24 - 36). All donors were healthy individuals and known not to have used medication at the moment of blood collection and had never been submitted to sanguineous transfusion and radiotherapy. Samples containing 40 mL of peripheral blood were collected and divided into four aliquots of 10 mL. Three aliquots were irradiated at room temperature using a  $^{60}\text{Co}$  gamma source (Theratron 780—Atomic Energy of Canada Limited) with a dose-rate of 196.67 cGy.min<sup>-1</sup> delivering doses of 0.5, 2 and 4 Gy, respectively. Non-irradiated blood aliquot was used as control.

### 2.2. Cell Culture

After irradiation, peripheral blood mononuclear cells (PBMCs) were isolated by centrifugation (at 400 g for 35 min) using Ficoll Paque Plus (GE Healthcare, USA). PBMCs were washed twice in phosphate-buffered saline (PBS 0.1M, pH 7.2 - 7.4) and resuspended in RPMI-1640 medium supplemented with 10% fetal bovine serum, 2 mM- L- glutamine, 50 mg/L of sulphate gentamicine and 2 mg/L of amphotericin B (Cultilab, Brazil) at the density of  $2 \times 10^6$  cells/ml. Cells were cultured in 96 U bottom well culture plates (TPP, Switzerland) at a density of  $2 \times 10^5$  cells/well for 72 hours in 5% CO<sub>2</sub> at 37°C. To each evaluated parameter, cultures were per-

formed in triplicate.

### 2.3. Cell Viability

Cell viability was scored at time zero and 72 hours after culture using trypan blue dye 0.4% (Sigma-Aldrich, USA) at 1 : 10 dilution (1 part of cells: 9 parts of dye). Viable and non-viable cells numbers were determined using a hemacytometer by optical microscopy. Viability was calculated as follows: (%) cell viability = total viable cells (unstained)/total cells (viable and non-viable cells)  $\times$  100. Each analysis was performed in triplicate.

### 2.4. Flow Cytometry

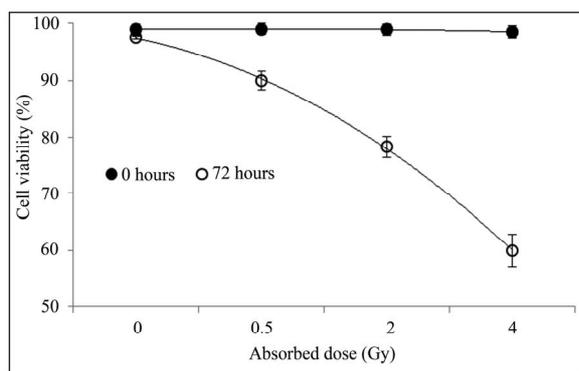
Following 72 hours culture, all cells were harvested and washed with RPMI-1640 medium supplemented with 10% fetal bovine serum and re-suspended in permeabilizing solution—1 mL of facs lysing solution (BD Pharmigen, USA), diluted water (1/10), per 9 mL of 4% paraformaldehyde in PBS—and incubated for 10 min at room temperature. After this period, cells were washed with PBS 0.05% Tween 20 (Merck, Germany) at 400 g for 5 min. Cell pellet was resuspended in 200  $\mu\text{L}$  of PBS 0.05% Tween 20 (Merck, Germany) and stained with PE-conjugated anti-p53 antibody (clone G59-12) (BD Pharmigen, USA) for 20 min at room temperature. The cells were then washed twice in PBS 0.05% Tween 20 (Merck, Germany) and fixed with 500  $\mu\text{L}$  of 1% paraformaldehyde in PBS. PE conjugated mouse IgG1 (clone MOPC-21) (BD Pharmigen, USA) was used as isotype control antibody. Fluorescence intensity of 50,000 events was acquired through FACScalibur flow cytometer (Becton Dickinson, USA), equipped with a 15 mW argon-ion laser (wavelength 488 nm). Data was processed using Cell Quest pro software (Becton Dickinson immunocytometry systems, USA).

### 2.5. Statistical Analysis

For this, it was performed using analysis of variance (ANOVA), differences were considered as statistically significant when  $p < 0.05$ .

## 3. RESULTS AND DISCUSSION

The **Figure 1** shows the percentage of viable PBMCs in non-irradiated and irradiated blood samples, emphasizing that the viability of PBMCs determined right after treatment with different absorbed doses did not differ from the total number of viable cells in non-irradiated cultures, suggesting that cell death induced by gamma radiation does not occur immediately. However, a decrease in cell viability was found in irradiated cells after 72 hours of culture, and it was directly proportional to



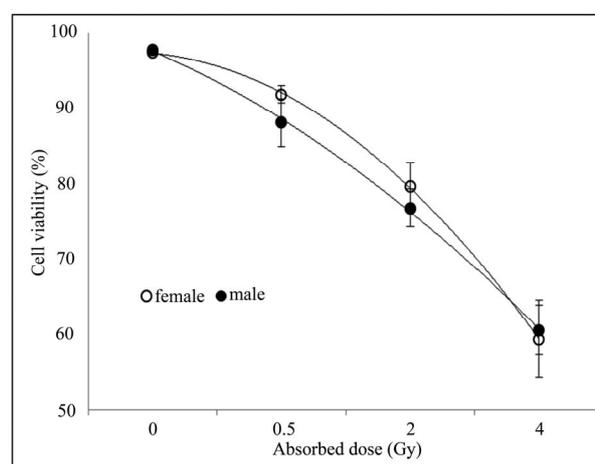
**Figure 1.** Percentage of viable cells in non-irradiated and irradiated PBMCs. Blood sample was divided into four aliquots of 10 mL each and irradiated with 0.5; 2; and 4 Gy; the remaining blood was used as non-irradiated control. Cellular viability was determined by trypan blue staining right after irradiation and 72 h after culture.

the absorbed dose. According to Boreham *et al.*, the ionizing radiation induces apoptosis in lymphocytes, which increases with time after exposure up to 72 hours [17]. The decline in PBMCs viability after irradiation occurred probably due to cell death by apoptosis, as the latter is the dominant mode of radiation-induced cell-killing mechanism in human lymphocytes [18,19], although we did not investigate apoptosis in this study. Trypan blue dye only stains cells that are already dead with plasmatic membrane damage (either by necrosis or apoptosis). Cells that are still undergoing the early events of death such as apoptosis cannot be identified using Trypan blue dye. Thus, further investigation of early and late phase of apoptosis is necessary to confirm this hypothesis.

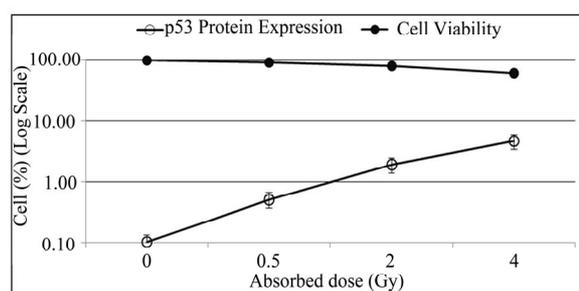
The **Figure 2** contrasts the percentage of viable PBMCs isolated from irradiated and non-irradiated blood samples from males and females subjects after 72 hours of cell culture.

Analyses of variance (ANOVA) of the cell viability in irradiated PBMCs isolated from blood samples of females and males subjects presented in the **Figure 2**, demonstrated that there are no statistically significant differences between the studied groups at the 99.5% confidence level, indicating that sensitivity of PBMCs to gamma radiation doesn't vary with gender. Borgmann *et al.* observed in literature remarkable differences in toxicity profiles after radiotherapy on clinical, molecular, and cellular levels between male and female [20]. However, further studies are required and should be focused on the aspect of individual radiosensitivity.

The **Figure 3** presents the perceptual of cell viability and the expression of p53 protein in non-irradiated and irradiated lymphocytes (with doses: 0.5, 2 and 4 Gy).



**Figure 2.** No difference was observed in cell viability of non-irradiated and irradiated PBMCs from female and male subjects. Blood sample was irradiated with 0.5; 2; and 4 Gy and the remaining was used as non-irradiated control. Cellular viability was determined by trypan blue staining right after irradiation and 72 h after culture.



**Figure 3.** Expression of p53 protein in lymphocytes increases with absorbed dose and the cell viability decrease. PBMCs were isolated from irradiated and non-irradiated human peripheral blood and p53 expression was determined by flow cytometry 72 h after culture.

In this study, an increase in the levels of p53 protein expression was observed in irradiated PBMCs after 72 hours in culture with no stimulation, which is not found in non-irradiated cultures (**Figure 3**). The increase of p53 expression was proportional to the absorbed dose and inversely proportional to the number of viable cells. In normal tissues, p53 plays a major role in maintaining genomic integrity. In response to radiation-induced DNA damage and other types of stress, p53 is stabilized, ensuring that cells carrying genomic damage are effectively eliminated. Activation of p53 gives rise to cell cycle arrest and apoptosis. When the damage is too extensive and cannot be repaired, apoptosis takes place [21]. Previous studies had correlated apoptotic response with the amount of p53 that the cell can accumulate as a result of radiosensitive stress, a high level of p53 mRNA was detected [13,22-24].

According to Dainiak, p53 is highly expressed in radiation-sensitive tissues such as the hematopoietic system [25]. MacCallum *et al.* studied p53 protein distribution among mouse tissues, and showed that radiation sensitive tissues expressed high levels of p53 protein (spleen, thymus, bone marrow, ependyma, intestine), while tissues that were resistant to radiation, such as skeletal muscle and brain, expressed very low levels of p53 protein [26]. Up to today, the sensitivity of normal tissue is not investigated prior to radiotherapy. Generally, the principal parameters for such treatments against cancer are tumor localization and volume as well as the disease staging. Individual radiosensitivity aspects are not considered, although it is important for the effectiveness of radiotherapy [1,27].

Despite its importance, p53 protein is not the only factor that determines radiosensitivity. Rare radiosensitivity syndromes which have genes involved in the detection of DNA damage or DNA repair (e.g. Ataxia-telangiectasia, Nijmegen breakage, Fanconi's anemia and Bloom syndrome) or excessive intoxication (e.g. alcohol and/or nicotine) can increase the sensitivity to ionizing radiation in patients undergoing radiotherapy [1,28]. Healthy individuals without genetic disorders also present adverse effects after irradiation of normal tissues probably, due to intrinsic genetic factors that determine the cellular response to the damage produced by IRs, in particular to the DNA molecule.

If an individual radiosensitivity could be predicted, side effects would be minimized after radiotherapy. In this scenario, the choice of the reference tissue for individual radiosensitivity is very important. Studies have suggested that the relative radiosensitivity of different types of cells from the same individual is similar, reflecting a common determinant of radiosensitivity [29-33]. Thus, on the basis of this assumption, one may expect that a shift in the level of p53 protein expression related to the radiosensitivity of lymphocytes could be similarly reflected in any other cells of the same subject.

#### 4. CONCLUSIONS

In this study, the rate of individual cell death assessed in irradiated PBMCs 72 h after culture *in vitro* without stimulation was proportional to the levels of p53 expression in lymphocytes and inversely proportional to the number of viable cells. The preliminary results presented in this work stressed the increase of p53 protein expression in lymphocytes with radiation doses. The use of p53 as a bioindicator to predict individual radiosensitivity could certainly improve the effectiveness of radiotherapy by personalizing treatment. However, further studies are needed, with a greater number of subjects and, as second step, clinical follow-up is necessary in order to compare

previous radiotherapy analysis of individual p53 expression levels with post-irradiation side effects.

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