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The Nephrotoxic Impact of Oseltamivir in Male Albino Rats after Repeated Exposure

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Abstract

The effects of oseltamivir administration, effectively against influenza viruses A and B, on some selected parameters of rat kidney function were investigated to evaluate its possible nephrotoxic effects. Eighteen (18) albino male Wistar rats with body weights ranging from 150 - 190 g were divided into three groups, the first group (T1) was treated orally with 1 mg/kg BW as a therapeutic dose of oseltamivir for 7 consecutive days. The second group (T2) was treated with the same dose for six weeks, while the control group was dosed with distilled water. The results revealed a significant increase (P < 0.05) in blood urea nitrogen (BUN), serum uric acid and serum creatinine in group T2, while the T1 group showed a significant increase (P < 0.05) in serum uric acid only. Kidney histopathological lesions in the T1 rats showed atrophy of the glomerular tufts, dilation of the Bowman's space, deposition of hyaline droplets within the lumen of the proximal and distal convoluted renal tubules, a cloudy swelling of the epithelial cells lining the renal tubules with perivascular lymphocytic cuffing and severe congestion of the interstitial renal blood vessels. The kidneys of the T2 rats showed necrosis of the epithelial cells lining the renal tubules and cystic dilation with complete dissolution of their epithelial linings. There was vacuolation of the glomerular tuft with proliferation of the parietal layer of the Bowman's capsule, and a serum protein presence in the glomerular space. Mild interstitial fibrosis and thickening of the Bowman's capsule were also observed. Similarly, there was fibrous thickening of the kidney capsule with mild medullary interstitial fibrosis. In conclusion, oseltamivir administered in repeated doses to rats induced some deleterious nephrotoxic effects in a time-dependent manner.

Keywords

Olseltamivir, Nephrotoxic, Albino Rats, Repeated Exposure

1. Introduction

Oseltamivir is an ester prodrug and a potent, selective inhibitor of neuraminidase that results in an inhibition of the release of the influenza virus from host cells and an inhibition of the growth of the influenza virus. Oseltamivir phosphate is used for the treatment of influenza virus infections [1]. It is used in the treatment and prophylaxis of both Influenza virus A and Influenza virus B [2]. Oseltamivir is the most widely used drug but is currently only available as an oral formulation and is eliminated primarily by tubular secretion [3]. The drug is readily absorbed and is converted by hepatic esterase to its active form, oseltamivir carboxylate (OC). The drug is excreted primarily by renal excretion with 60% - 70% of an oral dose appearing in the urine as an active metabolite; therefore, dosing must be adjusted in patients with renal impairments [4] [5]. The OC elimination halflife is approximately 6 to 8 h in adults and up to 13 h in children and 36 h in patients with renal failure [6]. The concentrations of a single dose of 14C radiolabeled [2-acetyl-14C]oseltamivir at 10 mg/kg in rats were widely distributed, with the highest tissue concentrations being observed in the small intestine, liver, stomach, and kidneys 1 h after dosing. The tissue radioactivity concentrations, with the exception of the large intestine, were lower at 6 h and were still detectable 24 h after dosing [7]. The kidney is highly susceptible to toxicants for two reasons: 1) a high volume of blood flows through it and 2) it filters large amounts of toxins, which can concentrate in the kidney tubules resulting in systemic toxicity causing a decreased ability to excrete bodily wastes [8]. This study was conducted to evaluate the possible nephrotoxic effects caused by the apeutic and prophylactic dose regimens of Oseltamivir available pharmaceutically as fluffy® produced by Julphar, Gulf Pharmaceutical Industries, Ras Al-Khaimah, UAE and imported by the medical authorities in Iraq. This study focused on some selected parameters of kidney function supported by a histopathological study.

2. Material and Methods

2.1. The Drugs

- 1) Oseltamivir (Flufly)[®] capsule 75 mg produced by *Julphar* Gulf Pharmaceutical Indestries, Ras A1-Khaimah, UAE.
- 2) Ketamine HCl (Ketamin10%)[®] obtained from KEPRO-HOLAND and xylazine HCl (XYLAZIN 2%) obtained from CEVA-GERMANY were used for animal anesthetization for blood collection.

Preparation of oseltamivir solution was achieved by dissolving one capsule (75 mg) in 75 milliliters of distilled water to achieve a concentration of 1 mg/ml

2.2. Oseltamivir Dosage and Dose

Each rat was dosed with 0.1 ml/100 gm of body weight of the drug solution, which contained 0.1 mg of oseltamivir at the therapeutic dose of 1 mg/Kg BW.

2.3. The Animals

Male albino Wistar rats with a body weight that ranged from 150 - 190 gm were procured from the animal house of the College of Veterinary Medicine, University of Baghdad and were maintained in an air-conditioned room $(25^{\circ}\text{C} \pm 1^{\circ}\text{C})$ with a 12 hours light: 12 hours dark cycle. Standard pellet diet and water were provided *ad libitum* [9].

2.4. Experimental Design

This work was performed with the approval of the College of Veterinary Medicine/Baghdad University in accordance with international ethical standards of research for work with laboratory animals.

2.5. Clinical Study

Eighteen male rats were divided equally into three groups (T1, T2 and control) and were subjected to a blood biochemical analysis. The two treatment groups T1 and T2 were dosed orally by gastric gavage at a dose of 1 mg/Kg BW for one and six weeks, respectively, while the control group was dosed with distilled water. Blood was collected from the animals in all the groups by heart puncture after anesthesia with 0.1 ml/100 gm BW of ketamine 10% and 0.1 ml/100 gm BW of xylazine 2% post treatment. The serum was obtained from all the

blood samples by centrifugation at 5000 r per minute for five minutes.

2.6. Histopathology

The animals in all the groups were sacrificed at the end of the dosing period. The kidneys were obtained and preserved in 10% formalin, then sent to the MEDICINAL CITY HOSPITAL laboratory for histopathology processing as previously described [10].

2.7. Blood Biochemical Analysis

Serum uric acid was assessed using a commercially available kit (Bio System-SPAIN). The principle of this method depends on the coupled quinonemine reaction results that are measured by spectrophotometry [11].

Blood urea nitrogen (BUN) was assessed using a commercially available kit (RANDOX-UK). The modified urease—Bertheholt method was used, and is based on the formation of a green complex of 2 - 2 dicarboxylindophenol [12].

Serum creatinine was assessed using a commercially available kit (BioSystem-SPAIN). The method depends on the reaction of serum creatinine with picrate in an alkaline medium, forming a colored complex, which is then measured by spectrophotometry [13].

Statistical analysis was performed using SPSS version 13.00. A one-way ANOVA was used for the assessment of the differences between the groups. A P-value < 0.05 was considered statistically significant. LSD multiple range tests were performed for comparisons between the means [14].

3. Results

3.1. Uric Acid

There were significant increases (P < 0.05) in the serum uric acid of the rats that had been given the therapeutic dose of 1 mg/kg BW for one week (4.56 ± 0.20 mg/dL) and six weeks (5.43 ± 0.34 mg/dL) in comparison with the serum uric acid of rats in the control group, which was (2.16 ± 0.09) mg/dL, as shown in Table 1.

3.2. Blood Urea Nitrogen (BUN)

There was a significant decrease (P < 0.05) in the blood urea nitrogen (BUN) of the rats given a therapeutic dose of oseltamivir at 1 mg/kg BW for six weeks recorded at 25.33 ± 2.24 mg/dL in comparison with 59.33 ± 2.70 mg/dL for the same therapeutic dose of oseltamivir given for one week; there was also a significant difference between T1 and the control group at 57.34 ± 3.73 mg/dL, see Table 2.

3.3. Serum Creatinine

There was a significant increase (P < 0.05) in serum creatinine in the rats dosed with oseltamivir at 1 mg/kg BW for six weeks (1.31 \pm 0.10 mg/dL) in comparison with the rats given the same therapeutic dose for one week (0.55 \pm 0.04 mg/dL) and the rats in the control group (0.60 \pm 0.10 mg/dL). No significant differences were observed between T1 and the control group, see **Table 3**.

3.4. Histopathology

The kidneys of the rats given the therapeutic dose of 1 m/kg BW of oseltamivir for one week showed atrophy of the glomerular tufts with dilation of the Bowman's space and deposition of hyaline droplets within the lumen of the proximal and distal convoluted renal tubules (**Figure 1**). There was also a cloudy swelling of the epithelial cells lining the renal tubules with perivascular lymphocytic cuffing (**Figure 2**). In addition, there was severe congestion of the interstitial renal blood vessels; many of them were dilated and contained large numbers of neutrophils.

The kidneys of rats given the therapeutic dose of 1 m/kg BW of oseltamivir for six weeks showed massive necrosis of the epithelial cells lining the renal tubules. Many of the renal tubules showed cystic dilation with a complete dissolution of their epithelial linings (**Figure 3**). Additionally, there was vacuolation of the glomerular tufts with proliferation of the parietal layer of the Bowman's capsule and a serum protein presence in the

Table 1. Serum uric acid in mg/dL in rats dosed orally with the therapeutic dose of 1 mg/kg BW of oseltamivir for different periods.

Cuona	Serum uric acid mg/dL			
Group	M	±	SE	
T1	4.56		0.20	A
T2	5.43		0.34	В
C	2.16		0.09	C

T1 = therapeutic dose of 1 mg/kg BW of oseltamivir for one week. T2 = therapeutic dose of 1 mg/kg BW of oseltamivir for six weeks. C = control dosed with distilled water. The capital letters denote significant differences between the groups at the P < 0.05 level.

Table 2. Blood urea nitrogen (BUN) in mg/dL of the rats given a therapeutic dose of oseltamivir orally at 1 mg/kg BW for different periods.

Canana	Seru				
Group	M	±	SE		
T1	59.33		2.70	A	
T2	25.33		2.24	В	
C	57.34		3.73	A	

T1 = therapeutic dose of 1 mg/kg BW of oseltamivir for one week. T2 = therapeutic dose of 1 mg/kg BW of oseltamivir for six weeks. C = control dosed with distilled water. The capital letters denote significant differences between the groups at the P < 0.05 level.

Table 3. Serum creatinine in mg/dL of the rats given the therapeutic dose of 1 mg/kg BW of oseltamivir orally for different periods.

Group	Serun	Serum creatinine mg/dL			
	M	±	SE		
T1	0.55		0.04	A	
T2	1.31		0.10	В	
C	0.60		0.10	A	

T1 = therapeutic dose of 1 mg/kg BW of oseltamivir for one week. T2 = therapeutic dose of 1 mg/kg BW of oseltamivir for six weeks. C = control dosed with distilled water. The capital letters denote significant differences between the groups at the P < 0.05 level.

glomerular space. Mild interstitial fibrosis leading to thickening of the Bowman's capsule was also observed (**Figure 4**). Similarly, there was a fibrous thickening of the kidney capsule that was infiltrated with chronic inflammatory cells. There was also mild medullary interstitial fibrosis (**Figure 5**).

4. Discussion

Biochemical markers play an important role in accurate diagnoses, assessing risks and adopting therapies that improve clinical outcomes. As markers of renal function, serum creatinine, blood urea nitrogen (BUN) and uric acid are routinely used for analysis [15]. Elevated uric acid is one of the most important risk factors for cardio-vascular disease; it plays a significant role in the development of renal disease and metabolic syndrome, and high serum urate levels are usually associated with defects in uric acid transport [16]. An elevated uric acid level often precedes kidney disease [17]. The repeated dosing of oseltamivir at 1 mg/kg BW throughout the period of the present study resulted in distinct pathological changes in the kidneys of the rats in both treated groups and was shown in Figures 1-5.

Urea is a major nitrogenous end product of the protein and amino acid catabolism produced by the liver and is distributed throughout the intracellular and extracellular fluid [15]. Low blood urea levels are seen in trauma and surgery and are associated with the use of some drugs, anabolic steroid use and malnutrition [18]. The significant reduction in the blood urea nitrogen concentration throughout the experimental period may be attributed to impairment of the urea cycle (in the liver) leading to the reduced production of metabolic products [19]. This is an indication of an abnormality in the physiological excretion of urea caused by a non-renal factor [20]. We considered that the reduction in the blood urea nitrogen level was due to the hepatotoxic effects of oseltamivir



Figure 1. The kidney of a male rat dosed with 1 mg/kg BW of oseltamivir for one week and showing atrophy of the glomerular tufts with the dilation of the Bowman's space (black arrow) and with the deposition of hyaline droplets in the lumina of the proximal and distal convoluted tubules (red arrow) (H&E 100×).

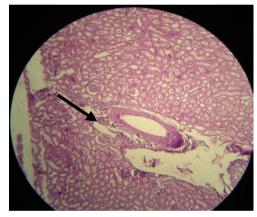


Figure 2. The kidney of a male rat dosed with 1 mg/kg BW of oseltamivir for one week and showing perivascular lymphocytic cuffing (black arrow) (H&E 100×).

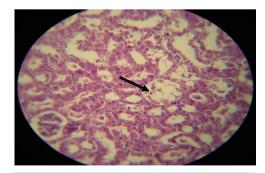


Figure 3. The kidney of a male rat dosed with 1 mg/kg BW of oseltamivir for six weeks showing cystic dilation of the cortical renal tubules with complete dissolution of their epithelial lining (black arrow) (H&E 100×).



Figure 4. The kidney of a male rat dosed with 1 mg/kg BW of oseltamivir for six weeks showing interstitial fibrosis leading to thickening of the Bowmans capsule (black arrow) (H&E 100×).

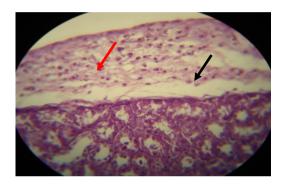


Figure 5. The kidney of a male rat dosed with 1 mg/kg BW of oseltamivir for six weeks showing fibrous thickening of the capsule (black arrow) with the infiltration of chronic inflammatory cells (red arrow) (H&E 100×).

dosed at 1 mg/kg BW for six weeks. We reached this conclusion by histopathological analysis and other liver function tests including alaninaminotrasferase (ALT), aspartate aminotrasferase (AST), alkaline phosphatase (AP), total bilirubin, conjugated and non-conjugated bilirubin and total serum protein [21]. The liver is the site of the urea cycle. However, the effects of the drug were not obvious at one week of dosing because the BUN level in the T1 group was similar to that in the control group.

Creatinine is commonly used as a measure of kidney function, and the calculation of the glomerular filtration rate may be derived from the serum creatinine level [22]. A diagnosis of renal failure is usually suspected when the serum creatinine is greater than the upper limit of the normal interval; in chronic renal failure and uremia, an eventual reduction in the excretion of creatinine by both glomeruli and tubules occurs [23]. These events are confirmed by histopathological findings in the kidneys of the rats treated with 1 mg/kg BW of oseltamivir for six weeks, which included cystic dilation of the cortical renal tubules with complete dissolution of their epithelial lining (Figure 3) and thickening of the Bowman's capsule (Figure 4).

Oseltamivir induced kidney injury in rats treated with 0.1 mg/kg BW for one and six weeks through the increased level of uric acid (**Table 1**). In other studies in rats, experimental hyperuremia (induced by oxonic acid) has also been associated with the development of mild renal disease, characterized by mild proteinuria, renal arteriolar changes, glomerular hypertrophy, tubulointerstitial fibrosis, and eventually glomerulosclerosis [24]. Interestingly, when hyperuremia was induced in rats with preexisting renal disease (*i.e.*, in which one entire kidney and two thirds of the other kidney had been removed), their renal lesions were dramatically worse than in similar rats without hyperuremia [25]. This suggests that the hyperuremia may not only cause renal disease but also exacerbate preexisting renal disease. The mechanism by which uric acid might cause renal disease was revealed by micropuncture studies that demonstrated that elevated uric acid $(3.1 \pm 0.2 \text{ mg/dL})$ caused glomerular

hypertension and cortical vasoconstriction [26]. These changes would be expected to induce glomerular damage and tubular ischemia. In addition, uric acid stimulated inflammatory mediators in vascularcells, including C-reactive protein and monocyte chemoattractant protein-1p [27] [28] and vasoconstrictive factors such as thromboxane [25]. After oral administration, oseltamivir is rapidly hydrolyzed to its active metabolite oseltamivir carboxylate (OC), which is then excreted by glomerular filtration and renal tubular secretion without further metabolism [29]. When rats are unable to hydrolyze the oseltamivir phosphate sufficiently to its metabolite, an excessive amount of phosphate will accumulate. This would negatively affect the dietary calcium/phosphate ratio in species known to be sensitive to this type of change and would consequently lead to mineralization of the kidney [30].

The results of this study suggest that the rats that had been exposed repeatedly to 1 mg/kg BW of oseltamivir experienced several deleterious effects on kidney function depending on the period of dosing, as shown by serum biochemical markers. Oseltamivir, when given at low therapeutic doses for prophylaxis (six weeks) caused renal damage; therefore, kidney and liver function tests must be performed during OM prophylaxis therapy.

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