Published Online May 2014 in SciRes. <a href="http://www.scirp.org/journal/ojss">http://www.scirp.org/journal/ojss</a> <a href="http://dx.doi.org/10.4236/ojss.2014.45019">http://dx.doi.org/10.4236/ojss.2014.45019</a>



# Calibration of CO<sub>2</sub> Trapping in Alkaline Solutions during Soil Incubation at Varying Temperatures Using a Respicond VI

Natalya Smirnova<sup>1,2</sup>, Michael Scott Demyan<sup>3</sup>, Frank Rasche<sup>3</sup>, Georg Cadisch<sup>3</sup>, Torsten Müller<sup>1\*</sup>

Email: \*torsten.mueller@uni-hohenheim.de

Received 17 March 2014; revised 17 April 2014; accepted 24 April 2014

Copyright © 2014 by authors and Scientific Research Publishing Inc.
This work is licensed under the Creative Commons Attribution International License (CC BY). http://creativecommons.org/licenses/by/4.0/



Open Access

#### **Abstract**

Measurements of carbon dioxide ( $CO_2$ )-evolution from soils are important in evaluating biomass and activity of soil microorganisms, as well as decomposition of soil organic matter. The Respicond VI is a fully computerized system allowing continuous measurement of  $CO_2$  evolution in short- and long-term soil incubation experiments in up to 96 incubation vessels. The measurement of  $CO_2$  evolution is based on the absorption of  $CO_2$  by an electrolyte (KOH solution) producing a change in the cell conductance measured using two electrodes. In this study, the Respicond VI was recalibrated yielding 174.5 mg  $CO_2$  as constant A expressing the theoretical maximum amount of  $CO_2$  absorbed in 10 ml 0.5 M KOH. This value of A corresponds to 34.9 mg  $CO_2$  ml<sup>-1</sup> 1 M KOH. The constant A does neither depend on the investigated incubation temperatures (5°C - 25°C) nor on the concentrations of the KOH solutions (0.5, 0.1, 0.05 M KOH). To eliminate any influence of changing incubation temperatures, either induced by uncertainties in temperature control or as a part of the experimental setup, on the conductance of KOH solution, a correction procedure was developed using a factor calculated from changing conductance of KOH solutions in incubation vessels without soil.

#### **Keywords**

Respirometer, Soil Respiration, Carbon Dioxide, Incubation

<sup>&</sup>lt;sup>1</sup>Institute of Crop Science (340i), University of Hohenheim, Stuttgart, Germany

<sup>&</sup>lt;sup>2</sup>Institute of Soil Science and Agrochemistry, Siberian Branch Russian Academy of Science, Novosibirsk, Russia <sup>3</sup>Institute of Plant Production and Agroecology in the Tropics and Subtropics (380a), University of Hohenheim, Stuttgart, Germany

<sup>\*</sup>Corresponding author.

## 1. Introduction

Measurements of carbon dioxide (CO<sub>2</sub>)-evolution from soils are important in evaluating biomass and activity of soil microorganisms, as well as decomposition of soil organic matter. Several methods have been developed for the determination of soil CO<sub>2</sub>-evolution, both under laboratory and field conditions [1]-[6] proposed a conductimetric method for measuring soil CO<sub>2</sub>-evolution under field and laboratory conditions. According to [5], the absorption of CO<sub>2</sub> by an electrolyte, such as potassium hydroxide (KOH), produces a change in the conductance of the solution. [7] improved this principle to a fully computerized sampling system (Respicond VI) allowing continuous measurements both in short- and long-term (less than one day up to several weeks) soil incubation experiments. However, the application of this method was limited to incubations at constant temperatures [8]-[10]. This is in contrast to natural field conditions with diurnal and weather determined fluctuations of soil temperatures. The objective of the present investigation was to develop a method and a calibration procedure for the Respicond VI allowing measurements of soil CO<sub>2</sub>-evolution under changing temperatures. The proposed temperature correction procedure will completely eliminate the influence of the temperature on the conductance during long-time incubation.

#### 2. Materials and Methods

## 2.1. Principles of the Respicond

The main principle of the Respicond respirometer has been described previously [7]. Briefly, it consists of water baths with inserted hermetically closed vessels for soil incubation. Each vessel includes a smaller open vessel with KOH solution and platinum electrodes inserted into the solution. Each set of electrodes is connected to a digital volt-/amperemeter and a source of constant current (**Figure 1**). The CO<sub>2</sub> released from the soil sample is absorbed by the KOH solution. The amount of absorbed CO<sub>2</sub> is determined by measuring changes in conductivity of the KOH solution. Data from each vessel are collected in hourly increments and analyzed using the software supplied with the Respicond VI. The chemical reaction of the absorption of CO<sub>2</sub> by the KOH solution is

$$KOH + CO_2 \rightarrow KHCO_3$$
  
 $KHCO_3 + KOH \rightarrow K_2CO_3 + H_2O$ 

The concentration of ions in solution decreases linearly in relation to the amount of absorbed  $CO_2$ . The amount of accumulated  $CO_2$  during incubation  $N(CO_2)$  can be calculated using the following equation (reformulated from [7]):

$$N(\text{CO}_2) = A \times \frac{C(t_0) - C(t_1)}{C(t_0)}$$
 (1)

where  $C(t_0)$  was the conductance at the beginning of the incubation (time  $t_0$ ) before any absorption of  $CO_2$  by KOH,  $C(t_1)$  was the conductance at time  $t_1$  and A was an empirically determined constant expressing the theoretical maximum amount of  $CO_2$  absorbed and it can be calculated as  $A = N(CO_2)$  if  $C(t_1) = 0$ . This calculation assumes that the temperature of the solution is constant during the experiment. The constant A is proportional to the volume and concentration of the solution. The default value of the constant A used by the software of the Respicond VI is 219 mg  $CO_2$  for 10 ml of 0.6 M KOH solution, corresponding to 36.5 mg  $CO_2$  ml<sup>-1</sup> 1 M KOH. Another published value for constant A is 9.94 mg  $CO_2$  for 5 ml of 0.05 M KOH solution [5], corresponding to 39.8 mg  $CO_2$  ml<sup>-1</sup> 1 M KOH. No further calibrations for other concentrations of KOH are available, although, a reliable detection of low  $CO_2$  evolution rates (<100  $\mu$ g  $CO_2$  h<sup>-1</sup>) is possible only with lower concentrations of KOH solution [7]. To meet the objective of this study, it was necessary to re-determine the value of the constant A for suitable concentrations of KOH, its dependence on the temperature and finally to develop a method to perform measurements under variable temperatures to allow for variations in experimental conditions or induced temperature treatments.

#### 2.2. Calibration

The determination of the constant A can be done using two different approaches [5] [6]:

1. Production of a defined amount of CO<sub>2</sub> in a closed vessel following the reaction

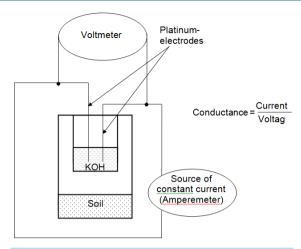


Figure 1. Principle design of a Respicond VI incubation vessel and the measuring unit.

$$2\text{NaCl} + \text{Na}_2\text{CO}_3 \rightarrow 2\text{NaCl} + \text{CO}_2 \uparrow + \text{H}_2\text{O}$$

The emitted CO<sub>2</sub> is absorbed by the KOH solution, resulting in a changing conductance of the KOH solution.

2. Replacing the KOH solution by mixtures with different ratios of KOH and potassium carbonate (K<sub>2</sub>CO<sub>3</sub>). According to Reaction 1, this imitates the absorption of defined amounts of CO<sub>2</sub>.

To apply the first calibration method, 10 ml 1.0 M hydrochloric acid (HCl) were added to empty vessels. After closing the vessels, sodium carbonate (Na<sub>2</sub>CO<sub>3</sub>) solution was injected through a septum into the HCl. Twelve injections were made, each of which with 1 ml 0.2 M Na<sub>2</sub>CO<sub>3</sub> producing 8.8 mg CO<sub>2</sub>. After each injection, the conductance of the KOH solution (10 ml, originally 0.5 M) was measured as soon as it stabilized (approx. 30 minutes). The whole experiment was repeated five times at 20°C.

The constant A was calculated from the linear relationship between the  $CO_2$  evolved and expected to be absorbed by KOH ( $N(CO_2)$ ), and the measured conductance after injection  $C(t_{ini})$ :

$$A = N(CO_2) \frac{C(t_0)}{C(t_0) - C(t_{ini})}$$
(2)

where  $C(t_0)$  and  $C(t_{inj})$  were the conductance of KOH solution before and after injection, respectively.

For the second calibration method, it is assumed that the absorption of one mole of  $CO_2$  converts two moles of KOH to one mole of  $K_2CO_3$  and one mole of  $H_2O$  (see reactions above). The absorption of  $CO_2$  could be simulated by replacing KOH with  $CO_2$ -equivalent amounts of  $K_2CO_3$  solution. We mixed 0.5 M, 0.1 M and 0.05 M KOH with 0.25 M, 0.05 M and 0.025 M  $K_2CO_3$ , respectively, each of which at 6 different ratios of KOH to  $K_2CO_3$ : 0/10, 2/8, 3/7, 5/5, 7/3, 9/1 at 20°C. Each combination of concentrations and ratios was replicated six to seven times.

To investigate the influence of temperature on the calibration of the constant A, the experiment with 0.5 M KOH and 0.25 M  $K_2CO_3$  was conducted again at six different temperatures (5°C, 10°C, 12°C, 15°C, 20°C and 25°C) with three experimental replicates for each temperature. According to Equation 2, constant A was calculated from the linear relationship between the equivalent amount of  $CO_2(Eq(CO_2))$ , and the corresponding measured conductance  $C_{K_2CO_2}$ :

$$A = Eq\left(\text{CO}_2\right) \frac{C_{\text{KOH}}}{C_{\text{KOH}} - C_{\text{K}_2\text{CO}_3}} \tag{3}$$

where  $C_{\text{KOH}}$  was the conductivity of the original KOH solution.

To detect a possible temperature dependency of the constant A, measurements at 6 temperatures from 5°C to 25°C were fitted to the following linear function:

$$A(T) = A_{20} + B(T - T_{20}) \tag{4}$$

where  $T_{20} = 20^{\circ}\text{C}$ , T is the actual temperature,  $A_{20}$  is the value of the constant A (mg CO<sub>2</sub>) at 20°C and B is the regression coefficient indicating temperature dependency.

Linear regressions and corresponding coefficients of determination were calculated. An analysis of variance was performed to identify significant differences between calibrations with different molarity of the KOH solution.

#### 2.3. Correction for Changing Incubation Temperatures

The conductance of the KOH solution is positively correlated with temperature. Equation (1) assumes that the temperature is constant during incubation. Thus, the conductance must be recalculated if the temperature changes. During incubation at a constant temperature (ideal condition), the conductance in vessels without soil does not change and any change in conductance measured in these control vessels is only related to changes in temperature during incubation. Fluctuations of conductance in vessels without soil can, thus, be used to calculate a correction factor  $(F_{\text{corr}}(t_0,t_1))$  for changes in temperature between times  $t_0$  and  $t_1$ :

$$F_{\text{corr}}\left(\mathbf{t}_{0}, \mathbf{t}_{1}\right) = \frac{C_{\text{empty}}\left(\mathbf{t}_{0}\right)}{C_{\text{empty}}\left(\mathbf{t}_{1}\right)} \tag{5}$$

where  $C_{\rm empty}(t_0)$  and  $C_{\rm empty}(t_1)$  are the conductances measured in the vessel without soil at time  $t_0$  and  $t_1$ , respectively. By assuming equal temperatures in vessels with and without soil, the temperature corrected conductance  $C_{\rm corr}(t_1)$  in the vessels with soil can be calculated for  $t_1$  using the following equation:

$$C_{\text{corr}}\left(\mathbf{t}_{1}\right) = C\left(\mathbf{t}_{1}\right) \cdot F_{\text{corr}}\left(\mathbf{t}_{0}, \mathbf{t}_{1}\right) \tag{6}$$

To verify this method, we performed the following experiment. Soils were placed into plastic vessels with hermetic lids and were pre-incubated at  $20^{\circ}$ C for 24 hours before starting the experiment. The temperature scheme of the incubation experiment in the Respicond VI water bath was ten days at  $20^{\circ}$ C, eight days at  $25^{\circ}$ C and another four days at  $20^{\circ}$ C. Two vessels without soil were used to calculate the correction factor  $F_{\text{corr}}(t_0,t_1)$ . Then, this correction factor was used to correct the conductance in vessels with soil. Five soil samples (0 - 60 cm) from two experimental sites in the state of Baden Württemberg (Southwest Germany) were used: one sample from a Calcaric Cambisol (silty loam, 0.7% organic C, pH(CaCl<sub>2</sub>) 6.4) and four samples from a Rendzic Leptosol (clay loam, 1.9% organic C, pH(CaCl<sub>2</sub>) 7.9). Soil samples were sieved (2 mm) and adjusted to water content of about 50% maximum water holding capacity. Three replicates were performed for each soil sample resulting in a total of 15 vessels.

#### 3. Results and Discussion

A nearly perfect linear relationship between the relative change of conductance and (equivalent) amounts of absorbed  $CO_2$  was found (Figure 2(a) and Figure 2(b)). Calculated values of constant A are shown in Table 1. For the 0.5 M KOH solution, both calibration methods gave nearly identical results ( $A = 173.3 \pm 1.4$  and  $175.8 \pm 1.6$  mg  $CO_2$ ). Since these results were statistically independent, we suggest using the average value A = 174.5 mg  $CO_2$ . This corresponds to 34.9 mg  $CO_2$  ml<sup>-1</sup>1 M KOH and is, thus, lower than the default value used by the Respicond VI software corresponding to 36.5 mg  $CO_2$  ml<sup>-1</sup>1 M KOH (see above). In calibration procedure 2, constants A for 0.1 and 0.05 M KOH corresponded to values of 34.0 and 33.4 mg  $CO_2$  ml<sup>-1</sup>1 M KOH, respectively. These values did not differ significantly from the value 33.4mg  $CO_2$  ml<sup>-1</sup>1 M KOH determined for 0.5 M KOH solution, indicating that the constant A must not be calibrated separately for KOH solutions differing in concentrations, at least within the range we investigated.

The temperature dependence of constant A is shown in Figure 3. Fitted values of  $A_{20}$  and B in Equation (4) were  $173.4 \pm 1.0$  and  $0.26 \pm 0.14$ , respectively. Since B was not significantly different from 0 (p > 0.05), it can be concluded that constant A does not depend on the temperature. The fitted value of  $A_{20}$  was again very close to the above-suggested mean value of constant A.

The method of temperature correction was validated in an incubation experiment with soils under two temperature conditions. The temperature correction factor,  $F_{corr}(20^{\circ}\text{C}, 25^{\circ}\text{C})$  was found to be  $1.083 \pm 0.001$ . The proposed temperature correction procedure completely eliminated the influence of the temperature on the conductance (Figure 4).

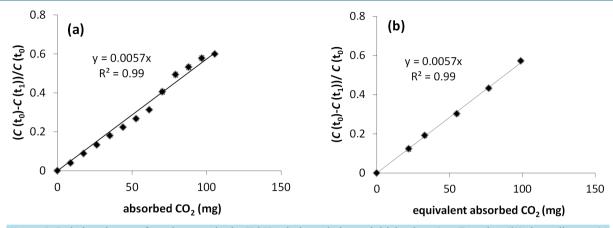


Figure 2. Relative change of conductance in the KOH solution relative to initial values (see Equation (2)) depending on (a) the amount of  $CO_2$  released after injection of  $Na_2CO_3$  into HCl and assumed to be absorbed by the KOH solution and (b) the amount of absorbed  $CO_2$  equivalent to the  $K_2CO_3$  replacing KOH in the solution. Data were fitted to linear functions forced through the origin. Error bars show standard errors ((a) n = 5, (b) n = 6) and are partly hidden by the symbols.

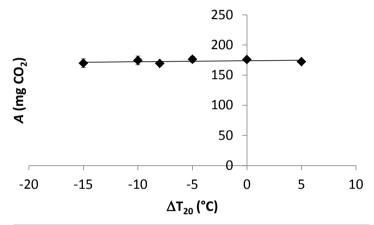
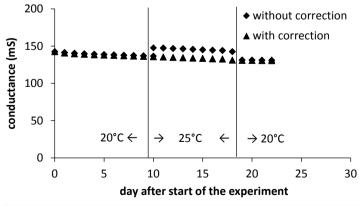


Figure 3. Relationship between the temperature in the water bath as the difference to the standard temperature of  $20^{\circ}\text{C}$  ( $\Delta T_{20} = 20^{\circ}\text{C}$ -actual temperature) in the water bath and the calculated constant A, where 0.5 M potassium hydroxide (KOH) was replaced by 0.25 M potassium carbonate ( $K_2\text{CO}_3$ ) to imitate carbon dioxide absorption. Error bars (standard deviations, n = 3) are partly hidden by the data symbols.



**Figure 4.** Conductance of 0.5 M KOH solution during incubation at 20°C/25°C/20°C with and without calculated temperature correction. Average of 15 vessels.

Table 1. Mean values ( $\pm$  standard deviations) of A calibrated using two different methods: Either production of a defined amount of CO<sub>2</sub> by injection of Na<sub>2</sub>CO<sub>3</sub> into HCl assumed to be absorbed by the KOH solution (Calibration 1) or replacing the KOH solution with mixtures of KOH and K<sub>2</sub>CO<sub>3</sub> in different ratios imitating absorption of definite amounts of CO<sub>2</sub>. Values in brackets show A converted to a KOH concentration of 0.5 M, assuming a linear relation to concentration.

| Molarity of reagent  | N | A (mg CO <sub>2</sub> ) | mg CO <sub>2</sub> ml <sup>-1</sup> 1 M KOH |
|--|---|-------------------------|---|
| Calibration 1 (injection of Na <sub>2</sub> CO <sub>3</sub> )      |   |                         |   |
| 0.5 M KOH  | 5 | $173.3 \pm 1.4$         | $34.7 \pm 0.3$                              |
| Calibration 2 (mixture of KOH and K <sub>2</sub> CO <sub>3</sub> ) |   |                         |   |
| $0.05 \text{ M KOH} + 0.025 \text{ M K}_2\text{CO}_3$              | 7 | $16.7\pm0.6$            | $33.4 \pm 1.2$                              |
| $0.1 \text{ M KOH} + 0.05 \text{ M K}_2\text{CO}_3$                | 6 | $34.0\pm1.0$            | $34.0 \pm 1.0$                              |
| $0.5 \text{ M KOH} + 0.25 \text{ M K}_2\text{CO}_3$                | 6 | $175.8 \pm 1.6$         | $35.2\pm0.3$                                |

## 4. Conclusion

We suggest using 174.5 mg CO<sub>2</sub> as constant A for the conversion of conductance to evolved CO<sub>2</sub> adsorbed in 10 ml 0.5 M KOH during soil incubation in the Respicond VI. This value of A corresponds to 34.9 mg CO<sub>2</sub> ml<sup>-1</sup> 1 M KOH. Within the ranges we investigated, constant A does neither depend on the incubation temperature nor on the concentrations of the KOH solutions. To eliminate the influence of changing temperatures in the Respicond VI during incubation on the conductance of the KOH solution, a correction factor calculated from changes in conductance of the KOH solution in incubation vessels without soil should be used. We recommend using the temperature correction procedure for incubation experiments with temperature instability of 1°C or more and for the measurement of the respiration rate even with stable temperature conditions. Our findings may also be applicable to earlier and later versions of the Respicond respirometer.

## **Acknowledgements**

We thank Anders Nordgren for cooperation and valuable comments, and we acknowledge the kind suggestions by Pavel Krokovny. This research was part of sub-project P3 within PAK 346, "Structure and Functions of Agricultural Landscapes under Global Climate Change-Processes and Projections on a Regional Scale" and FOR 1695 "Agricultural Landscapes under Global Climate Change-Processes and Feedbacks on a Regional Scale" with funding provided by the German Research Foundation (DFG). Natalya Smirnova was a collaborator via the "International Academic Mobility Network with Russia" (IAMONET-RU) as a part of the "Erasmus Mundus External Cooperation Window" funded by the European Commission.

## References

- [1] Anderson, J.P.E., Page, A.L., Miller, R.H. and Keeney, D.R. (1982) Soil Respiration. In: Page, A.L., Ed., *Methods of Soil Analysis*, Part 2, 2nd Edition, ASA and SSSA, Madison, 831-871.
- [2] Cropper, W.P., Ewel, K.C. and Raich, J.W. (1985) The Measurement of Soil CO<sub>2</sub> Evolution *In-Situ. Pedobiologia*, 28, 35-40.
- [3] Sparling, G.P. and West, A.W. (1990) A Comparison of Gas Chromatography and Differential Respirementer Methods to Measure Soil Respiration and to Estimate the Soil Microbial Biomass. *Pedobiologia*, **34**, 103-112.
- [4] Bachelor, P.P., McIntyre, J.I., Amonette, J.E., Hayes, J.C., Milbrath, B.D. and Saripalli, P. (2008) Potential Method for Measurement of CO<sub>2</sub> Leakage from Underground Sequestration Fields Using Radioactive Tracers. *Journal of Radioa-nalytical and Nuclear Chemistry*, 277, 85-89. <a href="http://dx.doi.org/10.1007/s10967-008-0713-8">http://dx.doi.org/10.1007/s10967-008-0713-8</a>
- [5] Chapman, S.B. (1971) A Simple Conductimetric Soil Respirometer for FIELD Use. Oikos, 22, 348-353. http://dx.doi.org/10.2307/3543857
- [6] Anderson, J.M. and Ineson, P. (1982) A Soil Microcosm System and Its Application to Measurements of Respiration and Nutrient Leaching. *Soil Biology and Biochemistry*, **14**, 415-416. <a href="http://dx.doi.org/10.1016/0038-0717(82)90015-3">http://dx.doi.org/10.1016/0038-0717(82)90015-3</a>
- [7] Nordgren, A. (1988.) Apparatus for Continuous, Long-Term Monitoring of Soil Respiration Rate in Large Numbers of Samples. Soil Biology and Biochemistry, 20, 955-957. http://dx.doi.org/10.1016/0038-0717(88)90110-1

- [8] Ilstedt, U., Nordgren, A. and Malmer, A. (2006) Soil Chemical and Microbial Properties after Disturbance by Crawler Tractors in a Malaysian Forest Plantation. Forest Ecology and Management, 225, 313-319. http://dx.doi.org/10.1016/j.foreco.2006.01.008
- [9] Gnankambary, Z., Ilstedt, U., Nyberg, G., Hien, V. and Malmer, A. (2008) Nitrogen and Phosphorus Limitation of Soil Microbial Respiration in Two Tropical Agroforestry Parklands in the South-Sudanese Zone of Burkina Faso: The Effects of Tree Canopy and Fertilization. Soil Biology and Biochemistry, 40, 350-359. http://dx.doi.org/10.1016/j.soilbio.2007.08.015
- [10] Hartley, I.P., Hopkins, D.W., Sommerkorn, M. and Wookey, P.A. (2010) The Response of Organic Matter Mineralisation to Nutrient and Substrate Additions in Sub-Arctic Soils. *Soil Biology and Biochemistry*, 42, 92-100. http://dx.doi.org/10.1016/j.soilbio.2009.10.004