

Rice Husk Biochar Influences Seedling Emergence of Junglerice (*Echinochloa colona*) and Herbicide Efficacy

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ABSTRACT

The use of carbonized rice husk biochar improves the fertility and productivity of poor soils in rice-based cropping systems. However, biochar may also influence weed seedling emergence and the efficacy of soil-applied herbicides. Experiments were conducted in a screenhouse to evaluate the effect of biochar rates (0, 20, 40, and 80 t \cdot ha⁻¹) and seed burial depth (0, 1, and 2 cm) on seedling emergence of junglerice (*Echinochloa colona*) and the effect of biochar rates and pendimethalin (0, 500, 1000, and 1500 g \cdot a.i. \cdot ha⁻¹) and pretilachlor doses (0, 300, 600, and 900 g \cdot a.i. \cdot ha⁻¹) on seedling emergence and seedling biomass of junglerice. Data were analyzed using nonlinear regression. The burial depth to inhibit 50% of maximum seedling emergence was 0.76 cm when biochar was not added to soil and the depth increased with an increase in biochar rates for soil. Similarly, compared with the soil with no biochar, the use of bichoar increased the pretilachlor dose to inhibit 50% of maximum emergence or biomass. The pretilachlor dose to inhibit 50% of maximum biomass of junglerice was 100, 130, 240, and 240 g \cdot ha⁻¹ when biochar was added at 0, 20, 40, and 80 t \cdot ha⁻¹. However, the efficacy of pendimethalin was not influenced by biochar rate. The results of this study suggest that rice husk biochar may increase weed seedling emergence from deeper burial depths and may decrease the efficacy of some soil-applied herbicides.

Keywords: Biochar; Emergence; Efficacy; Biomass; Rice; Asia

1. Introduction

Rice (Oryza sativa L.) is the most important crop in Asia and more than 90% of this crop is grown and consumed in Asia. Rice produces a considerable amount of residue (straw and husk). However, most of these residues are being burned in fields in many Asian regions (e.g., India, Sri Lanka, etc.), causing air pollution, human health problems, and considerable nutrient losses [1]. Furthermore, climate change throughout the world causes concern about rice residue management. Recently, there has been an interest among researchers to use carbonized rice husk, called biochar, as a soil amendment that can act as a carbon sink in agricultural soil and can improve soil fertility and organic matter [1,2]. In addition, biochar at high rates may increase soil water retention capacity [3]. Although biochar can increase soil fertility, it may also influence weed seedling emergence and the efficacy of soil-applied herbicides as the application of biochar decreases soil bulk density and increases organic matter [2, 4,5].

Because of labor and water shortages, direct-seeded rice systems are increasing in many Asian countries [6-9]. In direct-seeded rice systems, weeds are an important biological constraint and pre-emergence (PRE) herbicide application is essential for controlling weeds in these systems [6,7,10]. The use of biochar in such systems may inhibit or stimulate weed seedling emergence. Such information, however, is not available on rice weeds. In addition, the use of biochar may have either positive or negative impacts on weed management [4]. Strong adsorption of herbicides on biochar, for example, may decrease the efficacy of a soil-applied herbicide. Weed seedling emergence and herbicide efficacy may also be influenced by the rates of biochar. Such information is not available in Asia on rice weeds.

A study was conducted to evaluate the effect of biochar (carbonized rice husk) rates on seedling emergence of junglerice [*Echinochloa colona* (L.) Link], a common rice weed, from different seeding depths and the efficacy of two PRE herbicides on the weed.

2. Materials and Methods

Experiments were conducted in 2012 in a screenhouse at the International Rice Research Institute. Los Baños, Philippines, to evaluate the effect of burial depth on seedling emergence of junglerice and biochar rates on the efficacy of two PRE herbicides on junglerice. The soil used in the experiments was collected from an upland rice field. The soil had sand, silt, and clay contents of 31%, 37%, and 32%, respectively. Before using it, the soil was autoclaved and passed through a 3-mm sieve. In both experiments, 25 seeds of junglerice were sown on the soil surface in plastic pots and covered with a thin layer of soil. In the first experiment, biochar was mixed with soil at 0, 20 (1.5%), 40 (3%), and 80 (6%) t ha^{-1} (calculated as soil volume to 10-cm soil depth) and seed burial depths were 0, 1, and 2 cm. In the second experiment, biochar rates were similar to the rates used in the first experiment. In this experiment, pretilachlor and pendimethalin were used at four doses. Pretilachlor was used at 0, 300, 600, and 900 g \cdot a.i. \cdot ha⁻¹ and pendimethalin was used at 0, 500, 1000, and 1500 g a.i. ha^{-1} . Herbicides were sprayed 1 d after sowing (DAS) using a Research Track sprayer (De Vries Manufacturing, Hollandale, MN 56045) that delivered 210 L·ha⁻¹ of spray solution at a spray pressure of 140 kPa. Flat fan nozzles (Teejet E0015) were used in the sprayer. The pots were irrigated 1 d after herbicide spray. Thereafter, pots were irrigated daily with a sprinkler system.

In the first experiment, seedling emergence of junglerice from different seeding depths was determined at 7, 14, and 21 DAS. In the second experiment, seedling emergence (or survival) was determined at 21 DAS. The shoot (leaf and stem) of junglerice was dried in an oven at 70°C for 72 h to determine seedling biomass. Experiments were conducted in a randomized complete-block design with a factorial arrangement of treatments. In the first experiment, factors were biochar rates and seeding depths and factors in the second experiment were biochar rates and herbicide doses. Treatments were replicated four times and both experiments were conducted two times. The data from the two "runs" were combined for analysis because there was no interaction effect of treatment and runs. The data were analyzed using an exponential model (Sigma Plot 10.0):

$Y = a \times e^{-bx}$

where *Y* is the seedling emergence (%) or seedling biomass $(g \cdot \text{pot}^{-1})$ at seed burial depth or herbicide dose *x*, *a* is the maximum emergence or biomass, and *b* is the slope.

3. Results and Discussion

3.1. Effect of Biochar Rates and Burial Depths on Junglerice Emergence

Seedling emergence of junglerice as a function of seed burial depths is shown in **Figure 1** and parameter estimates of the fitted model are shown in **Table 1**. At 7 DAS, the greatest seedling emergence (*a*) from the soil surface was observed when biochar was added to soil at 80 t·ha⁻¹ (70% emergence), whereas seedling emergence from the soil surface was 58% when biochar was not added to the soil (**Figure 1** and **Table 1**). The burial depth (x_{50}) required for 50% inhibition of maximum emergence increased with an increase in biochar rates. This burial depth, for example, was 0.57 cm when biochar was not added to the soil and it increased to 0.83 cm when biochar was added to the soil at 80 t·ha⁻¹ (**Table 1**).

At 14 DAS, seedling emergence further increased; however, the response was similar to the previous observations (**Figure 1(b)** and **Table 1**). At the final observation (21 DAS), the response of seedling emergence was

Table 1. Effect of biochar rates and seed burial depths on seedling emergence of junglerice at 7, 14, and 21 d after sowing. Seedling emergence data were fitted to an exponential model: $Y = a \times e^{-bx}$. Y is the seedling emergence (%) at seed burial depth x, a is the maximum emergence, b is the slope, and x_{50} is the burial depth (cm) required to inhibit 50% of seedling emergence. Parameter estimates are followed by standard error in parentheses.

Biochar rates $t \cdot ha^{-1}$	а	b	R^2	<i>x</i> ⁵⁰ (cm)		
7 days after sowing						
0	57.7 (2.1)	1.20 (0.11)	0.99	0.57		
20	63.2 (2.3)	1.14 (0.10)	0.99	0.61		
40	65.6 (1.3)	1.08 (0.05)	0.99	0.64		
80	70.4 (6.2)	0.84 (0.17)	0.98	0.83		
14 days after sowing						
0	69.1 (0.6)	0.99 (0.02)	0.99	0.70		
20	71.7 (2.0)	0.93 (0.06)	0.99	0.74		
40	76.0 (3.6)	0.85 (0.09)	0.99	0.81		
80	81.6 (3.6)	0.71 (0.07)	0.99	0.97		
21 days after sowing						
0	71.1 (0.6)	0.91 (0.02)	0.99	0.76		
20	73.6 (0.7)	0.85 (0.02)	0.99	0.81		
40	78.8 (1.6)	0.72 (0.03)	0.99	0.96		
80	83.1 (2.9)	0.57 (0.05)	0.98	1.21		

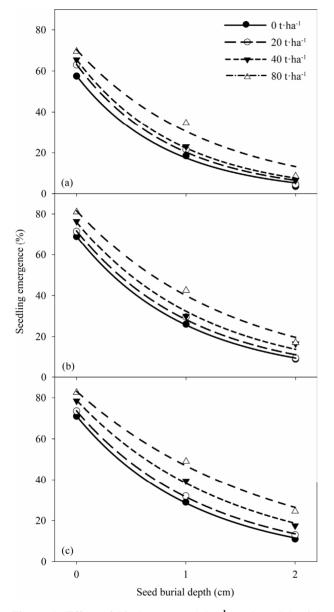


Figure 1. Effect of biochar rates $(t \cdot ha^{-1})$ and seed burial depths (cm) on seedling emergence (%) of junglerice at 7, 14, and 21 d after sowing (DAS). The lines represent an exponential model fitted to the seedling emergence data. (a) 7 DAS; (b) 14 DAS; (c) 21 DAS.

consistent with the previous two observations. Maximum seedling emergence (*a*) was 71% when biochar was not added to soil and it increased with an increase in biochar rates (**Figure 1(c)** and **Table 1**). Maximum seedling emergence was 74, 79, and 83% when biochar was added to the soil at 20, 40, and 80 t \cdot ha⁻¹, respectively. The fitted model also showed that the slope (*b*) decreased with an increase in biochar rates (**Table 1**).

The burial depth (x_{50}) required for 50% inhibition of maximum emergence was 0.76 cm when biochar was not added to the soil and it increased with an increase in

biochar rates. This burial depth was 0.81, 0.96, and 1.21 cm when biochar was added to the soil at 20, 40, and 80 t \cdot ha⁻¹, respectively (**Table 1**). The exponential model estimated that only 12% of the seedlings emerged from the seed burial depth of 2 cm when biochar was not added to the soil. When biochar was added at 20, 40, and 80 t \cdot ha⁻¹, seedling emergence from the 2-cm depth was 14%, 19%, and 26%, respectively.

The results of this study clearly suggest that seedling emergence of weeds from deeper depths may increase in biochar-amended soils compared with the soils to which biochar is not added. No such published information is available on weeds to compare our results; however, some information is available on seed germination of crops. Wheat (Triticum aestivum L.) seed germination increased with 10 t ha⁻¹ of papermill biochar [11], whereas maize (Zea mays L.) seed germination was not significantly affected by different biochars [12]. In a recent study, wheat seed germination was stimulated at the 10 t ha^{-1} biochar (rice husk) rate but not mungbean (Vigna mungo L.) [2]. The addition of biochar to soil may improve soil fertility by increasing soil pH, soil organic carbon, phosphorus, and potassium and this may help to stimulate seed germination [13]. However, these soil parameters were not estimated in my study. The reason for the greater emergence of junglerice from deeper depths (e.g., 2 cm) with the addition of biochar could be the increased bulk density of soil, and reduced soil compaction.

3.2. Effect of Biochar Rates and Herbicide Doses on Emergence and Biomass of Junglerice

Irrespective of the biochar rate and pendimethalin dose, no seedling survived when pendimethalin was applied (data not shown). However, biochar rates influenced pretilachlor efficacy on junglerice. Maximum seedling emergence (a) at different biochar rates was similar and ranged from 69 to 74% (Figure 2 and Table 2). However, the slope (b) of the fitted model decreased with an increase in biochar rates. The pretilachlor dose required for 50% inhibition of maximum emergence was 130 g ha⁻¹ when biochar was not added to the soil and this dose increased with an increase in biochar rates (Table 2). At 40 and 80 t ha^{-1} of biochar, the pretilachlor doses to inhibit 50% of maximum emergence were 270 and 290 g ha^{-1} , respectively. At the highest herbicide dose (900 g ha^{-1}), only 0.6% of the seedlings emerged when biochar was not added to the soil, whereas 7% to 9% of the seedlings emerged (or survived) when biochar was added at 40 to 80 t \cdot ha⁻¹.

Maximum seedling biomass (a) at different biochar rates ranged from 1.6 to 1.9 $g \cdot pot^{-1}$ (Figure 3 and Table 2). The slope (b) of the exponential model decreased

2.0

Table 2. Effect of biochar rates and pretilachlor doses on seedling emergence and seedling biomass of junglerice. Seedling emergence data were fitted to an exponential model: $Y = a \times e^{-bx}$. Y is the seedling emergence (%) or seedling biomass (g·pot⁻¹) at pretilachlor dose x, a is the maximum emergence or biomass, b is the slope, and x_{50} is the herbicide dose (g ha⁻¹) required to inhibit 50% of seedling emergence or seedling biomass. Parameter estimates are followed by standard error in parentheses.

Biochar rates (t·ha ⁻¹)	а	b	R^2	$x_{50} \left(\mathbf{g} \cdot \mathbf{ha}^{-1} \right)$		
Seedling emergence						
0	69.1 (6.0)	0.0053 (0.0013)	0.97	130		
20	72.3 (5.6)	0.0040 (0.0008)	0.98	170		
40	72.0 (5.3)	0.0026 (0.0004)	0.97	270		
80	73.9 (6.9)	0.0024 (0.0005)	0.96	290		
Seedling biomass						
0	1.57 (0.01)	0.0067 (0.0002)	0.99	100		
20	1.60 (0.07)	0.0053 (0.0006)	0.99	130		
40	1.69 (0.11)	0.0029 (0.0004)	0.98	240		
80	1.94 (0.14)	0.0029 (0.0004)	0.98	240		

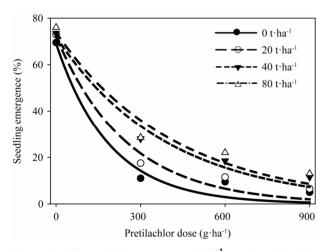
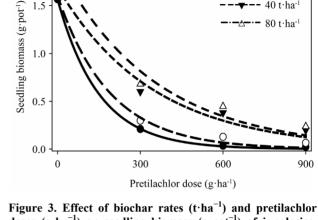


Figure 2. Effect of biochar rates (t ha⁻¹) and pretilachlor doses $(g \cdot ha^{-1})$ on seedling emergence (%) of junglerice. The lines represent an exponential model fitted to the seedling emergence data.

with an increase in biochar rates, suggesting that the rate of biomass reduction with increases in pretilachlor dose was slower when biochar was added to the soil. The pretilachlor dose required for 50% inhibition of maximum seedling biomass was 100 g ha⁻¹ when herbicide was applied in the absence of biochar (Table 2). However, this dose increased when herbicide was applied in the biochar-treated soil. The pretilachlor dose to inhibit 50% of maximum biomass was 240 g ha⁻¹ when biochar was



- 0 t·ha-1

20 t·ha-

- 40 t·ha-1

900

doses $(g \cdot ha^{-1})$ on seedling biomass $(g \cdot pot^{-1})$ of junglerice. The lines represent an exponential model fitted to the seedling biomass data.

added at 40 or 80 t ha⁻¹. At 900 g ha⁻¹ of pretilachlor (highest dose), the biomass of surviving seedlings was only 0.0004 $g \cdot pot^{-1}$ in the absence of biochar. With the addition of biochar at 20, 40, and 80 t ha⁻¹, however, the biomass of surviving seedlings increased to 0.0139, 0.1245, and 0.1445 $g \cdot pot^{-1}$, respectively.

In this study, the efficacy of pendimethalin was not influenced by the addition of biochar to the soil: however, biochar reduced the efficacy of pretilachlor. Pretilachlor is absorbed by roots only, whereas pendimethalin is absorbed by both roots and coleoptiles [14]. The most important absorption sites for pendimethalin in sensitive grass weeds are the coleoptiles. These observations suggest that, irrespective of the biochar rates, pendimethalin was readily absorbed by the emerging junglerice coleoptile as pendimethalin was applied on the soil surface. Organic matter in the biochar-treated soil might have bound pretilachlor and pretilachlor bioavailability to the roots of junglerice decreased, resulting in reduced efficacy in the biochar-treated soil. In a recent study, no seedlings of junglerice escaped pendimethalin when it was applied in the presence of rice residue (straw) cover [15]. However, some seedlings of barnyardgrass and rice flatsedge survived pendimethalin when it was applied in the presence of residue cover.

3.3. Implications for Weed Management

The addition of biochar can increase the quality, fertility, and productivity of poor soils, thereby providing an option to reduce greenhouse gas emissions and to sequester carbon in rice-based systems [1,16]. However, the use of biochar is not without its critics [5]. Some compounds in biochar have the potential to influence seedling emer-

gence of crops [2] as well as weeds. In this study, junglerice emergence was greater from 2-cm seed burial depth in the biochar-treated soil than in the soil without biochar. Similarly, the burial depth required to inhibit 50% of maximum emergence was greater when biochar was added to the soil. These results may have negative impacts on weed management. The addition of biochar may stimulate seedling emergence from deeper depths and, as seeds are present at different depths, seedling emergence may be prolonged. Because of small seed size, seedlings of some seeds, for example, Chinese sprangletop [Leptochloa chinensis (L.) Nees] and ludwigia [Ludwigia hyssopifolia (G. Don) Exell.], may not be able to emerge from a burial depth of 0.5 cm [17-19]. However, the addition of biochar to soils may decrease soil compaction and these weed species may easily emerge from such burial depths. There is a need to study the effect of biochar rates and seed burial depths on the seedling emergence of weeds with different seed sizes.

The high adsorption and retention capacity of biochar is not limited to nutrients only, but is exhibited toward pesticides also, including soil-applied herbicides [4,5]. Biochar addition to the soil can have positive or negative impacts on pest management [4]. Adsorption of herbicides can reduce leaching of soil-applied herbicides, and protect herbicides from degradation [5,20,21]. In terms of weed control, strong adsorption of soil-applied herbicides on biochar can decrease the efficacy of herbicides by influencing their bioavailability [4,5,22].

Reduced herbicidal efficacy of diuron in soils amended with wheat straw biochar and clomazone in soils amended with rice straw biochar was reported in recent studies [21]. With increasing biochar in soil, higher rates of application of herbicides may be needed to achieve the same weed control as in unamended soil [21]. The results of my study, alongside those of others, also suggest that biochar may reduce the efficacy of some soil-applied herbicides.

As results were different for two herbicides, pendimethalin and pretilachlor, there was a need for further research with several soil-applied herbicides and soil types. As biochar application is predicted to rise over the next decade in response to the need to improve the fertility of poor soils, it is important that herbicide companies provide strong guidance to farmers on herbicide rates in biochar-treated soils [5]. My study and most of the previous studies were conducted on freshly prepared biochar. The long-term effect of biochar on the persistence and efficacy of herbicides deserves urgent attention.

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