

The Distribution and Substrate Specificity of Extracellular Nuclease Activity in Marine Fungi

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ABSTRACT

The distribution and specificity of extracellular nucleases produced by marine fungi belonging to eleven genera, namely: *Alternaria*, *Aspergillus*, *Aureobasidium*, *Chaetomium*, *Fusarium*, *Glomastix*, *Humicola*, *Penicillium*, *Scopulariopsis*, *Wardomyces*, *Periconia*, have implied its important function in the organic phosphorus and nitrogen circle in the Ocean. The fungal nucleases of 64 isolates tested were more or less specific for single-stranded DNA with a high preferential specificity towards poly-U substrate with forming of 5'-phosphate mononucleotides. A couple of the nucleases were capable of RNA digesting. The highest level of extracellular nucleolytic ability was observed in *Penicillium* spp. isolates. The tight correlation found between extracellular nuclease activity and the rate of thymidine uptake by actively growing and sporulating marine fungus *Penicillium melinii* suggests that this nuclease is required for fulfilling the nucleotide pool of precursors of DNA biosynthesis during transformation of hyphae into the aerial mycelium and conidia in stressful environmental conditions.

Keywords: Marine Fungi; Marine Environment; Single-Strand-Specific Nuclease; DNase; RNase; ssDNA; Thymidine Uptake

1. Introduction

The main portion of the organic carbon, phosphorus and nitrogen is a direct result of enzymatic processing of dissolved macromolecules of DNA, RNA and proteins in aquatic environments due to their wide occurrence [1,2]. Aquatic microorganism populations utilize nucleic acids, enabling efficiently scavenging of organic matter from the dissolved pool, rapidly cycling the nucleic acids [3, 4]. Therefore, there is considerable interest in molecular utilization mechanism of nucleic acids in marine ecosystem. Some fields of fungal community studies have been described [5]. However, the contribution of marine mycobiota as a source and consumer of the dissolved organic matter such as nucleic acids has not been yet shown.

It is known that a large group of fungi and filamentous bacteria generate and disseminate spores for implement of survival program in stressful environmental conditions. These events call forth increase of nuclease activity dramatically that coincides with lysis of the substrate hyphae and formation of aerial mycelium [6]. It has analogy to the late steps of apoptosis of other eukaryotic cells [7]. Indeed, these programs may be activated by environmental stresses or by natural processes of senescence

[8,9]. Similarly, plant cells undergoing programmed cell death during tissue differentiation at the end of mitosis or under the influence of pathogens exhibit DNA degradation [10].

Only a few species of terrestrial fungi producing extracellular nucleolytic enzymes have been reported [11-13]. No information is available on the genus or species specificity of nucleolytic enzymes in marine fungi. However, the previous data have shown the substantial difference between the marine fungi and their terrestrial counterparts [14,15]. Moreover, most of marine fungi were found to be facultative with the potential for long-distance dispersal that might enable unusually rapid propagation of epidemics in marine systems [16].

The purpose of the present study was to evaluate extracellular nucleolytic ability in marine fungi using different natural and synthetic substrates and its possible biological and ecological role in marine environment.

2. Materials and Methods

2.1. Fungal Isolates and Cultural Conditions

The fungal strains were isolated from marine animals and sea bottom sediments collected in the shore water of the Kuril Islands and Primorskyi krai (**Table 1**) and identified as described previously [17,18]. Fungi cultures were

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Table 1. The distribution of extracellular nucleolytic activity in marine fungi.

Strain	Number of isolates	Marine source	Nuclease assay, OE ₂₂₆₀	Protein concentration, mg/ml	Specific activity, U/mg
<i>Aspergillus caespitosus</i>	1	<i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0.010	nd*	nd
<i>Aspergillus repens</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 75 m	0.010	nd	nd
<i>Aspergillus versicolor</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	≤0.010	nd	nd
<i>Aspergillus wentii</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 75 m	0.094	0.1	4.7
<i>Chaetomium globosum</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	0.163	0.25	3.3
<i>Chaetomium</i> sp.	3	<i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0.124	0.1	6.2
<i>Chaetomium</i> sp.	1	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0.146	0.1	7.3
<i>Fusarium solani</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	≤0.010	nd	nd
<i>Humicola</i> sp.	2	<i>Ophiura</i> sp., Kunashir Island, Sea of Okhotsk, 75 m	0.104	0.1	5.2
<i>Penicillium chrysogenum</i> (<i>P. notatum</i>)	14	<i>Antedon</i> sp., <i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m; <i>Soares</i> sp., <i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	0.138	0.05	13.8
<i>Penicillium commune</i>	1	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 75 m	0.290	0.2	7.3
<i>Penicillium melinii</i> (<i>P. estinogenum</i>)	1	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0.352	0.15	11.7
<i>Penicillium potalum</i> (<i>P. griseofulvum</i>)	1	Bottom of Peter the Great Bay, Sea of Japan, 46 m	0.454	0.08	28
<i>Penicillium herquej</i>	2	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	0.115	0.2	2.9
<i>Penicillium olivicolor</i>	1	<i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0.166	0.2	4.2
<i>Penicillium viridicatum</i>	1	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	≤0.01	nd	nd
<i>Scopulariopsis brevicaulis</i>	1	Bottom of Peter the Great Bay, Sea of Japan, 46 m	≤0.01	nd	nd
<i>Sterile mycelium</i> (<i>Dematiaceae</i>)	1	Bottom of Sea of Okhotsk, Iturup Island, 551 m	0.032	0.1	1.6
<i>Alternaria alternata</i>	1	<i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0	nd	nd
<i>Aspergillus candidus</i>	6	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0	nd	nd
<i>Aspergillus amilovorvus</i>	2	<i>Antoza</i> sp., Shikotan Island, Sea of Okhotsk, 68 m; <i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0	nd	nd
<i>Aspergillus amstelodami</i>	1	<i>Antoza</i> sp., Shikotan Island, Sea of Okhotsk, 68 m	0	nd	nd
<i>Aspergillus repens</i>	2	Bottom of Sea of Okhotsk, Kuriles, Iturup Island, 551 m	0	nd	nd
<i>Aspergillus varians</i>	9	<i>Spongia</i> sp., Kunashir Island, Sea of Okhotsk, 81 m; <i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m; <i>Antedon</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0	nd	nd
<i>Aureobasidium pullupans</i>	1	<i>Ophiura</i> sp., Kunashir Island, Sea of Okhotsk, 81 m	0	nd	nd
<i>Fusarium sporotrichiella</i>	1	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0	nd	nd
<i>Gliomastix cereales</i>	1	<i>Spongia</i> sp., Shikotan Island, Sea of Okhotsk, 68 m	0	nd	nd
<i>Penicillium citrinum</i>	1	<i>Spongia</i> sp., Iturup Island, Sea of Okhotsk, 270 m	0	nd	nd
<i>Penicillium jensenii</i>	2	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0	nd	nd
<i>Penicillium namyslowskii</i>	1	<i>Ascidium</i> sp., Shikotan Island, Sea of Okhotsk, 123 m	0	nd	nd
<i>Wardomyces anomalus</i>	1	Bottom of Peter the Great Bay, Sea of Japan, 46 m	0	nd	nd
<i>Periconia</i> sp.	2	<i>Alcyonaria</i> sp., Shikotan Island, Sea of Okhotsk, 68 m	0	nd	nd

*nd—not determined.

maintained on sterile seawater with Wort agar [19]. Each isolate was stationary grown in 100 ml of modified Ogata medium consisting of glucose 5%, pepton 1%, soybean flour 0.5%, $\text{MgSO}_4 \times 7\text{H}_2\text{O}$ – 0.05%, $\text{CaCl}_2 \times 2\text{H}_2\text{O}$ – 0.01%, KNO_3 – 0.2% and natural seawater without phosphates at 22°C for 7 days.

2.2. Extracellular Nuclease Activity Assay

Extracellular nucleolytic enzyme activity was determined in the fungal culture during the cells growth. For extracellular nucleolytic enzyme activity detection, cell-free supernatant after culture filtration was inoculated onto the agar plate. The agar plates were prepared with 3 g of liquefied agarose (“Hemapol”), 200 ml of 0.03 M sodium acetate buffer, pH 4.6, containing 1 mM ZnSO_4 and 0.05 M NaCl, and 2 ml heat-denatured DNA (ssDNA) at concentration of 1 - 2 mg/ml. The resulting mixture was immediately poured onto marked horizontal plates with the final volume about 20 ml (8×12 cm and 2 mm thick). Wells of 3 mm in diameter were cut in the agar plates. Each of two-microliter samples of fungal culture in equal protein concentrations was inoculated into the wells. After incubation at 37°C for 20 min, the result of the nuclease activity was observed at λ_{260} . The more active fungal isolates were selected by comparison of the dark circle zones produced by enzymatic cleavage of the substrate for the further quantitative analysis.

2.3. Nuclease Specificity

Nuclease specificity was evaluated in a mixture of 0.5 ml 0.03 M sodium acetate buffer, pH 4.6, containing 1 mM ZnSO_4 and 0.05 M NaCl, substrate in concentration of 10 OE_{260} and 0.02 ml of filtered fungal culture sample. The reaction mixture was incubated at 37°C for 20 min and terminated by adding 2.0 ml of 0.5 M HClO_4 and 0.1% $\text{La}(\text{NO}_3)_3$. The mixture was centrifuged at 2000 g for 10 min. The absorbance of supernatant at λ_{260} was measured. One unit of the enzyme was defined as the amount that catalyzes the production of acid-soluble nucleotides increased by 1.0 absorbance unit for 20 min. Protein concentration was defined by Bradford method [20]. ssDNA was prepared by incubating bovine spleen DNA (Biolar, Russia) solution in boiling water for 30 min and then in the ice bath for 10 min. Poly (A), Poly (C), and poly (U), Poly (A:U), Poly (C:G), Poly (I:C) were from “Reanal”. Poly (G), Poly (I), yeast RNA was from “Vector” (Russia).

2.4. [H^3]Thymidine Uptake and Conidia Productivity Assay

Penicillium melinii isolates were examined for the ability to take up [H^3]thymidine during cell growth. [H^3]Thy-

midine uptake was determined as a total cell-associated radioactivity by adding 1-ml samples to tubes containing 1 ml of 2 $\mu\text{Ci/ml}$ [H^3]-labeled thymidine in nutrient-free artificial seawater. Subsamples were filtered within 5 min onto 0.2-mm-pore-diameter GFC filters (Whatman) and were washed with 5 ml of 5% trichloroacetic acid with 1% sodium pyrophosphate and ethanol. Radioactivity in the samples was measured by liquid scintillation counting (Mark III). The conidia were direct counted immediately and again after every two days of incubation. Counting of conidia was carried out as described by Bilai [21]. An aliquot of 10 ml of cultural fluid with fungus was shaken for 30 min and centrifuged at 3000 g for 20 min. Sediment was weighed and 0.3 g of fungal biomass was suspended in 1 ml of distilled water. Concentration of conidia was calculated five times in Thom’s cell.

2.5. Statistical Analysis

Extracellular nucleolytic ability, [H^3]thymidine uptake and conidia productivity of fungi were evaluated as the average of three independent experiments. Statistical significance was analyzed by the Student’s t-test, the critical level of significance was 5%.

2.6. Molecular Cloning and Sequence Analysis

Recombinant DNA technique was performed using Ins T/Aclonem PCR Product Cloning Kit, restriction endonucleases, T4 DNA ligase, Long PCR mix (Fermentas), Smart Taq Polymerase (Topotili), automatic amplifier (Eppendorf). Two pair of primers (For-1 and Rev, sequences: 5’-CAYTTYATHGGNGAYATGAC-3’ and 5’-GCCANCKNGTNGCNGT-3’, respectively; For-2 and Rev, sequences: 5’-GAYTGGGAYACNTAYATGCC-3’ and 5’-GCCANCKNGTNGCNGT-3’, respectively) were constructed on the base of S1-like nuclease homologue sequence alignments to amplify the active site region of S1 type nuclease using chromosomal DNA of *P. melinii*, *P. patulum* and *P. chrysogenum* as templates. N- and C-termini region coding sequences were determined using two pair of primers Nuc-for1 and Nuc-rev, sequences: 5’-ATGGTCTCTCTATCCAAGATTG-3’ and 5’-ATGGCTATGCTCACGTCCTTCTGAGG-3’, respectively. PCR products were cloned and sequenced using the automated PE/ABI 310 DNA sequencer and PE/ABI-ABI PRISM BigDye Terminator cycle sequencing Ready Reaction Kit (PE Applied Biosystems). *E. coli* strains TOP10 or XL1 (Evrogen) were used for standard cloning procedures. Nucleotide and amino acid sequence analysis were performed with Chromas, GenRunner, SMART programs. Nucleotide and amino acid sequences homology and similarity searches and alignments were carried out by using BLAST and ClustalW, MOE 9.10 facilities.

3. Results

3.1. Screening of Extracellular Nuclease Activities

Fifty percent from more than 60 isolates tested was found to produce a different level of extracellular nuclease activity catalyzing the cleavage of ssDNA at pH 4.6 (**Table 1**). Of the isolates with the high level of extracellular nuclease activity (about 10.0 - 30.0 U/mg), 14 isolates belonged to the species *P. chrysogenum* (formerly *P. notatum*) isolated from various species of sponges collected near Iturup and Kunashir Islands at the depth of 270 m and 81 m, respectively. The isolates of *P. melinii* (formerly *P. estinogenum*) and *P. patulum* (formerly *P. griseofulvum*) isolated from colonial ascidium near Shikotan Island at the depth of 123 m and *P. patulum* isolated from Peter the Great Bay bottom sediments, Sea of Japan, at the depth of 46 m also showed a high level of extracellular nuclease activity (**Table 1**). Apparent nucleolytic activity was found in the isolates of *Chaetomium* sp., *A. wentii*, *Humicola* sp., *P. commune*, *P. herquej*, *P. olivicolor*, and *Dematiaceae* isolated from the bottom sediment of Sea of Okhotsk from 551 m depth (**Table 1**). The number of isolates belonging to genus *Penicillium* exhibited a highest level of extracellular nuclease activity accounted for 53% of the total. The majority of these isolates were identified as populations of *P. chrysogenum*. As shown in **Table 1**, *Penicillium* spp. coexists with the largest amount of the deep-sea inhabitants irrespectively of geographic area. Extracellular nucleolytic activity was not detected in any isolates belonging to the species *A. alternata*, *A. candidus*, *A. amilovor*, *A. amstelodami*, *A. am-*

stelodami, *A. varians*, *Aureobasidium pullupans*, *F. sporotrichiella*, *G. cereales*, *P. citrinum*, *P. jensenii*, *P. namyslowskii*, *W. anomalus*, *Periconia* sp. (**Table 1**).

3.2. Extracellular Nucleases Specificity

The marine fungi were found to produce non-specific extracellular ssDNA/RNA nucleases especially those, which was been habitable under the deep-sea conditions (**Table 1**). As shown in **Figure 1**, all of the strains, which were capable of ssDNA digestion, have a preference for Poly (U). The rate of Poly (U) hydrolysis was in 10 - 100-fold higher in comparison with the other substrates. The fungal extracellular nucleases acted both ssDNA and RNA at the same rate. Nuclease from a terrestrial strain of *A. oryzae* showed the similar specificity under the same reaction conditions. Nucleases from *A. wentii* (sponge, Kunashir Island, 75 m depth) and *Chaetomium* spp. isolates (colonial ascidium, Shikotan Island, 123 m depth) were found to be much more active with RNA than DNA as a substrate (**Figure 1**).

3.3. [³H]Thymidine Uptake versus the Extracellular Nuclease Activity and Conidia Productivity

As shown in **Figure 2**, the maximal nuclease productivity achieved when fungus colony started actively growing after 3 days of incubation on a nutrient-free medium and it was highest at 10 days. Since the fungus cells were capable of [³H]thymidine inclusion (**Figure 2**). The tight correlation ($r = 0.93$) found between extracellular nuclease activity and the rate of thymidine uptake. As ex-

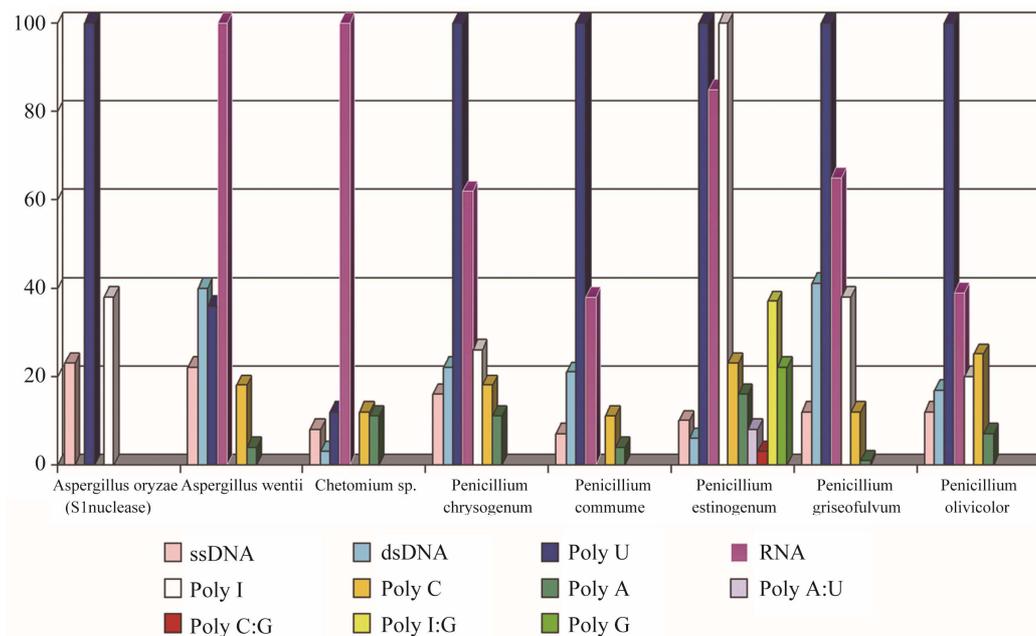


Figure 1. Specificity of extracellular nucleases of marine fungi.

pected, the amount of green conidia was highest at 11 days of incubation (**Figure 3**). The extracellular nucleolytic enzyme activity and $[H^3]$ thymidine uptake decreased when conidia productivity reached an exponential-stationary phase after 12 days of incubation (**Figures 2 and 3**). Shaking the fungal culture during the growth strongly repressed nuclease productivity, however, it was 2 - 5-fold higher when the fungus was cultivated without shaking (unpublished results).

3.4. Sequence Analysis

The structural and biochemical details of some ssDNA-preferential nucleases of class 3.1.30.1 isolated from the terrestrial sources are known [13,22,23]. We have cloned and analyzed ssDNA-preferential highly active extracellular nucleases from marine fungi, namely: *P. melinii* (GU331890), *P. patulum* (formerly *P. griseofulvum*) (GU331891), and *P. chrysogenum* (GU33182).

The presence of putative signal peptide indicates that these proteins are destined for secretion to the periplasm or excretion into the environment. Gene Bank databases searching by BLAST software disclosed a significant sequence homology of these enzymes with terrestrial *P. citrinum* (70% identity), *A. ozyae* (52% identity), and *N. crassa* (47% identity) S1 type nucleases that suggested the same mechanism of action (**Figure 4**).

4. Discussion

Our investigation demonstrated a large number of the fungal strains collected from the sea deepwater sources, producing highly active extracellular nucleases into marine environment. Extracellular nucleolytic ability was detected in the strains of the genera *Aspergillus*, *Chaetomium*, *Fusarium*, *Humicola*, *Penicillium*, *Scopulariopsis* that were found prolifically in the sea bottom sediments and marine animals inhabiting of the shelf waters of Japan Sea. The species of the genera *Penicillium*, *Aspergillus*, *Chaetomium*, *Fusarium* were of more frequent occurrence (**Table 1**). Some authors suggested the presence of interference competition among the fungi in mixed cultures on substrates, especially in congeneric species [24]. The ecological factor influences on the fungal population densities as well as their quality have also been reported [25]. The extracellular nuclease productivity among the marine fungi appears to be dependent on origin and depth as well as kind of host (**Table 1**). It might be related to taxonomic and ecological groups or/and biogeographic zones [26].

Of 64 isolates tested, two were capable of RNA digesting (**Figure 1**). Other fungal nucleases are more or less specific for ssDNA, preferentially cleaving Poly (U) substrate to leave 5'-phosphate mononucleotides. The high extracellular ability to cleave RNA indicates that

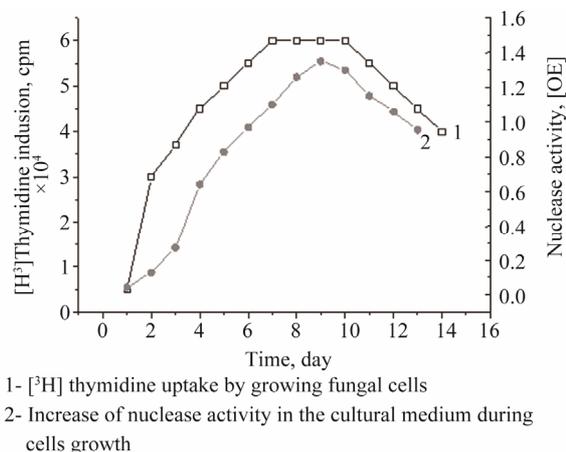


Figure 2. $[H^3]$ Thymidine uptake versus extracellular ssDNA-preferential nuclease production by *P. melinii*.

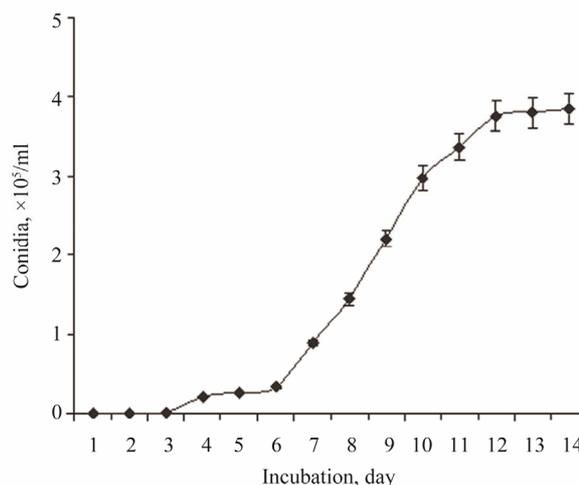


Figure 3. Conidia formation intensity in marine fungus *P. melinii* during incubation.

some marine fungi species have a potential to safe genetic information from attacking carrier of viruses resides providing participation in host defense [27].

Actively growing marine fungus *P. melinii* (formerly *P. estinogenum*) collected from colonial *Ascidium* spp. at the depth of 270 m was examined for $[H^3]$ thymidine uptake into the cells (**Figure 2**). The results obtained indicated that the extracellular nucleolytic enzyme activities in marine fungi were required for nucleotide inclusions into DNA of the newly developing cells. Its function proposed to have a key role in complete DNA degradation of exogenous substrates as well as putative apoptotic destruction of the fungal hyphae to supply building blocks for macromolecular biosynthesis in the aerial mycelium and conidia as the result of senescence or stressful environmental conditions [8,9]. The inhibition of either hydrolytic activity at least strongly impaired aerial mycelium formation [6]. In the other hand, we have found that

counterparts together with doubling of Arg content are essential in the marine fungi enzymes due to additional C-terminal peptide from 270 to 344 amino acid residues. An increase in polar intrinsic flexible Gly as well as non-polar small Ala residues and additional C-terminal sequence suggest that it is a functional adaptation molded by evolution under the deep-sea conditions [31].

In summary, we have shown that the marine fungi possess important extracellular nucleolytic enzymes, which are required for fulfilling the nucleotide pool of precursors of DNA biosynthesis and maintaining ecological balance. Their distribution in the deep-sea water and unique specificity confirmed the important function in the organic phosphorus and nitrogen circle in the Ocean. The potential of extracellular nucleolytic ability in marine fungi to estimate the marine environmental conditions as well as the biotechnology potential of the marine fungi nucleases will attract much attention.

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REFERENCES

- [1] J. H. Paul, W. H. Jeffrey and J. P. Cannon, "Production of Dissolved DNA, RNA and Protein by Microbial Populations in a Florida Reservoir," *Applied and Environmental Microbiology*, Vol. 56, No. 10, 1990, pp. 2957-2962.
- [2] A. Dell'Anno and R. Danavaro, "Extracellular DNA Plays a Key Role in Deep-Sea Ecosystem Functioning," *Science*, Vol. 309, No. 5744, 2005, p. 2179. [doi:10.1126/science.1117475](https://doi.org/10.1126/science.1117475)
- [3] J. H. Paul, A. W. Jeffrey and M. F. DeFlaun, "Dynamics of Extracellular DNA in the Marine Environment," *Applied and Environmental Microbiology*, Vol. 53, No. 1, 1987, pp. 170-179.
- [4] J. H. Paul, M. F. DeFlaun and W. H. Jeffrey, "Mechanisms of DNA Utilization by Estuarine Microbial Populations," *Applied and Environmental Microbiology*, Vol. 54, No. 7, 1988, pp. 1682-1688.
- [5] C. A. Shearer, "Fungal Competition," *Canadian Journal of Botany-Revue, Canadienne de Botanique*, Vol. 73, 1995, pp. S1259-S1264. [doi:10.1139/b95-386](https://doi.org/10.1139/b95-386)
- [6] R. G. Nieceza, J. Huergo, B. A. Connolly and J. Sánchez, "Purification, Characterization and Role of Nucleases and Serine Proteases in *Streptomyces* Differentiation: Analogies with the Biochemical Processes Described in Late-Steps of Eukaryotic Apoptosis," *Journal of Biological Chemistry*, Vol. 274, No. 29, 1999, pp. 20366-20375. [doi:10.1074/jbc.274.29.20366](https://doi.org/10.1074/jbc.274.29.20366)
- [7] S. Aoyagia, M. Sugiyama and H. Fukudac, "BEN1 and ZEN1 cDNAs Encoding S1-Type DNases That Are Associated with Programmed Cell Death in Plants," *FEBS Letters*, Vol. 429, 1998, pp. 134-138. [doi:10.1016/S0014-5793\(98\)00563-8](https://doi.org/10.1016/S0014-5793(98)00563-8)
- [8] T. Panavas and B. Rubinstein, "Oxidative Events during Programmed Cell Death of Daylily (*Hemerocallis* Hybrid) Petals," *Plant Science*, Vol. 133, No. 2, 1998, pp. 125-138. [doi:10.1016/S0168-9452\(98\)00034-X](https://doi.org/10.1016/S0168-9452(98)00034-X)
- [9] Y. Muramoto, A. Watanabe, T. Nakamura and T. Takabe, "Enhanced Expression of a Nuclease Gene in Leaves of Barley Plants under Salt Stress," *Gene*, Vol. 234, No. 2, 1999, pp. 315-321. [doi:10.1016/S0378-1119\(99\)00193-6](https://doi.org/10.1016/S0378-1119(99)00193-6)
- [10] M. Sugiyama, J. Ito, S. Aoyagi and H. Fukuda, "Endonucleases," *Plant Molecular Biology*, Vol. 44, No. 3, 2000, pp. 387-397. [doi:10.1023/A:1026504911786](https://doi.org/10.1023/A:1026504911786)
- [11] K. Shishido and T. Ando, "Single-Strand Specific Nucleases," In: S. M. Linn and R. J. Roberts, Eds., *Nucleases*, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, New York, 1982, pp. 155-185.
- [12] V. Vogt, "Purification and Future Properties of Single-Strand-Specific Nuclease from *Aspergillus oryzae*," *FEBS Journal*, Vol. 33, 2005, pp. 192-200.
- [13] N. A. Desai and V. Shankar, "Single-Strand-Specific Nuclease from *Basidiobolus haptosporus* Nuclease Bh1)," *Scientific Research and Essay*, Vol. 2, No. 5, 2007, pp. 139-146.
- [14] C. Osterhage, M. Schwibbe, G. M. König and A. D. Wright, "Difference between Marine and Terrestrial *Phoma* Species as Determined by HPLC-DAD and HPLC-MS," *Phytochemical Analysis*, Vol. 11, No. 5, 2000, pp. 288-294. [doi:10.1002/1099-1565\(200009/10\)11:5<288::AID-PCA528>3.0.CO;2-G](https://doi.org/10.1002/1099-1565(200009/10)11:5<288::AID-PCA528>3.0.CO;2-G)
- [15] A. Baakza, B. P. Yala and H. C. Dube, "A Comparative Study of Siderophore Production by Fungi from Marine and Terrestrial Habitats," *Journal of Experimental Marine Biology and Ecology*, Vol. 311, No. 1, 2004, pp. 1-9. [doi:10.1016/j.jembe.2003.12.028](https://doi.org/10.1016/j.jembe.2003.12.028)
- [16] H. McCallum, C. D. Harvell and A. Dobson, "Rates of Spread of Marine Pathogens," *Ecology Letters*, Vol. 6, No. 12, 2003, pp. 1062-1067. [doi:10.1046/j.1461-0248.2003.00545.x](https://doi.org/10.1046/j.1461-0248.2003.00545.x)
- [17] K. D. Hyde, C. A. Farrant and E. B. G. Jones, "Isolation of Marine Fungi," *Botanica Marina*, Vol. 30, 1987, pp. 291-303. [doi:10.1515/botm.1987.30.4.291](https://doi.org/10.1515/botm.1987.30.4.291)
- [18] Y. V. Khudyakova, M. V. Pivkin, T. A. Kuznetsova and V. I. Svetashev, "Fungi in Sediments of the Sea of Japan and Their Biologically Active Metabolites," *Microbiology*, Vol. 69, No. 5, 2000, pp. 608-611. [doi:10.1007/BF02756817](https://doi.org/10.1007/BF02756817)
- [19] N. Y. Artemchuk, "Microflora of USSR's Seas," Nauka, Moscow, 1981.
- [20] M. M. Bradford, "A Rapid and Sensitive Method for the Quantitation of Microgram Quantities of Protein-Dye Binding," *Analytical Biochemistry*, Vol. 72, No. 1-2, 1976, pp. 248-254. [doi:10.1016/0003-2697\(76\)90527-3](https://doi.org/10.1016/0003-2697(76)90527-3)
- [21] V. I. Bilai, "Definition of the Growth and Biosynthetic Activity of Fungi," In: V. I. Bilay, Ed., *Method of Experimental Mycology*, Naukova Dumka, Kiev, 1982, pp. 146-147.
- [22] S. A. Martin, R. S. Ullrich and W. L. Meyer, "A Com-

- parative Study of Nucleases Exhibiting Preference for Single-Stranded Nucleic Acid,” *Biochimica et Biophysica Acta*, Vol. 867, No. 1-2, 1986, pp. 76-80.
[doi:10.1016/0167-4781\(86\)90031-X](https://doi.org/10.1016/0167-4781(86)90031-X)
- [23] M. Pimkin, C. Miller, L. Blakesley, C. Oleykowski, N. Kodali and A. Yeung, “Characterization of a Periplasmic S1-Like Nuclease Coded by the Mesorhizobium Loti Symbiosis Island,” *Biochemical and Biophysical Research Communications*, Vol. 343, No. 1, 2006, pp. 77-84.
[doi:10.1016/j.bbrc.2006.02.117](https://doi.org/10.1016/j.bbrc.2006.02.117)
- [24] T. K. Tan, C. L. Teng and B. G. Jones, “Substrate Type and Microbial Interaction as Factors Affecting Ascocarp Formation by Mangrove Fungi,” *Hydrobiologia*, Vol. 295, 1995, pp. 127-134. [doi:10.1007/BF00029119](https://doi.org/10.1007/BF00029119)
- [25] M. Muntanola-Cvetkovic, “A Mycological Survey of the South Adriatic Sea,” *Journal of Experimental Marine Biology and Ecology*, Vol. 43, No. 3, 1980, pp. 193-206.
[doi:10.1016/0022-0981\(80\)90047-7](https://doi.org/10.1016/0022-0981(80)90047-7)
- [26] Y. Vishwakiran, N. L. Thakur, S. Raghukumar, P. L. Yennawar and A. C. Anil, “Spatial and Temporal Distribution of Fungi and Wood-Borers in the Coastal Tropical Waters of Goa, India,” *Botanica Marina*, Vol. 44, 2001, pp. 47-56. [doi:10.1515/BOT.2001.007](https://doi.org/10.1515/BOT.2001.007)
- [27] K. C. Hsia, C. L. Li and H. S. Yuan, “Structural and Functional Insight into Sugar-Nonspecific Nucleases in Host Defense,” *Current Opinion in Structural Biology*, Vol. 15, No. 1, 2005, pp. 193-206.
[doi:10.1016/j.sbi.2005.01.015](https://doi.org/10.1016/j.sbi.2005.01.015)
- [28] S. Y. Newell, “Established and Potential Impacts of Eukaryotic Mycelial Decomposers in Marine/Terrestrial Ecotones,” *Journal of Experimental Marine Biology and Ecology*, Vol. 200, 1996, pp. 187-206.
[doi:10.1016/S0022-0981\(96\)02643-3](https://doi.org/10.1016/S0022-0981(96)02643-3)
- [29] K. E. Gonga, D. Jendrossek and H. P. Molitoris, “Fungal Degradation of the Thermoplastic Polymer Poly-Beta-Hydroxybutyric Acid (PHB) under Simulated Deep Sea Pressure,” *Hydrobiologia*, Vol. 426, 2000, pp. 173-183.
[doi:10.1023/A:1003971925285](https://doi.org/10.1023/A:1003971925285)
- [30] J. Ito and H. Fukuda, “ZEN1 Is a Key Enzyme in the Degradation of Nuclear DNA during Programmed Cell Death of Tracheary Elements,” *Plant Cell*, Vol. 14, No. 12, 2002, pp. 3201-3211. [doi:10.1105/tpc.006411](https://doi.org/10.1105/tpc.006411)
- [31] L. A. Balabanova, Y. M. Gafurov, M. V. Pivkin, N. A. Terentyeva, G. N. Likhatskaya and V. A. Rasskazov, “An Extracellular S1-Type Nuclease of Marine Fungus *Penicillium melinii*,” *Marine Biotechnology*, Vol. 14, No. 1, 2012, pp. 87-95. [doi:10.1007/s10126-011-9392-5](https://doi.org/10.1007/s10126-011-9392-5)