

# Development of acidophilus milk via selected probiotics & prebiotics using artificial neural network

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## ABSTRACT

Commercial interest in functional foods containing probiotic strains has consistently increased due to the awareness of gut health. Recent advancements are leading to development of symbiotic foods, containing prebiotics and probiotics bearing synergistic effects of the two. Thus, in present study, symbiotic acidophilus milk was developed satisfying functional dairy food properties. Different sets of milk were fermented with probiotic cultures (*Lactobacillus acidophilus*, *Bifidobacterium bifidum*, *Lactobacillus casei*, bioyoghurt culture) singly or in combination, and prebiotics namely inulin (I), oat fibre (O) and honey (H). Obtained 20 symbiotic samples were organoleptically tested, physico-chemically (titrable acidity percentage (TA) & pH) and microbiologically (total viable count (TVC), coliform count and yeast & mold count) analyzed. The incorporation of honey and inulin led to development of sweetened and low calorie sweetened symbiotic acidophilus milk, respectively. Incorporation of *B. bifidum* increased the flavour of symbiotic acidophilus milk when compared to *L. acidophilus* as control, where as *L. casei* culture showed thinner consistency in the product. Addition of prebiotic affected only the sensory scores, whereas the probiotics addition resulted in a marginal variation of pH and TA. TVC of all symbiotic acidophilus milk samples obtained were more than desirable limits for harvesting probiotic effects ( $>10^8$  cfu/ml). Finally, a two layer feed-forward artificial neural network (ANN) was established to predict the sensory evaluation based on inputs of probiotic and prebiotic.

**Keywords:** Symbiotic Acidophilus Milk; Artificial Neural Network; Sensory Evaluation

## 1. INTRODUCTION

Notable fermented milks are buttermilk, acidophilus milk, bulgarian buttermilk, kumiss, kefir and yoghurt.

Acidophilus milk is a sour product that has been allowed to ferment under conditions that favor the growth and development of thermophilic lactic acid bacteria. This type of fermented milk is produced by development in milk of *Lactobacillus acidophilus*. It is claimed that acidophilus milk has therapeutic and health-promoting properties. It is also claimed that the growth of *Lactobacillus acidophilus* under the condition existing in the intestinal tract can replace undesirable putrefactive fermentation with a beneficial lactic fermentation [1].

Probiotics are technically defined as live microbial food ingredients that have a beneficial effect on human health. Some of the important beneficial effects are antimicrobial activity, immune system modulation, antitumorigenic activity, colonization resistance activity, maintenance of micro-ecology of bowel, stimulation of *Bifidobacteria*, deactivation of carcinogens etc. Commercially available probiotic strains belong to genera *Lactobacilli*, *Bifidobacterium*, *Streptococcus*, *Bacillus*, *Bacteroides*, *Pediococcus*, *Leuconostoc*, *Propionibacterium* [2-4], *Saccharomyces cerevisiae* and *Aspergillus oryzae* [5].

The other key component of functional dairy or other products is prebiotic ingredient. Prebiotics are non-digestible food ingredients that beneficially affect the host by selectively stimulating the growth and/or activity of one or a limited number of bacteria in colon that can improve the host health [6].

When prebiotics are used in combination with probiotics or live bacteria, the resultant has synergistic effects, referred to as "symbiotic". This is because in addition to the action of probiotics that promote the growth of existing strains of beneficial bacteria in the colon, prebiotics such as inulin and oligofructose also act to improve the survival, implantation and growth of newly added probiotics strains.

Inulin is a mixture of polymers consisting mainly of fructose unit; its partial enzymatic hydrolysis yields oligofructose. These are classified as prebiotic (dietary)

fibres, which help in stabilization of foams, assist in digestion, increase stool volume, stimulate *Bifidobacteria* and are used for formulation of low fat preparations as fat and sugar replacers, respectively [7-9].

As a prebiotic, honey contains carbohydrates called oligosaccharides, which may improve gastrointestinal health by stimulating the growth of good bacteria in the colon. Honey has been shown to enhance growth, activity of *Bifidobacteria* in fermented dairy food [10, 11]. Current market trends reveal that acidophilus milk could be a suitable vehicle to develop functional dairy foods. By incorporation of variations in terms of flavour, it could be easily used in wide range of nutritional and organoleptic qualities being demanded by consumers of all age, health status, and cultures.

In the food industry, end-products must achieve a compromise between several properties, including sensory, sanitary and technological properties. Among the latter, sensory property is essential because it influences consumer choice and performance. To produce a new product, if formulation of product is achieved automatically, production speed and efficiency can be improved in addition to the increased evaluation accuracy, with an accompanying reduction in production costs [12,13]. Artificial neural network (ANN) could be applied to evaluate its capability in predicting product composition involved in food formulation [14-19].

Scanty information is available on incorporation of prebiotic along with probiotic in acidophilus milk. Incorporation of prebiotic and probiotic for development of functional symbiotic acidophilus milk seem to be an effective area of research. In view of the above, present investigation was undertaken to develop quality of symbiotic acidophilus milk from toned cow's milk, by use of ANN technique.

## 2. MATERIALS AND METHODS

### 2.1. Materials and Chemicals

The freeze-dried cultures of *Lactobacillus acidophilus* NCDC No 015, *Bifidobacterium bifidum* NCDC No 255 and *Lactobacillus casei* NCDC No 063 were obtained from Culture Collection Centre, National Dairy Research Institute, Karnal (Haryana), India. Biogoghurt culture (*L. bulgaricus* and *S. thermophilus*) was obtained from K.C. Das Pvt. Ltd. (Bangalore). Inulin was obtained from Nutraingredients, Netherlands. Pasteurized toned cow's milk (3% fat and 8.5% Solid Non Fat), dietary fibre (oat), honey (Dabur India Ltd.), skim milk powder (Amul), polyethylene cups and aluminum foil were purchased from the local market. The chemicals and media used in the present investigation were of Analytical Reagent grade.

### 2.2. Activation of Cultures

The freeze-dried cultures were activated according to the recommendations of suppliers and grown in sterile skim milk at  $37 \pm 1^\circ\text{C}$  and then maintained by weekly transfers and stored at  $4 \pm 1^\circ\text{C}$  between transfers. These cultures were activated by sub culturing 3-4 times before use.

### 2.3. Preparation of Symbiotic Acidophilus Milk

Optimized level of skim milk powder (3% wt./v) based on sensory evaluation was added to pasteurized toned milk and reheated to at  $40\text{-}45^\circ\text{C}$ . Prebiotics were added at optimized level based on sensory evaluation namely inulin (10% wt./v), oat fibre (0.2% wt./v) and honey (7% wt./v). The milk was inoculated with starter culture (*L. acidophilus*) as a control, and other probiotics namely *L. casei*, *B. bifidum*, *L. bulgaricus*, *S. thermophilus* singly or in combination with same ratio (1:1 and 1:1:1) at 7 percent v./v., and then incubated at  $40^\circ\text{C}$  for 4-4½ hours. Different type of symbiotic acidophilus milk thus prepared were cooled to  $5^\circ\text{C}$  and stored for about 2 hours for inducing cold gelation, then stirred for better homogenous consistency. Thus, obtained symbiotic acidophilus milk samples were tested for pH, TA, microbial analysis and sensory evaluation.

### 2.4. Measurement of TA and pH of Samples

pH of the symbiotic acidophilus milk was measured directly using the digital pH meter (Digisun Electronics, Hyderabad). TA (as % lactic acid) of the samples was determined according to Ranganna [20].

### 2.5. Total Viable Count

1 ml of sample was diluted in 9 ml distilled water and then serial dilutions were prepared. Different dilutions ( $10^{-6}$ ,  $10^{-7}$  and  $10^{-8}$ ) were used to check total viable count present per ml on DE Man, Rogosa, Sharpe and All purpose medium with tween so agar (Hi-Media Laboratories Pvt. Ltd., Mumbai). After incubation the average count of colonies present on Petri plates were multiplied by dilution factor and expressed as colony forming units (cfu) per ml.

### 2.6. Yeast & Mold and Coliform Count

Potato dextrose agar and Acetamide media (Hi-Media Laboratories Pvt. Ltd., Mumbai) were used to enumerate the count of yeast & mold and coliform count, respectively. After incubation at  $22 \pm 1^\circ\text{C}$  for about a week and  $37^\circ\text{C}$  for 24-48 hrs, respectively, the average count of colonies present on petri plates, if any, were multiplied by dilution factor and expressed as colony forming units (cfu) per ml.

## 2.7. Sensory Evaluation

Synbiotic acidophilus milk samples were evaluated for their sensory characteristics namely colour, flavour, body texture and overall acceptability (OA) using semi-trained sensory panel consisting of 10 judges drawn from faculty members of Brindavan college Banglor. The judges were requested to record their degree of liking/disliking on a scorer and using hedonic scale ranging from 1 to 9, where 1 represented dislike extremely and 9 represented like extremely [20]. The samples were served to panelist randomly.

## 2.8. Hybrid Analytical Approach

An artificial neural network is usually defined as a network composed of a large number of processors (neurons) that are massively interconnected, operate in parallel, and learn from experience (examples). The ANN used in this research is a feed forward network that can be used to estimate a vector X from a measured vector V. The ANN “learns” by adjusting the interconnection weights between layers. The answers produced by the network are repeatedly compared with the correct answers, and each time the connecting weights are adjusted slightly in the direction of the correct answers. Eventually, if the problem is learned, a stable set of weights adaptively evolves which will provide good answers for all of the sample predictions. When a new vector V is entered into the network, it is subtracted from the stored vectors representing cluster centers. The squares of the differences are summed and fed into a non-linear activation function to recognize the pattern which is most similar to the entered one [21].

Neural network (NN) approach is an effective method for generating new combination of input vector PP. Because of inherent parallel structure, a multi layer ANN can be trained for generating new combinations of prebiotics and probiotics in a very short running time. The Training set is  $[(PP, UG), \quad PP \in R^n, UG \in R^4]$ , where PP is a sample random vector for the prebiotics and probiotics based on fractional factorial experiment. Generating new combinations by neural networks helps to approximate UG according to a given PP.

With a sigmoid activation function, a multi layer NN  $E[UG | PP]$  is trained and new interpolation combination can be obtained.

The Model  $OA \approx f(PP) + \varepsilon$  is a nonlinear equation that has been approximated by quadratic equation as given in Formula (1).

$$OA = \beta_0 + \sum_{i=1}^P \beta_i PP_i + \sum_{i=1}^P \sum_{r=i}^P \beta_{ir} PP_i PP_r + \varepsilon \quad (1)$$

where  $\beta_0$  is equal to the mean of OA and  $\beta_i$  and  $\beta_{ir}$  are called linear and quadratic coefficients, respect-

tively. Moreover, the expected value of  $\varepsilon$  and OA to PP can be recognized by Eqs.2 and 3 respectively:

$$E(\varepsilon | \overrightarrow{PP}) = 0 \quad (2)$$

$$E[OA | \overrightarrow{PP}] = f(\overrightarrow{PP}) \quad (3)$$

For obtaining the parameters of Formula (1), we minimize squared error by the following equations.

$$\varepsilon_j = OA_j - [\beta_0 + \sum_{i=1}^P \beta_i PP_{ij} + \sum_{i=1}^P \sum_{r=i}^P \beta_{ir} PP_{ij} PP_{jr}] \quad (4)$$

$$S(\beta) = \sum_{j=1}^n \varepsilon_j^2 = \sum_{j=1}^n \left( OA_j - \left[ \beta_0 + \sum_{i=1}^P \beta_i PP_{ij} + \sum_{i=1}^P \sum_{r=i}^P \beta_{ir} PP_{ij} PP_{jr} \right] \right)^2 \quad (5)$$

By using  $\frac{\partial S(\beta)}{\partial \beta} = 0$ , the corresponding parameters can be determined.

For higher polynomial degree of Eq.1, we used hypothesis test as follows:

$$\begin{cases} H_0 : \overrightarrow{\beta} = 0 \\ H_1 : \overrightarrow{\beta} \neq 0 \end{cases} \quad (6)$$

The t-test has been used to test the degree of nonlinear equation model. For validity of approved model, we apply F-test for the result of actual and approximated TD as Formula (7) by F statistic in Formula (8).

$$\begin{cases} H_0 : F_0 = F \\ H_1 : F_0 \neq F \end{cases} \quad (7)$$

$$F = \frac{SS_R / P_p}{SS_E / (n - P_p - 1)} \quad (8)$$

where  $SS_R$  is sum of squared regression results and  $SS_E$  is the sum of squared error while  $\frac{SS_R}{\sigma^2} \approx \chi^2_{P_p}$  and  $\frac{SS_E}{\sigma^2} \approx \chi^2_{(n - P_p - 1)}$ . Hence the F statistic has a Fisher distribution with safety interval of  $\alpha$ , and  $P_p$  and  $(n - P_p - 1)$  degree of freedom ( $F_{\alpha, P_p, (n - P_p - 1)}$ ).

If  $F_0 \geq F_{\alpha, P_p, (n - P_p - 1)}$ , then  $H_0$  is rejected and the model would be valid, otherwise the model should be modified.

## 3. RESULTS AND DISCUSSIONS

### 3.1. Changes in TA and pH

The pH and TA for samples is shown in Table 1. They remained unaffected by the type of additive used but the type of culture combination altered them. Maximum acidity and lowest pH were observed in case of C<sub>1</sub> culture (*Lactobacillus acidophilus*), followed by C<sub>2</sub> cultures (*Lactobacillus acidophilus* + *Bifidobacterium bifidum*), C<sub>3</sub> cultures (*Lactobacillus acidophilus* + *Lactobacillus*

**Table 1.** pH, TA and sensory score for colour, flavour, texture and OA of samples.

Index	C <sub>1</sub>	A <sub>1</sub>	B <sub>1</sub>	L <sub>1</sub>	C <sub>2</sub>	A <sub>2</sub>	B <sub>2</sub>	L <sub>2</sub>	C <sub>3</sub>	A <sub>3</sub>	B <sub>3</sub>	L <sub>3</sub>	C <sub>4</sub>	A <sub>4</sub>	B <sub>4</sub>	L <sub>4</sub>	C <sub>5</sub>	A <sub>5</sub>	B <sub>5</sub>	L <sub>5</sub>															
pH	4.70	4.70	4.70	4.70	4.73	4.73	4.73	4.73	4.79	4.79	4.79	4.79	4.81	4.81	4.81	4.81	4.84	4.84	4.84	4.84															
TA	1.23	1.23	1.23	1.23	1.20	1.20	1.20	1.20	1.14	1.14	1.14	1.14	1.00	1.00	1.00	1.00	0.88	0.88	0.88	0.88															
Colour	7	7	8	8	7	7.5	8	7	7	7	8	7	7	8	8	8	7	6	6	6															
Flavour	7	7.5	8	8	8	7.25	8	7	7	7.5	7.5	7	7.5	7.5	8	8.5	6	5.5	6	5.75															
Texture	6	7	8	8	8	7.5	7	7	6	7.5	7.5	7	7.5	7.5	8	8	6	5	5.5	5.75															
OA	7	8	8	8	8	7.25	7.25	7	7	7	7.5	7	7.5	7.5	8	8.5	5	6	6	6															
Combination	C <sub>1</sub> +Oat (0.2%): A <sub>1</sub> C <sub>2</sub> +Oat (0.2%): A <sub>2</sub> C <sub>3</sub> +Oat (0.2%): A <sub>3</sub> C <sub>4</sub> +Oat (0.2%): A <sub>4</sub> C <sub>5</sub> +Oat (0.2%): A <sub>5</sub>					C <sub>1</sub> +Inulin(10%): B <sub>1</sub> C <sub>2</sub> +Inulin (10%): B <sub>2</sub> C <sub>3</sub> +Inulin (10%): B <sub>3</sub> C <sub>4</sub> +Inulin (10%): B <sub>4</sub> C <sub>5</sub> +Inulin (10%): B <sub>5</sub>					C <sub>1</sub> +Honey (7%): L <sub>1</sub> C <sub>2</sub> +Honey (7%): L <sub>2</sub> C <sub>3</sub> +Honey (7%): L <sub>3</sub> C <sub>4</sub> +Honey (7%): L <sub>4</sub> C <sub>5</sub> +Honey (7%): L <sub>5</sub>																								
Probiotic	C <sub>1</sub> : <i>Lactobacillus acidophilus</i> ; C <sub>2</sub> : <i>Lactobacillus acidophilus + Bifidobacterium bifidum</i> ; C <sub>3</sub> : <i>Lactobacillus casei + Lactobacillus acidophilus</i> ; C <sub>4</sub> : <i>Lactobacillus acidophilus + Bifidobacterium bifidum + Lactobacillus casei</i> ; C <sub>5</sub> : <i>Lactobacillus acidophilus + Biyoghurt (Streptococcus thermophilus + Lactobacillus bulgaricus + Bifidobacterium bifidum)</i>																																		
Prebiotic	Inulin, Oat, Honey																																		

*casei*) cultures, C<sub>4</sub> cultures (*Lactobacillus acidophilus + Bifidobacterium bifidum + Lactobacillus casei*) and C<sub>5</sub> cultures (biyoghurt culture). The range of pH and titratable acidity in synbiotic acidophilus milk was 4.70-4.84 and 1.23-0.88, respectively. Lowest TA and highest pH was obtained when C<sub>5</sub> culture was used in preparation of synbiotic acidophilus milk, since it did not show acceptable quality. Similar observations were reported for different type of fermented milk by [20-24].

### 3.2. Changes in TVC, Coliform and Yeast & Mold Count

TVC of all synbiotic acidophilus milk samples obtained ranging from 10<sup>8</sup> to 10<sup>10</sup> cfu/ml, were higher than prescribed range of 10<sup>6</sup> to 10<sup>8</sup> cfu/ml at the end of storage period [25]. It can be inferred that the TVC obtained in synbiotic acidophilus milk samples in present investigation was equal to or higher than recommended range which were reported by [26,27]. No yeast& mold and coliform count appeared inferring that the product developed were of good quality.

### 3.3. Changes in Sensory Score

The sensory score for colour, flavour, texture and OA for samples is shown in **Table 1**. Synbiotic acidophilus milk prepared using starter culture (*Lactobacillus acidophilus*, *B. bifidum* and *L. casei*) and prebiotic additives (oat, inulin, honey) singly or in combination, significantly increased the colour, flavour, texture and OA of the synbiotic acidophilus milk. Addition of inulin (10%) or honey (7%) to C<sub>4</sub> increased the sensory score for colour, flavour, texture and OA of the product developed. From

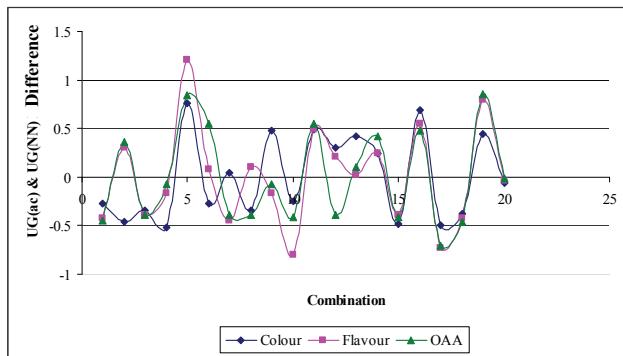
the experimental data, it was found that C<sub>4</sub> culture increased the sensory characteristic of the synbiotic acidophilus milk along with the honey or inulin; however addition of oat fibre did not affect the score of acidophilus milk sample much, and was not found to be as effective as inulin or honey. Thus, addition of inulin led to development of low calorie sweet acidophilus milk which is of value for recommendation to both diabetic and calorie conscious consumers.

A fractional factorial experiment combination of each element by experiment is obtained for the problem as actual UG. The training data (actual UG) are put into a neural network and the output of ANN is obtained as UG (NN). The structure of proposed ANN is as follows: An 8 element vector is considered as input layer of the network which are the most effective factors in the product specifications. And the output layer is included a 4 element vector. For instance, the input vector [1,0,1,0,0,0,0.002,0] indicates that one unit of L.c and one unite of L.a combined with 0.002 unit oat cause the output vector of [7,7.5,7.5,7] which represents a product with the score of 7, 7.5, 7.5, 7 for colour, flavour, texture and OA, respectively, in laboratory samples. Whilst the output of the proposed ANN is [7.3408, 7.3928, 7.1035, 7.3896]. The deviation between the output of NN and the observed actual UG ( $\Delta$ ) is reported in **Table 2**. Moreover the treatment of  $\Delta$  is shown in **Figure 1**. Two layer ANN has been developed to generate fractional factorial experiment of (PP,UG)

Two layer feed forward NN has been trained to generate new fractional factorial experiment of (PP,UG). The

**Table 2.** ANN Error ( $\Delta$ ), Actual and NN Output for different combinations.

Input	Actual Output	NN Output	NN Error
[1;0;0;0;0;0;0]	[7;7;6;7]	[7.2743;7.422;7.0387;7.4512]	[-0.27433;-0.422;-1.0387;-0.4512]
[1;1;0;0;0;0;0]	[7;8;8;8]	[7.4576;7.7008;7.4737;7.6375]	[-0.45762;0.29923;0.52627;0.36247]
[1;0;1;0;0;0;0]	[7;7;6;7]	[7.3409;7.3932;7.1037;7.3898]	[-0.34092;-0.39315;-1.1037;-0.38982]
[1;1;1;0;0;0;0]	[7;7.5;7.5;7.5]	[7.5242;7.6719;7.5386;7.5762]	[-0.5242;-0.17192;-0.038638;-0.07615]
[1;1;0;1;1;0;0]	[7;7.5;7;7.25]	[6.2442;6.298;5.7955;6.4128]	[0.75584;1.202;1.2045;0.83723]
[1;0;0;0;0;0;0.002;0]	[7;7.5;7;8]	[7.2742;7.4217;7.0386;7.451]	[-0.27421;0.078317;-0.038606;0.54898]
[1;1;0;0;0;0;0.002;0]	[7.5;7.25;7.5;7.25]	[7.4575;7.7005;7.4736;7.6373]	[0.042505;-0.45045;0.026408;-0.38735]
[1;0;1;0;0;0;0.002;0]	[7;7.5;7.5;7]	[7.3408;7.3928;7.1035;7.3896]	[-0.34079;0.10716;0.39649;-0.38964]
[1;1;1;0;0;0;0.002;0]	[8;7.5;7.5;7.5]	[7.5241;7.6716;7.5385;7.576]	[0.47592;-0.1716;-0.0385;-0.075966]
[1;1;0;1;1;0;0.002;0]	[6;5.5;5;6]	[6.244;6.2977;5.7953;6.4126]	[-0.24404;-0.79767;-0.79534;-0.41259]
[1;0;0;0;0;0.1;0;0]	[8;8;8;8]	[7.5089;7.5088;7.1325;7.4538]	[0.49114;0.49117;0.86746;0.54621]
[1;1;0;0;0;0.1;0;0]	[8;8;7;7.25]	[7.6921;7.7876;7.5675;7.6401]	[0.30786;0.2124;-0.56753;-0.39012]
[1;0;1;0;0;0.1;0;0]	[8;7;7.5;7.5]	[7.5754;7.48;7.1975;7.3924]	[0.42456;0.020018;0.30255;0.10759]
[1;1;1;0;0;0.1;0;0]	[8;8;8;8]	[7.7587;7.7587;7.6324;7.5787]	[0.24127;0.24125;0.36756;0.42126]
[1;1;0;1;1;0.1;0;0]	[6;6;5.5;6]	[6.4787;6.3848;5.8893;6.4154]	[-0.47868;-0.38481;-0.38928;-0.41536]
[1;0;0;0;0;0;0.07]	[8;8;8;8]	[7.3097;7.4554;7.1605;7.5169]	[0.69029;0.54465;0.83949;0.48314]
[1;1;0;0;0;0;0.07]	[7;7;7;7]	[7.493;7.7341;7.5955;7.7032]	[-0.493;-0.73412;-0.5955;-0.70319]
[1;0;1;0;0;0;0.07]	[7;7;7;7]	[7.3763;7.4265;7.2254;7.4555]	[-0.3763;-0.42651;-0.22542;-0.45548]
[1;1;1;0;0;0;0.07]	[8;8.5;8;8.5]	[7.5596;7.7053;7.6604;7.6418]	[0.44041;0.79473;0.33959;0.85819]
[1;1;0;1;0;0;0.07]	[6;5.75;5.75;6]	[6.0662;5.789;5.7895;6.0184]	[-0.066229;-0.039021;-0.039541;-0.018398]

**Figure 1.**  $\Delta$  Treatment.

block diagram of proposed NN has been illustrated in **Figure 3**. Two non-linear activation functions were dedicated to each layer as Logarithmic and Perlin for first and second layers, respectively. **Table 3** shows the simulated *OA* obtained by ANN for different combinations of probiotic and prebiotic. Block diagram of proposed ANN has been illustrated in **Figure 2**.

By using proposed approach, the equation  $OA = f(L.a, B.b, L.c, L.b, S.t, I, O, H)$  is approximated as follows:

$$\begin{aligned} OA = & 7.829(L.a) - 0.629(L.b) - 0.372(S.t) \\ & - 4428.58(O)^2 + 184.60(B.b)(O) \\ & + 4.80(L.c)(I) - 99.72(L.b)(O) - 5.14(S.t)(H) \end{aligned}$$

**Table 4** indicates the t-test analysis results for the coefficients of the above-mentioned quadratic equation.

With the equation which approximates OA in terms of probiotic and prebiotic, as it is observed from the results reported in **Table 4**, the p-values of  $B.b, I, O, H, I^2, H^2$ , and some of other components are greater than 0.360. This means that with a confidence of %64, these factors are not important in approximating OA and can be safely removed from further consideration.

For optimization, the following quadratic mathematical model has been solved by LINGO 8 software package.

$$\begin{aligned} \text{Max } OA = & 7.829(L.a) - 0.629(L.b) - 0.372(S.t) \\ & - 4428.58(O)^2 + 184.60(B.b)(O) + 4.80(L.c)(I) \\ & - 99.72(L.b)(O) - 5.14(S.t)(H) \end{aligned}$$

$$0 \leq I \leq 0.12$$

$$0 \leq O \leq 0.004$$

$$0 \leq H \leq 0.1$$

$$La, Bb, Lb, St, Lc \in \{0, 1\}$$

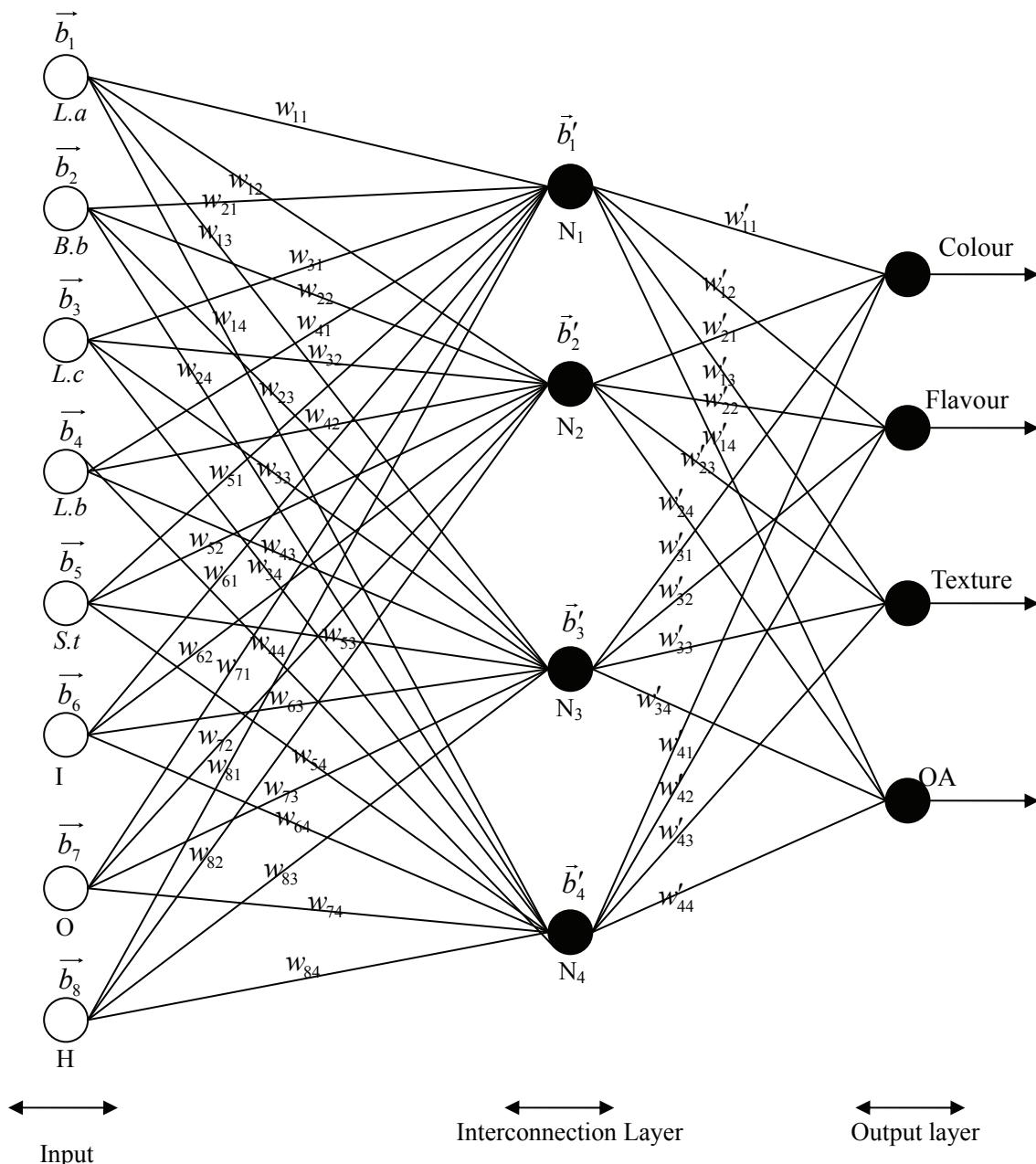
which lead to [1,1,1,0,0,0.09,0.002,0.075] as the optimum combination of prebiotic and probiotic.

#### 4. CONCLUSIONS

From the results of the experiments it can be concluded that the addition of inulin or honey had synergistic effect

on the physico-chemical and sensory quality of probiotic acidophilus milk. Also a satisfactorily good quality symbiotic acidophilus milk could be prepared by fermenting milk with combinations of all cultures ( $C_4$ , *Lactobacillus acidophilus*, *Bifidobacterium bifidum* and *Lactobacillus casei*) along with 7 percent honey (sweetened acidophilus milk) or 10 percent inulin (for low calorie sweetened acidophilus milk). Moreover, in this research an ar-

tificial neural network has been applied to create more experimental data. Then by developing a quadratic mathematical model, the optimum value of OA has been approximated with respect to probiotics and prebiotics, i.e. combination of *Lactobacillus acidophilus*, *Bifidobacterium bifidum* and *Lactobacillus casei* along with 7.5 percent honey, 9 percent inulin and 0.2 percent oat fibre.



**Figure 2.** Block diagram of proposed NN.

**Table 3.** ANN output.

<i>L.a</i>	<i>B.b</i>	<i>L.c</i>	<i>L.b</i>	<i>S.t</i>	I	O	H	OA	<i>L.a</i>	<i>B.b</i>	<i>L.c</i>	<i>L.b</i>	<i>S.t</i>	I	O	H	OA
1	1	0	1	0	0.12	0.003	0.05	6.451636804	1	1	1	1	0	0.08	0	0.05	7.484155
1	0	1	0	1	0.12	0.004	0.07	6.101418984	1	1	1	0	0	0.12	0.001	0.08	8.346724
1	0	0	1	1	0.06	0.004	0.01	5.751022455	1	1	0	0	0	0.02	0.003	0.02	7.528115
1	0	1	0	0	0.02	0.001	0.06	6.984832124	1	1	0	0	1	0.06	0.001	0.03	7.535261
1	0	0	0	1	0.06	0.004	0.05	6.435936602	1	0	0	1	0	0	0.002	0.09	6.791768
1	0	1	1	0	0.1	0.003	0	6.149605379	1	1	0	0	1	0.04	0.003	0.1	7.339067
1	1	0	1	1	0.1	0.002	0.03	5.752972025	1	1	1	0	0	0.12	0.002	0.09	8.346724
1	0	0	1	0	0.08	0.001	0.07	6.854626756	1	1	1	0	0	0.02	0.001	0.04	8.154417
1	0	1	1	1	0.06	0.004	0.08	4.65789474	1	1	0	1	0	0.06	0.002	0.04	7.423626
1	1	1	1	0	0.04	0.001	0.05	7.48414076	1	1	0	0	0	0.1	0.003	0.1	7.536114
1	1	0	1	1	0.06	0.003	0.05	5.75163863	1	1	0	0	1	0.1	0.002	0	7.532254
1	0	1	1	1	0.14	0.002	0.09	6.074391253	1	0	1	1	1	0.12	0.001	0.06	6.577422
1	0	1	0	0	0.12	0.003	0.01	7.006773357	1	1	0	1	1	0.08	0.001	0.05	5.757191
1	0	1	0	0	0.1	0.001	0.05	7.362838031	1	1	0	1	0	0.08	0.003	0.04	7.077645
1	0	1	1	0	0.08	0.001	0.04	6.633418746	1	0	1	1	0	0.06	0.002	0.06	6.145
1	0	1	1	1	0.08	0.001	0.04	6.28328269	1	1	1	1	0	0.02	0.003	0.1	7.480165
1	1	1	0	1	0.12	0	0.06	7.536177648	1	1	1	0	0	0.1	0.001	0.01	8.183134
1	0	0	0	1	0	0.003	0.1	6.936950573	1	1	1	1	1	0.14	0	0.04	7.568186
1	1	1	0	0	0.02	0.003	0.03	7.746431306	1	1	0	1	0	0.08	0.003	0.1	6.09158
1	1	1	0	1	0.04	0.002	0.09	7.536085487	1	1	1	0	1	0.12	0	0.09	7.536147
1	1	0	1	1	0.02	0.001	0.06	5.772025264	1	1	1	0	1	0.08	0.003	0.04	7.535996
1	1	0	1	0	0	0	0.03	7.483844669	1	1	0	1	0	0.12	0.001	0.07	7.26907
1	0	0	1	0	0	0.002	0.06	7.207824048	1	0	0	0	0	0.08	0.002	0.01	7.536163
1	0	0	0	1	0	0.001	0.06	7.52845751	1	1	1	1	1	0.1	0.001	0.03	7.549508
1	1	1	0	0	0.06	0.001	0.02	8.056513417	1	1	1	0	0	0.04	0.002	0.05	8.315503
1	1	1	0	0	0.08	0.001	0.08	8.346682	1	0	0	1	0	0.12	0.003	0.01	6.124803383
1	1	1	0	1	0.08	0.003	0.01	7.536134	1	0	0	0	1	0.06	0.003	0.06	6.935897712
1	0	0	0	1	0.12	0.002	0.1	6.198301	1	1	0	0	0	0.04	0.003	0.03	7.535469986
1	0	1	1	0	0.08	0.004	0.06	5.912921	1	1	1	1	1	0.12	0.004	0.01	6.504576323
1	0	0	0	0	0.06	0.002	0.06	7.536133	1	1	1	0	0	0.1	0.003	0.07	8.346595236
1	0	0	1	0	0.08	0.001	0.06	7.011749	1	1	1	1	0	0.06	0.001	0	7.484170893
1	1	0	1	1	0.04	0.002	0.03	5.760403	1	0	1	0	0	0.12	0.001	0.09	7.896582959
1	0	0	0	1	0.1	0.001	0.01	7.521084	1	1	1	0	1	0.06	0.002	0.02	7.536179995
1	0	0	0	1	0.1	0.001	0.03	7.501948	1	1	0	1	1	0.14	0.001	0.01	5.762277224
1	1	1	1	0	0.12	0.002	0.08	7.47813	1	1	1	0	1	0	0.02	0.02	7.536186589
1	1	0	0	1	0.1	0.002	0.05	7.505374	1	1	0	1	0	0.08	0.004	0.09	5.845121956
1	0	0	0	0	0.04	0.003	0.08	7.534859	1	0	0	0	0	0.02	0.002	0.07	7.534845748
1	1	1	0	1	0.12	0.004	0.02	7.534946	1	0	0	0	1	0.08	0.001	0.1	7.234711688
1	1	1	1	1	0.04	0.002	0.02	7.363935	1	1	0	1	1	0.02	0	0.03	6.090998447
1	1	0	0	1	0.12	0.001	0.03	7.531196	1	0	0	1	0	0	0.003	0.05	6.822632654
1	1	0	1	1	0.12	0.001	0.05	5.753154	1	0	0	0	1	0.12	0.001	0.06	7.346438551
1	0	1	1	0	0.06	0.003	0.04	6.112471	1	0	1	0	1	0.12	0.002	0.1	7.210684753
1	0	0	0	1	0.02	0.002	0.01	7.526696	1	1	0	1	0	0.08	0.003	0.08	6.399538417
1	1	0	1	0	0.08	0.004	0.03	6.622472	1	0	0	0	1	0.02	0	0.08	7.530073062
1	1	0	0	0	0.14	0	0.03	7.536974	1	1	0	0	0	0.06	0.002	0.01	7.535503254
1	1	0	0	0	0.1	0.004	0.08	7.53603	1	1	0	0	0	0.02	0.004	0.03	7.533041896
1	1	0	0	0	0.04	0.003	0.1	7.536176	1	1	0	0	1	0.02	0.002	0.06	7.531179784
1	0	1	1	1	0.04	0.003	0.03	5.24282	1	0	1	1	1	0.06	0	0.08	6.651366866
1	1	1	0	1	0.08	0.003	0.02	7.536106	1	0	0	0	0	0.12	0.003	0.03	7.536097338
1	1	0	1	1	0.1	0.003	0.09	5.751219	1	1	0	1	1	0.1	0.002	0.07	5.751497642

**Table 4.** The t-test and P-value analysis results for the coefficients.

Coefficients	<i>B.b</i>	<i>L.b</i>	<i>S.t</i>	I	O	H	<i>I</i> <sup>2</sup>	<i>O</i> <sup>2</sup>	<i>H</i> <sup>2</sup>	IO	IH	OH
T Stat	-0.298	-2.084	-1.247	-0.811	0.158	0.418	0.781	-1.388	0.223	-0.347	-0.813	-0.758
P-value	0.76	0.04	0.21	0.42	0.87	0.67	0.43	0.16	0.82	0.72	0.41	0.45
Coefficients	<i>B.b.I</i>	<i>B.b.O</i>	<i>B.b.H</i>	<i>L.c.I</i>	<i>L.c.O</i>	<i>L.c.H</i>	<i>L.b.I</i>	<i>L.b.O</i>	<i>L.b.H</i>	<i>S.t.I</i>	<i>S.t.O</i>	<i>S.t.H</i>
T Stat	0.477	2.045	0.217	1.974	-0.963	0.442	-0.669	-1.285	-0.661	0.187	0.235	-1.59
P-value	0.63	0.04	0.82	0.05	0.33	0.65	0.5	0.2	0.51	0.85	0.81	0.11

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