

Testosterone Levels Do Not Decline with Age in Healthy Men

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ABSTRACT

Aim: To establish norms for reproductive hormones in healthy males with normal urologic and sexual function, and to assess the change in reproductive hormone levels with respect to age among healthy males. **Methods:** Healthy volunteers aged 18 - 29 (group 1) and 45 - 65 (group 2) were recruited for enrollment in a prospective study. Inclusion criteria comprised normal urinary, ejaculatory, orgasmic, and erectile function, as determined by IPSS, MSHQ, and IEFF-15. Men with history of chronic medical illnesses or chronic medication use were excluded. Fifty men met the study criteria. Each participant underwent physical examination and standardized serum hormone evaluation using ELISA and EIA methods. Statistical analysis was performed using JMP 8.0 software (SAS Institute Inc., Cary, NC), to compare hormone levels between the two groups of men. **Results:** There were 25 men each in groups 1 (mean age 26) and 2 (mean age 51). Overall, 46% were Caucasian, 31% African American, 15% Hispanic, and 8% Asian. There was no statistically significant difference in serum total T, SHBG, E2, or LH between groups 1 and 2. Notably, the 95% CI for serum T levels for both groups of men were considerably higher than commonly accepted lower-end cut-off limits. **Conclusion:** Serum T levels do not vary significantly with age, in otherwise healthy men with normal sexual and urologic function. Furthermore, healthy men have much higher serum total and free T levels than the lower-end reference limits provided by commercial laboratories.

Keywords: Testosterone; Aging; Hypogonadism; Reference Values

1. Introduction

Hypogonadism is a clinical condition characterized by low levels of serum testosterone (T), in association with specific signs and symptoms of low T, which may include physical, sexual, reproductive, and cognitive effects. Hypogonadism affects an estimated two to four million men in the United States; its prevalence increases with age [1]. When hypogonadism occurs in older men, the condition is often called andropause, or androgen deficiency of the aging male [2].

Testosterone supplementation for the treatment of hypogonadism has increased substantially in the United States over the past few years, with an increase of more than 500% in prescription sales of testosterone products since 1993 [3]. Testosterone replacement therapy (TRT) results in a demonstrable improvement in symptoms of hypogonadism, including diminished libido, energy and

well-being, impaired cognition, decreased muscle mass/ strength, anemia, osteoporosis, erectile dysfunction, and visceral obesity [4-13].

Because the diagnosis of hypogonadism requires laboratory confirmation of a low level of testosterone, clinicians rely heavily on laboratory reference ranges to determine whether a patient may be a candidate for TRT. However, at present there is no universally accepted definition of hypogonadism, no universally used assay for the analysis of serum T, and no universally agreed-upon indications for the initiation and use of TRT.

Guidelines from the Endocrine Society maintain that T levels below 300 ng/dL are diagnostic of hypogonadism, while higher levels are normal [14]. Accordingly, the Society only recommends TRT in men with unequivocally low testosterone concentrations and symptoms associated with androgen deficiency [14]. Meanwhile, a consensus statement from the ISA, ISSAM, EAU, EAA, and ASA recommends that T levels above 350 ng/dL do

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not require treatment, while levels below 230 ng/dL do [15]. Yet other authorities emphasize the importance of the symptoms of androgen deficiency over serum biochemistry, suggesting that men may be "hypogonadal" at the tissue level while having a normal T level [16,17].

The confusion stems, in part, from the variability in testosterone assays and their interpretation. Historically, the reference range for serum testosterone reported by scientific publications was based on traditional radio-immunoassay (RIA) methods, and ranged from 300 - 1000 ng/dl (10.4 - 34.7 nmol/liter) [18]. Over the last decade, techniques for testosterone measurement have evolved from RIA, to automated enzyme-linked immunoassays (EIA) available in most laboratories, and liquid chromatography-mass spectroscopy methods in reference laboratories. As a result, widely variable reference ranges have been reported for different assays, spanning lowend clinical detection limits of 170 - 200 ng/dL to upper-end limits of 700 - 800 ng/dL [18]. However, these reference ranges are not specific for healthy men with normal sexual and reproductive function, and are rarely adequately adjusted for age [19,20]. Consequently, an objective comparison of results from different published studies, using different assays, is fraught with error.

The objective of this study was two-fold. Firstly, we aimed to establish norms for reproductive hormones based on a diverse population of healthy males with normal urologic and sexual function. Secondly, we aimed to assess the change in reproductive hormone levels with respect to age, in a cross sectional analysis of healthy men from two different age groups.

2. Materials and Methods

2.1. Study Population

Two groups of healthy male volunteers were recruited for enrollment into this prospective study via informational study flyers distributed amongst New York Presbyterian Hospital-affiliated medical facilities, and advertisements in local newspapers. Group 1, the younger group, consisted of men between the ages of 18 - 29 years, while group 2, the older group, consisted of men aged 45 - 65 years. All individuals who expressed interest in the study were pre-screened by the study coordinator (JK). Individuals with any history of a chronic medical condition or chronic medication use were excluded. Others were invited for enrollment on a first-come first-served basis.

Following pre-screening, all eligible volunteers completed the International Prostate Symptom Score (IPSS), International Index of Erectile Dysfunction (IIEF-15), and Male Sexual Health Questionnaire (MSHQ). All volunteers also underwent a complete physical examination by an attending physician (DAP). Only men with a normal physical exam, and normal erectile, ejaculatory, orgasmic, and voiding function, were included in the study. A total of 50 men were enrolled into the study, divided equally between groups 1 and 2. All study participants were provided a modest inconvenience fee.

Approval for this study was provided by the Weill Cornell Medical College Institutional Review Board.

2.2. Measurement of Hormone Levels

All subjects were scheduled for baseline hormone evaluation between the hours of 7 and 10 a.m. A peripheral blood sample was obtained from each study participant (25 mL), centrifuged for 30 minutes, and the serum transferred to a collection tube. All samples were maintained at -20° C prior to completion of hormonal assays, following which samples were transferred to -80°C. The hormones measured included: total testosterone (tT), free testosterone (fT), sex-hormone binding globulin (SHBG), estradiol (E2), follicle stimulating hormone (FSH), luteinizing hormone (LH), dehydroepiandrosterone (DHEA) and dehydroepiandrosterone sulfate (DHEA-S). All hormones were measured in duplicate, on the day of collection, by the same technician (AB), using commercially available enzyme-linked immunosorbent assays (ELISA) and enzyme immunoassays (EIA).

For each assay, micro plates were read using Spectra-Max 340 PC micro-plate spectrophotometer, controlled by Soft Max Pro data analysis software (Molecular Devices, Sunnyvale, CA), and concentrations calculated using four-parameter curve fit. Absorbance was measured at 450 nm, and dual wavelength correction at 620 nm was used in all assays. Standards and controls provided by manufacturer were included with each run and on each plate.

Biorad Lyphochek Assayed Chemistry Control Bilevel packs (Bio-Rad Laboratories, Hercules, CA) were used for quality control of assays. All measures were performed in duplicates. Samples with results exceeding dynamic range were diluted using charcoal stripped steroids free serum, and re-analyzed. In order to calculate inter-assay variability, ten randomly-selected subjects from each group underwent repeat hormone evaluation per the study protocol, within 6 months of the initiation of the study.

2.3. Statistical Analysis

All patients in group 1 completed the study. Three patients from group 2, aged 56, 57, and 59 years, were unable to complete the study, and were excluded from the final analysis. All results were entered in a study specific database and analyzed using JMP 8.0 statistical software (SAS Institute Inc., Cary, NC), to compare the various hormone levels between the two groups of men using Student's t-test. A p-value < 0.05 was considered statistically significant. GraphPad Prism 5.0 (GraphPad Software Inc., La Jolla, CA) was used to test for normal distribution of hormonal results and to verify the calculated hormone concentrations obtained from SoftMax Pro (Molecular Devices, Sunnyvale, CA).

3. Results

Twenty-five men each were enrolled into group 1 (mean age: 26) and group 2 (mean age 51). Demographic data was available for 48 subjects. Overall, 46% (22/48) of men were Caucasian, 31% (15/48) African American, 15% (7/48) Hispanic, and 8% (4/48) Asian. Ninety-five percent of study subjects identified themselves as being heterosexual.

Details of the manufacturers, catalog numbers, reported reference ranges, minimum and maximum limits of detection, and inter-assay variability for each of the hormone assays used in this study is summarized in **Table 1**. The laboratory co-efficient of variance was $\leq 6.2\%$ for each assay, indicating good reproducibility with limited inter-assay variability.

Serum hormone levels of tT, fT, SHBG, E2, FSH, and LH, DHEA, and DHEA-S in the two groups of men are listed in **Table 2**. There was no statistically significant difference noted in serum tT (766 vs. 599 ng/dL), SHBG (25.1 vs. 32.5 nmol/L), E2 (17.1 vs. 22.7 pg/mL), or LH (5.37 vs. 7.56 mIU/mL) between groups 1 and 2, respectively. Men in group 1 had higher levels of serum fT (74.5 vs. 50.9 pg/mL, p = 0.004), DHEA (11.4 vs. 7.58 ng/mL, p = 0.04), and DHEA-S (2.84 vs. 1.35 ug/mL, p < 0.001), compared to the older cohort of men in group 2. Serum FSH levels were lower in group 1 compared to group 2 (1.97 vs. 5.00 mIU/L, p < 0.001).

Compared to manufacturer-provided reference ranges (**Table 1**), the 95% CI values for serum tT, fT, DHEA, and DHEA-S were higher among the study subjects in both groups 1 and 2 (**Table 2**). Interestingly, the 95% CI range for serum FSH was both lower and tighter among the study subjects, who represent healthy men, than the reference ranges that have traditionally been reported by manufacturers.

4. Discussion

Our results demonstrate that serum testosterone levels are not significantly different in healthy men between the second and sixth decades of life. Furthermore, the range of distribution of serum testosterone levels in the study subjects was considerably higher than what is commonly reported by manufactures of commercially available testosterone assays, indicating that reference ranges for healthy men with normal sexual and urologic function may be different than the references ranges for the general population of men. These findings are in contrast to recent reports from longitudinal studies of aging men, which show that serum testosterone levels decline with each decade of life [21-23]. However, these population-based studies included men over the age of 65, and did not specifically select for healthy men without any medical comorbidities.

To our knowledge, to date, serum reference ranges of reproductive hormones have not been studied in a diverse population of healthy men with documented normal urologic and sexual function. The evaluation of orgasmic and ejaculatory function in healthy men with normal sexual function is difficult because of the ethical issues involved, concerns for privacy and confidentiality, increased scrutiny by internal review boards towards studies of human sexuality, and complex logistical issues involved in screening and enrolling patients for such studies. Furthermore, studies of healthy subjects are expensive, and difficult to obtain funding for, as no clinical benefit of "pharmacological" treatment can be proven in studies of normal subjects. Nevertheless, the importance of such studies, to establish a baseline reference range, cannot be emphasized enough.

Assays for the measurement of serum testosterone have been criticized for their unreliability [24,25], and variable limits of detection. Taieb et al. compared testosterone levels for 116 samples (50 males, 55 females and 11 children) measured by 10 different commercial kits with results obtained by isotope dilution-mass spectrometry, and concluded that none of the immunoassays tested was sufficiently reliable for the investigation of sera from children and women, in whom very low testosterone concentrations are expected [26]. There are significant intra-individual fluctuations in serum testosterone levels, and a wide range of testosterone levels in the general male population. Thus, it is likely that the variability between assays derives in large part from failure to establish specific norms for a healthy, young to middle-aged population of men. Additionally, many manufacturer-reported references ranges assume the serum testosterone follows a normal distribution, which is not, in fact, the case. As a result, manufacturer-quoted reference ranges are often too low, making them less sensitive in detecting hypogonadism [27]. This assertion is certainly supported by our data, which demonstrates a higher range of distribution of serum testosterone levels in the study subjects, compared to published reference ranges (Table 1).

While several cross sectional studies have demonstrated lower concentrations of circulating total and/or free T in older men [28-37], other have shown that T levels do not fall significantly with age in exceptionally healthy men [38,39], questioning the relative roles of age-related illness versus aging per se in producing the

Hormono	Type of Assay	Manufacturer	Catalog No.	Manufacturer Reported	Limits of	Lab Co-efficient
none				Reference Range	Detection	of Variance (%)
Т	EIA	Beckman Coulter; Webster, TX	DSL-10-4000	290 - 990 ng/dL (20 - 30 yr old); 160 - 660 ng/dL (50 - 60 yr old)	100 - 2500 ng/dL	6.2
Free T	EIA	Beckman Coulter; Webster, TX	DSL-10-49100	6.2 - 28.1 pg/mL	0.25 - 100 pg/mL	1.9
LH	ELISA	Beckman Coulter; Webster, TX	DSL-10-4600	1.08 - 8.34 mIU/mL	1 - 100 mIU/mL	3.5
FSH	EIA	ALPCO; Salem, NH	11-FSHHU-E01	1.0 - 18.0 mIU/mL	5 - 100 mIU/mL	2.8
SHBG	EIA	ALPCO; Salem, NH	11-SHBHU-E01	7.0 - 70.0 nmol/L	3.3 - 295 nmol/L	5.1
E2	EIA	ALPCO; Salem, NH	20-DR-4399	10.0 - 36.0 pg/mL	3 - 200 pg/mL	2.8
DHEA	EIA	Beckman Coulter; Webster, TX	DSL-10-9000	1.8 - 12.5 ng/mL	0.2 - 27 ng/mL	1.7
DHEA-S	EIA	ALPCO; Salem, NH	11-DHEHU-E01	0.39 - 4.63 ug/mL	0.005 - 10 ug/mL	2.9

Table 1. Manufacturer reported reference ranges, limits of detection, and laboratory co-efficient of variance for the assays used.

EIA = Enzyme immunoassay; ELISA = Enzyme-linked immunosorbent assay; T = Testosterone; LH = Luteinizing hormone; FSH = Follicle stimulating hormone; SHBG = Sex-hormone binding globulin; E2 = Estradiol; DHEA = Dehydroepiandrosterone; DHEA-S = Dehydroepiandrosterone sulfate.

Table 2. Means and distribution of serum hormone levels (reported as 95% C	CI) in healthy men of different age groups.
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	Group 1 (Age 18 - 29)	Group 2 (Age 45 - 65)	Statistical significance
Total T (95% CI)	766 (569 - 963) ng/dL	599 (497 - 700) ng/dL	p = 0.11
Free T (95% CI)	74.5 (60.6 - 88.4) pg/mL	50.9 (44.2 - 57.6) pg/mL	p = 0.004
SHBG (95% CI)	25.1 (15.8 - 34.4) nmol/L	32.5 (26.4–38.7) nmol/L	p = 0.16
E2 (95% CI)	17.1 (11.4 - 22.7) pg/mL	22.7 (9.67 - 35.8) pg/mL	p = 0.42
FSH (95% CI)	1.97 (1.32 - 2.61) mIU/mL	5.00 (3.67 - 6.34) mIU/mL	p < 0.001
LH (95% CI)	5.37 (2.62 - 8.13) mIU/mL	7.56 (3.42 - 11.7) mIU/mL	p = 0.36
DHEA (95% CI)	11.4 (7.97 - 14.8) ng/mL	7.58 (6.15 - 9.02) ng/mL	p = 0.04
DHEA-S (95% CI)	2.84 (2.26 - 3.42) ug/mL	1.35 (1.04 - 6.66) ug/mL	p < 0.001

observed decreases in serum testosterone. In the present study, which strictly excluded study subjects on the basis of any history of chronic medical illness or medication use, no significant decrease in serum testosterone was noted with increasing age. The utility of age-adjusted norms for serum testosterone levels has long been debated in the literature [27,40,41]. However, the present data suggests that establishing norms based on healthy men may be more important and utile than using age adjusted norms.

Lazarou *et al.* [19] point out that if hypogonadism affects 15% - 35% for men over the age of 50 years [1], but only 2.5% of values are categorized as "low," then a large majority of affected men will fail to be correctly identified as hypogonadal. Men who stand to benefit from TRT may not be offered treatment. The use of age-adjusted values may decrease the test sensitivity for detecting androgen deficiency in aging males [42]. While the use of age-adjusted norms may make sense for the statistical representation of a population, it has no clinical justification for the diagnosis of hypogonadism [19]. Our data lends supports the possibility that the use of norms based on healthy men with normal sexual function may

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optimize clinical care of both older and younger men.

One limitation of this study is the small sample size of healthy volunteers recruited for participation. Validation of our results in a larger and equally diverse population of healthy men would be beneficial. Secondly, the enrollment criteria did not specifically require documented paternity or other evidence of normal reproductive function, such as semen analyses. While normal reproductive function was assumed among the study subjects, based on a normal urologic, medical, and sexual history, confirmation of paternity or normal semen analysis would further strengthen the findings of this study. The gold standard assay for the measurement of serum testosterone is liquid chromatography-mass spectroscopy, used primarily in reference and research laboratories [43]. Nevertheless, for this study, we chose to use EIA and ELISA assays for the measurement of reproductive hormones in our study population, because these methods are commercially available and, therefore, more widely used in investigative laboratories.

In summary, healthy men with normal sexual function have much higher total and free testosterone levels, than the "normal" reference ranges commonly reported by commercial assays. In healthy men, with normal urologic and sexual function, serum testosterone levels do not appear to vary significantly with age. These findings are important for the diagnosis and treatment of hypogonadism in the general male population.

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