Dynamic Mechanical Properties of Tissue after Long-Term Implantation of Collagen and Polypropylene Meshes in Animal Models

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ABSTRACT

Purpose: Pelvic floor reconstructive surgery has grown significantly in recent years. A wide variety of available types of meshes exist but the safety and success has not been adequately proven. We sought to evaluate the effects on dynamic biomechanical properties of tissue after long-term implantation of synthetic and biological grafts. **Methods:** A total of 96 New Zealand white female rabbits (approximately 3 kg) were used, 72 of which were surgically implanted with acellular, collagen mesh (n = 36) or nonabsorbable monofilament polypropylene mesh (n = 36). There was a no mesh-rupture of fascia group (n = 12) and a second, no-mesh, no-fascia rupture control group (n = 12). In the 59 rabbits, of 72 (13 died) tissue was harvested 3 months (n = 24), 6 months (n = 23) and 9 months (n = 12) later, while in the fascia rupture group, tissue was harvested 6 months later. Tissue samples (2×2 cm) underwent dynamic mechanical analysis (DMA) testing during which the dynamic rigidity and tissue damping capacities were measured. The statistical analysis was performed with General Linear Model with Tukeys post hoc testing (sPss v.17.0). **Results:** With respect to mesh type, the rabbit tissue in which polypropylene mesh was used showed the greatest dynamic rigidity. Those with biological mesh delivered the lowest rigidity results, while the two other groups had almost similar behavior. The meshes exhibited their highest relative dynamic tissue stiffening effect at 9 months. **Conclusions:** Biological mesh causes lower tissue rigidity, resulting in inferior mechanical response and thus seems to be inferior to polypropylene.

Keywords: Biomechanical Properties; Tissue Stiffening; Collagen-Polypropylene Meshes

1. Introduction

Disorders of the pelvic floor encompass a wide spectrum of interrelated clinical entities, including pelvic organ prolapse and incontinence. In addition to the physical symptoms that accompany these disorders, there is an important emotional effect, which includes social isolation, anxiety and depression [1]. Pelvic organ prolapse affects almost one third of premenopausal and approximately one half of postmenopausal women [2,3]. Every year, approximately 135,000 women undergo surgery for urinary incontinence [4] and 225,000 for pelvic organ prolapse at a cost of greater than \$1 billion per year in the United States [4,5].

Mechanical failure of the tissues in these anatomical

areas is one of the causes [6] as the muscles and ligaments of the pelvis cannot provide adequate support, for which sufficient connective tissue is paramount [7]. On the other hand, reconstructive procedures for the pelvic floor have increased exponentially in the past 20 years and the recent literature has shown increasing interest in the use of biological and synthetic mesh [8,9]. The principle behind using such grafts in reconstructive surgery is reinforcement of existing tissues with materials that are safe, biologically compatible and provide suitable anatomical and functional results. An ideal mesh must be inert, non-carcinogenic, resistant to tension, sterilizable, non-allergenic and non-inflammatory, unaffected by the body's tissues and of course affordable [10]. In the effort



to find an ideal mesh, biomechanics, *i.e.* the study of mechanical properties of tissue and ligaments, could allow us a better understanding of the normal function of the tissues and of the effect of the treatment being applied.

The goal of this study was the appraisal of the biomechanical properties of rabbit tissues after the implantation of biological and synthetic mesh and their long-term stay in the abdominal rectus muscle.

2. Material and Methods

Permission for this study was obtained from the Veterinary Service of Eastern Attica Prefecture and from the Institutional Review Board of Aretaion Hospital (Reference number: N-52/20-12-2005). A total of 96 white, female New Zealand rabbits weighing 3 - 3.5 kg were used. All established rules regarding the humane handling of animals and animal products were followed and they were physically sheltered at the Experimental Research Center of ELPEN. The rabbits, in which mesh was placed, were divided into two groups. In the first group (n = 36)porous collagen mesh (Pelvisoft Acellular Collagen Biomesh C. R. Bard Inc, code number 481812) was placed into the rectus muscle, while in the second group (n = 36)it was a monofilament polypropylene mesh (Avaulta Solo Synthetic mesh C. R. Bard Inc, code number 486100). The meshes measuring 2×2 cm were sutured tensionfree with four 5 - 0 Prolene stitches. Subsequently, each group was divided into subsets of 12 each and sacrificed at 3, 6 and 9 months (post-operative) respectively. There were also two other groups; in one (n = 12) there was only surgical division of the fascia of the rectus abdominis muscle without any mesh placement (fascia rupture), while in the other (n = 12) neither mesh was placed nor surgical division of the fascia was done (control group). These groups (fascia rupture and control group) were sacrificed at 6 months and 0 months respectively (Table 1).

Subsequently, a 2×2 cm sample of tissue in the region of the mesh placement (without the mesh) or fasciotomy was removed and then preserved at -80° C until the biomechanical study with the Dynamic Mechanical Analyzer, DMA Q800 (by TA Instruments). The tissues were isothermally maintained at 37° C and were subjected to sine-wave dynamic tension under a spectrum of low

Table 1. Time period and number of rabbits being sacrificed from the 4 different groups.

Four groups/ sacrifice time (months)	0 month	3 months	6 months	9 months
1. Collagen mesh		12 rabbits	12 rabbits	6 rabbits
2. polypropylene mesh		12 rabbits	11 rabbits	6 rabbits
3. Fascia rupture			12 rabbits	
4. No mesh-no rupture	12 rabbits			

frequencies (0.1 - 15 Hz). DMA testing allows materials to be tested within ranges of frequencies which correspond to the actual loading conditions during their operation. Therefore frequencies of 0.1 - 15 Hz were used in order to simulate actual loading of tissue under static, e.g. standing (low frequencies in the area of 0.1 - 1 Hz) or dynamic conditions like running, jumping etc (frequencies > 1 - 15 Hz). Tissue was loaded under such strain and stress levels so as to correspond to linear elastic viscoelastic regime. The static force (preload) was 0.3 Newton and the oscillation amplitude 80 micrometers (=0.08 mm). The main three viscoelastic parameters were recorded, *i.e.* the capacity of the material to store elastic energy (storage modulus) or else its dynamic stiffness, the loss of elastic energy per load cycle (loss modulus) and the ability of the material to absorb oscillation and impact $(\tan \delta)$. For each group three different measurements were made by dividing each specimen into three equal parts (Number of cycles: 4 until data stabilization set by the DMA test set-up). The rationale behind employing DMA is the great sensitivity of the method to detect changes and the ability of the instrument to impose dynamic loading on the tissue specimens at various strains and frequencies.

The statistical analysis was performed with General Linear Model with Tukeys post hoc testing (sPss v.17.0). Differences were considered statistically significant when they reached a P value of <0.05 (P < 0.05).

3. Results

During the study, 10 rabbits showed mild subcutaneous inflammation (3 with biological mesh, 7 with synthetic), which was successfully treated with antibiotics, whereas 13 rabbits died and were excluded from analysis. In those 13 rabbits biological mesh had been placed in 6 of them and they belonged to the group to be sacrificed at 9 months. Polypropylene mesh had been placed in the remaining 7, six of which were to be sacrificed at 9 months, and one at 6 months. As for the 13 excluded participants, a possible connection to the mesh arose in only two (a subcutaneous tumor and an abscess, with biological and synthetic mesh respectively) on autopsy.

Considering the independent variables of the viscoelastic parameters (type of mesh, time, frequency), it became evident that primarily the type of mesh, and also time, affected the biomechanical properties of the tissues to a statistically significant degree (**Table 2**).

The tissue from rabbits that received polypropylene mesh showed the greatest dynamic stiffness (storage moduli). This implies that this tissue is resisting straining at a higher level than other samples, or in plain words that the corresponding specimens exhibited the highest rigidity. The collagen mesh showed the least rigidity among the four groups (synthetic mesh, biological mesh, fascia

 Table 2. Percentage of total variation explained by correlation factor (%).

s Tan δ
35.13b
0.98b
-
3.55a
52.53b

(-) Not statistically significant impact. Levels: a: 0.05 Significance, b: 0.001 Significance. Storage modulus: The capacity of the material to store elastic energy. Loss modulus: The loss of elastic energy per load cycle. Tan δ : The ability of the material to absorb oscillation and impact.

rupture, control group), while the 2 groups, in which no mesh was placed, showed no significant difference between them (**Table 3** and **Figure 1**). The greatest stiffening of tissue, irrespectively of mesh type, was achieved at 9 months (**Table 3** and **Figure 2**). At 3 and 6 months, the increase in dynamic stiffness for the three groups (biological mesh, synthetic mesh, fasciotomy) was not statistically significant.

4. Discussion

It is known that pelvic floor reconstructive surgery has grown significantly in recent years and several studies have been performed trying to determine which mesh is most suitable for implantation. These have been in animal models and focused on the dynamic properties of the mesh itself and not the tissue. In this study, the biomechanical properties of the tissue from rabbits were assessed, after the use of biological and synthetic mesh, and it appears that synthetic mesh is superior, as it provokes tissue stiffening to a greater degree. This is supported in the current literature with randomized controlled studies [10], which have stood the test of time.

Deprest *et al.* [11] found that collagen mesh causes less inflammatory reaction, delayed deposition of collagen and slower strengthening of tension than synthetic mesh. They believed this difference was due to polypropylene mesh being porous, whereas collagen mesh is not. In a more recent study of theirs [12] they placed such pores preoperatively in the biological mesh and observed more rapid neovascularization, deposition of collagen and strengthening of tissue. In our study, porous biological mesh was used, which proved inferior to the synthetic one in reinforcing the tissue of the animal model, based on our results.

In an effort to find a mesh that would be associated with fewer complications, such as erosion, infection and pain, as compared to synthetic ones, several clinical studies have been carried out regarding pelvic floor surgery. Altman *et al.* [13] reported that there were no complications when using biological mesh, Quiroz *et al.* [14] reported a complication rate of about 24% with collagen

	Sample type	Meana	Std. Error	Indw. Time [months]	Meana	Std. Error
Storage Modulus [MPa]	1	0.134	0.048	0	0.373	0.071
	2	0.933	0.049	3	0.520	0.051
	3	0.551	0.073	6	0.441	0.042
	4	0.373	0.071	9	0.695	0.073
Loss Modulus [MPa]	1	0.025	0.010	0	0.076	0.014
	2	0.208	0.010	3	0.109	0.010
	3	0.109	0.015	6	0.097	0.008
	4	0.076	0.014	9	0.151	0.015
Tan∂	1	0.141	0.004	0	0.154	0.006
	2	0.410	0.004	3	0.246	0.004
	3	0.262	0.006	6	0.280	0.004
	4	0.154	0.006	9	0.291	0.006

Table 3. Estimated marginal means for DMA parameters

with respect to mesh type and indwell time.

Sample type: 1. Collagen mesh; 2. Polypropylene mesh; 3. Fascia rupture; 4. No mesh-no rupture. MPa: megapascal [=1 N/mm²].



Figure 1. Dynamic stiffness related to 4 groups.



Figure 2. Dynamic stiffness related to time (irrespectively of mesh type).

mesh, while Deprest *et al.* [15] observed that biological mesh not only does not reduce complications compared to synthetic, but also has high, almost unacceptable rates of failure. In our study, mild subcutaneous inflammation was observed in 10 rabbits, which was treated successfully with antibiotics, while in the 13 subjects that died and were excluded from the study, there was a possible association with the mesh noted only in two (a subcutaneous tumor and an abscess) on autopsy.

In our study, the collagen mesh was inferior in tissue reinforcement, not only compared to the synthetic mesh, but also compared to the two groups in which no mesh was placed. This paradoxical finding may be explained by the fact that collagen mesh has been proven to undergo, to varying degrees, premature decomposition, for reasons unclear to date [10,16-18], and this might lead to a reduction in its biomechanical properties. Another explanation may be that the higher concentration of collagen in the area might decrease the micromechanical properties of the surrounding tissue.

There are limitations in our study as well. The biomechanical properties of the tissues were observed under dynamic loading, which does not occur to such a degree (frequencies up to 15 Hz) under normal conditions and therefore in future studies, the range of physiological tension of the vaginal wall must be considered [16,19-21]. In addition, rabbits exhibit increased collagenolytic activity and thereby perhaps cause more rapid decomposition of collagen mesh as compared to humans [16,17], although there is a study in the literature reporting similar decomposition of collagen mesh in women [18].

In conclusion, despite a wide variety of available types of mesh, the safety and success of any one of these has not been adequately proven. Further studies are needed to evaluate these parameters in order to show which mesh is ideal. We believe that our study contributes to better evaluating the biomechanical properties of mesh and that this knowledge may assist in the improved design of future studies.

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