

Adsorption and Leaching Potential of Imidacloprid Pesticide through Alluvial Soil

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ABSTRACT

This study was aimed to assess the adsorption and leaching potential of imidacloprid pesticide in column and field soil. To understand mechanism of adsorption and factors affecting, the experiments were carried out under laboratory and field conditions. Adsorption study was divided into kinetic and equilibrium sections. The evaluation of kinetic data was done through pseudo first and second order models. It was found that kinetic adsorption of imidacloprid on soil followed pseudo second order with rate constant value of 4.333 mg/g/h. Langmuir and Freundlich isotherms were used to explain equilibrium adsorption, from these isotherms it was evaluated that Freundlich isotherm was obeyed well with adsorption capacity of 2.190 - 4.573 mol/g. Leaching study was performed in laboratory using column made of polyvinyl chloride having 30 cm length. Known amount of imidacloprid pesticide was applied to column left for adsorption and then eluted with 500 mL water in five equal portions. These water portions and soil of column which was divided into three sections were analyzed by HPLC. The result revealed that the concentration of imidacloprid was decreased from 0.481 ppm in first portion of water to 0.327 ppm in last portion of water while 0.783 ppm in first section of column soil to 0.038 ppm in last section of column soil. In field the leaching power of imidacloprid was observed up to 60 cm depth, its concentration decreased with soil depth. It was 3.311 ppm in first portion of soil and 0.357 ppm in last portion of soil. The leaching potential of imidacloprid pesticide up to 60 cm soil depth was due to less organic matter, sandy texture, alkaline pH, and low cation exchange capacity.

Keywords: Pesticide; Adsorption; Leaching; Imidacloprid; Soil

1. Introduction

The utilization and production of pesticides are increasing worldwide day by day. It is important to know that when pesticides are applied to field, as the only small portion reaches to its target and remaining a large part is released into the environment. That may leads to some problems, such as toxicity to non-target organisms, leaching and accumulation. Polluted soil, surface and ground waters involve risk to the environment and also to human health due to possible direct or indirect exposures.

Imidacloprid, 1-(6-chloro-3-pyridinylmethyl)-N-nitroimidazolidin-2-ylideneamine, is a systemic chloronicotinoid insecticide which is used for soils, seeds and foliar applications for the control of sucking insects, including rice hoppers, aphids, thrips, whiteflies, termites, turf insects, soil insects, chewing insects, some beetles, scales, psyllids, plant bugs, and other various harmful pest species including resistant strains. It is most commonly used on rice, cereal, maize, sunflowers, potatoes and vegeta-

bles. It is applied as a seed and soil treatment, crop and structural treatment, and a topical flea control treatment on domestic pets [1-5]. It is also used for animal health applications to control parasites like fleas of dogs and cats [6]. It kills insects via ingestion or contact by disrupting the nervous system of an insect pest through interfering with the transmission of stimuli in the insect's nervous system. This pesticide causes an obstruction in the nicotineric neuronal pathway which is more abundant in insects than in warm-blooded animals, making the pesticide much more toxic to insects than to warm-blooded animals. It is sold in market with various brand names such as Pestidor, Concord, Pencidor, Ningo, Imicon, Confidor, Crown, Launcher, Jozer, Simida and Agro-tech.

The environmental fate of pesticides is mainly regulated by their behaviour in soil where various physico-chemical and biological processes control their dissipation and movement towards other environmental compartments like air, water and biota [7,8]. The mobility of a pesticide in soil is determined by the extent and strength

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of sorption, which is influenced by various soil physicochemical properties [9,10]. Sorption is one of the most important processes that affects the fate of pesticides in the soil and determines their distribution in the soil/water environment, it is widely used to describe the process of a pesticide partitioning between water solution and soil [11]. Sorption also determines availability of pesticides in the soil solution that governs the amount of pesticide that is available for uptake by plants and the effectiveness of pesticides [12]. It is reported that adsorption is one of the most important processes which controls all other processes such as their movement, persistence and degradation and determines the fate of pesticides in soil systems [13]. Extensive research has been done on imidacloprid in relation to its mobility and leaching potential in soils [14-16].

But above studies have been conducted in acidic soils whereas Sindh soils are sodic and alkaline in nature. There was no any such studies have been conducted on its adsorption behavior in this type of soil. The present study was undertaken to evaluate the adsorption, and leaching of imidacloprid in sodic and alkaline soil.

2. Materials and Methods

2.1. Chemicals

Imidacloprid standard of high purity (99.4%) was obtained from Bayer Crop Science and commercial imidacloprid was purchased from local market Hyderabad-Sindh, Pakistan. Imidacloprid stock solution was prepared in acetonitrile and other working solutions were prepared by diluting the stock solution.

2.2. Study Area

The study area was selected at the field of National Centre of Excellence in Analytical Chemistry, University of Sindh, Jamshoro, Sindh-Pakistan. Okra crop was sown in first week of November 2010 on the experimental field; plot size was 60 × 60 feet. One month after sowing okra was sprayed with imidacloprid at the recommended dose. Subsequently imidacloprid was sprayed two times per week and this makes a total of sixteen sprays until final harvest.

2.3. Soil Sampling

For laboratory study the inert soil samples were collected from the 0 to 15 cm soil depth, air-dried, and ground to pass through a 2 mm sieve. These soil samples were stored in plastic bags at room temperature. The standard methods were used to determine physicochemical properties of the soil, which are given in **Table 1**. The pH of soil was determined in a 1:1.25 soil to water suspension using a glass electrode, organic matter by Walkley-Black

Table 1. Physicochemical properties of the soil.

| Parameter | Value |
|--------------------------------------|---------------|
| pH | 8.46 ± 0.01 |
| Electrical Conductivity (µS/cm) | 560 ± 0.25 |
| Cation Exchange Capacity (meq/100 g) | 18 ± 0.21 |
| Organic Matter (%) | 0.899 ± 0.01 |
| Total Organic Carbon (%) | 0.522 ± 0.001 |
| Sand (%) | 74.75 ± 0.68 |
| Silt (%) | 18.15 ± 0.24 |
| Clay (%) | 7.1 ± 0.17 |
| Classification | Sandy Loam |

method and soil texture by hydrometer method.

For field study sampling was done from four locations, selected at random, of okra plot. From each sampling location, four soil samples were collected; first soil sample was collected from 0 to 10 cm, second from 11 to 20 cm, third from 21 to 30 cm and fourth from 31 to 60 cm depth. These samples were properly labeled and stored in plastic bags in a freezer at -10°C until extraction. In laboratory study, the experiment was carried out on leaching of pesticide through column and adsorption of pesticide on soil. While in field study the pesticide residue and leaching of pesticide in soil was known.

2.4. Adsorption Study

2.4.1. Kinetic Adsorption Study

For kinetic adsorption study, 5 g of soil was mixed with 100 mL of 100 ppm standard imidacloprid solution in a 250 mL conical flask. This mixture was shaken on an orbital shaker at the speed of 150 rpm for a period of 24 hours at room temperature. 5 mL of sample was collected from flask at time intervals of 0, 0.25, 0.5, 1, 2, 4, 8, 16 and 24 hours. The collected samples were extracted with 10 mL of methylene chloride solution in separating funnel. The water layer was decanted carefully and the supernatant was extracted with 5 mL of methylene chloride for two more times. Finally, extracted samples were filtered and analyzed for residual pesticide concentration using HPLC.

2.4.2. Equilibrium Adsorption Study

For equilibrium adsorption study, 5 g of soil was mixed with different imidacloprid concentrations as 0.15, 0.25, 1, 2, 5, 10, 25, 50 and 100 ppm in 250 mL conical flasks. Then reaction mixtures were shaken on an orbital shaker at the speed of 150 rpm for 6 hours (equilibrium time) at room temperature. After 6 hours, 5 mL of sample was taken from each conical flask and centrifuged at 3000 rpm for 15 min; the supernatant was extracted with methylene chloride (as described above). Extracted samples

were filtered and analyzed by HPLC. Same way a blank solution was also prepared to know the effect of imidacloprid adsorption on walls of conical flask.

2.5. Column Leaching Study

The column (30-cm length, 6-cm i.d.) was made from polyvinyl chloride pipe, fitted with 0.60 μm nylon membrane. To prevent the soil loss from column to outside, wool was placed on nylon membrane. On first day column was filled with soil, soaked with distilled water and left for overnight. Next day imidacloprid (5 mg) dissolved in 5 mL acetonitrile was applied to the surface of column and left for 24 hours. Then column was eluted with distilled water of 500 mL quantity at the flow rate of 100 mL/h with the help of peristaltic pump into five portions of 100 mL each. After elution was over the soil of column was divided into three sections of 10 cm each to determine adsorbed pesticide on soil.

2.6. Extraction of Soil and Water Samples

50 g of air dried soil was mixed with 50 mL acetonitrile in a 250 mL stoppered flask. The sample was equilibrated on a rotary shaker for 2 hour and the extract was filtered. Each 100 mL water portion eluted from column was mixed with 50 mL methylene chloride in a 250 mL separating funnel. The separating funnel was shaken vigorously for 5 minutes to ensure the complete mixing of two layers. After separation of two layers, methylene chloride layer containing pesticide residue was collected. These extracts (soil and water) were concentrated to 5 mL with stream of nitrogen and analyzed with HPLC instrument to know pesticide concentration present.

2.7. Analytical Technique

Analysis of imidacloprid was performed by HPLC Hitachi model L-6200 with UV-Vis detector model Hitachi L-4200. The selected wavelength was 254 nm. The solvent compositions were acetonitrile and water as 70:30 using reverse phase C18 inertsil ODS-3 (250 mm \times 4.6 mm) column with flow rate of 1 mL \cdot min⁻¹ and injection volume was 20 μL . The retention time for imidacloprid was 5.22 min.

2.8. Percent Recovery

Blank soil of 50 g weight was spiked with 0.1, 0.5 and 1 ppm imidacloprid standard solutions. The soil was mixed thoroughly for even distribution of pesticide and left for overnight so that the pesticide can be adsorbed completely. Then imidacloprid was extracted and analyzed as above. For calibration the imidacloprid standards from 0.01 to 10 ppm concentration were run on HPLC. The percent recoveries of imidacloprid found in soil and wa-

ter were 95.33% \pm 1.2% and 80.15% \pm 1.09% respectively.

2.9. Limits of Detection and Quantification

Limit of detection (LOD) of imidacloprid was calculated at a signal-to-signal ratio of 3, while the limit of quantification (LOQ) was obtained at a signal-to-signal ratio of 10. The LOD and LOQ for imidacloprid were 0.4 \pm 0.002 $\mu\text{g}/\text{kg}$ and 5 \pm 0.06 $\mu\text{g}/\text{kg}$ respectively.

3. Results and Discussion

3.1. Affect of Physicochemical Properties of Soil on Adsorption and Leaching of Imidacloprid Pesticide

Sorption is one of the most important processes affecting the leaching of pesticides through soil because it controls the amount of pesticide available for transport. Physicochemical properties of soil can significantly affect pesticide transport and the potential for groundwater contamination [17]. These properties include soil texture, organic matter, pH, cation exchange capacity, electrical conductance and moisture. The pesticide is more adsorbed to soil containing higher content of clay in its texture because the clay possesses a larger surface area [18,19]. As the texture of our soil was sandy loam containing 74.75% sand, so adsorption of imidacloprid was less because sand particles have less surface area than clay or slit. Therefore higher leaching power of imidacloprid was seen in column and field study. In column experiment the imidacloprid was adsorbed on soil filled in column as well as leached with water by passing through 30 cm column length. While in field study the imidacloprid leached upto 60 cm depth showing its strong leaching power as reported in literature [20]. This leaching tendency of imidacloprid was observed in soil on which it was applied first time but in field where it is applied from many years the leaching power will be more as above researcher reported. Higher organic matter favors adsorption while reduces leaching capacity of imidacloprid pesticide [21,22]. The organic matter content of our studied soil was less (0.899 \pm 0.01); therefore limited adsorption and more leaching of imidacloprid were seen. Organic matter is closely related to cation exchange capacity as it has the highest cation exchange capacity of all the soil constituents and a larger surface area, therefore higher amount of organic matter and cation exchange capacity increases adsorption and reduces leaching. Bansal in his study of three carbamate pesticides sorption had observed that, the addition of organic matter to soil increases soil organic carbon content and electrical conductivity while decreases pH [23]. He noticed that the adsorption was positively correlated with soil organic carbon and cation exchange capacity while negatively

correlated with soil pH. The effect of pH on adsorption of pesticide is that in acidic soils due to presence of a salt the stronger cation exchange mechanisms of chemisorption is favored hence adsorption is more as compared to basic soils having higher pH [24,25]. The same trend was observed by Ping *et al.* that imidacloprid adsorption was usually higher under lower pH and/or lower temperature at same condition [26]. The pH of our soil was 8.46 (in basic range), hence adsorption was low and leaching was more. The higher electrical conductivity value of soil enhances water retention by increasing the osmotic potential of the soil, thereby increasing the contact-time for adsorption. Leaching and adsorption are inversely related to each other, the soil in which leaching is more and adsorption will be less means that soil possess less organic matter and light texture. The physicochemical properties of soil are shown in **Table 1**.

3.2. Kinetic Adsorption Study

Kinetic adsorption revealed the removal rate of solute which controls the residence time of the pesticide in the solid-solution interface. The kinetic adsorption has shown two clear regions as given in **Figure 1**; initially a very rapid adsorption up to four hour followed a slow adsorption region. The pseudo equilibrium was achieved within six hour and then a uniform adsorption of imidacloprid in soil was observed.

The imidacloprid was removed from solution during initial four hours, as reported in literature for different pesticides by various researchers [27,28]. The imidacloprid was adsorbed rapidly during first stage because the vacant sites are available in soil initially, followed by a slow migration and diffusion of imidacloprid molecules in soil [29].

The pseudo first order and pseudo second order kinetic models were used to observe that which model was best

fitted to kinetic adsorption.

3.2.1. Pseudo First Order Model

First order rate constant was calculated by Equation (1).

$$\log(q_e - q_t) = \log q_e - k_1 \cdot t/2.303 \quad (1)$$

where q_e and q_t are the amount of imidacloprid adsorbed (mg/g) at equilibrium and at time t (min) respectively while k_1 (1/h) is the first order rate constant. Therefore first order rate constant k_1 and q_e were calculated from slope and intercept of plot $\log(q_e - q_t)$ versus t . This model was not followed by adsorption of imidacloprid.

3.2.2. Pseudo Second Order Model

Data was also examined through pseudo second order model given by Ho and McKay. Second order rate constant was calculated by Equation (2).

$$t/q_t = 1/k_2 \cdot q_e^2 + t/q_e \quad (2)$$

Plot of t/q_t versus t had shown straight line with R^2 value 0.999 for imidacloprid as shown in **Figure 2**, from which it was concluded that kinetic adsorption of imidacloprid had closely correlate with pseudo second order model [30]. In chemisorption process, the pseudo second order is superior to pseudo-first order model because it deals with interaction of adsorbent-adsorbate through their valency forces. The rate constant values of both orders are shown in **Table 2**.

3.3. Equilibrium Adsorption Study

Adsorption isotherm studies were conducted at 28°C with an equilibrium time of 6 h. Langmuir and Freundlich isotherm models were applied to calculate the isotherm constants and the maximum adsorption capacity of imidacloprid at nine different concentrations to evaluate its behavior clearly. These concentrations were 0.25, 0.5, 1,

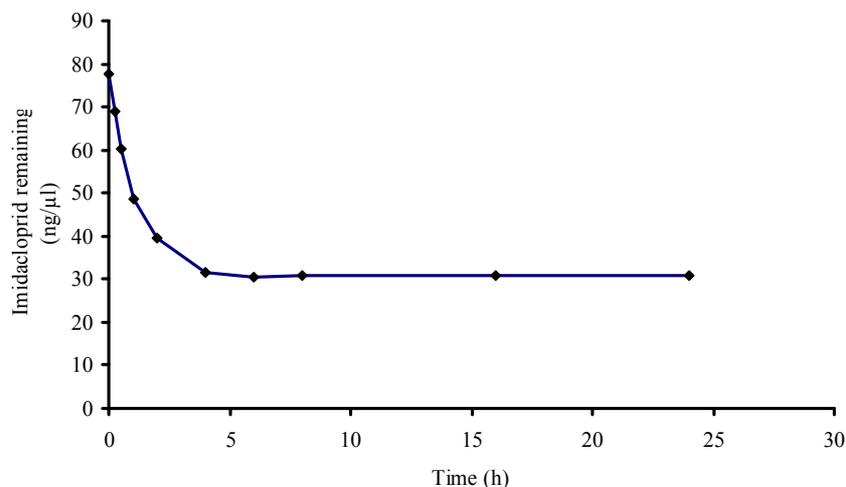


Figure 1. Kinetic adsorption of imidacloprid.

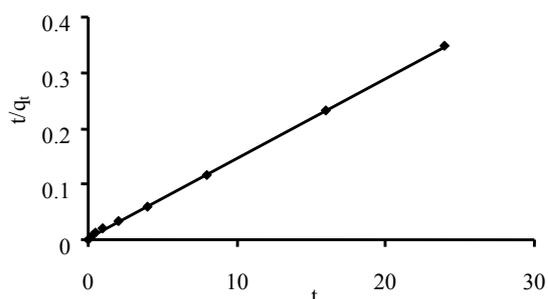


Figure 2. Pseudo second order plot for imidacloprid.

Table 2. Adsorption rates of imidacloprid.

| Models applied | Adsorption rate of imidacloprid |
|------------------------------|---------------------------------|
| Pseudo first order (1/h) | 0.845 ± 0.011 |
| Pseudo second order (mg/g/h) | 4.333 ± 0.02 |

2, 5, 10, 25, 50 and 100 ppm.

3.3.1. Langmuir Isotherm Model

This model deals with monolayer and homogeneous adsorption because the adsorbed layer is one molecule in thickness, with adsorption occurring at fixed sites, which are identical and equivalent. Linear form of this model is given in Equation (3).

$$C_e/C_{ads} = 1/Q_b + C_e/Q \quad (3)$$

where C_e is the equilibrium concentration of imidacloprid in solution (mol/L) and C_{ads} is the amount of imidacloprid (mol/g) on soil surface, Q is the monolayer adsorption capacity, and b is Langmuir constant. The plot of C_e/C_{ads} against C_e gives langmuir isotherm which was not obeyed by adsorption of imidacloprid.

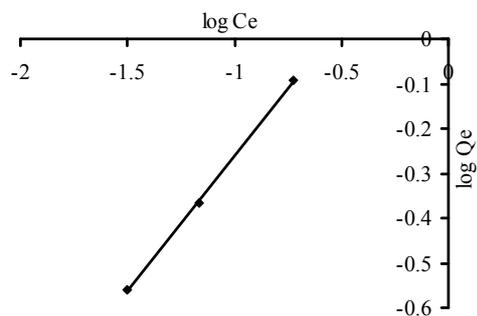
3.3.2. Freundlich Isotherm Model

Freundlich isotherm is related to the non-ideal and reversible adsorption, not limited to monolayer formation. Therefore it is applied to multilayer adsorption, with non-uniform distribution of adsorption heat and affinities over the heterogeneous surface. Linear form of this model is given as in Equation (4).

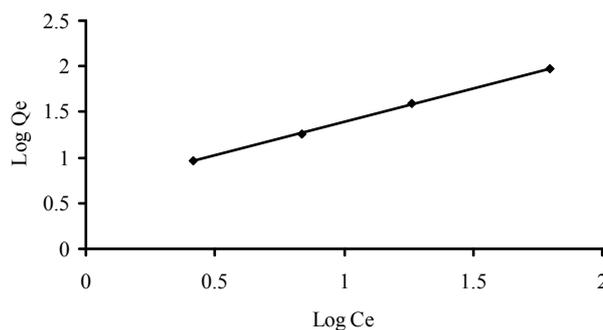
$$\log C_{ads} = \log A + (1/n) \log C_e \quad (4)$$

where A is multilayer adsorption capacity and n is adsorption intensity. A plot of $\log C_{ads}$ against C_e gives straight line showing that this model was obeyed by adsorption of imidacloprid [31] as shown in Figure 3. Freundlich isotherm was applied at both, lower (0.25, 0.5, 1, 2 ppm) and higher concentrations (5, 10, 25, 50 and 100 ppm) as shown in Figures 3(a) and (b).

The parameters of both models are shown in Table 3. The n value is 1.664 and 1.366 for imidacloprid in Freundlich isotherm, showing that adsorption process was favorable because $1 < n < 10$ represents the favorability of



(a)



(b)

Figure 3. Freundlich isotherm for imidacloprid at (a) lower and (b) higher concentrations.

Table 3. Equilibrium adsorption isotherm values for imidacloprid pesticide.

| Isotherm | Isotherm parameter | Value |
|--------------------------------------|--------------------|------------------------|
| Langmuir | Q (mol/g) | 6.459×10^{-4} |
| | b | 0.179 |
| | R^2 | 0.743 |
| Freundlich (at lower concentration) | A (mol/g) | 2.190 |
| | n | 1.664 |
| Freundlich (at higher concentration) | R^2 | 0.999 |
| | A (mol/g) | 4.573 |
| | N | 1.366 |
| | R^2 | 0.999 |

adsorption [32].

3.4. Column Leaching Study

From water portions of column it was observed that the amount of imidacloprid pesticide eluted was more as compared to adsorbed on soil filled in column due to more water solubility of imidacloprid. The amount of imidacloprid was eluted in each portion of water but it was decreased from 0.481 ppm in first portion of water to 0.327 ppm in last portion of water. During this elution the pesticide has transported 30 cm column length due to

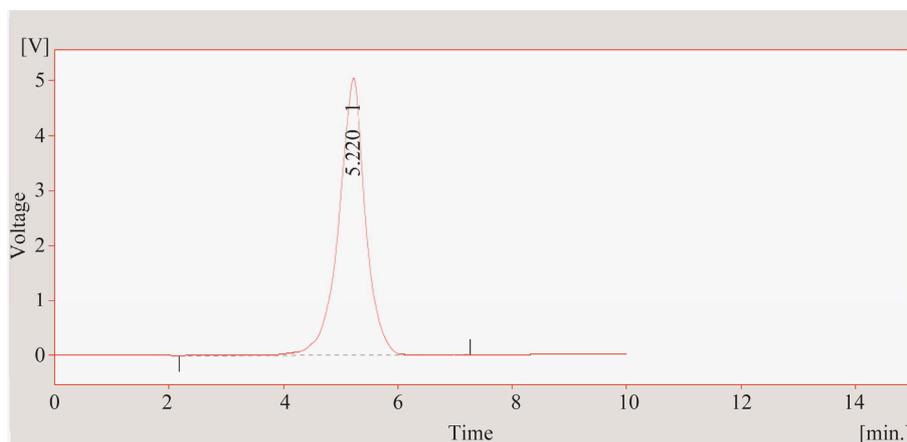


Figure 4. HPLC chromatogram showing peak for imidacloprid at retention time 5.201 min.

less organic matter content present in our soil, sandy texture and forcing power of water. The concentration of imidacloprid has decreased from 0.783 ppm in first section of column soil to 0.038 ppm in third section of column soil, showing both leaching and adsorption power of imidacloprid in column. The result is shown in **Table 4**. A representative chromatogram of imidacloprid is shown in **Figure 4**.

3.5. Field Study

Imidacloprid is mobile in the environment due to its weak adsorption on soil [33]; therefore in field it was leached upto 60 cm due to less organic matter content, sandy texture, low cation exchange capacity and basic pH. The concentration of imidacloprid was decreased as we go down. It was 3.311 ppm in first portion of soil having depth 0 - 10 cm, 1.276 ppm in second portion of soil having depth 11 - 20 cm, 0.923 ppm in third portion of soil having depth 21 - 30 cm and 0.357 ppm in fourth portion of soil having depth 31 - 60 cm. The results are shown in **Table 5**.

4. Conclusion

Present study suggests that the kinetic adsorption of imidacloprid on soil follows the second order kinetic model and the rate limiting step may be chemical adsorption, while equilibrium adsorption follows Freundlich isotherm describing the reversible, non-ideal and multilayer adsorption over the heterogeneous surface. From laboratory and field studies it was concluded that the leaching and adsorption power of imidacloprid in soil were dependent upon physicochemical properties of soil. In our soil the imidacloprid pesticide was leached up to 60 cm soil depth due to less organic matter, sandy texture, basic pH, and low cation exchange capacity, otherwise it may be adsorbed on surface of soil. The soils in Sindh, Pakistan are alkaline and have low organic matter, while

Table 4. Imidacloprid leaching and adsorption through column.

| Sample (cm) | Imidacloprid concentration found (ppm) |
|--------------|----------------------------------------|
| Soil 0 - 10 | 0.783 ± 0.02 |
| Soil 11 - 20 | 0.102 ± 0.011 |
| Soil 21 - 30 | 0.038 ± 0.002 |
| Water-01 | 0.481 ± 0.002 |
| Water-02 | 0.464 ± 0.001 |
| Water-03 | 0.362 ± 0.001 |
| Water-04 | 0.357 ± 0.003 |
| Water-05 | 0.327 ± 0.001 |

Table 5. Imidacloprid residue in okra field soil.

| Soil depth (cm) | Imidacloprid concentration found (ppm) | % of imidacloprid in each layer |
|-----------------|----------------------------------------|---------------------------------|
| Soil 0 - 10 | 3.311 ± 0.08 | 56.434 ± 0.09 |
| Soil 11 - 20 | 1.276 ± 0.09 | 21.749 ± 0.35 |
| Soil 21 - 30 | 0.923 ± 0.07 | 15.732 ± 0.04 |
| Soil 31 - 60 | 0.357 ± 0.02 | 6.084 ± 0.03 |
| Grand Total | 5.867 | 100 |

pesticide adsorption studies reported in literature have been conducted in soils having acidic pH and rich in organic matter. In basic soils with low organic matter, adsorption is low and leaching is more that we have pointed out here. More leaching is a potential threat to environment and whole food chain.

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