Impact of past HBV exposure on virological response to combined interferon ribavirin therapy in patients with chronic HCV genotype 4

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ABSTRACT

Aim: is to investigate if past exposure to HBV in chronic HCV infection promotes the progression of liver disease and influences Interferon (INF) response or not. Methods: 185 patients with chronic HCV were recruited and randomized into 2 groups, group I was treated with PEG-IFN alfa-2b plus ribavirin and group II with 3 MU/TIW IFN alfa-2b plus ribavirin. All patients had HBsAg negative & patients had HBc Ab positive tested for HBsAb. Patients with HBc Ab (n = 37) were tested for serum HBV DNA. Pretreatment liver biopsy and Immunohistochemistry was performed for detection of HBsAg and HBcAg using monoclonal antibodies. Demographic and laboratory data were collected. Results: 70/185 (38%) patients had Anti-HBc, 37 (20%) had isolated anti-HBc and none of the 37 subjects with isolated anti-HBc had serum HBV DNA. Sustained Viral Response (SVR) was achieved in 32/70 (46%) and 46/115 (40%) in anti-HBc seropositive and seronegative patients respectively, in conventional IFN group SVR was 37% and 39% in past HBV exposed patients and unexposed respectively, in PEG IFN group SVR was 59% and 41% in past HBV exposed patients and unexposed respectively. Among anti-HBc and anti-HBs seropositive patients SVR was 10/12 [83%] and 4/21 [19%] p = 0.004 in PEG and conventional IFN respectively. Marked fibrosis was diagnosed in 20% past HBV exposed patients versus 10% in unexposed patients (p = 0.01). Conclusion: past HBV exposure promotes the progression of liver disease but has no effect on the SVR to antiviral therapy in chronic **HCV** genotype 4 patients.

Keywords: HCV Genotype 4; Past HBV Exposure; Response to Interferon Therapy

1. INTRODUCTION

Hepatitis C virus (HCV) infection is a chronic disease affecting 270 - 300 million people worldwide [1]. About 85% of those exposed to HCV become chronically infected and only 15% clear the virus. Chronic hepatitis and liver cirrhosis have been documented in 70% and 20%, respectively, of those chronically infected [2,3]. Egypt has an overall HCV antibody prevalence around 15% in the general population [4-7] and genotype 4 has been identified in about 90% of HCV isolates [8-12]. Despite recent improvements in antiviral therapy with interferon and ribavirin, only about half of those who meet the criteria for treatment will respond to therapy. The mechanism responsible for the resistance to treatment and/or relapse of hepatitis after withdrawal of antiviral therapy may be host and/or virus related [13-15]. A possible factor influencing responsiveness of HCVpositive patients to antiviral therapy may be co-infection with HBV [16]. Concurrent infection with the two viruses may worsen the prognosis and result in a less favorable outcome to antiviral treatment [17-19]. Moreover, some patients who are HBsAg negative but hepatitis B core antibody (anti-HBc) positive are usually regarded as having occult HBV infection [20,21]. Some investigators have suggested that occult (in apparent) HBV infection correlates with a lack of response to interferon treatment in patients with chronic hepatitis C [22,23].

Hepatitis B virus (HBV) is endemic in Egypt. The HBV markers (presence of either anti-HBc and/or HBs Ag) were present in 24% of villagers in a study conducted in the Nile delta [24]. In a more recent serosur-



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veillance among Egyptian nationals working in the tourism industry the prevalence of serologic markers for currently asymptomatic or past HBV infection alone was 21% and of markers for both HBV and HCV was 6.9% [25].

To our knowledge, no studies have evaluated the role of past exposure to hepatitis B in HCV genotype 4 infected individuals. We therefore investigated whether past exposure to hepatitis B virus in individuals infected with HCV genotype 4 influences the response to interferon therapy.

2. PATIENTS AND METHODS

2.1. Selection of Patients

Chronically infected HCV naïve patients (>18 years old and <60 years) who had detectable levels of serum HCV-RNA, HCV antibodies and undetectable HbsAg, elevated serum transaminase for at least six months and a histological diagnosis of chronic hepatitis with or without liver cirrhosis were eligible for the study. Exclusion criteria included: decompensated liver disease, hemoglobin $< 13 \text{ g}\cdot\text{dL}^{-1}$ for men and $< 12 \text{ g}\cdot\text{dL}^{-1}$ for women, white blood cell count of <3000/mm³, neutrophil count of <1500/mm³, or platelet count of <100,000/ mm³. Patients who had had hepatitis B surface antigen (HBsAg), autoimmune hepatitis, hemochromatosis, Wilson's disease, α1-antitrypsin deficiency, aalcoholic liver disease, and drug induced liver disease, active Schistosomiasis, or hepatic tumors were excluded. Patients with poorly controlled diabetes mellitus or hypertension or under care of a psychiatrist were also excluded.

2.2. Study Design and Treatment Regimen

Two hundred and fifty patients were recruited in randomized, open label clinical trial to receive standard or pegylated interferon (alfa-2b & alfa-2a) with ribavirin; it was conducted at the National Hepatology and Tropical Medicine Research Institute (NHTMRI), Cairo Egypt in collaboration with the International Health Division, University of Maryland, and School of Medicine. The study was approved by the institutional review board at both institutions and all study subjects provided written informed consent. The protocol conformed to the ethical guidelines of the 1975 Declaration of Helsinki. It was found that severe fibrosis, presence of steatosis, treatment with standard interferon and higher serum AFP were all independently and negatively associated with SVR. Level of pretreatment AFP, being a biological and noninvasive marker, could be incorporated into the routine assessment of factors predictive of treatment response [26].

To investigate whether past exposure to hepatitis B among the HCV infected study cohort has influenced the

responsiveness to antiviral therapy, we performed a secondary analysis on a 185 out of 250 patients previously recruited who were HbsAg negative subjects. Anti-HBc was detected in 70 (38%) of these 185 subjects and remaining 115 (62%) were HBV seronegative with no evidence of past exposure to HBV.

2.3. Clinical Evaluation

The patients were evaluated as outpatients after 1, 2, 4, 6, 8 and 12 weeks of initiation of treatment, every 4 weeks thereafter during treatment, and 4, 12, and 24 weeks after treatment was completed. Patients were followed up until week 72 to assess sustained responses.

2.4. Laboratory Evaluation

Biochemical and hematological testing were performed at the Viral Hepatitis Research Laboratory (VHRL) at the NHTMRI. Virological studies included a third-generation screening test for the detection of antibody to hepatitis C virus (anti-HCV) (Abbott HCV EIA 3.0/Ortho HCV ELISA 3.0). Testing for hepatitis B markers included Hepatitis B Surface Antigen (HBsAg), Hepatitis B Surface Antibody (anti-HBs), and Total Hepatitis B Core Antibody (anti-HBc). Hepatitis C virus-RNA tests, viral genotyping, and histological evaluation of biopsy specimens were performed as well. Serum HCV-RNA tests were determined before and during treatment in the patients who were treated for 48 weeks; and after therapy weeks at week 72 to assess sustained responses. Serum HCV RNA was measured by an in-house reverse-transcripttion-polymerase-chain-reaction (RT)-PCR assay [27]. The assay has a lower limit of detection of 100 viral copies mL⁻¹. (RT)-PCR is a modification of the standard PCR that was developed to amplify RNA templates. The major steps of the (RT)-PCR assay are isolation of RNA, reverse transcription of RNA into cDNA, and amplification of the cDNA by PCR. Primer sets used in the PCR reaction are highly conserved and can detect genotypes 4, as previously published [27]. Quantitative RT-PCR (real-time PCR) was performed to test for the HCV viral load using the Cobas Amplicor Kits from Roche (Germany). Conventional qualitative PCR detection of HBV DNA was performed on sera from patients positive for isolated anti-HBc (Cobas Amplicor HBV Monitor Test (Roche Diagnostic systems, Pleasanton, CA). The lower limit of detection was 200 copies/ml. HBV DNA extraction from tissues was done by PCR for 9 patients with isolated anti-HBc and with lower limit of detection of 200 copies ml⁻¹.

Liver biopsy was performed at the beginning and the end of follow-up. Pretreatment liver biopsy was performed and evaluated in parallel with those obtained at week 72. Determination of the stage of fibrosis and the grade of inflammation was accomplished using the Modified Knodell score in the form of 18 points for

Histopathology Activity Index (HAI) and six points for fibrosis. Immunohistochemistry (IHC) was performed for the detection of hepatitis B surface and core antigens in liver biopsies using monoclonal antibodies.

2.5. Assessment of Efficacy and End Point

The primary end point was a sustained virologic response, defined as the absence of serum HCV RNA 24 weeks after treatment was completed. Secondary end points were normalization of the serum alanine aminotransferase concentration at week 24 and histologic improvement. The degree of hepatic inflammation and fibrosis was graded with a modified Knodell Histologic Activity Index. The inflammation score was obtained by combining the scores for the first three components of this index: portal, periportal, and lobular inflammation. The scores could range from 0 to 18, with higher scores indicating more severe abnormalities. The degree of fibrosis was graded as 0, no fibrosis; 1, portal fibrosis; 3, bridging fibrosis; or 4, cirrhosis. Histologic improvement was defined as a decrease of at least two points in the inflammation score, as compared with the score for the pretreatment biopsy specimen. The biochemical response and the sustained combined biochemical and virologic response were also assessed.

2.6. Statistical Analysis

Intention-to-treat analysis was used as the primary

analysis for all measures of efficacy. Quantitative variables were expressed as means and standard deviation, compared using the student's *t*-test, Mann-Whitney U test or ANOVA test when appropriate. Fisher exact test was used to compare categorical variables, with stratification by center for comparisons between treatment groups and stratification by center and treatment group for other categorical variables. Pearson correlation was used to compare quantitative variables between the groups. All statistical tests were 2-tailed; P < 0.05 was considered statistically significant. We used SPSS, version 10.0 for windows.

3. RESULTS

The enrolled 185 HCV chronically infected and HbsAg negative subjects were tested for Hepatitis B Core antibody. Anti-HBc was detected in 70 (38%) of the185 subjects, 37 (20%) were isolated anti-HBc, and 33 (18%) were anti-HBs and anti-HBc seropositive. The remaining 115 (62%) were HBV seronegative with no evidence of past exposure to HBV. None of the 37 subjects with isolated anti-HBc had HBV DNA detected in their sera by conventional qualitative PCR. The pretreatment characteristics of the patients were similar among patients with or without past exposure to hepatitis B in HCV genotype 4 infected individuals (**Table 1**).

Virological response: An overall sustained virological response (SVRs) (primary end point) of 42% (78/185)

Table 1. Baseline characteristics of patients with chronic hepatitis C with or without positive markers of past HBV.

Characteristics of chronically in-	HBV sero-negative	Isolated anti-HBc	Anti-HBs & anti-HBc seropositive
fected HCV naive patients	N (%) = 115 (62)	N (%) = 37 (20)	N (%) = 33 (18)
Age mean (SD), y	37.6 (8.5)	41.2 (8.6)	43.4 (8.2)
Sex, No (%)	00 (55)	21 (0.0)	20 (05)
Men	89 (77)	31 (84)	28 (85)
Women	26 (23)	6 (16)	5 (15)
Residence, No (%)			
Rural	55 (48)	18 (49)	19 (58)
Urban	60 (52)	19 (51)	14 (42)
No (%) with active schistosomiasis	18 (16)	8 (22)	5 (15)
Serum ALT mean (SD)	104 (70)	117 (62)	115 (72)
Serum AST mean (SD)	64 (39)	69 (29)	72 (45)
HCV RNA mean (IU/ml)	2×10^{6}	0.4×10^{6}	0.5×10^{6}
No (%) HBeAb	-	23 (62.2)	20 (60.6)
Serum HBV DNA	NA	NEGATIVE†	NA
Tissue HBV DNA	NA	NEGATIVE‡	NA
Abd. Ultrasound, No (%)			
Hepatomegaly	27 (24)	9 (24)	9 (27)
Spenomegaly	18 (16)	5 (14)	4 (12)
Liver Histology§ HAI No (%) Minimal	11 (10)	4 (11)	3 (9)
Mild	76 (66)	20 (54)	20 (61)
Moderate	27 (23)	12 (32)	7 (21)
Severe	1(1)	1 (3)	3 (9)
Fibrosis No (%) Minimal	15 (13)	88 (77)	12 (10)
Moderate	-	29 (78)	8 (22)
Marked	2 (6)	25 (76)	6 (18)

†No Serum HBV DNA was detected in subjects with isolated anti-HBc (n=37); ‡No Tissue HBV DNA was detected in 9 patients with isolated anti-HBc; § One hundred and eighty five patients had pretreatment liver biopsy.

was achieved using combination therapy with interferon and ribavirin. SVR was achieved in 32 (46%) of the 70 patients with laboratory evidence of prior exposure to HBV and 46 patients (40%) of the 115 patients without prior exposure (p = 0.5). Among patients treated with standard interferon the SVR was 37% in the HBV exposed and 39% in the HBV unexposed patients. Also, among the pegylated interferon treated group no statistically significant differences in the SVR of patients with past exposure compared to those without HBV exposure 59% (17/29) vs. 41% (26/63) respectively (**Table 2**). Whereas, among the 33 patients who were Anti-HBs & anti-HBc seropositive significantly more patients in the pegylated interferon treated group achieved a SVR compared to those treated with standard interferon group (10/12 [83%] versus 4/21 [19%] p = 0.004) respectively.

Virological relapse was similar in both arms of the study. A total of the 11 (12%) of the 93 patients in the standard interferon group and 13 (14%) of the 92 patients in the pegylated interferon group relapsed after having undetectable HCV RNA in their sera. Of the seventy HBV exposed patients four (5.7%) had virological relapse versus 20 (17%) of the 115 HBV unexposed patients.

Among patients with history of past exposure HBV infection (anti-HBc with or without anti-HBs), 3%, 77%, and 20% were diagnosed as having minimal, moderated or marked fibrosis respectively while 13%, 77%, 10% of HBV seronegative patients were diagnosed as having minimal, moderated or marked fibrosis respectively (**Table 3**).

Table 2. Percent of patients with sustained virological response (SVR) after 72 weeks.

Characteristics	Anti-HBs and/or anti-HBC seropositive N (%) = 70 (38)	e seronegative	
Standard Interferon	37%	39%	
PEG INF	59%	41%	
Total N (%)	32 (46%)	46 (40%)	

Table 3. Stages of fibrosis in chronically infected HCV naive patients with or without past exposure to hepatitis B virus.

Characteristics	Anti-HBC ∨ anti-HBs seropositive N (%) = 70 (38)	HBV seronegative N (%) = 115 (62)
Minimal (stages 0-1)	2 (3)	15 (13)
Moderate (stages 2-4)	54 (77)	88 (77)
Marked* (stages 4-6)	14 (20)	12 (10)

^{*}P = 0.01.

4. DISCUSSION

The primary goal of therapy for chronic HCV infection is viral eradication. Treatment of chronic HCV infection has significantly improved over the past several years with combination PEG-IFN and ribavirin therapy. SVR is achievable in nearly half of patients with genotype 1 and about 80% of those with genotypes 2 and 3 [28-30]. In a published meta-analysis of randomized, controlled, open-label treatment trials of naive patients with chronic HCV genotype 4 [31,32] the overall SVR to combination therapy using PEG-IFN and ribavirin therapy was 55%. In another study to evaluate the treatment response and duration of therapy in patients with HCV genotype 4, a randomized, double-blind study of PEG-IFN alfa-2b and ribavirin was performed [33] and SVR was achieved in 29%, 66%, and 69% of patients treated for 24, 36, and 48 weeks, respectively. Thus, the data available suggest that treatment response to genotype 4 lies somewhere between that for genotype 1 and genotypes 2 and 3.

To date, the reason for the differences in response of various genotypes to antiviral therapy remains unclear as it is not known why certain genotypes respond more favorably to antiviral therapy than other genotypes. The outcome of therapy may be influenced by a dynamic complex relationship that exists between the pharmacological characteristics of the therapeutic regimen, viral kinetics, and host immune responses, or concurrent co-infection with HBV. It was observed that the presence of markers indicating past HBV infection (anti HBV antibodies) was associated with a reduced rate of treatment response [23], in more recent study past history of HBV infection had no influence on the efficacy of treatment with IFN-alpha and ribavirin through Clearance of HCV from serum and from PBMC [34].

We evaluated the effectiveness of interferon treatment in HCV genotype 4 chronically infected naive patients with past exposure to hepatitis B virus infection. We found more than one third (38%) of our study participants had laboratory evidence of past exposure to HBV. The percentage of SVR achieved was similar in the group of patients with or without evidence of past exposure to HBV infection. SVRs were also similar in the pegylated interferon treated and the standard interferon treated groups. However, patients treated with peg interferon and having both hepatitis B core and surface antibodies had a significantly better SVR than that observed in the standard interferon treated patients (p = 0.004).

Although virological relapse was similar in both treatment arms of the study, yet a significantly lower virological relapse rate was observed in the HBV exposed patients compared to the HBV unexposed patients.

Finally our results showed that the prevalence of anti-HBV antibodies was parallel to the severity of liver

disease, with significantly more HBV exposed patients (anti-HBc with or without anti-HBs) with marked fibrosis and this was still true after adjustment for age, sex and history of schistosomiasis, the presence of positive anti-HBc was an important independent risk factor for the development of severe fibrosis in patients with HCV related chronic liver disease. Conclusion: these data suggest that past exposure to HBV infection may promote the progression of liver disease but has no effect on the SVR to combination therapy in chronic hepatitis C serotype 4 patients residing in areas where HBV infection is endemic.

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REFERENCES

- [1] Ryan, K.J. and Ray, C.G. (Eds). (2004) Sherris medical microbiology. 4th Edition, McGraw Hill, 551-552.
- [2] Lauer, G.M. and Walker, B.D. (2001) Hepatitis C virus infection. New England Journal of Medicine, 345, 41-52. doi:10.1056/NEJM200107053450107
- [3] Seeff, L.B. (2002) Natural history of chronic hepatitis. Hepatology, 36, 35-46. doi:10.1053/jhep.2002.36806
- [4] Abdel-Aziz, F., Habib, M., Mohamed, M.K., Abdel-Hamid, M., Gamil, F., Madkour, S. *et al.* (2000) Hepatitis C virus (HCV) infection in a community in the Nile Delta: Population description and HCV prevalence. *Hepatology*, **32**, 111-115. doi:10.1053/jhep.2000.8438
- [5] Habib, M., Mohamed, M.K., Abdel-Aziz, F., Magder, L.S., Abdel-Hamid, M., Gamil, F. et al. (2001) Hepatitis C virus infection in a community in the Nile Delta: Risk factors for seropositivity. Hepatology, 33, 248-253. doi:10.1053/jhep.2001.20797
- [6] Nafeh, M.A., Medhat, A., Shehata, M., Mikhail, N.N., Swifee, Y., Abdel-Hamid, M. et al. (2000) Hepatitis C in a community in Upper Egypt: I. Cross-sectional survey. American Journal of Tropical Medicine and Hygiene, 63, 236-241.
- [7] Medhat, A., Shehata, M., Magder, L.S., Mikhail, N., Abdel-Baki, L., Nafeh, M. et al. (2002) Hepatitis C in a community in Upper Egypt: Risk factors for infection. American Journal of Tropical Medicine and Hygiene, 66, 633-638.
- [8] Dusheiko, G.M. (1994) Progress in hepatitis C research. Lancet, 344, 605-606.

doi:10.1016/S0140-6736(94)91984-4

- [9] McOmish, F., Yap, P.L., Dow, B.C., Follett, E.A., Seed, C., Keller, A.J. et al. (1994) Geographical distribution of hepatitis C virus genotypes in blood donors: An international collaborative survey. *Journal of Clinical Mi*crobiology, 32, 884-892.
- [10] Mellor, J., Holmes, E.C., Jarvis, L.M., Yap, P.L. and Simmonds, P. (1995) Investigation of the pattern of hepatitis C virus sequence diversity in different geographical regions: Implications for virus classification. The International HCV Collaborative Study Group. *Jour*nal of General Virology, 76, 2493-2507.
- [11] Quinti, I., El-Salman, D., Monier, M.K., Hackbart, B.G., Darwish, M.S., El-Zamiaty, D. et al. (1997) HCV infection in Egyptian patients with acute hepatitis. *Di*gestive Diseases and Sciences, 42, 2017-2023. doi:10.1023/A:1018897813268
- [12] Zekri, A.R., Bahnassy, A.A., Ramadan, A.S., El-Bassuoni, M., Badran, A., Madwar, M.A. (2001) Hepatitis C virus genotyping versus serotyping in Egyptian patients. *Infection*, 29, 24-26. doi:10.1007/s15010-001-0010-8
- [13] Farci, P., Alter, H.J., Wong, D.C., Miller, R.H., Govindarajan, S., Engle, R. et al. (1994) Prevention of hepatitis C virus infection in chimpanzees after antibodymediated in vitro neutralization. Proceedings of the National Academy of Sciences of the United States of America, 1, 7792-7796. doi:10.1073/pnas.91.16.7792
- [14] Weltman, M.D., Brotodihardjo, A., Crewe, E.B., Farrell, G.C., Bilous, M., Grierson, J.M. et al. (1995) Coinfection with hepatitis B and C or B, C and delta viruses results in severe chronic liver disease and responds poorly to interferon-alpha treatment. *Journal of Viral Hepatitis*, 2, 39-45. doi:10.1111/j.1365-2893.1995.tb00070.x
- [15] Liaw, Y.F. (1995) Role of hepatitis C virus in dual and triple hepatitis virus infection. *Hepatology*, 22, 1101-1108.
- [16] Fukuda, R., Ishimura, N., Niigaki, M., Hamamoto, S., Satoh, S., Tanaka, S. et al. (1999) Serologically silent hepatitis B virus coinfection in patients with hepatitis C virus-associated chronic liver disease: Clinical and virological significance. *Journal of Medical Virology*, 58, 201-207. doi:10.1002/(SICI)1096-9071(199907)58:3<201::AID-J MV3>3.0.CO;2-2
- [17] Mrani, S., Chemin, I., Menouar, K., Guillaud, O., Pradat, P., Borghi, G. et al. (2007) Occult HBV infection may represent a major risk factor of non-response to antiviral therapy of chronic hepatitis C. Journal of Medical Virology, 79, 1075-1081. doi:10.1002/jmv.20943
- [18] Miura, Y., Shibuya, A., Adachi, S., Takeuchi, A., Tsuchihashi, T., Nakazawa, T. *et al.* (2008) Occult hepatitis B virus infection as a risk factor for hepatocellular carcinoma in patients with chronic hepatitis C in whom viral eradication fails. *Hepatology Research*, **38**, 546-556. doi:10.1111/j.1872-034X.2007.00316.x
- [19] Branco, F., Mattos, A.A., Coral, G.P., Vanderborght, B., Santos, D.E., Franca, P. et al. (2007) Occult hepatitis B virus infection in patients with chronic liver disease due to hepatitis C virus and hepatocellular carcinoma in Brazil. Arquivos de Gastroenterologia, 44, 58-63. doi:10.1590/S0004-28032007000100013
- [20] Liang, T.J., Blum, H.E. and Wands, J.R. (1990) Ch-

- aracterization and biological properties of a hepatitis B virus isolated from a patient without hepatitis B virus serologic markers. *Hepatology*, **12**, 204-212.
- [21] Zhang, Y.Y., Hansson, B.G., Kuo, L.S., Widell, A. and Nordenfelt, E. (1993) Hepatitis B virus DNA in serum and liver is commonly found in Chinese patients with chronic liver disease despite the presence of antibodies to HBsAg. *Hepatology*, 17, 538-544. doi:10.1002/hep.1840170403
- [22] Cacciola, I., Pollicino, T., Squadrito, G., Cerenzia, G., Orlando, M.E. and Raimondo, G. (1999) Occult hepatitis B virus infection in patients with chronic hepatitis C liver disease. New England Journal of Medicine, 341, 22-26. doi:10.1056/NEJM199907013410104
- [23] Zignego, A.L., Fontana, R., Puliti, S., Barbagli, S., Monti, M., Careccia, G. et al. (1997) Relevance of inapparent coinfection by hepatitis B virus in alpha interferontreated patients with hepatitis C virus chronic hepatitis. *Journal of Medical Virology*, 51, 313-318.
- [24] Kamel, M.A., Miller, F.D., El Masry, A.G., Zakaria, S., Khattab, M., Essmat, G., et al. (1994) The epidemiology of Schistosoma mansoni, hepatitis B and hepatitis C infection in Egypt. Annals of Tropical Medicine and Parasitology, 88, 501-509.
- [25] El-Sayed, N.M., Gomatos, P.J., Rodier, G.R., Wierzba, T.F., Darwish, A., Khashaba, S. et al. (1996) Seroprevalence survey of Egyptian tourism workers for hepatitis B virus, hepatitis C virus, human immunodeficiency virus, and Treponema pallidum infections: Association of hepatitis C virus infections with specific regions of Egypt. American Journal of Tropical Medicine and Hygiene, 55, 179-184
- [26] Gad, R.R., Males, S., El Makhzangy, H., Shouman, S., Hasan, A., Attala, A. et al. (2008) Predictors of a sustained virological response in patients with genotype 4 chronic hepatitis C. Liver International, 28, 1478-3223.
- [27] Abdel-Hamid, M., Edelman, D.C., Highsmith, W.E. et al. (1997) Optimization, assessment, and proposed use of a direct nested reverse transcribtion-polymease chain re-

- action protocol for the detection of hepatitis C virus. *Journal of Human Virology*, **1**, 58-65.
- [28] Manns, M.P., McHutchison, J.G., Gordon, S.C., Rustgi, V.K., Shiffman, M., Reindollar, R. et al. (2001) Peginterferon alfa-2b plus ribavirin compared with interferon alfa-2b plus ribavirin for initial treatment of chronic hepatitis C: A randomised trial. *Lancet*, 358, 958-965. doi:10.1016/S0140-6736(01)06102-5
- [29] Fried, M.W., Shiffman, M.L., Reddy, K.R., Smith, C., Marinos, G., Gonçales, F.L. Jr et al. (2002) Peginterferon alfa-2a plus ribavirin for chronic hepatitis C virus infection. New England Journal of Medicine, 347, 975-982. doi:10.1056/NEJMoa020047
- [30] Hadziyannis, S.J. and Koskinas, J.S. (2004) Differences in epidemiology, liver disease and treatment response among HCV genotypes. *Hepatology Research*, 29, 129-135. doi:10.1016/j.hepres.2004.02.011
- [31] Shobokshi, O.A. (2003) Hepatis C virus genotype 4. Saudi Medical Journal, 24, 79-80.
- [32] Alfaleh, F.Z., Hadad, Q., Khuroo, M.S., Algamedi, A., Alashgar, H., *et al.* (2004) Peginterferon alpha-2b plus ribavirin compared with interferon alpha-2b plus ribavirin for initial treatment of chronic hepatitis C in Saudi patients commonly infected with genotype 4. *Liver International*, **24**, 568-574. doi:10.1111/j.1478-3231.2004.0976.x
- [33] Kamal, S.M., El Tawil, A.A., Nakano, T., He, Q., Rasenack, J., Hakam, S.A. et al. (2005) Peginterferon {alpha}-2b and ribavirin therapy in chronic hepatitis C genotype 4: Impact of treatment duration and viral kinetics on sustained virological response. Gut, 54, 858-866. doi:10.1136/gut.2004.057182
- [34] Majda-Stanisławska, E., Bednarek, M., Jóźwiak, B., Sidorkiewicz, M., Piekarska, A. and Kuydowicz, J.E. (2006) Effect of interferon alfa and ribavirin treatment on hepatitis C virus RNA in serum and peripheral blood mononuclear cells in children with chronic hepatitis C. Acta Gastroenterol Belg, 69, 187-190.