

Effect of Grains of Paradise (GP) Extract Intake on Obesity and Sympathetic Nerve Activity

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Abstract

The methanol extract of grains of paradise (GP), the seed of *Aframomum melegueta*, which is distributed throughout West Africa, was administered during an animal breeding test. The extract suppressed body weight gain and decreased the weight of adipose tissues in breeding mice, with a greater effect on mice fed a high-fat diet (HFD) than on those fed a normal diet (ND). Other significant effects of GP intake included increased serum triglyceride (TG) concentration and reduced hepatic total cholesterol (TC) and TG concentrations. GP intake markedly prevented fat accumulation and improved hepatic lipid metabolism in HFD-fed mice. In addition, GP extract at a dosage of 5 mg/kg body weight decreased sympathetic nerve activity (SNA) in brown adipose tissue (BAT), while capsaicin, a major component of chili pepper, activated BAT SNA. This suggested that GP exerts a potential anti-obesity effect by a different mechanism from that of capsaicin.

Keywords

Anti-Obesity Effect, Grains of Paradise, *Aframomum melegueta*, Vanilloid, BAT SNA

1. Introduction

Obesity is defined as an abnormal or excessive fat accumulation and is recognized as a major risk factor for diabetes, cardiovascular disease, and cancer by the World Health Organization (WHO). The number of obese people increased more than two times between 1980 and 2014. Reducing lipid accumulation causing body weight loss is crucial to management of obesity. The development of effective treatments for overweight and obese patients has become very desirable in recent years.

Adipose tissue is a major metabolic organ, and roughly divided into two types: one is white adipose tissue (WAT) that stores energy in the form of triglyceride; the other is brown adipose tissue (BAT). BAT is involved in the dissipation and expenditure of energy as heat. This process of thermogenesis is induced by high-fat diets and cold exposure [1] [2]. Therefore, adipocytes play a key role in energy homeostasis. Activating BAT by stimulating sympathetic nerve activity (SNA) is one of the principal strategies to enhance energy expenditure and lipolysis [3] [4] [5]. This is an effective and practical approach to treating obesity-related diseases.

Aframomum melegueta is an herbaceous plant, widely distributed throughout Nigeria, Ghana, Guinea, and other countries in West Africa [6]. Its seeds are called grains of paradise (GP), Guinea pepper, alligator pepper, or melegueta pepper. It has been traditionally used as a spice for flavouring food and as a remedy for digestive and intestinal health, dysentery, migraine, and fever [7]. Recently, a number of studies have reported that GP extract has a range of activities such as antibacterial, repellent [8] [9], antioxidant, anti-inflammatory [10] [11], and hypoglycemic [12] effects. Moreover, GP contains many non-volatile pungent compounds such as 6-paradol, 6-gingerol, and 6-shogaol [13]. These compounds possess a vanillyl moiety, and are structurally similar to capsaicin and capsiate, which are found in chili pepper [14]. It is well known that capsinoids, including capsaicin and capsiate, exert an anti-obesity effect by stimulating SNA and hence BAT thermogenesis [15]. Similarly, it might be expected that GP extract containing vanilloids would activate BAT SNA and BAT thermogenesis. In the present study, GP extract was administered during an animal breeding test to investigate its efficacy in obesity prevention. Furthermore, electrical activity in BAT interscapular nerves was recorded using an electrophysiological method to investigate the anti-obesity mechanism of GP.

2. Materials and Methods

2.1. General Experimental Procedures

Tween 80 was purchased from MP Biomedicals, LLC (Santa Ana, CA, USA). Saline was purchased from Otsuka Pharmaceutical Co., Ltd. (Tokyo, Japan). A Bioelectric Amplifier ER-1 (Bio Research Center Co., Ltd., Nag) was used for amplifying and filtering sympathetic efferent nerve impulses. A PowerLab (AD Instruments Japan Inc., Nagoya, Japan) was used for converting the amplified signals, which were then recorded on a computer using Chart 5 software (AD Instruments Japan Inc.). Other commercially available products, including urethane for anesthesia, were purchased from Wako Chemicals (Richmond, VA, USA).

2.2. Sample Preparation

GP was provided by Share Trade Inc (Tokyo, Japan). Dried GP seed powder (about 5 kg) underwent methanol extraction all night at room temperature ($20^{\circ}\text{C} \pm 2^{\circ}\text{C}$) and the extract was obtained in 5.6% yield based on the powder.

The extract was dissolved in 10% Tween 80 saline solution containing 10% ethanol, and was used for oral administration during neural recording [16] [17].

2.3. Animal Breeding

Five-week-old male mice (24.8 ± 0.9 g) were purchased from Japan SLC Inc. (Hamamatsu, Japan) and placed in a breeding environment ($25^\circ\text{C} \pm 1^\circ\text{C}$, 12 h light-dark cycle). The mice were fed a normal diet (ND) for a period of one week. Both feed and water were available ad libitum. After preliminary breeding, they were divided into five groups. Three of the groups were fed a high-fat diet (HFD), composed of ND, 20% lard, 1% cholesterol powder, and 0.25% sodium cholate, containing either 2% GP seed powder, 0.3% GP extract, or 1% GP extract [18]. Two control groups were fed either an ND or HFD only. At 11 weeks of age, the mice were dissected to obtain samples of serum, the liver, and fat.

All experimental procedures were approved by the Gifu University Animal Care and Use Committee.

2.4. Determination of Total Cholesterol (TC), Triglyceride (TG), and High-Density Lipoprotein-Cholesterol (HDL-C) Accumulation in Serum and the Liver

Blood samples were stored at room temperature ($20^\circ\text{C} \pm 2^\circ\text{C}$) for 1 h, and then centrifuged at 3500 rpm for 15 min at room temperature ($20^\circ\text{C} \pm 2^\circ\text{C}$). Liver tissue (40 mg) was homogenized with 0.1 M ethyl acetate, methanol, and chloroform (4:10:5) at 4000 rpm for 1 min, and the homogenates were centrifuged at 5800 rpm for 10 min. The TC, TG, and HDL-C analyses were performed using TC, TG, and HDL-C E-test kits, respectively (Wako Pure Chemical Industries, Ltd.).

2.5. Neural Activity Measurement

The electrical activity of interscapular nerves innervating BAT was recorded as previously described [19]. In brief, a rat was anesthetized using urethane solution (1 g urethane/1kg body weight), and a small incision was made above the scapula to find sympathetic nerves entering BAT. Four sympathetic nerves were isolated and separated from the connective tissues. The isolated nerves were each placed on a pair of silver electrodes (0.3 mm, AG 401325, Nilaco Corp., Tokyo, Japan) to detect SNA, and the electrodes were immersed in mineral oil to prevent drying and for electrical insulation. Sympathetic efferent discharges were amplified and filtered, with the amplified impulses being converted to digital signals using PowerLab, and recorded using a computer software. Spikes above a threshold voltage, set just above background levels, were counted by spike histogram. Baseline BAT SNA was recorded for at least 30 min. Following oral administration of GP extract (5 mg/kg body weight) *via* a gastric tube, BAT SNA was recorded for every five minutes.

2.6. Statistical Analysis

All data were expressed as means \pm SD values. Statistical significance of differ-

ences was evaluated using the Student’s t-test. The difference was considered to be significant if $p < 0.05$.

3. Results

3.1. GP Intake Suppresses Body Weight Gain and Lipid Accumulation

To investigate the anti-obesity effect of GP and GP extract, the mice were divided into five groups after preliminary breeding. Mice willingly ate the feed provided in five meals, including those in the group receiving 2% GP. Despite this, the 2% GP group exhibited significantly lower body weight gain over the five weeks of feeding compared with the HFD group ($p < 0.01$) (Table 1 and Figure 1). The body weight gain of the groups fed GP extract with the HFD notably decreased, to a similar level as those of the mice in the ND control group

Table 1. Food intake, body weight gain, and liver, epididymal fat, and mesenteric fat weight of mice (n = 9) fed a normal diet (ND), high-fat diet (HFD), HFD including 0.3% GP extract, HFD including 1% GP extract, and HFD including 2% GP.

Group	Food intake (g)	Body weight (g)	Liver weight (g)	Epididymal fat (g)	Mesenteric fat (g)
ND (Normal Diet)	198.3 ± 21.0	44.0 ± 4.7	1.89 ± 0.23	1.13 ± 0.47	0.82 ± 0.19
HFD (High Fat Diet)	195.3 ± 22.2	52.3 ± 5.8 ^c	2.59 ± 0.48 ^c	2.16 ± 0.75 ^c	1.36 ± 0.51 ^c
HFD + 0.3% GP ext.	200.2 ± 26.5	42.1 ± 3.2 ^f	2.63 ± 0.29	0.80 ± 0.15 ^f	0.68 ± 0.10 ^d
HFD + 1% GP ext.	220.6 ± 27.3	40.6 ± 3.1 ^f	2.69 ± 0.32	0.90 ± 0.29 ^f	0.69 ± 0.20 ^d
HFD + 2% GP seed	245.8 ± 15.6 ^a	42.5 ± 4.6 ^d	2.63 ± 0.53	0.90 ± 0.28 ^f	0.53 ± 0.24 ^f

All data were expressed as the mean ± S.D. Differences were examined for statistical significance using Student’s t-test. n = 9. ^{a,c}: $p < 0.05$, 0.01 compared with ND values respectively. ^{d,f}: $p < 0.01$, 0.001 compared with HFD values respectively.

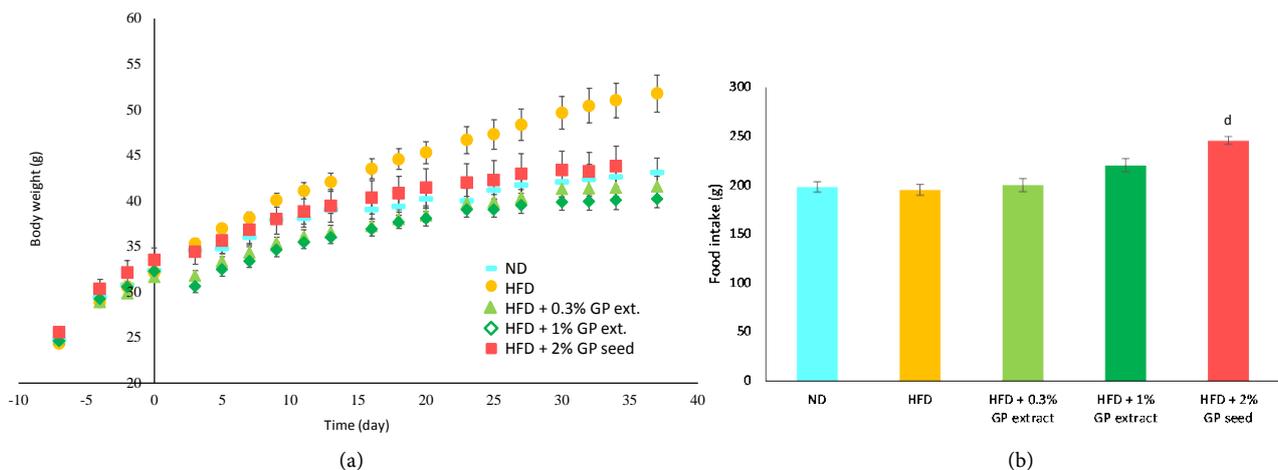


Figure 1. Body weight over time in mice (n = 9) fed a normal diet (ND), high-fat diet (HFD), HFD including 0.3% GP extract, HFD including 1% GP extract, and HFD including 2% GP (a) and the total food intake per mouse (b). d: $p < 0.01$ compared with HFD values.

(0.3% GP extract group: $p < 0.001$, 1% GP extract group: $p < 0.001$). Epididymal fat and mesenteric fat weights significantly decreased in the groups receiving GP in HFD when compared with those in the ND control group (0.3% GP extract group: $p < 0.001$ and < 0.01 , respectively; 1% GP extract group: $p < 0.001$ and < 0.01 , respectively; 2% GP group: $p < 0.01$ and < 0.001 , respectively).

Lipid accumulation was further investigated to explore the anti-obesity effect of GP on mice. HFD intake significantly increased the total cholesterol (TC) and triglyceride (TG) concentration in the liver. GP intake had no significant impact on liver weight (Table 2 and Figure 2). However, the GP and GP extract groups had significantly decreased levels of hepatic TC and TG. This effect was most marked in the 1% GP extract group, whose TC and TG levels (190 and 260 mg/dL, respectively) were far lower than those of the HFD group (257 and 388 mg/dL, respectively). In contrast, there was no significant difference in serum levels of either TC or high-density lipoprotein-cholesterol (HDL-C) among any of the groups on the HFD. Serum TG concentrations significantly increased in the GP intake group (0.3% GP extract group: $p < 0.05$; 1% GP extract group: $p < 0.01$) (Table 2 and Figure 3).

Table 2. Total cholesterol (TC), triglyceride (TG), and high-density lipoprotein-cholesterol (HDL-C) concentrations in serum and the livers of mice (n = 9) fed a normal diet (ND), high-fat diet (HFD), HFD including 0.3% GP extract, HFD including 1% GP extract, and HFD including 2% GP.

Group	Serum			Liver	
	TC (mg/dl)	TG (mg/dl)	HDL-C (mg/dl)	TC (mg/dl)	TG (mg/dl)
ND (Normal Diet)	127.9 ± 23.2	126.4 ± 51.7	88.7 ± 27.8	91.4 ± 34.1	96.5 ± 28.8
HFD (High Fat Diet)	199.9 ± 37.9 ^e	66.6 ± 11.0 ^e	88.1 ± 16.1	256.5 ± 47.4 ^e	388.4 ± 92.2 ^e
HFD + 0.3% GP ext.	207.1 ± 29.9	82.8 ± 23.6 ^d	88.0 ± 23.6	204.5 ± 39.7 ^f	251.9 ± 106.0 ^f
HFD + 1% GP ext.	230.4 ± 33.5	107.3 ± 83.8 ^b	87.5 ± 19.7	189.5 ± 58.7 ^f	258.8 ± 80.2 ^f
HFD + 2% GP seed	224.7 ± 39.5	75.3 ± 15.6	84.6 ± 7.9	198.0 ± 85.6 ^b	236.9 ± 51.8 ^f

All data were expressed as the mean ± S.D. Differences were examined for statistical significance using Student's t-test. n = 9. ^e: $p < 0.001$ compared with ND values. ^{b,d,f}: $p < 0.05, 0.01, 0.001$ compared with HFD values, respectively.

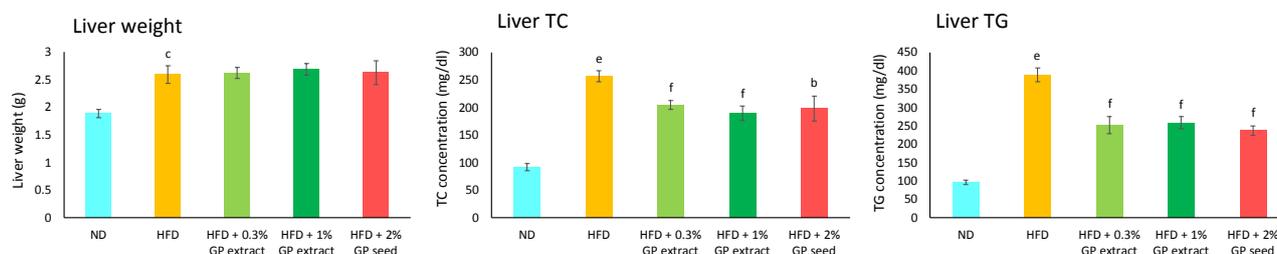


Figure 2. Liver weight, total cholesterol (TC), and triglyceride (TG) concentrations in mice (n = 9) fed a normal diet (ND), high-fat diet (HFD), HFD including 0.3% GP extract, HFD including 1% GP extract, and HFD including 2% GP. (c, e): $p < 0.01, 0.001$ compared with ND values respectively. (b, f): $p < 0.05, 0.001$ compared with HFD values, respectively.

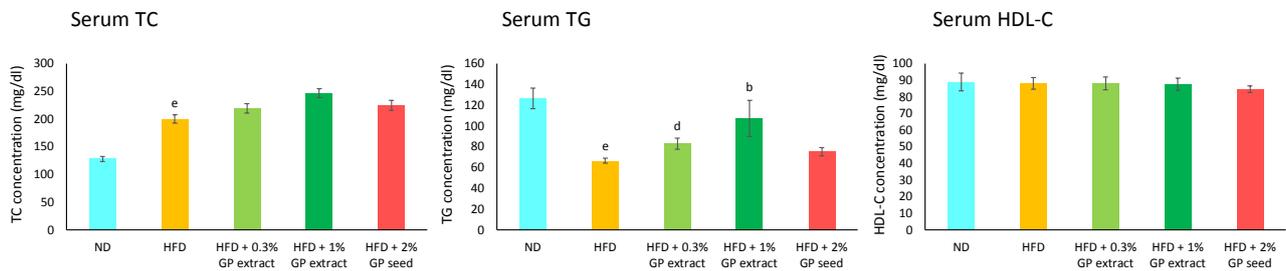


Figure 3. Total cholesterol (TC), triglyceride (TG), and high density lipoprotein-cholesterol (HDL-C) concentrations in serum of mice ($n = 9$) fed a normal diet (ND), high-fat diet (HFD), HFD including 0.3% GP extract, HFD including 1% GP extract, and HFD including 2% GP. (e): $p < 0.001$ compared with ND values. (b, d): $p < 0.05, 0.01$ compared with HFD values, respectively.

3.2. GP Extract Decreases Brown Adipose Tissue Sympathetic Nerve Activity (BAT SNA)

To clarify the anti-obesity effect of GP extract in mice, BAT SNA was determined using an electrophysiological method. An initial intragastric infusion of GP extract (5 mg/kg body weight) immediately decreased BAT SNA by around 10%, and this decrease lasted for at least 1 h. After BAT SNA recovery, a second intragastric infusion of GP extract at same concentration produced the same effect on BAT SNA (Figure 4). This procedure was performed on three rats, and produced similar results for each mouse.

4. Discussion

The animal breeding experiment in this study was performed to investigate the anti-obesity effects of GP and GP extract. The results are shown in Figures 1-3 and Table 1 & Table 2. The feeding of HFD containing 1% GP extract for five weeks greatly suppressed body weight gain and fat accumulation in mice. This suggested that GP has the potential to inhibit lipid accumulation. GP intake significantly decreased TC and TG concentrations in liver tissue, and prevented the pale discoloration of the liver, which was observed in the HFD group; however, it had no significant impact on liver weight. The serum TC and HDL-C concentrations in the GP intake groups were unchanged compared with those in the HFD group mice. Serum TG concentrations in the GP intake groups significantly increased. These results suggested that hepatic lipid metabolism was improved by GP intake. Consequently, GP intake potentially decreases fat accumulation in HFD mice. De Creamer *et al.* (1994) reported that hepatomegaly was observed in mice fed a diet rich in fish oil, with no significant changes in body, heart, or kidney weights. They concluded that this could be caused by induction of a liver peroxisome, which regulates β -oxidation and biosynthesis of bile acid. This metabolic change occurs via peroxisome proliferator-activated receptors (PPAR), which are involved in adipocyte differentiation [20] [21] [22] [23]. One of the components of GP, 6-shogaol, has been proven to activate PPAR [24]. Hence, the unchanged liver weight of mice in the GP groups compared with those of mice in the HFD group observed in this present study might be due to increased PPAR-stimulated hepatic peroxisome induction.

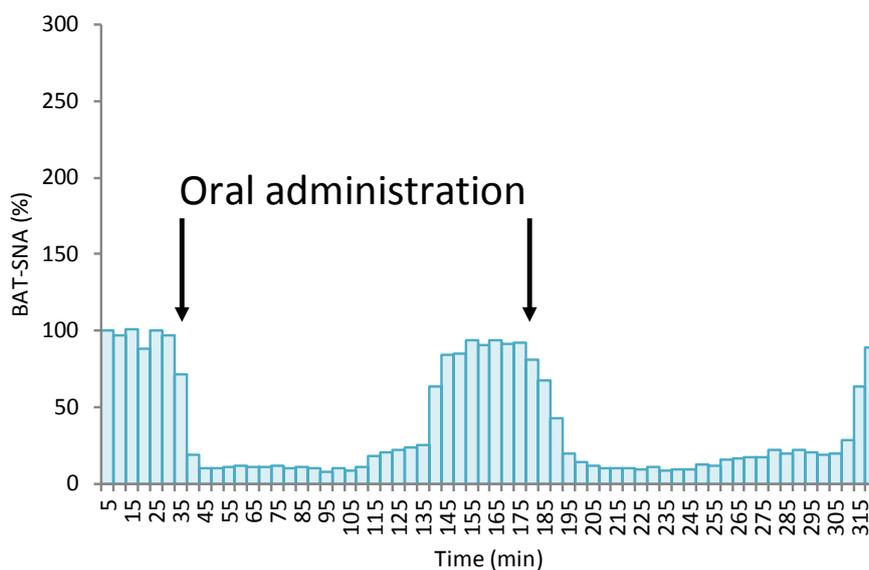


Figure 4. Effect of GP extract on sympathetic nerve activity in the brown adipose tissue. After recording stable sympathetic nerve activity for 30 min, the GP extract of 5 mg/kg body weight was administered using gastric tube. Similar results were observed from three individuals of rat.

Adipose tissues are classified as either WAT or BAT, which have different functions and morphology [25]. BAT is found in infants, young children, and rodents. Recently, BAT has been observed around the scapula, axilla, and vertebral column of adult humans using ^{18}F -fluorodeoxyglucose (^{18}F -FDG) and positron-emission tomography (PET) [3] [26] [27]. Thermogenesis in BAT is mediated by sympathetic stimulation of BAT mitochondrial uncoupling protein 1 (UCP1). We hypothesized that the anti-obesity activity of GP intake in this mouse breeding test was due to this mechanism, and BAT SNA was involved in the effect of GP extract.

Capsaicin, which is the major ingredient in chili pepper, has the potential to increase energy expenditure and decrease body fat by activating BAT in humans and rodents [28] [29] [30]. This results in stimulation of SNA by transient receptor potential vanilloid 1 (TRPV1), which is expressed in the gastrointestinal tract and sensory nervous system and specifically combines with capsaicin at the vanillyl moiety [15] [31]. The results of the present study were consistent with data obtained from experiments with capsaicin and capsiate, which is also found in chili peppers. Surprisingly however, GP extract exhibits a different mode of action from that of capsaicin and capsiate; GP extract decreases SNA, rather than increasing it as seen with capsaicin and capsiate.

Capsaicin and capsiate bear structural similarities to the non-volatile vanilloids found in GP [32] [33] [34]. It has also been reported that 6-paradol and 6-shogaol in GP extract were found to activate TRPV1 through non-covalent bonding [35]. The HPLC analysis showed that GP extract contains many vanilloid compounds possessing hydrocarbon side chains with a double bond and/or carbonyl group (data not shown). These substructures of vanilloids may be re-

sponsible for activating BAT SNA. This might suggest that the vanilloids in GP contribute to the anti-obesity effect by stimulating BAT SNA. However, contrary to expectations, BAT SNA was decreased by oral administration of GP extract.

Crocin, ezetimibe, catechins, and caffeine inhibit intestinal absorption of cholesterol and fat by inactivating pancreatic lipase [36] [37] [38]. The aqueous extract of *Zingiber officinale* Roscoe also prevents intestinal lipid absorption [39] through the actions of vanilloid compounds such as 6-gingerol and 6-shogaol [40]. These reports suggest that GP may inhibit body weight gain in mice by suppressing lipid absorption rather than by activating BAT SNA.

5. Conclusion

In summary, the present study demonstrated that GP and its extract exert an anti-obesity effect in HFD-fed mice. Prevention of body weight gain and fat accumulation occurs through improved hepatic lipid metabolism. In addition, BAT SNA decreased with intragastric infusion of GP extract. In general, decreasing BAT SNA would increase parasympathetic nerve activity, thus having a relaxation effect in humans. Therefore, GP or GP extract may have the additional benefit of promoting relaxation while controlling obesity in both humans and animals. Further research is required to clarify the mechanisms of the anti-obesity properties of GP extract; this may require investigation of lipid absorption in the small intestine.

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