

# Performance and Muscular Development of Nile Tilapia Larvae (*Oreochromis niloticus*) Fed Increasing Concentrations of Phenylalanine

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## Abstract

Even though tilapia is of great economic interest, data on the nutritional requirements of amino acids during commercial breeding stages are scarce. So, the aim of this study was to analyse the performance and muscular growth of Nile tilapia larvae (*Oreochromis niloticus*) fed diets containing increasing concentrations of phenylalanine. The experiment took place at the Laboratory of Aquaculture of the Grupo de Estudos de Manejo na Aquicultura (GEMAQ) of the Universidade Estadual do Oeste do Paraná (Unioeste), Toledo, PR, Brazil, for 30 days. A total of 450 larvae, with mean initial length of  $1.5 \pm 0.14$  cm and mean initial weight of  $0.04 \pm 0.004$  g, were randomly distributed into 6 treatment groups with 5 repetitions (30 tanks). Each experimental unit consisted of a 30 L tank containing 15 larvae. Six diets were formulated with increasing concentrations of phenylalanine (1.09, 1.24, 1.39, 1.54, 1.69 and 1.84%) and offered four times a day until apparent satiation. By the end of the experimental period, the following productivity indexes were analysed: length, weight, survival, weight gain, uniformity, feed conversion, specific growth rate and protein efficiency ratio. Three fish from each tank were collected for muscular growth analysis and the frequency of small ( $<20 \mu\text{m}$  in diameter) and large ( $20 - 50 \mu\text{m}$ ) muscle fibres recorded. Small fibres corresponded to 60% - 90% of the fibres observed, suggesting that muscular growth at this stage occurred mainly by hyperplasia. No significant difference ( $P > 0.05$ ) was observed in the productive performance parameters between the treatment groups. Thus, it can be concluded that the levels of phenylalanine contained in the experimental diets did not influence the performance of Nile tilapia larvae. However, there was more hyperplasia when we put 1.39% of phenylalanine in the diet.

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## Keywords

Amino Acids, Amino Acid Requirements, Larval Fish Nutrition

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### 1. Introduction

The Nile tilapia (*Oreochromis niloticus*) is one of the fresh water species with the greatest technological package due to its rusticity, climate adaptation, prolificacy and excellent flesh quality [1]. Thus, the farming of this species has been growing and approximately 169.453 thousand tonnes of Nile tilapia were produced in Brazil in 2013 [2].

Studies aimed at increasing productivity while reducing production costs often use plant proteins to replace those of animal origin [3] [4] or include by-products of processing industries [5]. In recent years, the addition of amino acids to fish diets has proven popular and new information on their effects and requirements has been detailed.

In general, fish does not have protein requirements per-se but needs a balance between essential and non-essential amino acids [6], which must be present in adequate quantities and ratios or be supplemented in their synthetic form in the feed [7]. Like all other animals, fish requires ten essential amino acids in their diet: arginine, phenylalanine, histadine, isoleucine, lysine, methionine, threonine, tryptophan and valine, out of which lysine and methionine are the most limiting [8].

Santiago and Lovell [9] established the phenylalanine + tyrosine requirement of Nile tilapia larvae to be 1.05% of the diet or 3.75% of the protein content. Based on the essential amino acid: digestible lysine ratio, Furuya [10] estimated the requirement of phenylalanine + tyrosine to be 2.38% for the sexual inversion phase, 1.65% for fish up to 100 g of live weight and 1.50% for those above 100 g of live weight. However, more information can help to determine the best supplementation levels of these amino acids in each phase of development.

Phenylalanine is an aromatic amino acid and its requirement is influenced by the concentration of tyrosine in the diet. Phenylalanine is the sole precursor of tyrosine, which in turn is the precursor of T3 (triiodothyronine) and T4 (thyroxine) hormones that are essential for normal metabolic processes and growth [11]. The NRC [12] highlights that the aromatic amino acid requirements of salmon, channel catfish, common carp and Japanese eel are 5.1%, 5.0%, 6.5% and 5.8% of the protein, respectively.

Even though tilapia is of great economic interest, data on the nutritional requirements of amino acids during commercial breeding stages are scarce. Great economic losses could result if these requirements were to be based only in relation to what is found in the fillet, as these requirements might vary with each stage of development.

Studies aimed at determining the percentage of essential amino acids inclusion in the diet of Nile tilapia are of great importance for the development of a balanced diet that

provides sufficient nutrients for successful fish production. Thus, the aim of this study was to analyse the influence of phenylalanine on the productive performance and muscular growth of Nile tilapia larvae (*Oreochromis niloticus*).

## 2. Materials and Methods

The experiment took place at the Laboratory of Aquaculture of the Grupo de Estudos de Manejo na Aquicultura (GEMAQ), Universidade Estadual do Oeste do Paraná (Unioeste), Toledo-PR, Brazil. The experiment lasted for 30 days and was approved by the Ethics Committee on the Use of Animals for Research (protocol number 24/2014).

Four hundred and fifty Nile tilapia larvae were used in this study. The larvae had mean initial length and weight of  $1.5 \pm 0.14$  cm and  $0.04 \pm 0.004$  g, respectively. The fish were randomly distributed into six treatments groups with five repetitions and placed into thirty 30L tanks containing 15 fish each (experimental unit). The tanks were individually equipped with a constant aeration system and syphoned twice daily (40% of aquarium volume) before the first and after the last feed, in order to remove leftover food and residues from fish excretion.

Six diets were formulated with increasing concentrations of phenylalanine: 1.09, 1.24, 1.39, 1.54, 1.69 and 1.84% (**Table 1**), based on Furuya [10]. The ingredients used were selected according to the quantity of phenylalanine in their composition [13] and ground using a hammer mill with 0.5 mm in diameter screen holes. The ground ingredients were weighed, homogenized, humidified and extruded using an extrusion equipment (Ex-Micro® extruder, ExTeec Company, Ribeirão Preto, Brazil). Subsequently, they were dried in a kiln (TE-394/3, Tecnal, Piracicaba, Brazil) by forced ventilation for 72 hours at 55°C. The pellets were ground in order to obtain a meal ration and alpha-methyl-testosterone (60 mg/kg of feed) added to finalize larval sexual inversion. Fish were fed four times a day at 8:00 hr, 11:00 hr, 14:00 hr and 17:00 hr until apparent satiation.

The physical and chemical parameters of the water, such as pH (7.3) and dissolved oxygen ( $4.32 \text{ mg}\cdot\text{L}^{-1}$ ), were measured weekly using a multi-parameter digital equipment (YSI®) and the temperature ( $24.2^\circ\text{C} \pm 1.5^\circ\text{C}$ ) recorded daily.

At the end of the experiment, food was withheld for 24 hours to allow emptying of the digestive tract. The fish were removed from the tanks, desensitized with benzocaine (ethyl-aminobenzoate) at  $240 \text{ mg}\cdot\text{L}^{-1}$  of water, according to the protocol by Okamura *et al.* [14], and subsequently euthanized.

The productivity indexes analysed were: final length (cm), final weight (g), survival rate (%), weight gain (g), specific growth rate (SGR), feed conversion (FCR), protein efficiency rate (weight gain/consumed protein) (%) and uniformity (%). The survival (Sur) (1), SGR (2) and FCR (3) rates were calculated using the following equations:

$$Sur(\%) = \left( \frac{nf}{ni} \right) \times 100 \quad (1)$$

**Table 1.** Percentage and nutritional composition of the diets containing increasing concentrations of phenylalanine fed to Nile tilapia larvae (*Oreochromis niloticus*).

| Ingredient (%)                        | Phenylalanine concentration (%) |       |       |       |       |       |
|---------------------------------------|---------------------------------|-------|-------|-------|-------|-------|
|                                       | 1.09                            | 1.24  | 1.39  | 1.54  | 1.69  | 1.84  |
| Meat and bone meal                    | 26.23                           | 26.23 | 26.23 | 26.23 | 26.23 | 26.23 |
| Fishmeal                              | 24.90                           | 24.90 | 24.90 | 24.90 | 24.90 | 24.90 |
| Wheat Bran                            | 23.75                           | 23.75 | 23.75 | 23.75 | 23.75 | 23.75 |
| L. Alanine                            | 7.50                            | 7.40  | 7.29  | 7.19  | 7.08  | 6.98  |
| L. Glutamic Acid                      | 7.50                            | 7.55  | 7.60  | 7.65  | 7.70  | 7.75  |
| Soybean oil                           | 6.88                            | 6.78  | 6.68  | 6.58  | 6.48  | 6.39  |
| Premix <sup>a</sup>                   | 1.00                            | 1.00  | 1.00  | 1.00  | 1.00  | 1.00  |
| L-lysine                              | 0.79                            | 0.79  | 0.79  | 0.79  | 0.79  | 0.79  |
| L-threonine                           | 0.63                            | 0.63  | 0.63  | 0.63  | 0.63  | 0.63  |
| Salt                                  | 0.30                            | 0.30  | 0.30  | 0.30  | 0.30  | 0.30  |
| L-tryptophan                          | 0.21                            | 0.21  | 0.21  | 0.21  | 0.21  | 0.21  |
| DL-methionine                         | 0.20                            | 0.20  | 0.20  | 0.20  | 0.20  | 0.20  |
| Antifungal                            | 0.10                            | 0.10  | 0.10  | 0.10  | 0.10  | 0.10  |
| Antioxidant (BHT)                     | 0.02                            | 0.02  | 0.02  | 0.02  | 0.02  | 0.02  |
| L-phenylalanine                       | 0.00                            | 0.15  | 0.30  | 0.45  | 0.61  | 0.76  |
| Nutrients (%)                         |                                 |       |       |       |       |       |
| Linoleic acid                         | 4.18                            | 4.13  | 4.07  | 4.02  | 3.96  | 3.91  |
| Starch                                | 7.44                            | 7.44  | 7.44  | 7.45  | 7.45  | 7.45  |
| Calcium                               | 4.46                            | 4.46  | 4.46  | 4.46  | 4.46  | 4.46  |
| Digestible energy (Kcal) <sup>b</sup> | 3,250                           | 3,250 | 3,250 | 3,250 | 3,250 | 3,250 |
| Total Phenylalanine                   | 1.09                            | 1.24  | 1.39  | 1.54  | 1.69  | 1.84  |
| Total phenylalanine + tyrosine        | 1.89                            | 2.04  | 2.19  | 2.34  | 2.49  | 2.64  |
| Total Phosphorus                      | 2.59                            | 2.59  | 2.59  | 2.59  | 2.59  | 2.59  |
| Available phosphorus                  | 2.50                            | 2.50  | 2.50  | 2.50  | 2.50  | 2.50  |
| Fat                                   | 14.29                           | 14.19 | 14.09 | 13.40 | 13.90 | 13.80 |
| Lysine                                | 2.20                            | 2.20  | 2.20  | 2.20  | 2.20  | 2.20  |
| Methionine                            | 0.75                            | 0.75  | 0.75  | 0.75  | 0.75  | 0.75  |
| Crude Protein                         | 42.79                           | 42.79 | 42.79 | 42.79 | 42.79 | 42.79 |
| Digestible Protein <sup>c</sup>       | 38.60                           | 38.60 | 38.60 | 38.60 | 38.60 | 38.60 |
| Threonine                             | 1.70                            | 1.70  | 1.70  | 1.70  | 1.70  | 1.70  |
| Tryptophan                            | 0.43                            | 0.43  | 0.43  | 0.43  | 0.43  | 0.43  |

Based on the concentrations recommended by Furuya [10]; The diet was formulated with the assistance of software Supercrac (TD Software, Viçosa, MG, Brazil); <sup>a</sup>Nutrition facts (minimum): Vitamin A = 1,000,000 UI/kg; Vitamin D3 = 500,000 UI/kg; Vitamin E = 20,000 UI/kg; Vitamin K3 = 500 mg/kg; Vitamin B1 = 1,900 mg/kg; Vitamin B2 = 2000 mg/kg; Vitamin B6 = 2400 mg/kg; Vitamin B12 = 3500 mcg/kg; Vitamin C = 25 g/kg; Niacin = 5,000 mg/kg; Pantothenic acid = 4800 mg/kg; Folic acid = 200 mg/kg; Biotin = 40 mg/kg; Manganese = 7500 mg/kg; Zinc = 25.0 g/kg; Iron = 12.50 g/kg; Copper = 2000 mg/kg; Iodine = 200 mg/kg; Selenium = 70 mg/kg; <sup>b</sup>Level of digestible energy recommended by Boscolo *et al.* [15]; <sup>c</sup>Level of digestible protein recommended by Hayashi *et al.* [16].

$$SGR(\% \text{ day}) = \left( \frac{\ln wf - \ln wi}{t} \right) \times 100 \quad (2)$$

$$FCR = \frac{FI}{WG} \quad (3)$$

where  $nf$  = number of fish in the tanks at the end of the experiment;  $ni$  = number of fish at the start of the experiment;  $wf$  = final weight (g);  $wi$  = initial weight (g);  $t$  = length of experiment (days);  $FI$  = feed intake (g);  $WG$  = weight gain (g).

Three fish from each tank were used to analyse muscular growth. A vertical sample (dorso-ventral) was removed with the aid of a blade and fixed in 10% buffered formalin containing a descaler, for 24 hours, prior to paraffin embedding. Transversal sections (6  $\mu\text{m}$ ) were stained with haematoxylin-eosin and a system of image analysis used for morphometry (Image-Pro Plus), in which the smallest diameter of the muscle fibres was determined. A total of 200 muscle fibres were analysed per animal and then classified according to their diameter (<20  $\mu\text{m}$  or 20 - 50  $\mu\text{m}$ ) in order to determine the contribution of hyperplasia and hypertrophy to muscular growth [17]. These analysis took place at the Laboratory of Histology of the Grupo de Estudos de Manejo na Aquicultura (GEMAQ), Universidade Estadual do Oeste do Paraná (Unioeste), Toledo, PR, Brazil. The data was ranked (<20  $\mu\text{m}$ ) and logarithmized (20 - 50  $\mu\text{m}$ ) so that parametric analysis could be applied.

Statistical analysis was performed by ANOVA followed by Tukey test using the programme Statistic "Statistic 7.1" [18]. Significance was considered at 5%.

### 3. Results

No significant difference ( $P > 0.05$ ) was observed in mean final length (MFL), mean final weight (MFW), survival rate (Sur), weight gain (WG), specific growth rate (SGR), uniformity (Unif) or feed conversion (FCR) (Table 2).

**Table 2.** Productive performance of Nile tilapia larvae fed diets containing increasing concentrations of phenylalanine.

| Variables <sup>a,b</sup> | Phenylalanine concentration (%) |               |               |               |               |               |
|--------------------------|---------------------------------|---------------|---------------|---------------|---------------|---------------|
|                          | 1.09                            | 1.24          | 1.39          | 1.54          | 1.69          | 1.84          |
| MFW (g)                  | 0.70 ± 0.03                     | 0.73 ± 0.26   | 0.70 ± 0.07   | 0.81 ± 0.23   | 0.77 ± 0.18   | 0.86 ± 0.16   |
| MFL (cm)                 | 3.40 ± 0.15                     | 3.54 ± 0.32   | 3.36 ± 0.14   | 3.49 ± 0.28   | 3.64 ± 0.31   | 3.57 ± 0.26   |
| WG (g)                   | 0.64 ± 0.03                     | 0.66 ± 0.26   | 0.64 ± 0.06   | 0.74 ± 0.23   | 0.70 ± 0.18   | 0.79 ± 0.16   |
| FCR                      | 1.93 ± 1.28                     | 3.01 ± 1.15   | 1.72 ± 0.69   | 2.01 ± 0.35   | 2.05 ± 1.55   | 1.67 ± 0.19   |
| Sur (%)                  | 75.00 ± 24.49                   | 65.00 ± 16.60 | 85.00 ± 22.41 | 66.67 ± 10.54 | 57.33 ± 25.65 | 73.33 ± 20.00 |
| Unif (%)                 | 60.75 ± 11.00                   | 79.60 ± 10.33 | 76.60 ± 10.88 | 71.50 ± 16.93 | 82.62 ± 19.27 | 73.12 ± 12.20 |
| SGR                      | 7.96 ± 0.26                     | 8.63 ± 1.16   | 7.81 ± 0.29   | 8.22 ± 0.91   | 8.09 ± 0.52   | 8.56 ± 0.73   |
| PER (%)                  | 3.17 ± 3.04                     | 5.53 ± 2.42   | 2.78 ± 1.38   | 3.28 ± 0.64   | 3.42 ± 3.89   | 2.80 ± 0.32   |

<sup>a</sup>Data is presented as mean ± standard deviation; final weight (FW); mean final length (MFL); weight gain (WG) = final weight-initial weight; survival (Sur) = 100 (final number of fish/initial number of fish); specific growth rate (SGR) = 100 \* ((ln final weight-ln initial weight)/experimental days); feed conversion (FCR) = diet consumed/weight gain; protein efficiency rate (PER) = (weight gain/protein consumed); Uniformity (Unif) = 100 \* (number of fish with body weight within the mean ± standard deviation /total number of fish); <sup>b</sup>No significant difference ( $P > 0.05$ ).

Muscle fibres with diameter smaller than 20  $\mu\text{m}$  corresponded to 60% - 90% of the fibres observed while those with diameter between 20 - 50  $\mu\text{m}$  corresponded only to 10% - 40% (**Table 3**).

#### 4. Discussion

The lack of significant differences in the productive performance parameters of the larvae fed diets containing increasing concentrations of phenylalanine is probably due to the fact that the amino acid requirements for this stage of development was met by the diet with the lowest concentration of phenylalanine tested (1.09%). Santiago and Lovell [9] established the requirement of phenylalanine by Nile tilapia larvae to be 1.05% of phenylalanine + tyrosine; however, these authors used purified diets while the diets used in the current study were semi-purified.

Furuya [10] stated that diets for Nile tilapia at the sexual inversion stage must contain 2.38% phenylalanine + tyrosine. In the present study, the concentrations of phenylalanine + tyrosine used were 1.89%, 2.04%, 2.19%, 2.34%, 2.49% and 2.69%; and the lowest concentration of both amino acids did not cause loss in the performance of larval stage. However, differently from the study by Furuya [10], the present study used dose-response testing, which explains the results obtained.

Borlogan [19], when determining the requirements of phenylalanine + tyrosine by juvenile "milkfish" (*Chanos chanos*), obtained better weight gain results with diets containing 1.3 and 1.2% of phenylalanine and tyrosine, respectively (phenylalanine + tyrosine = 2.5%). This difference to the Nile tilapia can be explained by its feeding habit (omnivore), as carnivore fish have greater amino acid and protein requirements.

Tyrosine is a non-essential amino acid that fish can synthesize from phenylalanine [20]. Thus, when tyrosine is added to the diet it is possible to reduce the concentration of phenylalanine by 40% - 60% [12]. In the present study, all diets were formulated to contain levels of tyrosine (0.8%) similar to those found in food.

Borlogan [19] states that diets with high concentrations of phenylalanine can be harmful to fish as its accumulation and oxidation generate toxic metabolites and, thus, hinder growth. Furthermore, a good balance of amino acids is required for maximum absorption. An imbalance of amino acids can affect productive performance and cause disproportionate absorption and use of other amino acids [21] [22]. In the present study, productive analysis did not suggest any toxic effect or amino acid imbalance as

**Table 3.** Frequency of muscle fibre distribution according to their diameter (<20  $\mu\text{m}$  and 20 - 50  $\mu\text{m}$ ) in Nile tilapia larvae fed diets containing increasing concentrations of phenylalanine.

| Diameter <sup>a</sup> | Phenylalanine concentration (%) |                               |                               |                                |                                |                                 |
|-----------------------|---------------------------------|-------------------------------|-------------------------------|--------------------------------|--------------------------------|---------------------------------|
|                       | 1.09                            | 1.24                          | 1.39                          | 1.54                           | 1.69                           | 1.84                            |
| <20 $\mu\text{m}$     | 60.41 $\pm$ 13.02 <sup>b</sup>  | 64.16 $\pm$ 4.98 <sup>b</sup> | 80.07 $\pm$ 8.95 <sup>a</sup> | 72.96 $\pm$ 5.89 <sup>ab</sup> | 59.81 $\pm$ 10.77 <sup>b</sup> | 75.02 $\pm$ 11.18 <sup>ab</sup> |
| 20 - 50 $\mu\text{m}$ | 39.59 $\pm$ 13.02 <sup>a</sup>  | 35.90 $\pm$ 4.85 <sup>a</sup> | 19.92 $\pm$ 8.95 <sup>b</sup> | 27.04 $\pm$ 5.89 <sup>ab</sup> | 40.35 $\pm$ 10.64 <sup>a</sup> | 29.85 $\pm$ 11.99 <sup>ab</sup> |

<sup>a</sup>Data is represented as mean  $\pm$  standard deviation. Different letters on the same row indicate significant difference ( $P < 0.05$ ) in the frequency of muscle fibres distribution within a specific diameter.

there was no significant difference in fish productive performance between the different diets used.

The high feed conversion rate of the present study was probably due to the unrestricted feeding and to the fact that the ration used was ground to a meal after extrusion. High levels of feed conversion (1.9 to 2.6) have also been observed by Ngamsnae *et al.* [23] in juvenile *Bidyanus bidyanus* fed diets containing varying concentrations of arginine and phenylalanine and by Ahmed [24] who reported feed conversion levels of 1.34 to 2.42 in a study to determine the requirements of tyrosine and phenylalanine by *Cirrhinus mrigala* carp larvae. Besides, the diet has been continued to be offered until satiation even during the periods when the temperature of water was pleasant.

No significant difference ( $P > 0.05$ ) was observed in the survival rate of the larvae in the present study. Borlogan and Colosso [25], in a study to determine the amino acid requirements of juvenile “milkfish” (*Chanos chanos*), observed better survival rates in juveniles fed diets containing high concentrations of phenylalanine (1.9%, 2.20% and 2.50%) and established, through breakpoint equation, 2.8% as the optimal value to obtain the best survival rates and productive performance indexes. Ngamsnae *et al.* [23] did not observe any significant difference in the survival rates of juvenile *Bidyanus bidyanus*, which were similar to the ones obtained in this study. Woynarovich [26] related that can occur mortality from 40% to 50% in the early stage of fish life. It can occur due to several factors, one of them is the water temperature that stayed lower than recommended levels.

No significant difference ( $P > 0.05$ ) was observed in the uniformity of the fish, which may have been due to the favourable conditions the larvae were kept throughout the experiment. Stock density is known to be a determining factor in the uniformity of individuals, thus, the low stock density of the tanks enabled the larvae to be comfortable and have sufficient good quality feed and may have contributed to the uniformity observed. Zaminham [27], in a study with larvae of the same species fed diets containing increasing concentrations of tryptophan, obtained the best results when higher concentrations of this essential aromatic amino acid were used. Even though phenylalanine is an aromatic amino acid, there are currently no reports on whether it could lead to results similar to those obtained with tryptophan. Nevertheless, as there was no significant difference in uniformity between the treatments in this study, it can be assumed that the diet and environment provided were adequate for the species studied.

The specific growth rate in this study varied from 7.5% and 9% a day, similarly to those observed by Bomfim *et al.* [28] (8.7%) when diets containing 32% of crude protein were used. The diets from the current study contained 42% of crude protein [16], which may have resulted in the higher levels observed. Furthermore, the stage of larval development in which the experiment was conducted is characterized by being the stage with the greatest growth rates in fish.

The protein efficiency rate in this study was relatively high; however, no significant difference ( $P > 0.05$ ) was observed between the treatments. Furuya *et al.* [29] reported means of 2.60% (30% DP) to 3.22% (25.5% DP), with rates inversely proportional to the concentration of digestible protein. In the present study, digestible protein corresponded to 38%,

which may have directly contributed to the results obtained. It is important to note that the diets were formulated to maintain a constant level of the remaining amino acids and only increase the concentration of phenylalanine. Therefore, it can be said that the diets were balanced and that body protein synthesis would only be affected by the concentration of phenylalanine, a fact that was not observed even at the lowest level tested.

Rowlerson and Veggetti [30] determined that high frequency of muscle fibres smaller than 20  $\mu\text{m}$  characterized hyperplasia while fibres with diameters greater 50  $\mu\text{m}$  characterized hypertrophy. Analysis of the frequency of muscle fibres of different diameters suggest that the muscular development observed in this study occurred mainly by hyperplasia, albeit it seems that hypertrophy may have occurred concomitantly. However, it is possible that muscular development may have occurred only by hyperplasia in mosaic form, in which new muscle fibres originate from the fusion of satellite cells [30] and where small diameter fibres that recently formed around larger ones are notoriously present, as observed in this study.

Aguiar *et al.* [31] observed that in Nile tilapia the muscle fibres of greater diameter occur in fish that are 90 to 190 days old and that the fries from this species undergo muscular growth by hyperplasia in mosaic form, in accordance with the results obtained in the present study. Jhonston *et al.* [32] and Zimmerman & Lowery [33] state that muscular development by hyperplasia is replaced by hypertrophy of the muscular cells when a fish reaches 44% of its final size; however, this can vary with species. Furthermore, Rowlerson and Veggetti [30] reported that fish have an undetermined growth, which makes it difficult to establish the final weight of an adult.

As well as in our study, Almeida *et al.* [17] stated that in juvenile Pacu (*Piaractus mesopotamicus*) muscular development occurs predominantly by hyperplasia. Similarly, Aguiar *et al.* [34] observed that Nile tilapia larvae fed diets containing increasing concentrations of lysine showed mainly cells smaller than 20  $\mu\text{m}$  in diameter, characterizing a predominant development by hyperplasia. These reports are in accordance with the results observed in the current study.

The phenylalanine provides modifications in hyperplastic and hypertrophic muscle growth of the skeletal muscle fibers of tilapia larvae. The higher frequency of fibers with diameter lower than 20  $\mu\text{m}$  was observed in fish fed with diet containing 1.39% of phenylalanine. This result confirm the role of this amino acid in the protein synthesis, however very few studies have analysed the effects of fish nutrition on muscular growth and development and even fewer studies provide information on the effects of phenylalanine on muscular hypertrophy and hyperplasia in fish. Until recently, studies on phenylalanine have focused mainly on its effects as precursor to T3 (triiodotironine) and T4 (thyroxine) hormones, which are required for normal metabolic processes and growth [11]. However, the main response to amino acid consumption is protein synthesis [12], which consequently leads to greater weight gain.

In our study, we verified that the diet containing 1.69% of phenylalanine resulted in greater frequency of muscle fibres with 20 - 50  $\mu\text{m}$  in diameter, which could be related to the survival rate of this treatment group as it enabled the fish to consume more ra-

tion than those from the other treatment groups. According to Houlihan [35], muscular tissue growth can result from an increase in the metabolic rate or in the concentration of amino acids available through feeding. However, further studies using dose-response are necessary to determine the effects of phenylalanine for muscle development.

In summary, it was observed that phenylalanine can interfere with the frequency of muscular fibres of different diameters; however, a specific rate could not be determined as the lowest tested level of this amino acid met the nutritional requirements of the Nile tilapia larvae.

## 5. Conclusion

The levels of phenylalanine contained in the experimental diets did not influence the performance of Nile tilapia larvae. However, there was more hyperplasia when we put 1.39% of phenylalanine in the diet.

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